

# Package ‘MetabolomicsPipeline’

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**Title** Metabolomics Pipeline Tools

**Version** 0.99.0

## Description

This package provides analysis tools for analyzing metabolomics data from Metabolon. The tools in this package compliment the analysis from Metabolon. We provide functionality for hypothesis testing at the subpathway level, pairwise comparisons of metabolites, and tools for exploratory analysis.

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**VignetteBuilder** knitr

**URL** <https://github.com/JoelParkerUofA/MetabolomicsPipeline>

## BugReports

<https://github.com/JoelParkerUofA/MetabolomicsPipeline/issues>

**NeedsCompilation** no

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---

MetabolomicsPipeline-package

*MetabolomicsPipeline: Metabolomics Pipeline Tools*

---

## Description

This package provides analysis tools for analyzing metabolomics data from Metabolon. The tools in this package compliment the analysis from Metabolon. We provide functionality for hypothesis testing at the subpathway level, pairwise comparisons of metabolites, and tools for exploratory analysis.

## Author(s)

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Authors:

- Bonnie Lafleur <blafleur@arizona.edu>

## See Also

Useful links:

- <https://github.com/JoelParkerUofA/MetabolomicsPipeline>
- Report bugs at <https://github.com/JoelParkerUofA/MetabolomicsPipeline/issues>

---

all_sig_subpath	<i>Table of Significant Subpathways</i>
-----------------	---

---

**Description**

Create a table of all significant subpathways

**Usage**

```
all_sig_subpath(path_results)
```

**Arguments**

path\_results Results data frame generated by [subpathway\\_analysis](#)

**Value**

A table of all significant subpathways. Including the significant model type and model type p-value.

**Examples**

```
# Load data
data("demoDataSmall", package = "MetabolomicsPipeline")
dat <- demoDataSmall

# Runsubpathay analysis
sub_analysis <- subpathway_analysis(dat,
  treat_var = "GROUP_NAME",
  block_var = "TIME1",
  strat_var = NULL,
  Assay = "normalized"
)

#####
### Results Plots #####
#####

# significant subpathways by model type
subpath_by_model(sub_analysis)

# Percentage of significant subpathways within superpathways
subpath_within_superpath(sub_analysis)

met_within_sub(sub_analysis, subpathway = "Aminosugar Metabolism")

# All signifiicant subpathways
all_sig_subpath(sub_analysis)
```

---

`create_heatmap_Data`*Create metadata and matrices for metabolite heatmaps*

---

**Description**

This function creates the required matrices for the metabolite heatmaps.

**Usage**

```
create_heatmap_Data(data, heatmap_variables, Assay = "normalized", ...)
```

**Arguments**

<code>data</code>	A SummarizedExperiment containing Metabolon data.
<code>heatmap_variables</code>	A vector of variable names that are NOT metabolites.
<code>Assay</code>	Name of assay data to be used for heatmaps. Default="normalized".
<code>...</code>	Additional arguments that can be passed into the arrange function. This parameter will order the columns of the heatmap data.

**Value**

A list of matrices including the heatmap variable (meta data for heatmap) and the values for the heatmap.

---

`demoDat`*Demo data for the MetabolomicsPipeline,*

---

**Description**

Demo data consisting of 86 samples (42 males, 44 females), three treatment groups, and the samples were taken

**Usage**

```
demoDat
```

**Format**

SummarizedExperiment object

**Value**

A SummarizedExperiment object with 86 samples

---

demoDataSmall	<i>Subset of Demo data for the MetabolomicsPipeline ,</i>
---------------	---

---

**Description**

Demo data consisting of 86 samples (42 males, 44 females), three treatment groups, and the samples were taken at three different time points. We focus on a subset of 10 Subpathways.

**Usage**

```
demoDataSmall
```

**Format**

Rd

**Value**

A subset of the metabolites in the DemoData.

