

Scenic

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Contents

Loading packages	1
Loading data	1
SCENIC	2
Prepare cellinfo and expression matrix	2
Download binding motifs for mouse	2
Initialise settings	2
Gene filtering	2
Correlation	2
Genie3	3
Build and score the GRN	3
Plotting the results	3
Viewing markers based on specificity score	3
Plot on Heatmap the 10 markers the most specific of each cell type	3
plotting Mafk and c-Maf	4

Loading packages

```
suppressMessages(library(dplyr))
suppressMessages(library(Seurat))
suppressMessages(library(patchwork))
suppressMessages(library(ggplot2))
suppressMessages(library(SCENIC))
suppressMessages(library(ComplexHeatmap))
suppressMessages(library(AUCell))
```

Loading data

```
myeloid_cells_clustered <-
readRDS("../3-Visualisation_Clustering/Myeloid_cells_Final.rds")
```

SCENIC

Prepare cellinfo and expression matrix

```
exprMat <- myeloid_cells_clustered@assays$RNA@data
cellInfo <- data.frame(seuratCluster=Idents(myeloid_cells_clustered))
cellInfo$nGene <- colSums(exprMat>0)

dir.create ("int")
#saveRDS(exprMat, file = "int/exprMat.Rds")
#saveRDS(cellInfo, file = "int/cellInfo.Rds")
```

Download binding motifs for mouse

```
dbFiles <-
c("https://resources.aertslab.org/cistarget/databases/old/mus_musculus/mm9/refseq_r45/mc9nr/gene_based/mm9",
  "https://resources.aertslab.org/cistarget/databases/old/mus_musculus/mm9/refseq_r45/mc9nr/gene_based/mm10")

dir.create("cisTarget_databases");
setwd("cisTarget_databases")
for(featherURL in dbFiles)
{
  download.file(featherURL, destfile=basename(featherURL)) # saved in current dir
}
```

Initialise settings

```
org <- "mgi"
dbDir <- "cisTarget_databases"
dbDir <- path.expand(dbDir)
myDatasetTitle <- "SCENIC Analysis"
data(defaultDbNames)
dbs <- defaultDbNames[[org]]
scenicOptions <- initializeScenic(org=org, dbDir=dbDir, dbs=dbs, nCores=25)
```

Gene filtering

1. Filter by the total number of reads per gene. Keeps only the genes with at least 6 UMI counts across all samples.
2. Filter by the number of cells in which the gene is detected.

```
exprMat <- as.matrix(exprMat) # need a regulat matrix
genesKept <- geneFiltering(exprMat, scenicOptions=scenicOptions,
                           minCountsPerGene=3*.01*ncol(exprMat),
                           minSamples=ncol(exprMat)*.01)

exprMat_filtered <- exprMat[genesKept, ]
```

Correlation

```
runCorrelation(exprMat_filtered, scenicOptions)
```

Genie3

This step is time consuming

```
runGenie3(exprMat, scenicOptions)
```

Build and score the GRN

```
runGenie3(exprMat_filtered, scenicOptions)

scenicOptions@settings$verbose <- TRUE
scenicOptions@settings$nCores <- 10
scenicOptions@settings$seed <- 123

runSCENIC_1_coexNetwork2modules(scenicOptions)
runSCENIC_2_createRegulons(scenicOptions)
runSCENIC_3_scoreCells(scenicOptions, exprMat_filtered)
```

Plotting the results

Viewing markers based on specificity score

```
regulonAUC <- loadInt(scenicOptions, "aucell_regulonAUC") # require file
int/3.4_regulonAUC.Rds

rss <- calcRSS(AUC=getAUC(regulonAUC), cellAnnotation=cellInfo[colnames(regulonAUC),
"seuratCluster"])
#rssPlot <- plotRSS(rss["Mafb (44g)",])
rssPlot <- plotRSS(rss)
plotly::ggplotly(rssPlot$plot)

plotly::ggplotly(rssPlot$plot)
```

