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function [] = VolumeThresholdhelper(fileName)

VolumeThresholdhelper Information

VolumeThresholdhelper is used to help the user find parameters to use for batch processing with LC3B_Tandem_Puncta_QuantificationV2. The function takes in the fileName of a file in the current directory. It then asks the user to input parameters.

Function asks the user to enter the following information which is used for thresholding and identifying the cell boundaries. Three GUIs will be opened for the user to do this.

Three GUIs propompted to the user are:

1.Asks the user to input integers in the range [1,3] which is used to specify the order for the cell markers. This is, the channels for the nuclear marker, eGFP vacuoles and mCherry vacuoles.

2.Asks the user to input integers which are used to threshold the mature eGFP and mCherry vacuoles. Code assumes images are 16-bit so inputs should be in the range of [0, 65535] which specifies the intensity value for unit16 arrays.

3.Asks the user to input approximate volume sizes, in voxels, for the nuclear marker, eGFP vacuoles, mCherry vacuoles and the solidity for the vacuoles which is type float between [0,1].

Once the parameters are input by the user, the function will use the inputs to label each slice in the image data with yellow circles wherever it identifies a vacuole and presents the volume data using sliceViewer with the yellow circles overlaid for the users to check if the parameters worked or should be adjusted. A dialogue box is presented for the user to enter Y/N if they would like to try other parameters to improve the segmentation. Once satisfied the user can enter N and proceed to see what the Ridgelines and single cell containers look like to verify the SNR is high enough to perform segmentation with Otsu's method. Finally, subplots for the single cell data with the nuclear marker, eGFP and mCherry vacuoles are presented for viewing as isosurface plots. Note: user should place waitfor() after figures to stop the function at each plot for viewing in the functions WatershedOberserver and IsosurfaceVacuoles.

Once the user has identified appropriate parameters for their data they can use the parameters in LC3B_Tandem_Puncta_QuantificationV2 to perform batch processing for their data.

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Run while loop until user finds parameters for their data to use

```
while 1
%clear work space and cmd on each run
close all;
clc;
%store inputs in struct to return at the end so they know what they
decided
%worked for their data
Input.Nuclear channel = [];
Input.eGFP_channel = [];
Input.mCherry_channel = [];
Input.threshold_eGFP = [];
Input.threshold mCherry = [];
Input.minGFPvacVolume = [];
Input.minRFPvacVolume = [];
Input.minNucleusVolume = [];
Input.Solidity = [];
%get some info from the user
%ask user what order the channels are in
waitfor(msgbox('Please let us know what order the eGFP, mCherry and
nuclear marker are in'));
input = inputdlq({'Nuclear marker', 'eGFP', 'mCherry'},...
'Channels',1,{'1','2','3'},'on');
%convert char to double
Input.Nuclear_channel = str2double(input(1));
Input.eGFP channel = str2double(input(2));
Input.mCherry_channel = str2double(input(3));
%ask user to input thresholds for eGFP and mCherry channels
waitfor(msgbox('Please enter thresholds for developed puncta. This
 should be for developed eGFP and mCherry vacuoles which are used to
 identify neutral and acidified vacuoles, respectively.'));
input = inputdlg({'GFP Threshold Value 16-bit [0-65536]','RFP
 Threshold Value 16-bit [0-65536]'},...
'Thresholds',1,{'25000','20000'},'on');
%convert char to double
Input.threshold_eGFP = str2double(input(1));
Input.threshold mCherry = str2double(input(2));
%get min sizes in voxels for cell markers and solidity
input = inputdlg({'Minimum volume of GFP vacuole in voxels is >',...
    'Minimum volume of RFP vacuole in voxels is >',...
    'Minimum volume of Nucleus vacuole in voxels is >',...
    'Solidity Thershold Vacuoles [0,1]'},...
```

```
'Thresholds',1,{'1','15','15000','0.6'},'on');
%convert char to double
Input.minGFPvacVolume = str2double(input(1));
Input.minRFPvacVolume = str2double(input(2));
Input.minNucleusVolume = str2double(input(3));
Input.Solidity = str2double(input(4));
%unpack user inputs for passing to functions
Nuclear_channel = Input.Nuclear_channel;
eGFP_channel = Input.eGFP_channel;
mCherry_channel = Input.mCherry_channel;
eGFP threshold = Input.threshold eGFP;
mCherry_threshold = Input.threshold_mCherry;
MinvacVoxeGFP = Input.minGFPvacVolume;
MinvacVoxmCherry = Input.minRFPvacVolume;
MinNucVox = Input.minNucleusVolume;
Solidity = Input.Solidity;
```

