# Package 'MetabolomicsPipeline'

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**Title** Metabolomics Pipeline Tools

**Version** 0.99.0

## **Description**

This package provides analysis tools for analyzing metabolomics data from Metabolon. The tools in this package compliment the analysis from Metabolon. We provide functionality for hypothesis testing at

the subpathway level, pairwise comparisons of metabolites, and tools for exploratory analysis.

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MetabolomicsPipeline-package

MetabolomicsPipeline: Metabolomics Pipeline Tools

## **Description**

This package provides analysis tools for analyzing metabolomics data from Metabolon. The tools in this package compliment the analysis from Metabolon. We provide functionality for hypothesis testing at the subpathway level, pairwise comparisons of metabolites, and tools for exploratory analysis.

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## See Also

Useful links:

- https://github.com/datalifecycle-ua/MetabolomicsPipeline
- $\bullet \ \ Report \ bugs \ at \ https://github.com/datalifecycle-ua/MetabolomicsPipeline/issues$

all\_sig\_subpath 3

 $all\_sig\_subpath$ 

Table of Significant Subpathways

## **Description**

Create a table of all significant subpathways

## Usage

```
all_sig_subpath(path_results)
```

## **Arguments**

```
path_results Results data frame generated by subpathway_analysis
```

#### Value

A table of all significant subpathways. Including the significant model type and model type p-value.

```
# Load data
data("demoDataSmall", package = "MetabolomicsPipeline")
dat <- demoDataSmall</pre>
# Runsubpathay analysis
sub_analysis <- subpathway_analysis(dat,</pre>
  treat_var = "GROUP_NAME",
  block_var = "TIME1",
  strat_var = NULL,
  Assay = "normalized"
)
# significant subpathways by model type
subpath_by_model(sub_analysis)
# Percentage of signficant subpathways within superpathways
subpath_within_superpath(sub_analysis)
met_within_sub(sub_analysis, subpathway = "Aminosugar Metabolism")
# All signifiicant subpathways
all_sig_subpath(sub_analysis)
```

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create\_heatmap\_Data

Create metadata and matricies for metabolite heatmaps

## **Description**

This function creates the required matrices for the metabolite heatmaps.

## Usage

```
create_heatmap_Data(data, heatmap_variables, Assay = "normalized", ...)
```

## **Arguments**

data A SummarizedExperiment containing Metabolon data.

heatmap\_variables

A vector of variable names that are NOT metabolites.

Assay Name of assay data to be used for heatmaps. Default="normalized".

... Additional arguments that can be passed into the arrange function. This param-

eter will order the columns of the heatmap data.

#### Value

A list of matrices including the heatmap variable (meta data for heatmap) and the values for the heatmap.

demoDat

Demo data for the MetabolomicsPipeline,

## **Description**

Demo data consisting of 86 samples (42 males, 44 females), three treatment groups, and the samples were taken

# Usage

```
data(demoDat)
```

## **Format**

SummarizedExperiment object

## Value

A SummarizedExperiment object with 86 samples

demoDataSmall 5

demoDataSmall	Subset of Demo data	for the MetabolomicsPipeline,
acmobacaomarr	Subset of Denie date	joi me mendo connest ipenne,

## **Description**

Demo data consisting of 86 samples (42 males, 44 females), three treatment groups, and the samples were taken at three different time points. We focus on a subset of 10 Subpathways.

## Usage

```
data(demoDataSmall)
```

## **Format**

Rd

#### Value

A subset of the metabolites in the DemoData.

loadMetabolon Load Metabolomic Data as SummarizedExperiment

## **Description**

Automatically load metabolomic data from Metabolon

# Usage

```
loadMetabolon(
  path,
  raw_sheet = "Peak Area Data",
  chemical_sheet = "Chemical Annotation",
  sample_meta = "Sample Meta Data",
  normalized_peak = "Log Transformed Data",
  sample_names = "PARENT_SAMPLE_NAME",
  chemicalID = "CHEM_ID"
)
```

## **Arguments**

path Path to Metabolon .xlsx file containg peak data, chemical annotations, sample

meta data, and (optionally) the normalized peak counts

raw\_sheet Sheet name for the raw peak data.
chemical\_sheet Sheet name for chemical annotation.
sample\_meta Sheet name for sample meta data.
normalized\_peak

Sheet name for the normalized peak data. If you are not adding the normalized data from the excel file then set normalized\_peak=NA.

