

Abstract

Gut microbiota, composed by bacteria, fungi and viruses, play an essential role in the health of human and animals. Gut microbiota in infant can take effort to the gut development in adult. There is a dynamic process between bacteria and bacteriophages in early life, however, the interaction between the two important composition is unclear. In this project, we will use computational methods to study if CRISPR and Anti-CRISPR take part in the process and what the role it takes based on data from public database.

Keywords: Infant gut microbiota CRISPR Anti-CRISPR

1 Introduction

The mammalian microbiome consists of bacteria, archaea, fungi and viruses(Lynch and Pedersen, 2016). They have a profound influence on human physiology, nutrition and disease(Qin et al., 2010). They form a bioreactor and produce bioactive compounds. These compounds signal to distant organs including brain, liver, heart, lung and so forth(Schroeder and Bäckhed, 2016). In heart, gut micro-derived metabolites are recognized as contributors to atherogenesis(Wang et al., 2011). Microbiomes play a role in neurodevelopmental and mood disorders(De Theije et al., 2014). Differences in species richness and their diversity between autism spectrum disorder (ASD) and controls have been reported(Williams et al., 2012). As one part of the gut microbiome, the virome has a significant implication in health and disease(Virgin, 2014). during inflammatory bowel diseases(IBD) the intestinal phage population is altered and transitions from an ordered state to a stochastic dysbiosis in murine(Duerkop et al., 2018).

The microbiota of infants is a key factor for the development of the microbiome(Stewart et al., 2018). Epidemiological studies have shown that factors that alter bacterial communities in infants during childhood increase the risk for several diseases(Tamburini et al., 2016). Childhood overweight is associated with the microbiota of early life in Denmark(Ajslev et al., 2011) and Norway(Stanislawski et al., 2018). Early gut microbiota was associated with infant growth rates.(White et al., 2013) As for immune system, a diverse early life intestinal microbiota can inhibit the pathways that lead to allergic sensitization by multiple mechanisms(Reynolds and Finlay, 2017). Besides, researches over the past few years reveal that the gut microbiome plays a role in basic neurogenerative processes and modulates many aspects of animal behavior in early life(Sharon et al., 2016).

The ecology of the bacterial microbiome increases in richness and diversity towards an adult-like composition(Yatsunenko et al., 2012). Soon after birth, the bacterial microbiome rapidly switches from predominantly facultative anaerobic bacteria to a diverse community of anaerobes(Koenig et al., 2011). During the first months of life, the early infant gut bacteriophage virome is composed of a rich community of bacteriophages, the majority of which derive from the Caudovirales order.

Subsequently, the bacteriophages decrease in richness and shifts towards a Microviridae-dominated community over the first 2 years of life. Thus, the infant virome and bacterial microbiome evolves in a dynamic trajectory during the early years of life (Lim et al., 2015).

As the Red Queen hypothesis proposes that organisms must continually evolve new mechanisms of resistance to parasites to avoid extinction (Liow et al., 2011). Bacteria have evolved a great of diverse strategies to defend themselves against phage, including restriction modification enzymes that inactivate target DNA by cleavage, toxin-antitoxin modules that lead to phage abortive infection and CRISPR-Cas systems that target and inactivate specific nucleic acid sequences by cleavage. In response, phages have evolved various mechanisms to overcome these defenses, including expression of proteins that modify restriction sites (Krüger and Bickle, 1983) or degrade restriction modification cofactors (Studier and Movva, 1976), antitoxin molecules that inhibit the activity of toxin-antitoxin abortive infection systems (Otsuka and Yonesaki, 2012) and proteins that directly bind to and inactivate CRISPR-Cas machinery (Pawluk et al., 2016).

Thus, CRISPR-Cas and anti-CRISPR of the metagenomic and 16S rRNA sequences from eight healthy infants (four twin pairs) between 0-2 years old (Lim et al., 2015) will be studied. Computational approaches will be used to detect CRISPR-Cas and anti-CRISPR sequences. What kind of CRISPR-Cas and anti-CRISPR are contained and during the dynamic process of bacteria-bacteriophage, if the CRISPR and Anti-CRISPR systems change, if they play a role will be answered. If the switch of lysogenic and lytic life style drives the dynamic process will be studied. Besides, in terms of previous researches, there is no CRISPR system in Bacteroidetes and no anti-CRISPR in Caudovirales⁶² which are dominant in gut, new insight is wished.