Use bfopen to read file

```
img = bfopen(fileName);
name = fileName;
```

Create subvolumes for each channel

```
[r,g,b] =
imreadVolume(img,Nuclear_channel,eGFP_channel,mCherry_channel);
```

Label objects (vacuoles) with yellow circles and present to user with a slice viewer to check if parameters are ideal for segmentation

```
%set thresholds from user inputs for developed vacuoles.
%THIS IS ALL REDUNDANT TO THE VACUOLE FINDER AND IS JUST BEING USED TO
    UPDATE AND
%PERFECT THE VACUOLE SEGMENTATION

%convert input thresholds to double precision
max = 65536;min = 0;
Green_Thresh = (eGFP_threshold - min)/(max - min);
Red_Thresh = (mCherry_threshold - min)/(max - min);
```

```
%segment vacuoles
[pre green Vacs, green Vacs CC] = VacuoleCleansing(q, Green Thresh,...
   MinvacVoxeGFP, Solidity);
[red Vacs, red Vacs CC] =
VacuoleCleansing(r,Red_Thresh,MinvacVoxmCherry...
    ,Solidity);
%get intial stats for vacuoles
red_vacuoles_stats = regionprops3(red_Vacs_CC, red_Vacs, 'all');
pre_green_stats = regionprops3(green_Vacs_CC,pre_green_Vacs,'all');
%only keeps green vacoles if a red vacuole is associated with it. Due
to the
%nature of the probe this should be the case. It helps as a precheck
%ensuring the vacuole is indeed a LC3B vacuole.
[green_vacuoles_stats] = OverlappingChannels(pre_green_stats,...
   red_vacuoles_stats);
%now create the final binary for green vacuoles which overlap mcherry.
[~,idx] = intersect(pre_green_stats.EquivDiameter,...
   green_vacuoles_stats.EquivDiameter);
%return the updated indexes for eGFP binary image
bi_img = ismember(labelmatrix(green_Vacs_CC),idx);
%make sure to get the new connected components for updated eGFP stats
green_Vacs_CC = bwconncomp(bi_img);
%label the found vacuoles
label r = labelmatrix(red Vacs CC);
label_g = labelmatrix(green_Vacs_CC);
%display current stats for vacuoles. Will probably reduce the outputs
%the future but as of now everyhting from regionprops3 is returned
disp('Stats for eGFP vacuoles using current parameters')
disp(green_vacuoles_stats)
disp('Stats for mCherry vacuoles using current parameters')
disp(red_vacuoles_stats)
Stats for eGFP vacuoles using current parameters
   Volume
                     Centroid
                                 BoundingBox
             SubarrayIdx
                                                  Image
                 Extent VoxelIdxList
 EquivDiameter
                                                VoxelList
    PrincipalAxisLength
                                       Orientation
EigenVectors EigenValues
                               ConvexHull
                                                   ConvexImage
    ConvexVolume Solidity
                               SurfaceArea
                                              VoxelValues
      WeightedCentroid MeanIntensity MinIntensity
MaxIntensity
```

```
82 135.76 165.87 7.3659 [1×6 double] {1×7 double} {1×6 double} {1×4 double} {7×6×4 logical}
  0.84536 95.657 { 82×1 double} 135.78 165.84
7.3363 0.41188 0.20423 0.76669
  119 153.5 78.739 9.8319 [1×6 double] {1×7 double} {1×8 double} {1×5 double} {7×8×5 logical}
  80 153.5 95.713 9.65 [1x6 double] {1x6 double} {1x6 double} {1x4 double} {6x6x4 logical} 

5.346 0.55556 {80x1 double} {80x3 double} 5.6377 

5.5397 3.9046 -7.4139 -16 175.95 {3x3 double} 

{3x1 double} {41x3 double} {6x6x4 logical} 91 

0.87912 92.061 {80x1 double} 153.52 95.764 

9.7042 0.38502 0.18906 0.68329 

208 99.433 47 221 13 163 [1x6 double] {1x8
 208 99.433 47.221 13.163 [1x6 double] {1x8 double} {1x13 double} {1x8 double} {8x13x8 logical} 7.3511 0.25 {208x1 double} {208x3 double} 13.001 6.614 5.1231 85.642 15.116 -31.639 {3x3 double}
       \{3\times1\ double\} \{48\times3\ double\} \{8\times13\times8\ logical\} 331
 {3x1 double} {48x3 double} {8x13x8 logical} 331
0.6284 218.72 {208x1 double} 99.135 47.194
13.299 0.35992 0.18873 0.67758
75 166.73 221.29 12.947 [1x6 double] {1x8
double} {1x6 double} {1x3 double} {8x6x3 logical}
5.2322 0.52083 {75x1 double} {75x3 double} 7.2159
4.8882 3.221 -17.635 -2.074 -7.6291 {3x3 double}
{3x1 double} {39x3 double} {8x6x3 logical} 84
0.89286 91.13 {75x1 double} 166.7 221.32
12.974 0.3774 0.19127 0.65256
157 77.637 98.516 17.325 [1x6 double] {1x8
double} {1x8 double} {1x5 double} {8x8x5 logical}
 double} {1×8 double} {1×5 double} {8×8×5 logical} 6.6932 0.49062 {157×1 double} {157×3 double} 7.8519 6.6215 4.4466 -61.264 -4.5475 -3.587 {3×3 double}
  {3×1 double} {53×3 double} {8×8×5 logical} 181
0.8674 148.33 {157×1 double} 77.693 98.504
17.342 0.48249 0.18855 0.91596
Stats for mCherry vacuoles using current parameters
     Volume Centroid BoundingBox
                       SubarrayIdx
                                                                                                    Image
 EquivDiameter Extent VoxelIdxList VoxelList
PrincipalAxisLength Orientation
EigenVectors EigenValues ConvexHull ConvexImage
```