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sample_names	Column name in the meta data containing the sample names. This must correspond to the row names of the raw peak data in the excel file.
chemicalID	Column name in the meta data containing the sample names. This must correspond to the column names of the raw peak data.

#### **Details**

The Metabolon experiment data are stored in a SummarizedExperiment.

#### Value

A SummarizedExperiment containing Metabolon expirement data.

#### See Also

SummarizedExperiment::SummarizedExperiment

# **Description**

This function log transforms each metabolite in the Metabolon data.

## Usage

```
log_transformation(peak_data)
```

#### **Arguments**

peak\_data A matrix of peak data with metabolites in the columns

#### Value

log transformed peak data

```
data("demoDataSmall", package = "MetabolomicsPipeline")
peak <- SummarizedExperiment::assay(demoDataSmall, "peak")

# Median standardization
peak_med <- median_standardization(peak_data = peak)

# Min value imputation
peakImpute <- min_val_impute(peak_data = peak_med)

# log transformation
peak_log <- log_transformation(peak_data = peakImpute)</pre>
```

median\_standardization 7

```
median_standardization
```

Median standardization for metabolite data

## **Description**

This function standardizes the metabolites by the median of the metabolite.

## Usage

```
median_standardization(peak_data)
```

#### **Arguments**

peak\_data

Peak data with metabolites in the columns. The data also must include the "PARENT\_SAMPLE\_NAME".

## Value

Median standardized peak data.

# **Examples**

```
data("demoDataSmall", package = "MetabolomicsPipeline")
peak <- SummarizedExperiment::assay(demoDataSmall, "peak")

# Median standardization
peak_med <- median_standardization(peak_data = peak)

# Min value imputation
peakImpute <- min_val_impute(peak_data = peak_med)

# log transformation
peak_log <- log_transformation(peak_data = peakImpute)</pre>
```

metabolite\_heatmap

Create metabolite heatmap

# Description

Create heatmaps which are arranged by the experimental conditions.

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## Usage

```
metabolite_heatmap(
  data,
  top_mets = 50,
  group_vars,
  strat_var = NULL,
  caption = NULL,
  Assay = "normalized",
  sample_names = "PARENT_SAMPLE_NAME",
  ...
)
```

# **Arguments**

data	A SummarizedExperiment containing the Metabolon experiment data.
top_mets	Number of metabolites to include in the heatmap. Metabolites are chosen based on the highest variability.
group_vars	Vector of variables to annotate heatmap with. Columns will be grouped by these variables.
strat_var	Variable to stratify the heatmap by.
caption	A title for the heatmap. If strat_var is used, the title will automatically include the stratum with the tile.
Assay	Which assay data to use for the heatmap (default="normalized").
sample_names	Column name in the meta data containing the sample names. This must correspond to the row names of the raw peak data in the excel file.
	Additional arguments can be passed into the arrange function. This parameter will order the columns of the heatmap.

# Value

A gtable class with all of the information to build the heatmap. To view the heatmap use ggplotify::as.ggplot().

```
# load data
data("demoDat", package = "MetabolomicsPipeline")
dat <- demoDat

# Heatmap with one group
treat_heatmap <- metabolite_heatmap(dat,
    top_mets = 50,
    group_vars = "GROUP_NAME",
    strat_var = NULL,
    caption = "Heatmap Arranged By Group",
    Assay = "normalized",
    GROUP_NAME
)</pre>
```

metabolite\_pairwise 9

metabolite\_pairwise M

Metabolite Pairwise Comparisons.

## **Description**

Computes the pairwise comparison estimates and p-values for each metabolite.

## Usage

```
metabolite_pairwise(
  data,
  form,
  Assay = "normalized",
  strat_var = NULL,
  mets = NULL
)
```

## **Arguments**

data	SummarizedExperiment with Metabolon experiment data.
form	This is a character string the resembles the right hand side of a simple linear regression model in R. For example form = " $Group1 + Group2$ ".
Assay	Name of the assay to be used for the pairwise analysis (default='normalized')
strat_var	A variable in the analysis data to stratify the model by. If this is specified, a list of results will be returned.
mets	Chemical ID for the metabolites of interest. If NULL then the pairwise analysis is completed for all metabololites.

