

Clustering and Visualisation

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2024-10-18 11:47:17 +0200

Loading packages

```
suppressMessages(library(dplyr))
suppressMessages(library(Seurat))
suppressMessages(library(patchwork))
suppressMessages(library(ggplot2))
suppressMessages(library(dittoSeq))
suppressMessages(library(formatR))
suppressMessages(library(ComplexHeatmap))
```

Loading data

```
myeloid_cells <- readRDS("../2-Integration_workflow/Myeloid_cells_Part2.rds")
```

The Condition Myeloid PR8 1 and Myeloid PR8 2 are merged into one condition IAV

and the IM cells are renamed Steady-State CD64

```
myeloid_cells$orig.ident <- as.factor(myeloid_cells$orig.ident)
levels(myeloid_cells$orig.ident) <-
c("myeloid_PR8", "cell_myeloid_ctrl", "myeloid_PR8", "Steady-State CD64")
```

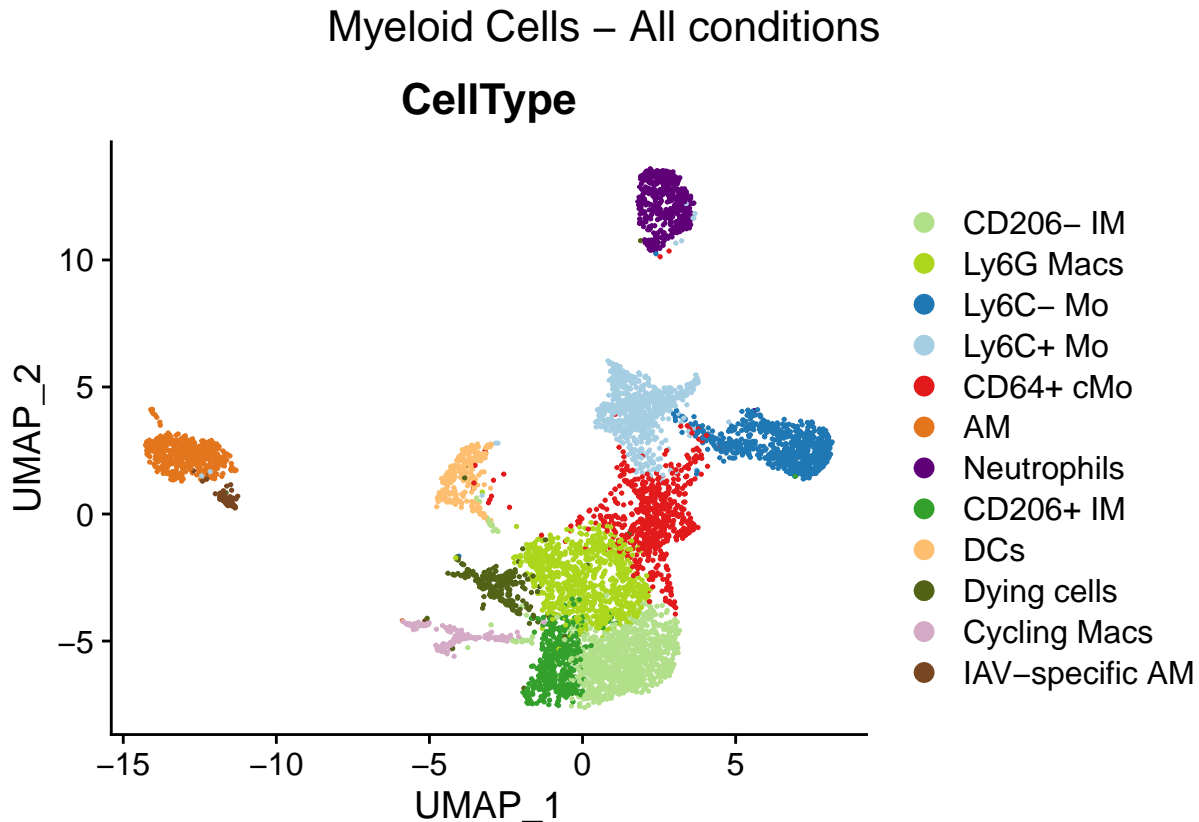
Adding the cell type to metadata

```
labels <- c("CD206- IM", "Ly6G Macs", "Ly6C- Mo", "Ly6C+ Mo", "CD64+
cMo", "AM", "Neutrophils", "CD206+ IM", "DCs", "Dying cells", "Cycling Macs", "IAV-specific AM")
colors <- c("#B2DF8A", "#ABD61C", "#1F78B4", "#A6CEE3", "#E31A1C", "#E3751C", "#600078",
"#33A02C", "#FDBF6F", "#526317", "#D4AAC6", "#784620")

# create a new column in metadata
myeloid_cells$CellType <- myeloid_cells$seurat_clusters

# replace cluster number by cell type
levels(myeloid_cells$CellType) <- labels
```

```
DimPlot(myeloid_cells, reduction = "umap", cols = colors, group.by = "CellType") +
plot_annotation(
  title = 'Myeloid Cells - All conditions',
  theme = theme(plot.title = element_text(size = 16, hjust = 0.5)))
```



```
#ggsave("graph_Umap.pdf", height = 12, width = 19, units = "cm")
```