5

```
ConvexVolume Solidity SurfaceArea VoxelValues WeightedCentroid MeanIntensity MinIntensity
MaxIntensity
20 154.85 191.85 5.3 [1×6 double] {1×5 double} {1×3 double} {1×3 double} {5×3×3 logical} 3.3678 0.44444 {20×1 double} {20×3 double} 4.436 3.1484 2.4186 6.6043 -6.0081 -15.642 {3×3 double}
                                                                                                                                   21
   \{3\times1 \text{ double}\} \{21\times3 \text{ double}\} \{5\times3\times3 \text{ logical}\}
{3x1 double} {21x3 double} {5x3x3 logical} 21
0.95238 35.488 { 20x1 double} 154.87 191.83
5.2978 0.19315 0.13134 0.28973
100 135.58 166.7 6.5 [1x6 double] {1x7
double} {1x6 double} {1x4 double} {7x6x4 logical}
5.7588 0.59524 {100x1 double} {100x3 double} 6.825
5.6103 3.9092 -1.2438 15.362 178.19 {3x3 double}
5.6103 3.9092 -1.2438 15.362 178.19 {3x3 double} {3x1 double} {40x3 double} {7x6x4 logical} 108 0.92593 108.12 {100x1 double} 135.65 166.72 6.5692 0.26936 0.1177 0.48489 33 154.64 199.7 5.8788 [1x6 double] {1x5 double} {1x6 double} {1x3 double} {5x6x3 logical} 3.9796 0.36667 {33x1 double} {33x3 double} 6.0842 4.5438 2.7361 -83.854 9.9176 -11.34 {3x3 double}
4.5438 2.7361 -83.854 9.9176 -11.34 {3x3 double} {3x1 double} {28x3 double} {5x6x3 logical} 49 0.67347 61.071 { 33x1 double} 154.73 199.73 5.8806 0.15794 0.11916 0.21847 25 146.52 240.88 7.44 [1x6 double] {1x5 double} {1x4 double} {1x3 double} {5x4x3 logical} 3.6278 0.41667 { 25x1 double} { 25x3 double} 5.2829 3.8198 2.3845 -2.3753 -9.2406 169.34 {3x3 double}
{3\times1 \text{ double}} {37\times3 \text{ double}} {6\times6\times5 \text{ logical}} 96
0.83333 93.427 { 80×1 double} 153.61 79.822

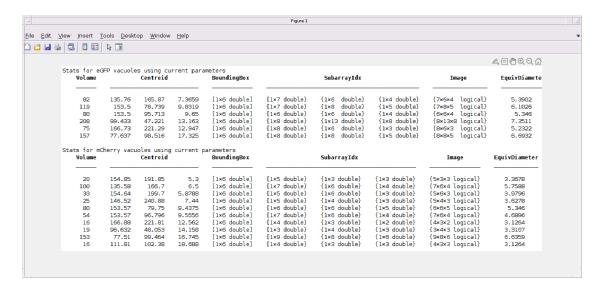
9.4625 0.24282 0.11853 0.42815

54 153.57 96.796 9.5556 [1×6 double] {1×7
double} {1x6 double} {1x4 double} {7x6x4 logical}
4.6896 0.32143 {54x1 double} {54x3 double} 6.02
4.412 3.3901 -23.32 -14.219 -0.83694 {3x3 double}
{3×1 double} {36×3 double} {7×6×4 logical} 66
0.81818 73.87 { 54×1 double} 153.56 96.863
9.6123 0.24218 0.11795 0.40777
```