# Details

This function will analyze each metabolite individually. For each metabolite, the metabolite\_pairwise function will first test whether the model explained a significant proportion of the variance in the metabolite using an F-test. Since we will be looking at multiple comparisons for the metabolite, it is good practice to first look at the overall p-value from the F-test before looking at the pairwise comparisons. The metabolite\_pairwise function then looks at all pairwise comparisons utilizing the emmeans package. The metabolite\_pairwise function returns a data frame with the metabolite overall p-value, log fold change for each group, and the p-value for each comparison.

## Value

The overall F-test p-value, and the estimate and pvalue for each pairwise comparison.

```
# Load data
data("demoDataSmall", package = "MetabolomicsPipeline")
dat <- demoDataSmall

# Run pairwise analysis
strat_pairwise <- metabolite_pairwise(dat,
    form = "GROUP_NAME*TIME1",</pre>
```

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```
strat_var = "Gender"
met_est_heatmap(strat_pairwise$Female, dat,
      interactive = FALSE.
      CHEM_ID = "CHEM_ID", SUB_PATHWAY = "SUB_PATHWAY",
      CHEMICAL_NAME = "CHEMICAL_NAME",
      main = "Log fold change heatmap", show_rownames = FALSE
)
# Female
met_p_heatmap(strat_pairwise$Female, dat,
     interactive = FALSE, show_rownames = FALSE,
     main = "Pvalue Heatmap"
)
```

metabolite\_pca

Metabolite PCA

# **Description**

Computes and plots the first two components of the PCA from the metabolite data.

## Usage

```
metabolite_pca(data, Assay = "normalized", meta_var)
```

# **Arguments**

data SummarizedExperiment with Metabolon experiment data.

Assay Name of the assay to be used for the pairwise analysis (default='normalized')

meta\_var A metadata variable to color code the PCA plot by.

## Value

A PCA plot of the first two principal components, colored by the metadata variable.

```
# load data
data("demoDat", package = "MetabolomicsPipeline")
dat <- demoDat
# Define PCA label from metadata</pre>
```

met\_est\_heatmap 11

```
meta_var <- "Gender"

# Run PCA
pca <- metabolite_pca(dat,
    meta_var = meta_var
)

# Show PCA
pca</pre>
```

met\_est\_heatmap

Metabolite Pairwise Estimate Interactive Heatmap.

# Description

Produce an interactive heatmap of the estimates produced in metabolite\_pairwise.

# Usage

```
met_est_heatmap(
  results_data,
  data,
  interactive = FALSE,
  SUB_PATHWAY = "SUB_PATHWAY",
  CHEMICAL_NAME = "CHEMICAL_NAME",
  plotlyTitle = "Metabolite log fold change",
  ...
)
```

# Arguments

results_data	Results data frame of the pairwise comparisons produced by metabolite_pairwise.
data	A SummarizedExperiment containing the Metabolon experiment data.
interactive	boolean (T/F) for whether or not the plot should be interactive. Use interactive=T to produce an interactive plot using plotly. Use interactive=F to produce a static heatmap using pheatmap.
SUB_PATHWAY	Column name in the chemical annotation worksheet which contains the subpathway information.
CHEMICAL_NAME	Column name in the chemical annotation worksheet which contains the chemical name.
plotlyTitle	Title for the interactive heatmap.
	Additional arguments that can be passed to pheatmap.

#### **Details**

This function will produce a heatmap of the log fold changes for the metabolites with a significant overall p-value (which tested if the treatment group means were equal under the null hypothesis). The heatmap colors will only show if the log fold-change is greater than log(2) or less than log(.5). Therefore, this heatmap will only focus on comparisons with a fold change of two or greater.

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#### Value

An interactive heatmap of pairwise estimates.

# **Examples**

```
# Load data
data("demoDataSmall", package = "MetabolomicsPipeline")
dat <- demoDataSmall</pre>
# Run pairwise analysis
strat_pairwise <- metabolite_pairwise(dat,</pre>
  form = "GROUP_NAME*TIME1",
  strat_var = "Gender"
)
met_est_heatmap(strat_pairwise$Female, dat,
        interactive = FALSE,
         SUB_PATHWAY = "SUB_PATHWAY",
        CHEMICAL_NAME = "CHEMICAL_NAME",
        plotlyTitle = "Metabolite log fold change",
        main = "Log fold change heatmap", show_rownames = FALSE
)
# Female
met_p_heatmap(strat_pairwise$Female, dat,
       interactive = FALSE, show_rownames = FALSE,
       plotlyTitle = "P-Value Heatmap",
       main = "Pvalue Heatmap"
)
```

met\_p\_heatmap

Metabolite Pairwise P-Value Interactive Heatmap.

## **Description**

Produce an interactive heatmap of the p-values produced in metabolite\_pairwise.

#### Usage

