Analysing short tandem repeats in genomic sequences

What, why, how?

Short tandem repeats (a.k.a. microsatellites)

Repetitions of 1-6 nt DNA motifs

Short tandem repeat (STR)

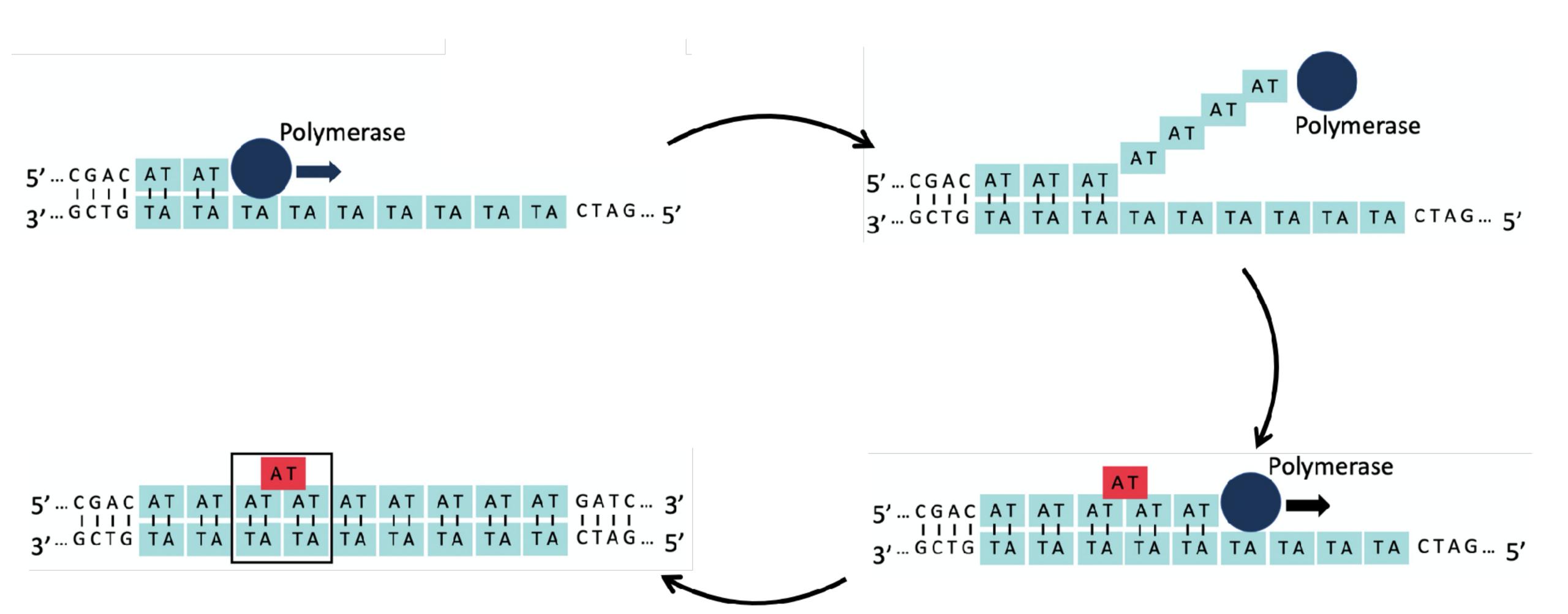
Motif: the repeating unit ('AT' in this case)