Visualisation of the frequency graph.

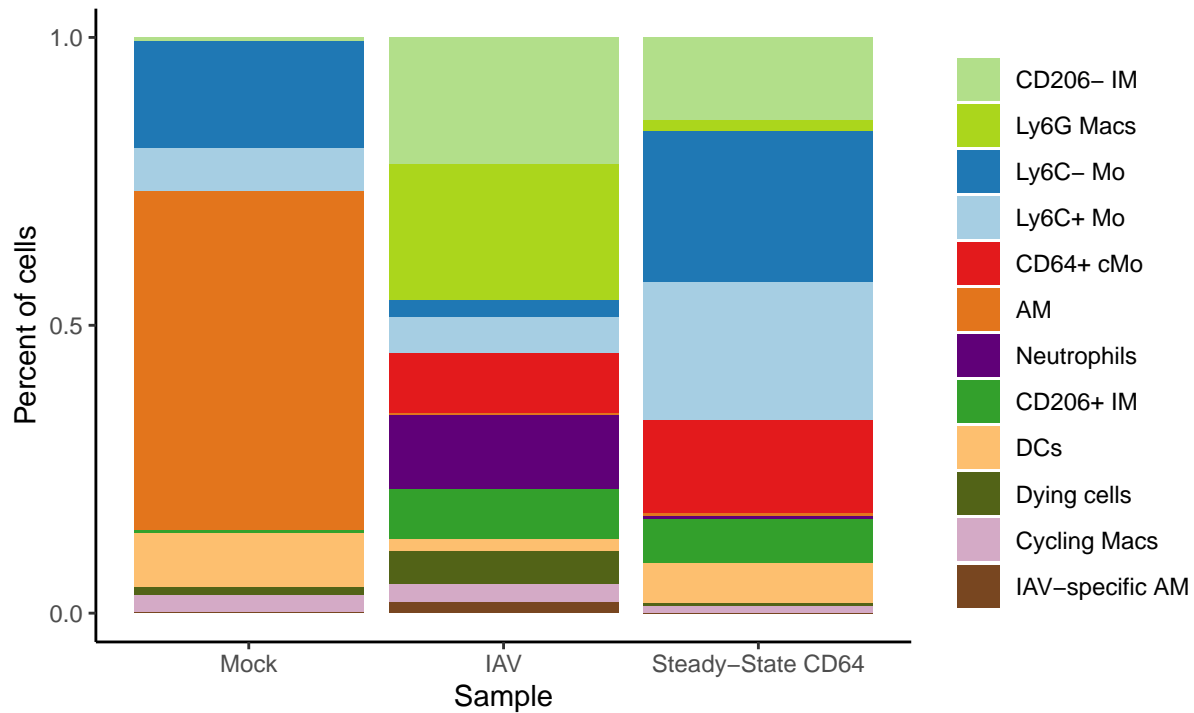
```
colors <- c("#B2DF8A", "#ABD61C", "#1F78B4", "#A6CEE3", "#E31A1C", "#E3751C", "#600078",
"#33A02C", "#FDBF6F", "#526317", "#D4AAC6", "#784620")

labels <- c("CD206- IM", "Ly6G Macs", "Ly6C- Mo", "Ly6C+ Mo", "CD64+
cMo", "AM", "Neutrophils", "CD206+ IM", "DCs", "Dying cells", "Cycling Macs", "IAV-specific AM")

#var.labels.rename = labels
var_order <- c(1, 2, 5, 6, 7, 8, 9, 10, 11, 12, 3, 4)

dittoBarPlot(myeloid_cells, "seurat_clusters", group.by = "orig.ident", color.panel =
colors, main = "", var.labels.reorder = var_order, var.labels.rename = labels,
x.labels.rotate = F, xlab = "Sample", x.labels = c("Mock", "IAV", "Steady-State CD64")) +
plot_annotation(
  title = 'Myeloid Cells - Cluster frequency',
  theme = theme(plot.title = element_text(size = 16, hjust = 0.5)))
```