Plot on Heatmap the 10 markers the most specific of each cell type

```
rss <- rss[,c(8,11,2,3,7,1,12,9,4,6,5,10)]

top_tf <- c()
for (i in 1:length(colnames(rss))) {

  TFs <- sort(rss[,i], decreasing = T)
  top_10 <- head(TFs, 10)
  top_tf <- c(top_tf, top_10)
}

length(top_tf)
```

```

top_tf

# Creation of the Heatmap

regulonAUC.mat <- regulonAUC@assays@data@listData$AUC
Subset_regulonActivity <-regulonAUC.mat[names(top_tf),]

regulonActivity_byCellType_Scaled <- t(scale(t(Subset_regulonActivity), center = T,
scale=T))

colors <- c("#B2DF8A", "#ABD61C", "#1F78B4", "#A6CEE3", "#E31A1C", "#E3751C", "#600078",
"#33A02C", "#FDBF6F", "#526317", "#D4AAC6", "#784620")

colors <- c("#B2DF8A", "#ABD61C", "#1F78B4", "#A6CEE3", "#E31A1C", "#E3751C", "#600078",
"#33A02C", "#FDBF6F", "#526317", "#D4AAC6", "#784620")
names(colors) <- levels(myeloid_cells_clustered$CellType)

df <- as.data.frame(myeloid_cells_clustered$CellType)
colnames(df) <- "CellType"

color_df <- list(CellType =
  c("CD206- IM" = "#B2DF8A",
    "Ly6G Macs" = "#ABD61C",
    "Ly6C- Mo" = "#1F78B4",
    "Ly6C+ Mo" = "#A6CEE3",
    "CD64+ cMo" = "#E31A1C",
    "AM" = "#E3751C",
    "Neutrophils" = "#600078",
    "CD206+ IM" = "#33A02C",
    "DCs" = "#FDBF6F",
    "Dying cells" = "#526317",
    "Cycling Macs" = "#D4AAC6",
    "IAV-specific AM" = "#784620"))

png(file="/home/qiang/Documents/Travail_cell/Travail_V3/Cécilia_project_2/4Scenic/Heatmap.png",
width = 2000, height = 2500)

Heatmap(Subset_regulonActivity[, name="Regulon activity", show_column_names = FALSE,
  column_split = factor(myeloid_cells_clustered$CellType),
  cluster_column_slices = F,
  cluster_rows = F,
  bottom_annotation = HeatmapAnnotation(df = df, col = color_df))
dev.off()

```

plotting Mafb and c-Maf

```

TF <- c("Mafb (44g)", "Maf (220g)")

df <- as.data.frame(myeloid_cells_clustered$CellType)
colnames(df) <- "Clusters"

color_df <- list(Clusters =

```

```

        c("CD206 IM" = "#B2DF8A",
          "PR8 Mac1" = "#ABD61C",
          "pMo" = "#1F78B4",
          "cMo" = "#A6CEE3",
          "CD64+ cMo" = "#E31A1C",
          "AM" = "#E377C2",
          "Neutro" = "#600078",
          "CD206+ IM" = "#33A02C",
          "DCs" = "#FDBF6F",
          "PR8 Mac2" = "#526317",
          "Cycling Mac" = "#D4AAC6",
          "PR8 AM" = "#784620"))

#pdf(file="/home/qiang/Documents/Travail_cell/Travail_V3/Cécilia_project_2/4Scenic/Heatmap_cMafMafbstat2.pdf",
width = 2000, height = 1000)
heatmap_maf <- Heatmap(regulonAUC.mat[TF,], name="Regulon activity", show_column_names =
FALSE,
      column_split = factor(myeloid_cells_clustered$CellType),
      cluster_column_slices = F,
      cluster_rows = F,
      bottom_annotation = HeatmapAnnotation(df = df, col = color_df))
#dev.off()

#setequal(levels(df), names(color_df$test))

```

saving heatmap as pdf

```

tidyHeatmap::save_pdf(heatmap_maf,
"/home/qiang/Documents/Travail_cell/Travail_V3/Cécilia_project_2/4Scenic/Heatmap_cMafMafbstat2.pdf",
width = 30, height = 10, units = "cm")

```

```
sessionInfo()
```

```

## R version 4.3.1 (2023-06-16)
## Platform: x86_64-pc-linux-gnu (64-bit)
## Running under: Ubuntu 22.04.3 LTS
##
## Matrix products: default
## BLAS:   /usr/lib/x86_64-linux-gnu/openblas-pthread/libblas.so.3
## LAPACK: /usr/lib/x86_64-linux-gnu/openblas-pthread/libopenblas-p0.3.20.so; LAPACK version 3.10.0
##
## locale:
##  [1] LC_CTYPE=en_US.UTF-8      LC_NUMERIC=C
##  [3] LC_TIME=fr_BE.UTF-8      LC_COLLATE=en_US.UTF-8
##  [5] LC_MONETARY=fr_BE.UTF-8  LC_MESSAGES=en_US.UTF-8
##  [7] LC_PAPER=fr_BE.UTF-8     LC_NAME=C
##  [9] LC_ADDRESS=C             LC_TELEPHONE=C
## [11] LC_MEASUREMENT=fr_BE.UTF-8 LC_IDENTIFICATION=C
##
## time zone: Europe/Brussels
## tzcode source: system (glibc)
##
## attached base packages:
## [1] grid      stats      graphics  grDevices  utils      datasets  methods