---

loadMetabolon	<i>Load Metabolomic Data as SummarizedExperiment</i>
---------------	--

---

**Description**

Automatically load metabolomic data from Metabolon

**Usage**

```
loadMetabolon(  
  path,  
  raw_sheet = "Peak Area Data",  
  chemical_sheet = "Chemical Annotation",  
  sample_meta = "Sample Meta Data",  
  normalized_peak = "Log Transformed Data",  
  sample_names = "PARENT_SAMPLE_NAME",  
  chemicalID = "CHEM_ID"  
)
```

**Arguments**

path	Path to Metabolon .xlsx file containg peak data, chemical annotations, sample meta data, and (optionally) the normalized peak counts
raw_sheet	Sheet name for the raw peak data.
chemical_sheet	Sheet name for chemical annotation.
sample_meta	Sheet name for sample meta data.

normalized_peak	Sheet name for the normalized peak data. If you are not adding the normalized data from the excel file then set normalized_peak=NA.
sample_names	Column name in the meta data containing the sample names. This must correspond to the row names of the raw peak data in the excel file.
chemicalID	Column name in the meta data containing the sample names. This must correspond to the column names of the raw peak data.

### Details

The Metabolon experiment data are stored in a SummarizedExperiment.

### Value

A SummarizedExperiment containing Metabolon experiment data.

### See Also

[SummarizedExperiment::SummarizedExperiment](#)

---

log\_transformation *Log transformation of metabolite data*

---

### Description

This function log transforms each metabolite in the Metabolon data.

### Usage

```
log_transformation(peak_data)
```

### Arguments

peak\_data      A matrix of peak data with metabolites in the columns

### Value

log transformed peak data

### Examples

```
data("demoDataSmall", package = "MetabolomicsPipeline")
peak <- SummarizedExperiment::assay(demoDataSmall, "peak")

# Median standardization
peak_med <- median_standardization(peak_data = peak)

# Min value imputation
peakImpute <- min_val_impute(peak_data = peak_med)

# log transformation
peak_log <- log_transformation(peak_data = peakImpute)
```

---

`median_standardization`*Median standardization for metabolite data*

---

### Description

This function standardizes the metabolites by the median of the metabolite.

### Usage

```
median_standardization(peak_data)
```

### Arguments

<code>peak_data</code>	Peak data with metabolites in the columns. The data also must include the "PARENT_SAMPLE_NAME".
------------------------	---

### Value

Median standardized peak data.

### Examples

```
data("demoDataSmall", package = "MetabolomicsPipeline")
peak <- SummarizedExperiment::assay(demoDataSmall, "peak")

# Median standardization
peak_med <- median_standardization(peak_data = peak)

# Min value imputation
peakImpute <- min_val_impute(peak_data = peak_med)

# log transformation
peak_log <- log_transformation(peak_data = peakImpute)
```

---

`metabolite_heatmap` *Create metabolite heatmap*

---

### Description

Create heatmaps which are arranged by the experimental conditions.

**Usage**

```
metabolite_heatmap(
  data,
  top_mets = 50,
  group_vars,
  strat_var = NULL,
  caption = NULL,
  Assay = "normalized",
  ...
)
```

**Arguments**

<code>data</code>	A SummarizedExperiment containing the Metabolon experiment data.
<code>top_mets</code>	Number of metabolites to include in the heatmap. Metabolites are chosen based on the highest variability.
<code>group_vars</code>	Vector of variables to annotate heatmap with. Columns will be grouped by these variables.
<code>strat_var</code>	Variable to stratify the heatmap by.
<code>caption</code>	A title for the heatmap. If <code>strat_var</code> is used, the title will automatically include the stratum with the tile.
<code>Assay</code>	Which assay data to use for the heatmap (default="normalized").
<code>...</code>	Additional arguments can be passed into the arrange function. This parameter will order the columns of the heatmap.

**Value**

A `gtable` class with all of the information to build the heatmap. To view the heatmap use `ggplotify::as.ggplot()`.