```
met_p_heatmap(
  results_data,
  data,
  interactive = FALSE,
```

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```
SUB_PATHWAY = "SUB_PATHWAY",
CHEMICAL_NAME = "CHEMICAL_NAME",
plotlyTitle = "P-Value Heatmap",
...
)
```

#### **Arguments**

results\_data Results data frame of the pairwise comparisons produced by metabolite\_pairwise. A SummarizedExperiment containing Metabolon experiment data. data interactive boolean (T/F) for whether or not the plot should be interactive. Use interactive=T to produce an interactive plot using plotly. Use interactive=F to produce a static heatmap using pheatmap. SUB\_PATHWAY Column name in the chemical annotation worksheet which contains the subpathway information. Column name in the chemical annotation worksheet which contains the chemi-CHEMICAL\_NAME cal name. plotlyTitle Title for the interactive heatmap. Additional arguments that can be passed to pheatmap.

#### **Details**

For the metabolites which had a significant overall p-value (which tested if the treatment group means were equal under the null hypothesis), we will produce a heatmap of the p-values.

#### Value

An interactive heatmap of pairwise p-values.

```
# Load data
data("demoDataSmall", package = "MetabolomicsPipeline")
dat <- demoDataSmall</pre>
# Run pairwise analysis
strat_pairwise <- metabolite_pairwise(dat,</pre>
  form = "GROUP_NAME*TIME1",
  strat_var = "Gender"
)
met_est_heatmap(strat_pairwise$Female, dat,
          interactive = FALSE,
          CHEM_ID = "CHEM_ID", SUB_PATHWAY = "SUB_PATHWAY",
          CHEMICAL_NAME = "CHEMICAL_NAME",
          plotlyTitle = "Estimate Heatmap",
          main = "Log fold change heatmap", show_rownames = FALSE
)
```

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met\_within\_sub

Metabolites within Subpathway Table

## **Description**

Return the model results for each metabolite within a subpathway.

## Usage

```
met_within_sub(
   subpath_results,
   subpathway,
   mod = c("interaction", "parallel", "single")
)
```

## **Arguments**

subpath\_results

Results data frame generated by subpathway\_analysis

subpathway

Character string of the subpathway of interest. This is case sensitive and must

be listed in the subpath\_results.

mod

Model of interest. This can be a single model or a vector of model types that

can take on the values "interaction", "parallel", or "single".

# Value

A table with the results from the model types specified and for each metabolite within the superpathway specified.

```
data("demoDataSmall", package = "MetabolomicsPipeline")
dat <- demoDataSmall

# Runsubpathay analysis
sub_analysis <- subpathway_analysis(dat,
    treat_var = "GROUP_NAME",
    block_var = "TIME1",
    strat_var = NULL,</pre>
```

min\_val\_impute 15

min\_val\_impute

Minimum Value Imputation

## **Description**

Imputes the minimum value for each metabolite

## Usage

```
min_val_impute(peak_data)
```

# **Arguments**

peak\_data

Peak data matrix with metabolites in the columns.

#### Value

Metabolite imputed peak data.

```
data("demoDataSmall", package = "MetabolomicsPipeline")
peak <- SummarizedExperiment::assay(demoDataSmall, "peak")

# Median standardization
peak_med <- median_standardization(peak_data = peak)

# Min value imputation
peakImpute <- min_val_impute(peak_data = peak_med)

# log transformation
peak_log <- log_transformation(peak_data = peakImpute)</pre>
```

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pairwise	Pairwise function
----------	-------------------

## **Description**

This is the main function for metabolite\_pairwise

## Usage

```
pairwise(out, form, data)
```

#### **Arguments**

out Outcome used as reponse form form of the model data data used for modeling

#### Value

Pairwise comparisons for a single metabolite.

```
subpathway_analysis
Subpathway Analysis
```

# Description

Subpathway analysis for metabolite data.