```

```

## [8] base
##
## other attached packages:
## [1] AUCell_1.22.0          ComplexHeatmap_2.16.0 SCENIC_1.3.1
## [4] ggplot2_3.4.2          patchwork_1.1.2       SeuratObject_4.1.3
## [7] Seurat_4.3.0           dplyr_1.1.2
##
## loaded via a namespace (and not attached):
## [1] RcppAnnoy_0.0.21       splines_4.3.1
## [3] later_1.3.1            bitops_1.0-7
## [5] tibble_3.2.1           R.oo_1.25.0
## [7] polyclip_1.10-4        graph_1.78.0
## [9] XML_3.99-0.14          lifecycle_1.0.3
## [11] doParallel_1.0.17      globals_0.16.2
## [13] lattice_0.21-8         MASS_7.3-60
## [15] magrittr_2.0.3         plotly_4.10.2
## [17] rmarkdown_2.23         yaml_2.3.7
## [19] httpuv_1.6.11          sctransform_0.3.5
## [21] spam_2.9-1             sp_2.0-0
## [23] spatstat.sparse_3.0-2  reticulate_1.30
## [25] cowplot_1.1.1          pbapply_1.7-2
## [27] DBI_1.1.3              RColorBrewer_1.1-3
## [29] abind_1.4-5            zlibbioc_1.46.0
## [31] Rtsne_0.16             GenomicRanges_1.52.0
## [33] purrr_1.0.1           R.utils_2.12.2
## [35] BiocGenerics_0.46.0    RCurl_1.98-1.12
## [37] circlize_0.4.15        GenomeInfoDbData_1.2.10
## [39] IRanges_2.34.0         S4Vectors_0.38.1
## [41] ggrepel_0.9.3          irlba_2.3.5.1
## [43] listenv_0.9.0          spatstat.utils_3.0-3
## [45] goftest_1.2-3          spatstat.random_3.1-5
## [47] annotate_1.78.0         fitdistrplus_1.1-11
## [49] parallelly_1.36.0      DelayedMatrixStats_1.22.1
## [51] leiden_0.4.3           codetools_0.2-19
## [53] DelayedArray_0.26.3    shape_1.4.6
## [55] tidyselect_1.2.0       matrixStats_1.0.0
## [57] stats4_4.3.1           spatstat.explore_3.2-1
## [59] jsonlite_1.8.7         GetoptLong_1.0.5
## [61] ellipsis_0.3.2         progressr_0.13.0
## [63] iterators_1.0.14       ggribes_0.5.4
## [65] survival_3.5-5         foreach_1.5.2
## [67] tools_4.3.1            ica_1.0-3
## [69] Rcpp_1.0.11            glue_1.6.2
## [71] gridExtra_2.3          xfun_0.39
## [73] MatrixGenerics_1.12.2  GenomeInfoDb_1.36.0
## [75] withr_2.5.0            fastmap_1.1.1
## [77] fansi_1.0.4            digest_0.6.33
## [79] R6_2.5.1              mime_0.12
## [81] colorspace_2.1-0       scattermore_1.2
## [83] tensor_1.5             spatstat.data_3.0-1
## [85] RSQLite_2.3.1          R.methodsS3_1.8.2
## [87] utf8_1.2.3            tidyr_1.3.0
## [89] generics_0.1.3         data.table_1.14.8
## [91] httr_1.4.6            htmlwidgets_1.6.2

```

## [93] S4Arrays_1.0.4	uwot_0.1.16
## [95] pkgconfig_2.0.3	gtable_0.3.3
## [97] blob_1.2.4	lmtest_0.9-40
## [99] XVector_0.40.0	htmltools_0.5.5
## [101] dotCall64_1.0-2	clue_0.3-64
## [103] GSEABase_1.62.0	scales_1.2.1
## [105] Biobase_2.60.0	png_0.1-8
## [107] knitr_1.43	rstudioapi_0.14
## [109] rjson_0.2.21	reshape2_1.4.4
## [111] nlme_3.1-162	GlobalOptions_0.1.2
## [113] zoo_1.8-12	cachem_1.0.8
## [115] stringr_1.5.0	KernSmooth_2.23-22
## [117] parallel_4.3.1	miniUI_0.1.1.1
## [119] AnnotationDbi_1.62.1	pillar_1.9.0
## [121] vctrs_0.6.3	RANN_2.6.1
## [123] promises_1.2.0.1	xtable_1.8-4
## [125] cluster_2.1.4	evaluate_0.21
## [127] cli_3.6.1	compiler_4.3.1
## [129] rlang_1.1.1	crayon_1.5.2
## [131] future.apply_1.11.0	plyr_1.8.8
## [133] stringi_1.7.12	viridisLite_0.4.2
## [135] deldir_1.0-9	munsell_0.5.0
## [137] Biostrings_2.68.1	lazyeval_0.2.2
## [139] spatstat.geom_3.2-4	Matrix_1.6-0
## [141] sparseMatrixStats_1.12.0	bit64_4.0.5
## [143] future_1.33.0	KEGGREST_1.40.0
## [145] shiny_1.7.4.1	SummarizedExperiment_1.30.2
## [147] ROCR_1.0-11	igraph_1.5.0.1
## [149] memoise_2.0.1	bit_4.0.5