**Examples**

```
# load data
data("demoDat", package = "MetabolomicsPipeline")
dat <- demoDat

# Heatmap with one group
treat_heatmap <- metabolite_heatmap(dat,
  top_mets = 50,
  group_vars = "GROUP_NAME",
  strat_var = NULL,
  caption = "Heatmap Arranged By Group",
  Assay = "normalized",
  GROUP_NAME
)
```



---

`metabolite_pairwise`*Metabolite Pairwise Comparisons.*

---

## Description

Computes the pairwise comparison estimates and p-values for each metabolite.

## Usage

```
metabolite_pairwise(  
  data,  
  form,  
  Assay = "normalized",  
  strat_var = NULL,  
  mets = NULL  
)
```

## Arguments

<code>data</code>	SummarizedExperiment with Metabolon experiment data.
<code>form</code>	This is a character string the resembles the right hand side of a simple linear regression model in R. For example <code>form = "Group1 + Group2"</code> .
<code>Assay</code>	Name of the assay to be used for the pairwise analysis (default='normalized')
<code>strat_var</code>	A variable in the analysis data to stratify the model by. If this is specified, a list of results will be returned.
<code>mets</code>	Chemical ID for the metabolites of interest. If NULL then the pairwise analysis is completed for all metabololites.

## Details

This function will analyze each metabolite individually. For each metabolite, the `metabolite_pairwise` function will first test whether the model explained a significant proportion of the variance in the metabolite using an F-test. Since we will be looking at multiple comparisons for the metabolite, it is good practice to first look at the overall p-value from the F-test before looking at the pairwise comparisons. The `metabolite_pairwise` function then looks at all pairwise comparisons utilizing the `emmeans` package. The `metabolite_pairwise` function returns a data frame with the metabolite overall p-value, log fold change for each group, and the p-value for each comparison.

## Value

The overall F-test p-value, and the estimate and pvalue for each pairwise comparison.

## Examples

```
# Load data  
data("demoDat", package = "MetabolomicsPipeline")  
dat <- demoDat  
  
# Run pairwise analysis  
strat_pairwise <- metabolite_pairwise(dat,
```

```
    form = "GROUP_NAME*TIME1",  
    strat_var = "Gender"  
  )
```

---

metabolite_pca	<i>Metabolite PCA</i>
----------------	-----------------------

---

## Description

Computes and plots the first two components of the PCA from the metabolite data.

## Usage

```
metabolite_pca(data, Assay = "normalized", meta_var)
```

## Arguments

data	SummarizedExperiment with Metabolon experiment data.
Assay	Name of the assay to be used for the pairwise analysis (default='normalized')
meta_var	A metadata variable to color code the PCA plot by.

## Value

A PCA plot of the first two principal components, colored by the metadata variable.

## Examples

```
# load data  
data("demoDat", package = "MetabolomicsPipeline")  
dat <- demoDat  
  
# Define PCA label from metadata  
meta_var <- "Gender"  
  
# Run PCA  
pca <- metabolite_pca(dat,  
  meta_var = meta_var  
)  
  
# Show PCA  
pca
```

---

met_est_heatmap	<i>Metabolite Pairwise Estimate Interactive Heatmap.</i>
-----------------	--

---

## Description

Produce an interactive heatmap of the estimates produced in [metabolite\\_pairwise](#).