Myeloid Cells – Cluster frequency



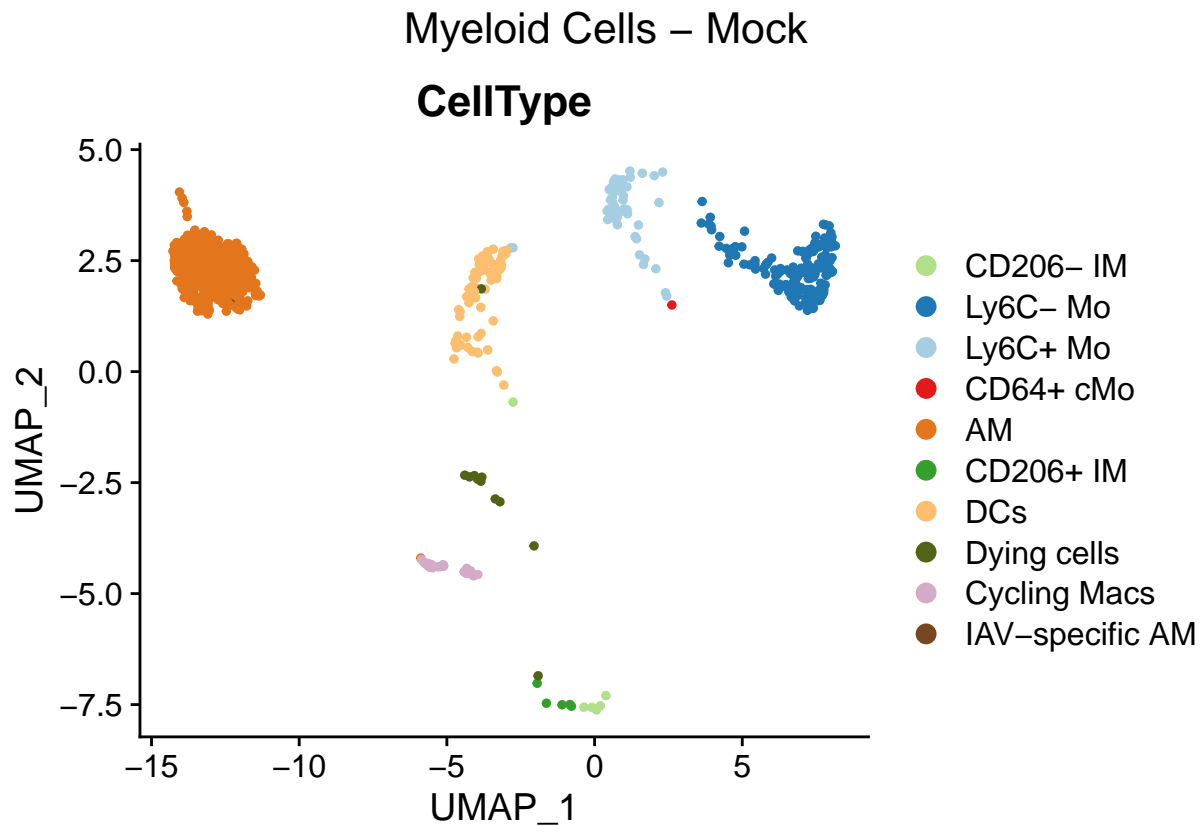
```
#ggsave("graphe_clusterFreq.pdf", height = 12 , width = 19, units = "cm")
```

Visualisation of the Mock cells

```
cell_myeloid_ctrl <- subset(myeloid_cells, orig.ident %in% "cell_myeloid_ctrl")

colors_mock <- c("#B2DF8A", "#1F78B4", "#A6CEE3", "#E31A1C", "#E377C2", "#33A02C",
"#FDBF6F", "#526317", "#D4AAC6", "#784620")

DimPlot(cell_myeloid_ctrl, reduction = "umap", cols = colors_mock, group.by = "CellType")
+ plot_annotation(
  title = 'Myeloid Cells - Mock',
  theme = theme(plot.title = element_text(size = 16, hjust = 0.5)))
```



```
#ggsave("UMAP_mock.pdf", height = 12 , width = 19, units = "cm")
```