Extrally, Maybe we can study the known interactions between anti-CRISPR proteins and their binding site of CRISPR-Cas, and then predict the novel anti-CRISPR proteins based on the numerous CRISPR-Cas data (Alipanahi et al., 2015).

2 Materials & Methods

Customized metagenomic and 16S rRNA sequences from eight healthy infants (four twin pairs) between 0-2 years old (Lim et al., 2015) can be downloaded. Some operations have been done previous researchers. And the methods of virome sequencing processing, virome analysis and 16S rRNA gene analysis have been described (Lim et al., 2015). And the Methods of CRISPR analysis including identification CRISPR spacers and identification of phage contigs have been described (Stern et al., 2012). As for Anti-CRISPR analysis, There are two strategies for detecting Anti-CRISPR. First, using the Anti-CRISPR database to query the metagenomic data using BLASTN with e-value threshold of $1E-4$. Second, using the aca gene sequences combined from all the previous researches to query the metagenomic data using the same parameters. There are

two principles(Stern et al., 2012) that can lead to the Python script for detection of the lysogenic phages. First, the lysogenic phages should be combined with integrase and recombinase. Second, the lysogenic phages should be integrated into bacteria with flanking sequences. And for the left phages, we regard them as lytic phages.

3 Budget

Probably, this project don't need budget.

Supervisor Tim G. Barraclough, Stineke Van Houte and Edze Westra has seen and accepted budget.

Signature:

References

- TA Ajslev, CS Andersen, M Gamborg, TIA Sørensen, and T Jess. Childhood overweight after establishment of the gut microbiota: the role of delivery mode, pre-pregnancy weight and early administration of antibiotics. *International journal of obesity*, 35(4):522, 2011.
- Babak Alipanahi, Andrew Delong, Matthew T Weirauch, and Brendan J Frey. Predicting the sequence specificities of dna-and rna-binding proteins by deep learning. *Nature biotechnology*, 33(8):831, 2015.
- Caroline GM De Theije, Bas M Bavelaar, Sofia Lopes da Silva, Sijmen Mechiel Korte, Berend Olivier, Johan Garssen, and Aletta D Kraneveld. Food allergy and food-based therapies in neurodevelopmental disorders. *Pediatric Allergy and Immunology*, 25(3):218–226, 2014.
- Breck A Duerkop, Manuel Kleiner, David Paez-Espino, Wenhan Zhu, Brian Bushnell, Brian Hassell, Sebastian E Winter, Nikos C Kyrpides, and Lora V Hooper. Murine colitis reveals a disease-associated bacteriophage community. *Nature microbiology*, 3(9):1023, 2018.
- Jeremy E Koenig, Aymé Spor, Nicholas Scalfone, Ashwana D Fricker, Jesse Stombaugh, Rob Knight, Largus T Angenent, and Ruth E Ley. Succession of microbial consortia in the developing infant gut microbiome. *Proceedings of the National Academy of Sciences*, 108(Supplement 1):4578–4585, 2011.
- DH Krüger and Thomas A Bickle. Bacteriophage survival: multiple mechanisms for avoiding the deoxyribonucleic acid restriction systems of their hosts. *Microbiological reviews*, 47(3):345, 1983.
- Efrem S Lim, Yanjiao Zhou, Guoyan Zhao, Irma K Bauer, Lindsay Droit, I Malick Ndao, Barbara B Warner, Phillip I Tarr, David Wang, and Lori R Holtz. Early life dynamics of the human gut virome and bacterial microbiome in infants. *Nature medicine*, 21(10):1228, 2015.
- Lee Hsiang Liow, Leigh Van Valen, and Nils Chr Stenseth. Red queen: from populations to taxa and communities. *Trends in ecology & evolution*, 26(7):349–358, 2011.
- Susan V Lynch and Oluf Pedersen. The human intestinal microbiome in health and disease. *New England Journal of Medicine*, 375(24):2369–2379, 2016.
- Yuichi Otsuka and Tetsuro Yonesaki. Dmd of bacteriophage t4 functions as an antitoxin against escherichia coli lsa and rnlA toxins. *Molecular microbiology*, 83(4):669–681, 2012.
- April Pawluk, Nadia Amrani, Yan Zhang, Bianca Garcia, Yurima Hidalgo-Reyes, Jooyoung Lee, Alireza Edraki, Megha Shah, Erik J Sontheimer, Karen L Maxwell, et al. Naturally occurring off-switches for crispr-cas9. *Cell*, 167(7):1829–1838, 2016.