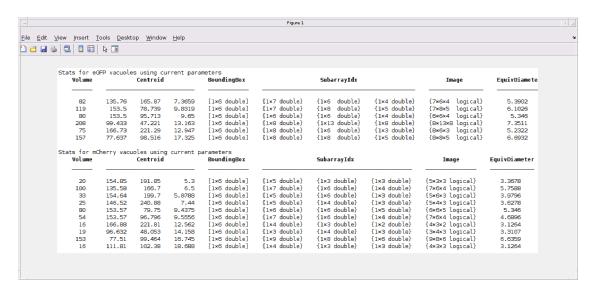
```
166.88 221.81 12.562 [1×6 double] {1×4 {1×3 double} {1×2 double} {4×3×2 logical} 0.66667 { 16×1 double} { 16×3 double} 4.2419
double }
3.1264
             2.0535 -8.3792 -1.668 25.333 {3×3 double}
   \{3\times1\ double\} \{23\times3\ double\} \{4\times3\times2\ logical\}
   0.94118 30.521 { 16×1 double} 166.85
                                                                 221.86
             0.18293 0.13857 0.24761
12.558
            96.632 48.053 14.158 [1×6 double] {1×3 {1×4 double} {1×3 double} {3×4×3 logical} 0.52778 { 19×1 double} { 19×3 double} 4.6708
   19
double }
3.3107
             2.6087 -59.719 -19.712 -1.4611 {3×3 double}
 2.6999
   {3\times1 \text{ double}} {26\times3 \text{ double}} {3\times4\times3 \text{ logical}}
   0.90476 34.425 { 19×1 double} 96.603
                                                                 48.081
                                                  0.3221
14.211
             0.20339 0.12047
           77.51 99.464 16.745 [1x6 double] {1x9 {1x8 double} {1x6 double} {9x8x6 logical} 0.35417 {153x1 double} {153x3 double} 7.8497
   153
double }
6.6359
   7.155 4.2759 -56.692 -9.258 4.9168 {3×3 double} {3×1 double} {44×3 double} {9×8×6 logical} 188
  7.155
   0.81383 153.03 {153×1 double} 77.569
                                                                 99.484
              0.3136 0.11792 0.60988
16.826
          111.81 102.38 18.688 [1×6 double] {1×4 {1×3 double} {1×3 double} {4×3×3 logical} 0.44444 { 16×1 double} { 16×3 double} 4.282
   16
double }
3.1264
 2.8013 2.3329 -31.766 10.015 39.377 {3x3 double}
  {3\times1 \text{ double}} {20\times3 \text{ double}} {4\times3\times3 \text{ logical}}
  0.94118 30.352 { 16×1 double} 111.81
                                                                 102.35
18.679 0.19713
                                 0.1436 0.29888
```

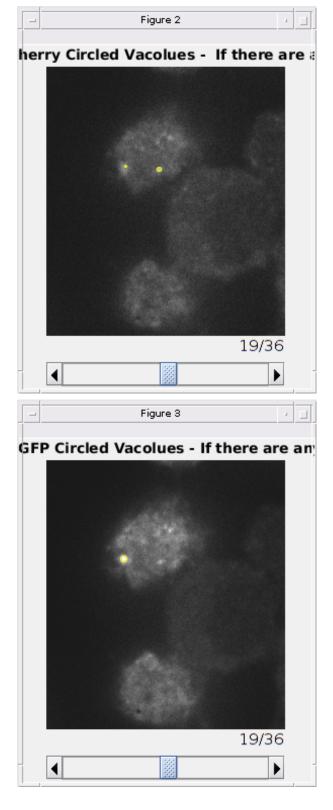
%The published out for disp looks horrible without using a Latex
wrapper