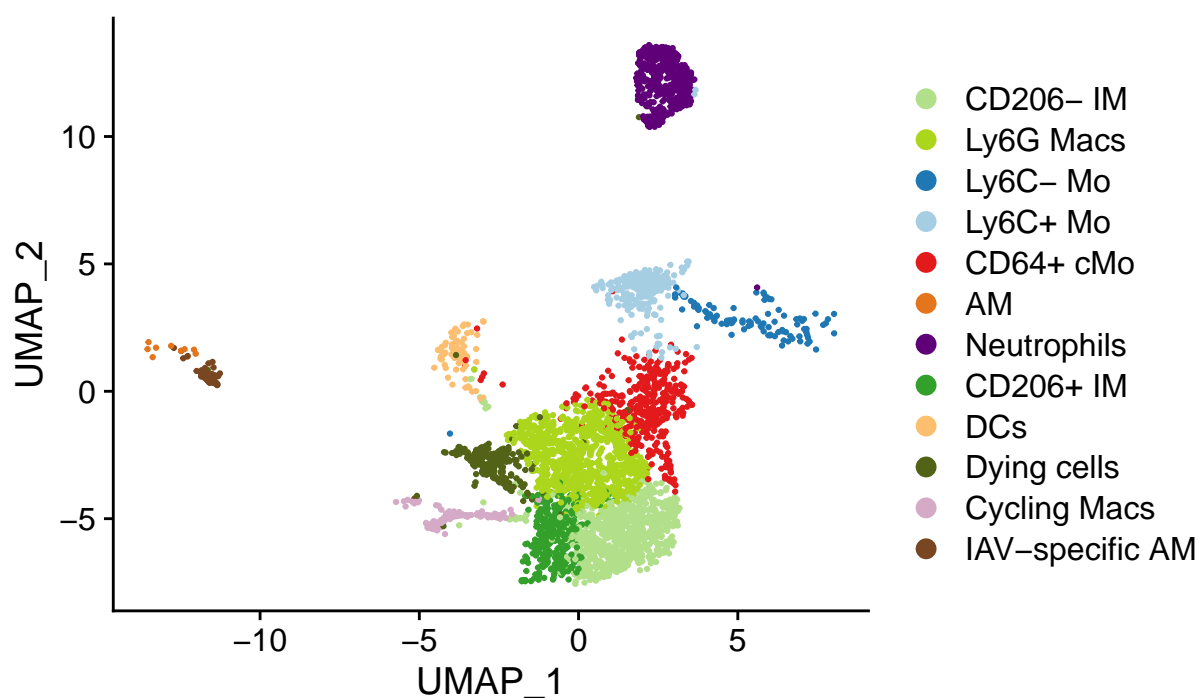
Visualisation of the PR8 cells

```
cell_myeloid_Pr8 <- subset(myeloid_cells, orig.ident %in% "myeloid_PR8")

DimPlot(cell_myeloid_Pr8, reduction = "umap", cols = colors, group.by = "CellType") +
plot_annotation(
  title = 'Myeloid Cells - PR8',
  theme = theme(plot.title = element_text(size = 16, hjust = 0.5)))
```

Myeloid Cells – PR8

CellType



```
#ggsave("UMAP_PR8.pdf", height = 12 , width = 19, units = "cm")
```