## Usage

```
met_est_heatmap(  
  results_data,  
  data,  
  interactive = FALSE,  
  CHEM_ID = "CHEM_ID",  
  SUB_PATHWAY = "SUB_PATHWAY",  
  CHEMICAL_NAME = "CHEMICAL_NAME",  
  ...  
)
```

## Arguments

<code>results_data</code>	Results data frame of the pairwise comparisons produced by <a href="#">metabolite_pairwise</a> .
<code>data</code>	A SummarizedExperiment containing the Metabolon experiment data.
<code>interactive</code>	boolean (T/F) for whether or not the plot should be interactive. Use <code>interactive=T</code> to produce an interactive plot using <code>plotly</code> . Use <code>interactive=F</code> to produce a static heatmap using <code>pheatmap</code> .
<code>CHEM_ID</code>	Column name in the chemical annotation worksheet that contains the chemical ID.
<code>SUB_PATHWAY</code>	Column name in the chemical annotation worksheet which contains the subpathway information.
<code>CHEMICAL_NAME</code>	Column name in the chemical annotation worksheet which contains the chemical name.
<code>...</code>	Additional arguments that can be passed to <code>pheatmap</code> .

## Details

This function will produce a heatmap of the log fold changes for the metabolites with a significant overall p-value (which tested if the treatment group means were equal under the null hypothesis). The heatmap colors will only show if the log fold-change is greater than  $\log(2)$  or less than  $\log(.5)$ . Therefore, this heatmap will only focus on comparisons with a fold change of two or greater.

## Value

An interactive heatmap of pairwise estimates.

---

met_p_heatmap	<i>Metabolite Pairwise P-Value Interactive Heatmap.</i>
---------------	---

---

## Description

Produce an interactive heatmap of the p-values produced in `metabolite_pairwise`.

## Usage

```
met_p_heatmap(  
  results_data,  
  data,  
  interactive = FALSE,  
  CHEM_ID = "CHEM_ID",  
  SUB_PATHWAY = "SUB_PATHWAY",  
  CHEMICAL_NAME = "CHEMICAL_NAME",  
  ...  
)
```

## Arguments

<code>results_data</code>	Results data frame of the pairwise comparisons produced by <code>metabolite_pairwise</code> .
<code>data</code>	A SummarizedExperiment containing Metabolon experiment data.
<code>interactive</code>	boolean (T/F) for whether or not the plot should be interactive. Use <code>interactive=T</code> to produce an interactive plot using <code>plotly</code> . Use <code>interactive=F</code> to produce a static heatmap using <code>pheatmap</code> .
<code>CHEM_ID</code>	Column name in the chemical annotation worksheet that contains the chemical ID.
<code>SUB_PATHWAY</code>	Column name in the chemical annotation worksheet which contains the subpathway information.
<code>CHEMICAL_NAME</code>	Column name in the chemical annotation worksheet which contains the chemical name.
<code>...</code>	Additional arguments that can be passed to <code>pheatmap</code> .

## Details

For the metabolites which had a significant overall p-value (which tested if the treatment group means were equal under the null hypothesis), we will produce a heatmap of the p-values.

## Value

An interactive heatmap of pairwise p-values.

---

met_within_sub	<i>Metabolites within Subpathway Table</i>
----------------	--

---

## Description

Return the model results for each metabolite within a subpathway.

## Usage

```
met_within_sub(
  subpath_results,
  subpathway,
  mod = c("interaction", "parallel", "single")
)
```

## Arguments

subpath_results	Results data frame generated by <a href="#">subpathway_analysis</a>
subpathway	Character string of the subpathway of interest. This is case sensitive and must be listed in the subpath_results.
mod	Model of interest. This can be a single model or a vector of model types that can take on the values "interaction", "parallel", or "single".

## Value

A table with the results from the model types specified and for each metabolite within the superpathway specified.

## Examples

```
data("demoDataSmall", package = "MetabolomicsPipeline")
dat <- demoDataSmall

# Runsubpathay analysis
sub_analysis <- subpathway_analysis(dat,
  treat_var = "GROUP_NAME",
  block_var = "TIME1",
  strat_var = NULL,
  Assay = "normalized"
)

#####
### Results Plots #####
#####

# significant subpathways by model type
subpath_by_model(sub_analysis)

# Percentage of significant subpathways within superpathways
subpath_within_superpath(sub_analysis)

met_within_sub(sub_analysis, subpathway = "Aminosugar Metabolism")
```

---

min_val_impute	<i>Minimum Value Imputation</i>
----------------	---------------------------------

---

**Description**

Imputes the minimum value for each metabolite

**Usage**

```
min_val_impute(peak_data)
```

**Arguments**

peak\_data      Peak data matrix with metabolites in the columns.