%so I've included an image of how it looks in the command window.
%I need to add a Latex wrapper for this
imshow('StatsCmdOutput.png')
snapnow

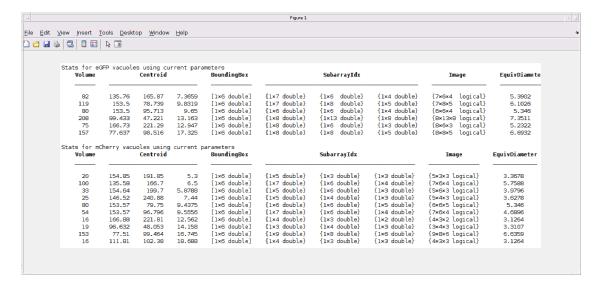


Label vacuoles with circles and display using sliceViewer





end



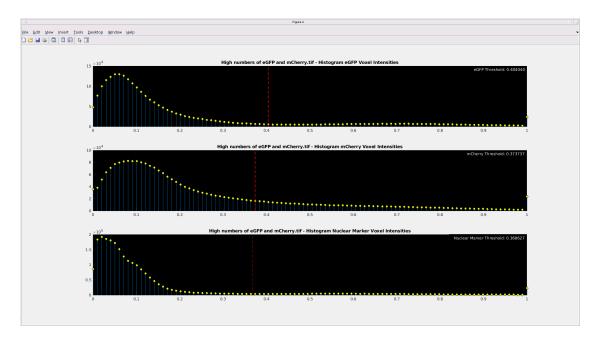
%if user likes the parameters used, proceed and show them what is created

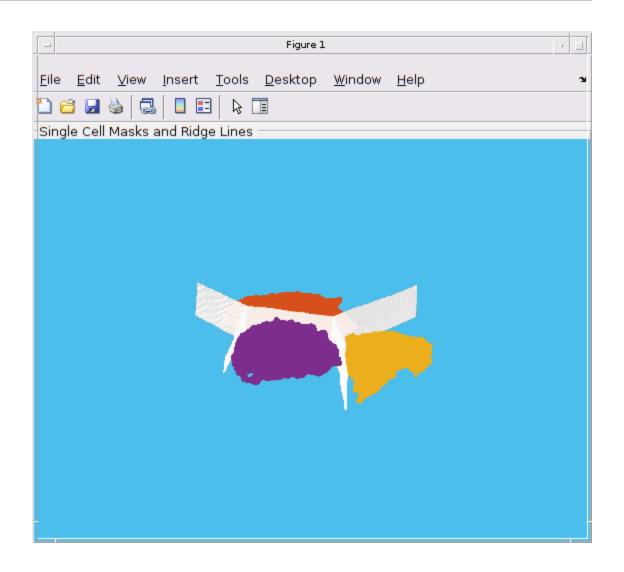
%using the rest of the script.

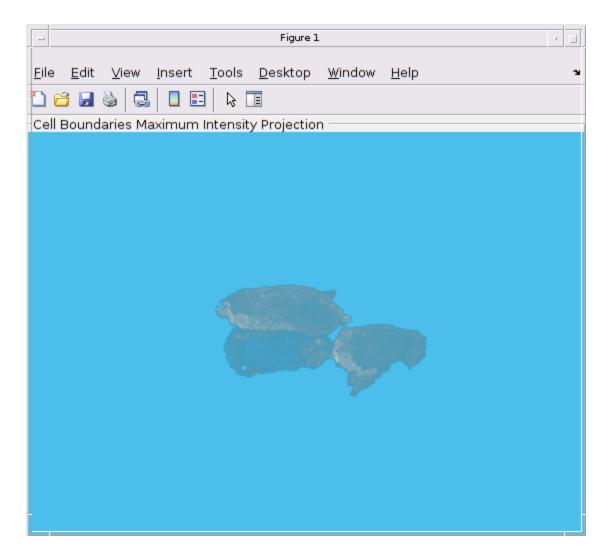
%segment cells and get ROIs

[bw_stats] = Watershed2(r,g,b,name,MinNucVox,true);