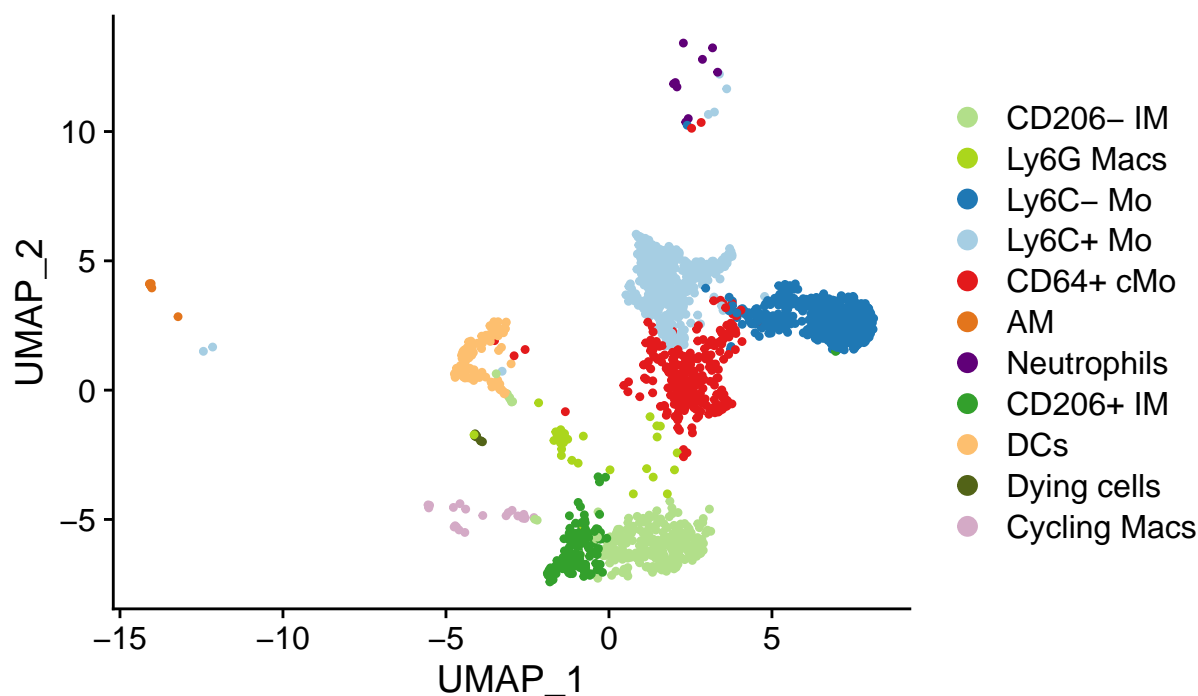
Visualisation IMcells

```
cell_myeloid_IMcells <- subset(myeloid_cells, orig.ident %in% "Steady-State CD64")

DimPlot(cell_myeloid_IMcells, reduction = "umap", cols = colors, group.by = "CellType") +
plot_annotation(
  title = 'Myeloid Cells - Mo-IM',
  theme = theme(plot.title = element_text(size = 16, hjust = 0.5)))
```

Myeloid Cells – Mo–IM

CellType



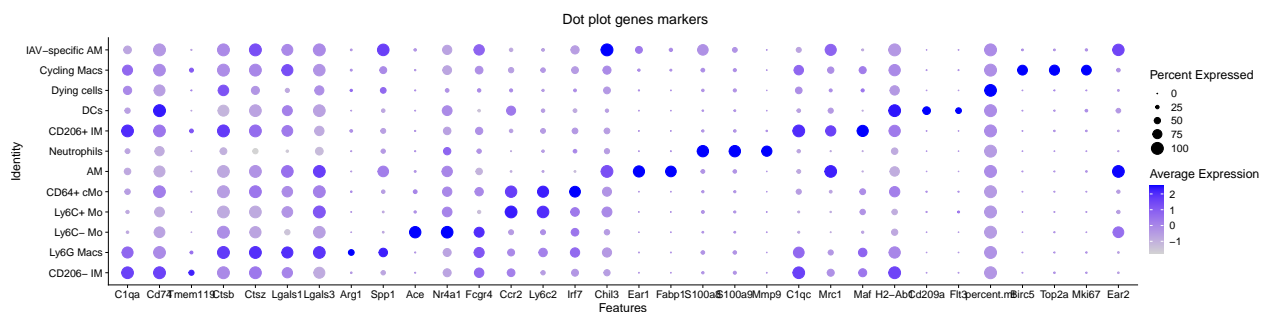
```
#ggsave("UMAP_PR8.pdf", height = 12 , width = 19, units = "cm")
```