**Value**

Metabolite imputed peak data.

**Examples**

```
data("demoDataSmall", package = "MetabolomicsPipeline")
peak <- SummarizedExperiment::assay(demoDataSmall, "peak")

# Median standardization
peak_med <- median_standardization(peak_data = peak)

# Min value imputation
peakImpute <- min_val_impute(peak_data = peak_med)

# log transformation
peak_log <- log_transformation(peak_data = peakImpute)
```

---

pairwise	<i>Pairwise function</i>
----------	--------------------------

---

**Description**

This is the main function for metabolite\_pairwise

**Usage**

```
pairwise(out, form, data)
```

**Arguments**

out	Outcome used as reponse
form	form of the model
data	data used for modeling

**Value**

Pairwise comparisons for a single metabolite.

---

subpathway\_analysis

*Subpathway Analysis*


---

**Description**

Subpathway analysis for metabolite data.

**Usage**

```
subpathway_analysis(
  data,
  treat_var,
  block_var = NULL,
  strat_var = NULL,
  Assay = "normalized",
  subPathwayName = "SUB_PATHWAY",
  chemName = "CHEMICAL_NAME",
  superPathwayName = "SUPER_PATHWAY"
)
```

**Arguments**

data	SummarizedExperiment with Metabolon experiment data.
treat_var	This is the name of the variable in the analysis data that is the main variable of interest.
block_var	This is the name of the blocking variable in the dataset. If the the experimental design does not include a blocking variable, then the value of block_var=NULL.
strat_var	Variable to stratify the subpathway analysis by. This is set to NULL by default and will not stratify the analysis unless specified.
Assay	Name of the assay to be used for the pairwise analysis (default='normalized')
subPathwayName	Column name for subpathway variable as defined in the chemical annotation worksheet.
chemName	Column name for chemical name variable as defined in the chemical annotation worksheet.
superPathwayName	Column name for super-pathway variable as defined in the chemical annotation worksheet.

**Details**

For each metabolite, we test three models using using ANOVA.

1. Interaction:  $\log Peak = Treatment + block + Treatment * block$
2. Parallel:  $\log Peak = Treatment + block$

### 3. Single: $\log Peak = Treatment$

For the interaction model, we are focusing only on the interaction term "Treatment\*block" to test if there is a significant interaction between our treatment and the block variable. The parallel model tests if the block variable explains a significant amount of the metabolite variance, and the treatment model tests if the treatment explains a significant proportion of the variance for each metabolite. Then, we use the Combined Fisher probability to test each model at the subpathway level.

$$\tilde{X} = -2 \sum_{i=1}^k \ln(p_i)$$

where  $k$  is the number of metabolites in the subpathway. We can get a p-value from  $P(X \geq \tilde{X})$ , knowing that  $\tilde{X} \sim \chi_{2k}^2$ . You will notice that smaller p-values will lead to a larger  $\tilde{X}$ .

#### Value

A data frame with "CHEM\_ID", "sub\_pathway", "chem\_name", "interaction\_pval", "interaction\_fisher", "parallel\_pval", "p", "single\_pval", "single\_fisher", and "model" for each metabolite.

#### See Also

Loughin, Thomas M. "A systematic comparison of methods for combining p-values from independent tests." *Computational statistics & data analysis* 47.3 (2004): 467-485.

