# Scoring cell Epithelial

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Loading package	
<pre>suppressMessages(library(dplyr)) suppressMessages(library(patchwork)) suppressMessages(library(ggplot2)) suppressMessages(library(readxl)) suppressMessages(library(singscore)) suppressMessages(library(pheatmap))</pre>	

# Loading the normalised count matrix

```
Norm_exprMat <- read_excel("../Data_DSPDA/Q3 - publication - V2.xlsx", sheet =
"TargetCountMatrix")
Norm_exprMat <- as.data.frame(Norm_exprMat)
rownames(Norm_exprMat) <- Norm_exprMat$TargetName
Norm_exprMat$TargetName <- NULL</pre>
```

#### Get score AT2

```
markers_AT2 <- read.csv("Signature/Markers_AT2.csv")
markers_AT2 <- markers_AT2$X[1:20]
rankData <- rankGenes(Norm_exprMat)</pre>
```

```
scoredf <- simpleScore(rankData, upSet = markers_AT2)
## Warning in checkGenes(upSet, rownames(rankData)): 2 genes missing: Sdpr, Malat1
AT2 <- scoredf$TotalScore</pre>
```

#### Get score PrimedAT2

```
markers.PrimedAT2 <- read.csv("Signature/Markers_PrimedAT2.csv")
markers.PrimedAT2 <- markers.PrimedAT2$X[1:20]

rankData <- rankGenes(Norm_exprMat)

scoredf <- simpleScore(rankData, upSet = markers.PrimedAT2)

## Warning in checkGenes(upSet, rownames(rankData)): 3 genes missing: Atp5f1,
## Wbp5, Ngfrap1

PrimedAT2 <- scoredf$TotalScore

cell_singscore <- data.frame(AT2, PrimedAT2)</pre>
```

#### Get score DATP

```
markers.DATP <- read.csv("Signature/Markers_DATP.csv")
markers.DATP <- markers.DATP$X[1:20]

rankData <- rankGenes(Norm_exprMat)

scoredf <- simpleScore(rankData, upSet = markers.DATP)

## Warning in checkGenes(upSet, rownames(rankData)): 2 genes missing: Gm26870,
## Hist1h2bc

cell_singscore <- cbind(cell_singscore, DATP = scoredf$TotalScore)</pre>
```

#### Get score AT1

```
markers.AT1 <- read.csv("Signature/Markers_AT1.csv")
markers.AT1 <- markers.AT1$X[1:20]

rankData <- rankGenes(Norm_exprMat)
scoredf <- simpleScore(rankData, upSet = markers.AT1)

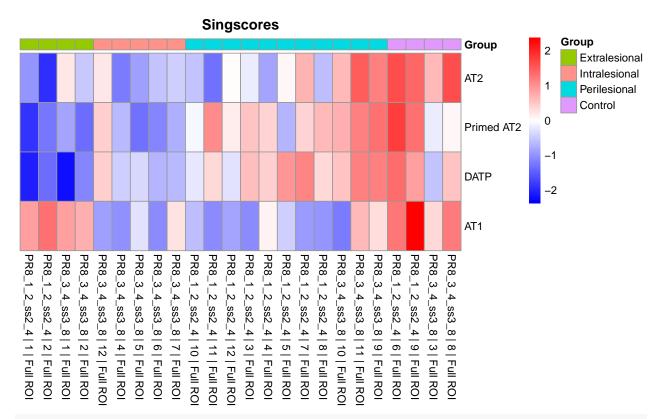
## Warning in checkGenes(upSet, rownames(rankData)): 2 genes missing: Ly6i, mt-Nd2
cell_singscore <- cbind(cell_singscore, AT1 = scoredf$TotalScore)</pre>
```

## Write csv file for singscore

```
rownames(cell_singscore) <- colnames(Norm_exprMat)
#write.csv(cell_singscore, "Score_Epithelial.csv")</pre>
```

## plotting score in pheatmap

```
annotation_sample <- read_excel("../Data_DSPDA/Q3 - publication - V2.xlsx", sheet =
"SegmentProperties")
singscores <- cell_singscore</pre>
singscores <- merge(singscores, annotation_sample[,c(5,25)], by x = "row.names", by y =
"SegmentDisplayName")
singscores <- singscores[order(singscores$Group),]</pre>
singscores$Group<- NULL</pre>
rownames(singscores) <- singscores$Row.names</pre>
singscores$Row.names <- NULL</pre>
df <- as.data.frame(annotation_sample[,c(5,25)])</pre>
df$Group <- as.factor(df$Group)</pre>
levels(df$Group) <- c( "Extralesional", "Intralesional", "Perilesional", "Control")</pre>
rownames(df)<-df$SegmentDisplayName</pre>
df$SegmentDisplayName <- NULL</pre>
pheatmap_singscore<- pheatmap(t(singscores), cluster_rows=FALSE, show_rownames=T,</pre>
         cluster_cols=F, scale = "row", annotation_col=df, main = "Singscores",
         labels_row = c("AT2", "Primed AT2", "DATP", "AT1"), color =
         colorRampPalette(c("blue", "white", "red"))(100))
```