## Usage

```
subpathway_analysis(
  data,
  treat_var,
  block_var = NULL,
  strat_var = NULL,
  Assay = "normalized",
  subPathwayName = "SUB_PATHWAY",
  chemName = "CHEMICAL_NAME",
  superPathwayName = "SUPER_PATHWAY"
)
```

## **Arguments**

data SummarizedExperiment with Metabolon experiment data.

treat\_var This is the name of the variable in the analysis data that is the main variable of

interest.

block\_var This is the name of the blocking variable in the dataset. If the the experimental

design does not include a blocking variable, then the value of block\_var=NULL.

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strat\_var Variable to stratify the subpathway analysis by. This is set to NULL by default

and will not stratify the analysis unless specified.

Assay Name of the assay to be used for the pairwise analysis (default='normalized')

subPathwayName Column name for subpathway variable as defined in the chemical annotation

worksheet.

chemName Column name for chemical name variable as defined in the chemical annotation

worksheet.

superPathwayName

Column name for super-pathway variable as defined in the chemical annotation worksheet.

#### **Details**

For each metabolite, we test three models using using ANOVA.

1. Interaction: logPeak = Treatment + block + Treatment \* block

2. Parallel: logPeak = Treatment + block

3. Single: logPeak = Treatment

For the interaction model, we are focusing only on the interaction term "Treatment\*block" to test if there is a significant interaction between our treatment and the block variable. The parallel model tests if the block variable explains a significant amount of the metabolite variance, and the treatment model tests if the treatment explains a significant proportion of the variance for each metabolite. Then, we use the Combined Fisher probability to test each model at the subpathway level.

$$\tilde{X} = -2\sum_{i=1}^{k} \ln(p_i)$$

where k is the number of metabolites in the subpathway. We can get a p-value from  $P(X \ge \tilde{X})$ , knowing that  $\tilde{X} \sim \chi^2_{2k}$ . You will notice that smaller p-values will lead to a larger  $\tilde{X}$ .

## Value

A data frame with "CHEM\_ID", "sub\_pathway", "chem\_name", "interaction\_pval", "interaction\_fisher", "parallel\_pval", "resingle\_pval", "single\_fisher", and "model" for each metabolite.

#### See Also

Loughin, Thomas M. "A systematic comparison of methods for combining p-values from independent tests." Computational statistics & data analysis 47.3 (2004): 467-485.

```
# Load data
data("demoDataSmall", package = "MetabolomicsPipeline")
dat <- demoDataSmall

# Runsubpathay analysis
sub_analysis <- subpathway_analysis(dat,
    treat_var = "GROUP_NAME",
    block_var = "TIME1",
    strat_var = NULL,
    Assay = "normalized"</pre>
```

subpathway\_boxplots

Subpathway Boxplots

## **Description**

Creates boxplots for each metabolite within a specified subpathway.

## Usage

```
subpathway_boxplots(
  data,
  subpathway,
  block_var,
  treat_var,
  Assay = "normalized",
  CHEMICAL_NAME = "CHEMICAL_NAME",
  SUB_PATHWAY = "SUB_PATHWAY",
  ...
)
```

## **Arguments**

data	SummarizedExperiment with Metabolon experiment data.
subpathway	Character value of the subpathway of interest. This is case sensitive and must be in the chemical annotation file.
block_var	This the the name of the variable in the meta data that is used for the $X$ axis of the box plots. We recommend using the "block_var" from the subpathway analysis.
treat_var	This is a grouping variable. As a recommendation the treatment groups should be used in the treat_var argument as this will provide a different color for each of the treatments making it easier to identify.
Assay	Name of the assay to be used for the pairwise analysis (default='normalized')

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CHEMICAL_NAME	Column name in the chemical annotation worksheet which contains the chemical name.
SUB_PATHWAY	Column name in chemical annotation file which contains the SUB_PATHWAY information
	Additional arguments to filter the analysis data by.

## **Details**

.