Save the result for further analysis

```
saveRDS(myeloid_cells, "Myeloid_cells_Final.rds")
```

```
DefaultAssay(myeloid_cells) <- "RNA"
```

```
DotPlot(myeloid_cells, features = c("C1qa", "Cd74", "Tmem119", "Ctsb", "Ctsz", "Lgals1",
  "Lgals3", "Arg1", "Spp1", "Ace", "Nr4a1", "Fcgr4", "Ccr2", "Ly6c2", "Irf7", "Chil3",
  "Ear1", "Fabp1", "S100a8", "S100a9", "Mmp9", "C1qc", "Mrc1", "Maf", "H2-Ab1", "Cd209a",
  "Flt3", "percent.mt", "Birc5", "Top2a", "Mki67", "Ear2"), group.by = "CellType") +
  plot_annotation(
    title = 'Dot plot genes markers',
    theme = theme(plot.title = element_text(size = 16, hjust = 0.5)))
```



Differential expression Analysis

```
# find markers for every cluster compared to all remaining cells, report only the positive ones
myeloids.markers <- FindAllMarkers(myeloid_cells, only.pos = TRUE, min.pct = 0.25,
logfc.threshold = 0.25, test.use = "LR", latent.vars = "orig.ident") # The execution is quite long
```

The results from DE can directly be load from a csv file

```
myeloids.markers <- read.csv("myeloids_markers.csv")
```

```
# select the top 10 markers the most differentially expressed of each clusters
myeloids.markers %>%
  group_by(cluster) %>%
  top_n(n = 20, wt = avg_log2FC) -> top10

mat <- as.matrix( GetAssayData(object = myeloid_cells, slot =
"data")[as.character(top10$gene),])

df <- as.data.frame(myeloid_cells$CellType)
colnames(df) <- "Clusters"

color_df <- list(Clusters =
  c("CD206- IM" = "#B2DF8A",
    "Ly6G Macs" = "#ABD61C",
    "Ly6C- Mo" = "#1F78B4",
    "Ly6C+ Mo" = "#A6CEE3",
    "CD64+ cMo" = "#E31A1C",
    "AM" = "#E377C2",
    "Neutrophils" = "#600078",
    "CD206+ IM" = "#33A02C",
    "DCs" = "#FDBF6F",
    "Dying cells" = "#526317",
    "Cycling Macs" = "#D4AAC6",
    "IAV-specific AM" = "#784620"))

heatmap_allGenes <- Heatmap(t(scale(t(mat))), name="expressop", show_column_names =
FALSE,
  column_split = factor(myeloid_cells$CellType),
  cluster_column_slices = F,
  cluster_rows = F,
  show_column_dend = F,
  top_annotation = HeatmapAnnotation(df = df, col = color_df),
  column_title_rot = 90,
  row_names_gp = gpar(fontsize = 8),
  row_names_side = "left",
  use_raster=F,
  show_heatmap_legend = F)

#tidyHeatmap::save_pdf(heatmap_allGenes, "Heatmap.pdf", width = 30, height = 55, units = "cm")
```

sessionInfo()

```
## R version 4.3.3 (2024-02-29)
## Platform: x86_64-pc-linux-gnu (64-bit)
## Running under: Ubuntu 22.04.4 LTS
##
## Matrix products: default
## BLAS:   /usr/lib/x86_64-linux-gnu/openblas-pthread/libblas.so.3
## LAPACK: /usr/lib/x86_64-linux-gnu/openblas-pthread/libopenblas-p-r0.3.20.so; LAPACK version 3.10.0
##
## locale:
##  [1] LC_CTYPE=en_US.UTF-8      LC_NUMERIC=C
##  [3] LC_TIME=fr_BE.UTF-8      LC_COLLATE=en_US.UTF-8
##  [5] LC_MONETARY=fr_BE.UTF-8  LC_MESSAGES=en_US.UTF-8
##  [7] LC_PAPER=fr_BE.UTF-8     LC_NAME=C
##  [9] LC_ADDRESS=C             LC_TELEPHONE=C
## [11] LC_MEASUREMENT=fr_BE.UTF-8 LC_IDENTIFICATION=C
##
## time zone: Europe/Brussels
## tzcode source: system (glibc)
##
## attached base packages:
## [1] grid      stats      graphics  grDevices  utils      datasets  methods
## [8] base
##
## other attached packages:
## [1] ComplexHeatmap_2.16.0 formatR_1.14      dittoSeq_1.12.0
## [4] ggplot2_3.4.2      patchwork_1.1.2   SeuratObject_4.1.3
## [7] Seurat_4.3.0       dplyr_1.1.2
##
## loaded via a namespace (and not attached):
##  [1] RColorBrewer_1.1-3      shape_1.4.6
##  [3] rstudioapi_0.14        jsonlite_1.8.7
##  [5] magrittr_2.0.3         magick_2.7.5
##  [7] spatstat.utils_3.0-3    farver_2.1.1
##  [9] rmarkdown_2.23         GlobalOptions_0.1.2
## [11] zlibbioc_1.46.0        vctrs_0.6.3
## [13] ROCR_1.0-11            Cairo_1.6-2
## [15] spatstat.explore_3.2-1  RCurl_1.98-1.12
## [17] S4Arrays_1.2.1         htmltools_0.5.5
## [19] sctransform_0.3.5      parallelly_1.36.0
## [21] KernSmooth_2.23-22     htmlwidgets_1.6.2
## [23] ica_1.0-3              plyr_1.8.8
## [25] plotly_4.10.2          zoo_1.8-12
## [27] igraph_1.5.0.1         iterators_1.0.14
## [29] mime_0.12              lifecycle_1.0.3
## [31] pkgconfig_2.0.3        Matrix_1.6-1
## [33] R6_2.5.1               fastmap_1.1.1
## [35] clue_0.3-64            GenomeInfoDbData_1.2.10
## [37] MatrixGenerics_1.12.2  fitdistrplus_1.1-11
## [39] future_1.33.0          shiny_1.7.4.1
## [41] digest_0.6.33          colorspace_2.1-0
## [43] S4Vectors_0.38.1      tensor_1.5
```