Data for plots generated from - High numbers of eGFP and mCherry.tif





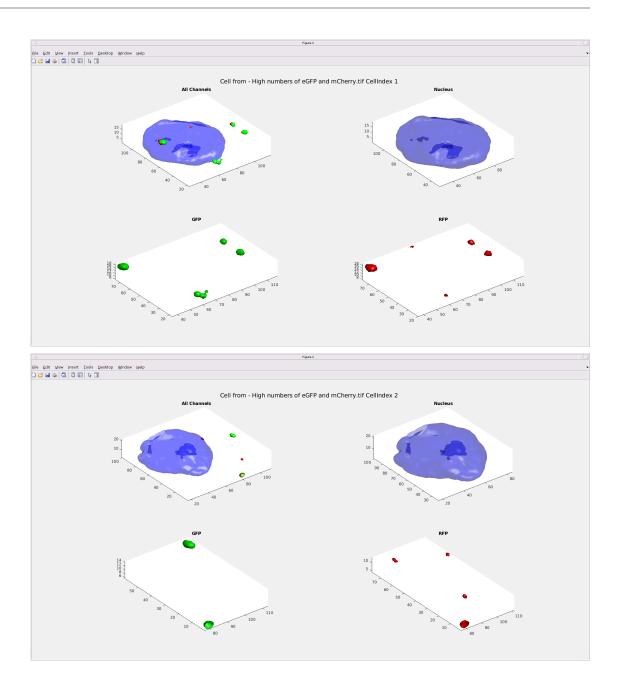


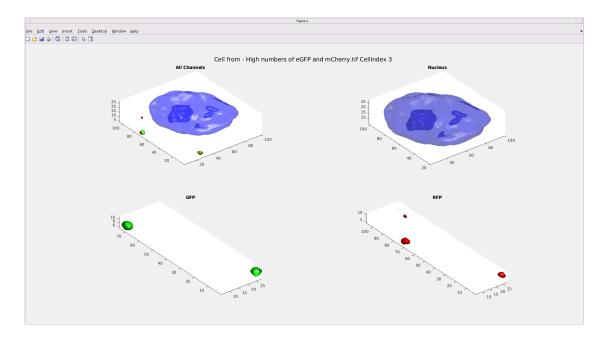
 $find\ vacuoles\ in\ single\ cells\ and\ return\ the\ results\ in\ a\ structure\ [SingleCellStructure] =$

FindVaculoes(r,g,b,bw_stats,name,eGFP_threshold,...

mCherry_threshold,MinvacVoxeGFP,MinvacVoxmCherry,MinNucVox,Solidity);

%close all figures and clear command window as we are done close all; clc;



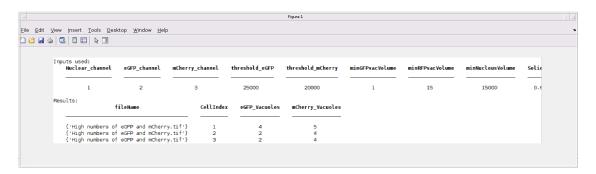


Accumulate results

```
dat.FileName = name;
dat.SingleCell = SingleCellStructure;
%return single cell structure containg counts as a table
[Results_Table] = TabularCountVacuoles(dat);
%report the parameters the user decided worked for their data
disp('Inputs used: ')
disp(struct2table(Input))
Inputs used:
   Nuclear channel eGFP channel mCherry channel
 threshold_eGFP threshold_mCherry
                                       minGFPvacVolume
minRFPvacVolume
                   minNucleusVolume
                                       Solidity
                                                              25000
             20000
                                                     15
 15000
                 0.6
%diplay the fruits of your labor for this image using the inputs
accepted
disp('Results: ')
disp(Results_Table)
Results:
                   fileName
                                               CellIndex
eGFP_Vacuoles
                 mCherry_Vacuoles
```

```
{'High numbers of eGFP and mCherry.tif'} 1 4
5
{'High numbers of eGFP and mCherry.tif'} 2 2
4
{'High numbers of eGFP and mCherry.tif'} 3 2
```

%The published display looks horrible without using a Latex wrapper so
I've
%included an image of how it looks in the command window.
%I need to add a Latex wrapper for this
imshow('Resultscmd.png')
snapnow



end

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