#### Value

Boxplots stratified by metabolites.

```
# load data
data("demoDat", package = "MetabolomicsPipeline")
dat <- demoDat
subpathway_boxplots(dat,
  subpathway = "Lactoyl Amino Acid", block_var = TIME1,
  treat_var = GROUP_NAME, Assay = "normalized",
  CHEMICAL_NAME = "CHEMICAL_NAME",
  SUB_PATHWAY = "SUB_PATHWAY", Gender == "Female"
)
# Set up data
dat$TIME1 <- as.numeric(factor(dat$TIME1,</pre>
  levels = c("PreSymp", "Onset", "End")
))
# Create line plots
subpathway_lineplots(dat,
           subpathway = "Lactoyl Amino Acid",
           block_var = TIME1, treat_var = GROUP_NAME,
           Assay = "normalized",
           CHEMICAL_NAME = "CHEMICAL_NAME",
           SUB_PATHWAY="SUB_PATHWAY",Gender == "Female")
```

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```
subpathway_lineplots Subpathway Lineplots
```

# Description

Create line plots for each metabolite within a subpathway.

# Usage

```
subpathway_lineplots(
  data,
  subpathway,
  block_var,
  treat_var,
  Assay = "normalized",
  CHEMICAL_NAME = "CHEMICAL_NAME",
  SUB_PATHWAY = "SUB_PATHWAY",
  ...
)
```

# **Arguments**

data	SummarizedExperiment with Metabolon experiment data.
subpathway	Character value of the subpathway of interest. This is case sensitive and must be in the chemical annotation file.
block_var	This the the name of the variable in the meta data that is used for the X axis of the line plots. We recommend using the "block_var" variable from the subpathway analyis.
treat_var	This is a grouping variable. As a recommendation the treatment groups should be used in the groupBy argument as this will provide a different color for each of the treatments making it easier to identify.
Assay	Name of the assay to be used for the pairwise analysis (default='normalized')
CHEMICAL_NAME	Column name in the chemical annotation worksheet which contains the chemical name.
SUB_PATHWAY	Column name in the chemical annotation worksheet which contains the subpathway information.
	Additional arguments to filter the analysis data by.

## Value

Line plots stratified by metabolite.

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```
subpathway_boxplots(dat,
  subpathway = "Lactoyl Amino Acid", block_var = TIME1,
  treat_var = GROUP_NAME, Assay = "normalized",
  CHEMICAL_NAME = "CHEMICAL_NAME",
  SUB_PATHWAY = "SUB_PATHWAY", Gender == "Female"
)
# Set up data
dat$TIME1 <- as.numeric(factor(dat$TIME1,</pre>
  levels = c("PreSymp", "Onset", "End")
))
# Create line plots
subpathway_lineplots(dat,
  subpathway = "Lactoyl Amino Acid",
  block_var = TIME1, treat_var = GROUP_NAME,
  Assay = "normalized",
  CHEMICAL_NAME = "CHEMICAL_NAME",
  SUB_PATHWAY = "SUB_PATHWAY", Gender == "Female"
```

subpath\_by\_model

Subpathway model type table

## Description

Create a table with the number of significant subpathways for each model type.

## Usage

```
subpath_by_model(subpath_results)
```

#### **Arguments**

subpath\_results

Results data frame generated by subpathway\_analysis

## **Details**

Each subpathway will only have one model type. We first test the interaction, and then the parallel and single models are tested last. Suppose a subpathway has a significant interaction model type. In that case, the table will count it as an interaction and not as a parallel or single.

## Value

A table of the number of significant subpathways by model type.

## **Examples**

```
# Load data
data("demoDataSmall", package = "MetabolomicsPipeline")
dat <- demoDataSmall</pre>
# Runsubpathay analysis
sub_analysis <- subpathway_analysis(dat,</pre>
  treat_var = "GROUP_NAME",
  block_var = "TIME1",
  strat_var = NULL,
  Assay = "normalized"
)
# significant subpathways by model type
subpath_by_model(sub_analysis)
# Percentage of signficant subpathways within superpathways
subpath_within_superpath(sub_analysis)
met_within_sub(sub_analysis, subpathway = "Aminosugar Metabolism")
```

subpath\_within\_superpath

Proportion of the Significant Subpathways Within Superpathways

## **Description**

Create a table that gives the percentage of significant subpathways within each superpathway.

## Usage

```
subpath_within_superpath(subpath_results)
```

#### **Arguments**

```
subpath_results
```

Results data frame generated by subpathway\_analysis

#### Value

A table with the proportion (and percent) of significant subpathways within superpathways.

```
# Load data
data("demoDataSmall", package = "MetabolomicsPipeline")
dat <- demoDataSmall</pre>
```

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