#### Examples

```
# Load data
data("demoDataSmall", package = "MetabolomicsPipeline")
dat <- demoDataSmall

# Runsubpathay analysis
sub_analysis <- subpathway_analysis(dat,
  treat_var = "GROUP_NAME",
  block_var = "TIME1",
  strat_var = NULL,
  Assay = "normalized"
)

#####
### Results Plots #####
#####

# significant subpathways by model type
subpath_by_model(sub_analysis)

# Percentage of significant subpathways within superpathways
subpath_within_superpath(sub_analysis)

met_within_sub(sub_analysis, subpathway = "Aminosugar Metabolism")

# All signifiicant subpathways
all_sig_subpath(sub_analysis)
```



---

`subpathway_boxplots`*Subpathway Boxplots*

---

## Description

Creates boxplots for each metabolite within a specified subpathway.

## Usage

```
subpathway_boxplots(  
  data,  
  subpathway,  
  block_var,  
  treat_var,  
  Assay = "normalized",  
  CHEMICAL_NAME = "CHEMICAL_NAME",  
  CHEM_ID = "CHEM_ID",  
  SUB_PATHWAY = "SUB_PATHWAY",  
  ...  
)
```

## Arguments

<code>data</code>	SummarizedExperiment with Metabolon experiment data.
<code>subpathway</code>	Character value of the subpathway of interest. This is case sensitive and must be in the chemical annotation file.
<code>block_var</code>	This the the name of the variable in the meta data that is used for the X axis of the box plots. We recommend using the "block_var" from the subpathway analysis.
<code>treat_var</code>	This is a grouping variable. As a recommendation the treatment groups should be used in the treat_var argument as this will provide a different color for each of the treatments making it easier to identify.
<code>Assay</code>	Name of the assay to be used for the pairwise analysis (default='normalized')
<code>CHEMICAL_NAME</code>	Column name in the chemical annotation worksheet which contains the chemical name.
<code>CHEM_ID</code>	Column name in the chemical annotation worksheet that contains the chemical ID.
<code>SUB_PATHWAY</code>	Column name in chemical annotation file which contains the SUB_PATHWAY information
<code>...</code>	Additional arguments to filter the analysis data by.

## Details

.

## Value

Boxplots stratified by metabolites.

**Examples**

```
# load data
data("demoDat", package = "MetabolomicsPipeline")
dat <- demoDat

#####
### BoxPlots #####
#####

subpathway_boxplots(dat,
  subpathway = "Lactoyl Amino Acid", block_var = TIME1,
  treat_var = GROUP_NAME, Assay = "normalized",
  CHEMICAL_NAME = "CHEMICAL_NAME",
  CHEM_ID="CHEM_ID",
  SUB_PATHWAY="SUB_PATHWAY", Gender == "Female")

#####
## Line plots #####
#####

# Set up data
dat$TIME1 <- as.numeric(factor(dat$TIME1,
  levels = c("PreSymp", "Onset", "End")
))
# Create line plots
subpathway_lineplots(dat,
  subpathway = "Lactoyl Amino Acid",
  block_var = TIME1, treat_var = GROUP_NAME,
  Assay = "normalized",
  CHEMICAL_NAME = "CHEMICAL_NAME",
  CHEM_ID="CHEM_ID",
  SUB_PATHWAY="SUB_PATHWAY", Gender == "Female")
```

---

subpathway\_lineplots

*Subpathway Lineplots*


---

**Description**

Create line plots for each metabolite within a subpathway.

**Usage**

```
subpathway_lineplots(
  data,
  subpathway,
  block_var,
  treat_var,
  Assay = "normalized",
```

```

    CHEMICAL_NAME = "CHEMICAL_NAME",
    CHEM_ID = "CHEM_ID",
    SUB_PATHWAY = "SUB_PATHWAY",
    ...
)

```

## Arguments

data	SummarizedExperiment with Metabolon experiment data.
subpathway	Character value of the subpathway of interest. This is case sensitive and must be in the chemical annotation file.
block_var	This the the name of the variable in the meta data that is used for the X axis of the line plots. We recommend using the "block_var" variable from the subpathway analysis.
treat_var	This is a grouping variable. As a recommendation the treatment groups should be used in the groupBy argument as this will provide a different color for each of the treatments making it easier to identify.
Assay	Name of the assay to be used for the pairwise analysis (default='normalized')
CHEMICAL_NAME	Column name in the chemical annotation worksheet which contains the chemical name.
CHEM_ID	Column name in the chemical annotation worksheet that contains the chemical ID.
SUB_PATHWAY	Column name in the chemical annotation worksheet which contains the subpathway information.
...	Additional arguments to filter the analysis data by.