## [45] irlba_2.3.5.1	GenomicRanges_1.52.0
## [47] labeling_0.4.2	progressr_0.13.0
## [49] fansi_1.0.4	spatstat.sparse_3.0-2
## [51] httr_1.4.6	polyclip_1.10-4
## [53] abind_1.4-5	compiler_4.3.3
## [55] doParallel_1.0.17	withr_2.5.0
## [57] highr_0.10	MASS_7.3-60.0.1
## [59] DelayedArray_0.26.3	rjson_0.2.21
## [61] tools_4.3.3	lmtest_0.9-40
## [63] httpuv_1.6.11	future.apply_1.11.0
## [65] goftest_1.2-3	glue_1.6.2
## [67] nlme_3.1-164	promises_1.2.0.1
## [69] Rtsne_0.16	cluster_2.1.6
## [71] reshape2_1.4.4	generics_0.1.3
## [73] gtable_0.3.3	spatstat.data_3.0-1
## [75] tidyr_1.3.0	data.table_1.14.8
## [77] XVector_0.40.0	sp_2.0-0
## [79] utf8_1.2.3	BiocGenerics_0.46.0
## [81] spatstat.geom_3.2-4	RcppAnnoy_0.0.21
## [83] foreach_1.5.2	ggrepel_0.9.3
## [85] RANN_2.6.1	pillar_1.9.0
## [87] stringr_1.5.0	spam_2.9-1
## [89] later_1.3.1	circlize_0.4.15
## [91] splines_4.3.3	lattice_0.22-5
## [93] survival_3.5-8	deldir_1.0-9
## [95] tidyselect_1.2.0	SingleCellExperiment_1.22.0
## [97] miniUI_0.1.1.1	pbapply_1.7-2
## [99] knitr_1.43	gridExtra_2.3
## [101] IRanges_2.34.0	SummarizedExperiment_1.30.2
## [103] scattermore_1.2	stats4_4.3.3
## [105] xfun_0.39	Biobase_2.60.0
## [107] matrixStats_1.0.0	pheatmap_1.0.12
## [109] stringi_1.7.12	lazyeval_0.2.2
## [111] yaml_2.3.7	evaluate_0.21
## [113] codetools_0.2-19	tibble_3.2.1
## [115] cli_3.6.1	uwot_0.1.16
## [117] xtable_1.8-4	reticulate_1.30
## [119] munsell_0.5.0	Rcpp_1.0.11
## [121] GenomeInfoDb_1.36.0	globals_0.16.2
## [123] spatstat.random_3.1-5	png_0.1-8
## [125] parallel_4.3.3	ellipsis_0.3.2
## [127] dotCall64_1.0-2	bitops_1.0-7
## [129] listenv_0.9.0	viridisLite_0.4.2
## [131] scales_1.2.1	ggribes_0.5.4
## [133] crayon_1.5.2	leiden_0.4.3
## [135] purrr_1.0.1	GetoptLong_1.0.5
## [137] rlang_1.1.1	cowplot_1.1.1