#### sessionInfo()

```
## R version 4.3.1 (2023-06-16)
## Platform: x86_64-pc-linux-gnu (64-bit)
## Running under: Ubuntu 22.04.3 LTS
##
## Matrix products: default
          /usr/lib/x86_64-linux-gnu/openblas-pthread/libblas.so.3
## LAPACK: /usr/lib/x86_64-linux-gnu/openblas-pthread/libopenblasp-r0.3.20.so; LAPACK version 3.10.0
##
## locale:
  [1] LC_CTYPE=en_US.UTF-8
                                  LC NUMERIC=C
  [3] LC_TIME=fr_BE.UTF-8
                                  LC_COLLATE=en_US.UTF-8
   [5] LC_MONETARY=fr_BE.UTF-8
                                  LC_MESSAGES=en_US.UTF-8
##
##
   [7] LC_PAPER=fr_BE.UTF-8
                                  LC NAME=C
   [9] LC_ADDRESS=C
                                  LC_TELEPHONE=C
##
## [11] LC_MEASUREMENT=fr_BE.UTF-8 LC_IDENTIFICATION=C
## time zone: Europe/Brussels
## tzcode source: system (glibc)
##
## attached base packages:
## [1] stats
                graphics grDevices utils
                                              datasets methods
                                                                   base
## other attached packages:
## [1] pheatmap_1.0.12 singscore_1.20.0 readxl_1.4.2
                                                         ggplot2_3.4.2
## [5] patchwork_1.1.2 dplyr_1.1.2
## loaded via a namespace (and not attached):
```

```
## [1] KEGGREST_1.40.0
                                    SummarizedExperiment_1.30.2
## [3] gtable_0.3.3
                                    xfun_0.39
## [5] lattice 0.21-8
                                    Biobase 2.60.0
## [7] vctrs_0.6.3
                                    tools_4.3.1
## [9] bitops_1.0-7
                                    generics_0.1.3
## [11] stats4 4.3.1
                                    tibble 3.2.1
                                    AnnotationDbi_1.62.1
## [13] fansi 1.0.4
## [15] RSQLite_2.3.1
                                    highr_0.10
## [17] blob_1.2.4
                                    pkgconfig_2.0.3
## [19] Matrix_1.6-0
                                    RColorBrewer_1.1-3
## [21] S4Vectors_0.38.1
                                    graph_1.78.0
                                    GenomeInfoDbData_1.2.10
## [23] lifecycle_1.0.3
                                    stringr_1.5.0
## [25] farver_2.1.1
## [27] compiler_4.3.1
                                    Biostrings_2.68.1
## [29] munsell_0.5.0
                                    GenomeInfoDb_1.36.0
## [31] htmltools_0.5.5
                                    RCurl_1.98-1.12
## [33] yaml_2.3.7
                                    tidyr_1.3.0
## [35] pillar 1.9.0
                                    crayon 1.5.2
## [37] limma_3.56.2
                                    DelayedArray_0.26.3
## [39] cachem 1.0.8
                                    locfit 1.5-9.8
## [41] tidyselect_1.2.0
                                    digest_0.6.33
## [43] stringi_1.7.12
                                    reshape2_1.4.4
                                    fastmap_1.1.1
## [45] purrr_1.0.1
## [47] grid 4.3.1
                                    colorspace 2.1-0
## [49] cli 3.6.1
                                    magrittr_2.0.3
## [51] S4Arrays_1.0.4
                                    XML_3.99-0.14
## [53] utf8_1.2.3
                                    GSEABase_1.62.0
## [55] edgeR_3.42.4
                                    withr_2.5.0
## [57] scales_1.2.1
                                    bit64_4.0.5
                                    XVector_0.40.0
## [59] rmarkdown_2.23
## [61] httr_1.4.6
                                    matrixStats_1.0.0
## [63] bit_4.0.5
                                    cellranger_1.1.0
## [65] png_0.1-8
                                    memoise_2.0.1
## [67] evaluate_0.21
                                    knitr_1.43
## [69] GenomicRanges_1.52.0
                                    IRanges_2.34.0
## [71] rlang_1.1.1
                                    Rcpp_1.0.11
## [73] xtable 1.8-4
                                    glue 1.6.2
## [75] DBI_1.1.3
                                    BiocGenerics_0.46.0
## [77] rstudioapi_0.14
                                    annotate_1.78.0
## [79] plyr_1.8.8
                                    R6_2.5.1
## [81] MatrixGenerics_1.12.2
                                    zlibbioc 1.46.0
```