## Value

Line plots stratified by metabolite.

## Examples

```

data("demoDat", package = "MetabolomicsPipeline")
dat <- demoDat

#####
### BoxPlots #####
#####

subpathway_boxplots(dat,
  subpathway = "Lactoyl Amino Acid", block_var = TIME1,
  treat_var = GROUP_NAME, Assay = "normalized",
  CHEMICAL_NAME = "CHEMICAL_NAME",
  CHEM_ID="CHEM_ID",
  SUB_PATHWAY="SUB_PATHWAY",Gender == "Female")

#####
## Line plots #####
#####

# Set up data

```

```

dat$TIME1 <- as.numeric(factor(dat$TIME1,
  levels = c("PreSymp", "Onset", "End")
))

# Create line plots
subpathway_lineplots(dat,
  subpathway = "Lactoyl Amino Acid",
  block_var = TIME1, treat_var = GROUP_NAME,
  Assay = "normalized",
  CHEMICAL_NAME = "CHEMICAL_NAME",
  CHEM_ID="CHEM_ID",
  SUB_PATHWAY="SUB_PATHWAY", Gender == "Female")

```

---

subpath_by_model	<i>Subpathway model type table</i>
------------------	------------------------------------

---

### Description

Create a table with the number of significant subpathways for each model type.

### Usage

```
subpath_by_model(subpath_results)
```

### Arguments

subpath\_results  
Results data frame generated by [subpathway\\_analysis](#)

### Details

Each subpathway will only have one model type. We first test the interaction, and then the parallel and single models are tested last. Suppose a subpathway has a significant interaction model type. In that case, the table will count it as an interaction and not as a parallel or single.

### Value

A table of the number of significant subpathways by model type.

### Examples

```

# Load data
data("demoDataSmall", package = "MetabolomicsPipeline")
dat <- demoDataSmall

# Runsubpathay analysis
sub_analysis <- subpathway_analysis(dat,
  treat_var = "GROUP_NAME",
  block_var = "TIME1",
  strat_var = NULL,

```

```

    Assay = "normalized"
  )

#####
### Results Plots #####
#####

# significant subpathways by model type
subpath_by_model(sub_analysis)

# Percentage of significant subpathways within superpathways
subpath_within_superpath(sub_analysis)

met_within_sub(sub_analysis, subpathway = "Aminosugar Metabolism")

```

---

subpath\_within\_superpath  
*Proportion of the Significant Subpathways Within Superpathways*

---

## Description

Create a table that gives the percentage of significant subpathways within each superpathway.

## Usage

```
subpath_within_superpath(subpath_results)
```

## Arguments

```
subpath_results
```

Results data frame generated by [subpathway\\_analysis](#)

## Value

A table with the proportion (and percent) of significant subpathways within superpathways.

## Examples

```

# Load data
data("demoDataSmall", package = "MetabolomicsPipeline")
dat <- demoDataSmall

# Runsubpathay analysis
sub_analysis <- subpathway_analysis(dat,
  treat_var = "GROUP_NAME",
  block_var = "TIME1",
  strat_var = NULL,
  Assay = "normalized"
)

#####
### Results Plots #####
#####

```

```
# significant subpathways by model type
subpath_by_model(sub_analysis)

# Percentage of significant subpathways within superpathways
subpath_within_superpath(sub_analysis)

met_within_sub(sub_analysis, subpathway = "Aminosugar Metabolism")
```