- Junjie Qin, Ruiqiang Li, Jeroen Raes, Manimozhiyan Arumugam, Kristoffer Solvsten Burgdorf, Chaysavanh Manichanh, Trine Nielsen, Nicolas Pons, Florence Levenez, Takuji Yamada, et al. A human gut microbial gene catalogue established by metagenomic sequencing. *nature*, 464(7285): 59, 2010.
- Lisa A Reynolds and B Brett Finlay. Early life factors that affect allergy development. *Nature Reviews Immunology*, 17(8):518, 2017.
- Bjoern O Schroeder and Fredrik Bäckhed. Signals from the gut microbiota to distant organs in physiology and disease. *Nature medicine*, 22(10):1079, 2016.
- Gil Sharon, Timothy R Sampson, Daniel H Geschwind, and Sarkis K Mazmanian. The central nervous system and the gut microbiome. *Cell*, 167(4):915–932, 2016.
- Maggie A Stanislawski, Dana Dabelea, Brandie D Wagner, Nina Iszatt, Cecilie Dahl, Marci K Sontag, Rob Knight, Catherine A Lozupone, and Merete Eggesbø. Gut microbiota in the first 2 years of life and the association with body mass index at age 12 in a norwegian birth cohort. *mBio*, 9(5):e01751–18, 2018.
- Adi Stern, Eran Mick, Itay Tirosh, Or Sagy, and Rotem Sorek. Crispr targeting reveals a reservoir of common phages associated with the human gut microbiome. *Genome research*, pages gr-138297, 2012.
- Christopher J Stewart, Nadim J Ajami, Jacqueline L O'Brien, Diane S Hutchinson, Daniel P Smith, Matthew C Wong, Matthew C Ross, Richard E Lloyd, HarshaVardhan Doddapaneni, Ginger A Metcalf, et al. Temporal development of the gut microbiome in early childhood from the teddy study. *Nature*, 562(7728):583, 2018.
- F WILLIAM Studier and NR Movva. Samase gene of bacteriophage t3 is responsible for overcoming host restriction. *Journal of virology*, 19(1):136–145, 1976.
- Sabrina Tamburini, Nan Shen, Han Chih Wu, and Jose C Clemente. The microbiome in early life: implications for health outcomes. *Nature medicine*, 22(7):713, 2016.
- Herbert W Virgin. The virome in mammalian physiology and disease. *Cell*, 157(1):142–150, 2014.
- Zeneng Wang, Elizabeth Klipfell, Brian J Bennett, Robert Koeth, Bruce S Levison, Brandon DuGar, Ariel E Feldstein, Earl B Britt, Xiaoming Fu, Yoon-Mi Chung, et al. Gut flora metabolism of phosphatidylcholine promotes cardiovascular disease. *Nature*, 472(7341):57, 2011.
- Richard A White, Jørgen V Bjørnholt, Donna D Baird, Tore Midtvedt, Jennifer R Harris, Marcello Pagano, Winston Hide, Knut Rudi, Birgitte Moen, Nina Iszatt, et al. Novel developmental

- analyses identify longitudinal patterns of early gut microbiota that affect infant growth. *PLoS computational biology*, 9(5):e1003042, 2013.
- Brent L Williams, Mady Hornig, Tanmay Parekh, and W Ian Lipkin. Application of novel pcr-based methods for detection, quantitation, and phylogenetic characterization of *sutterella* species in intestinal biopsy samples from children with autism and gastrointestinal disturbances. *MBio*, 3(1):e00261–11, 2012.
- Tanya Yatsunenko, Federico E Rey, Mark J Manary, Indi Trehan, Maria Gloria Dominguez-Bello, Monica Contreras, Magda Magris, Glida Hidalgo, Robert N Baldassano, Andrey P Anokhin, et al. Human gut microbiome viewed across age and geography. *nature*, 486(7402):222, 2012.