Motif length: length of the repeating unit

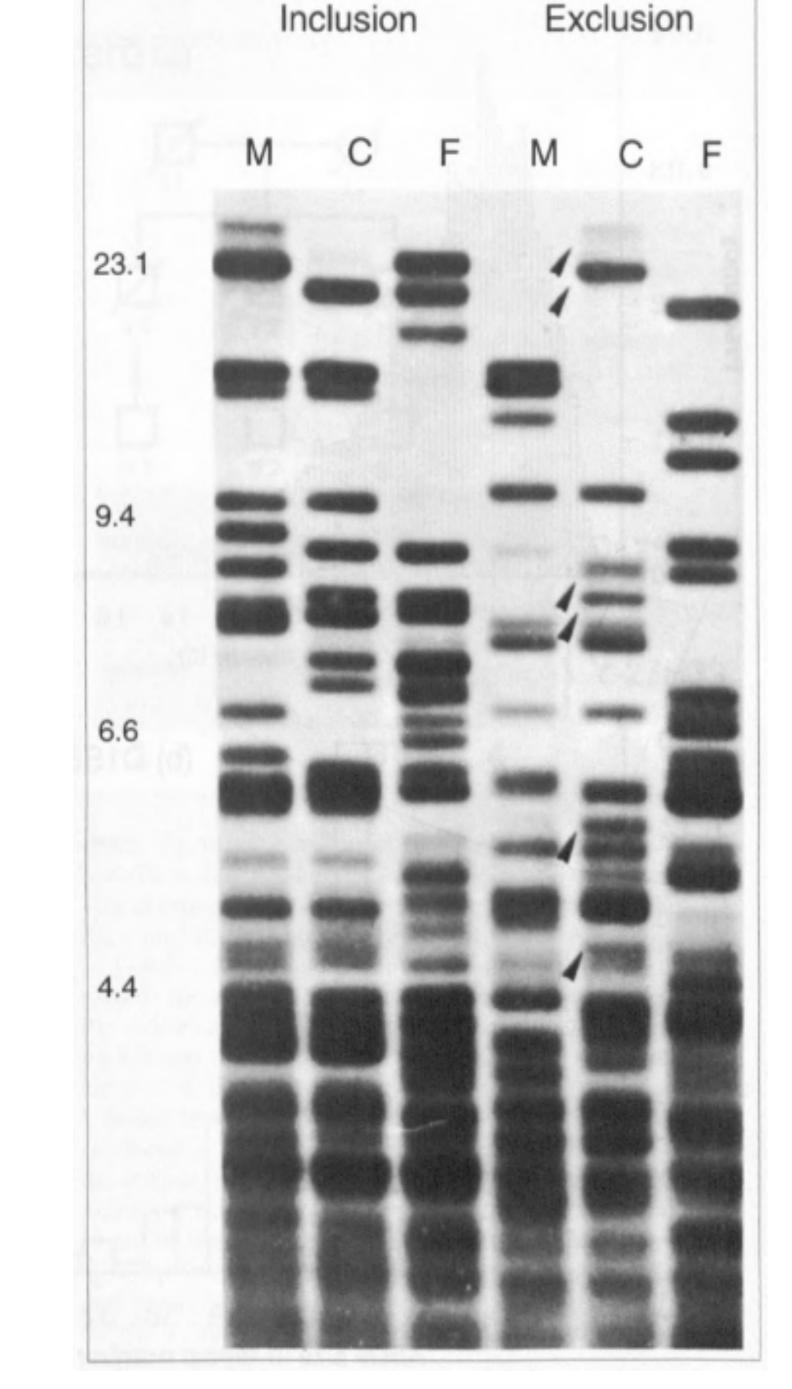
Copy number: number of times the unit is repeated

STRs have mutation rates up to 10'000 times higher than point mutations

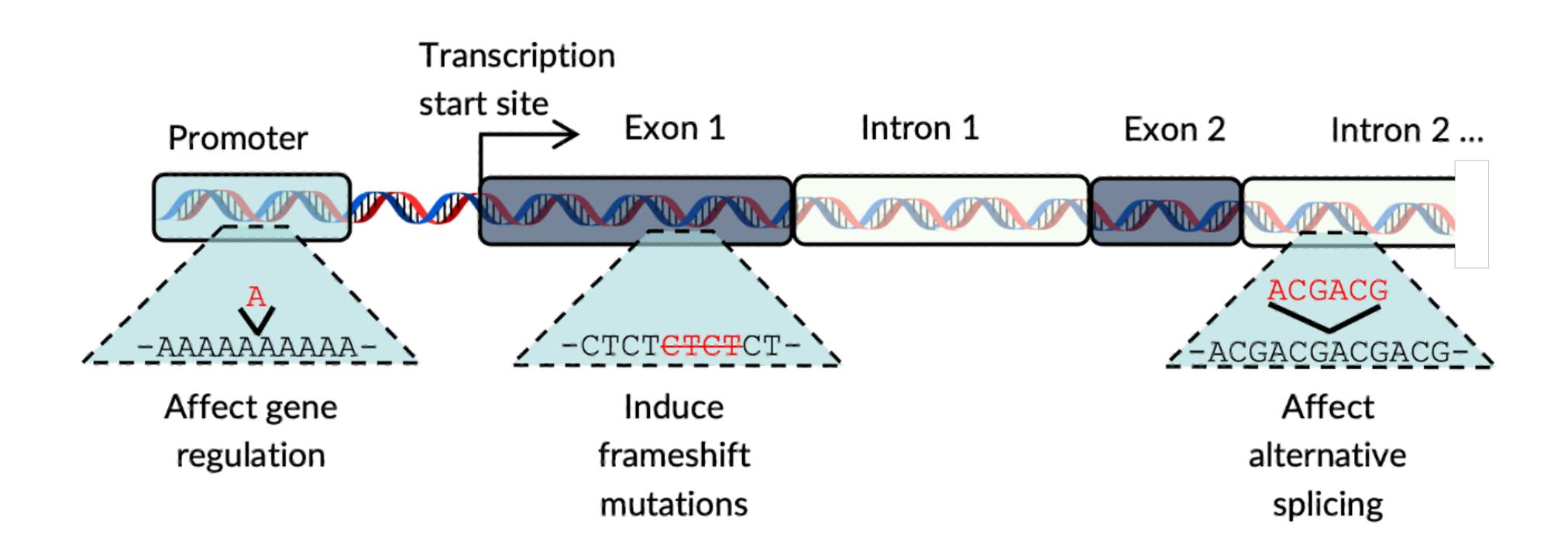
Slipped-strand mispairing

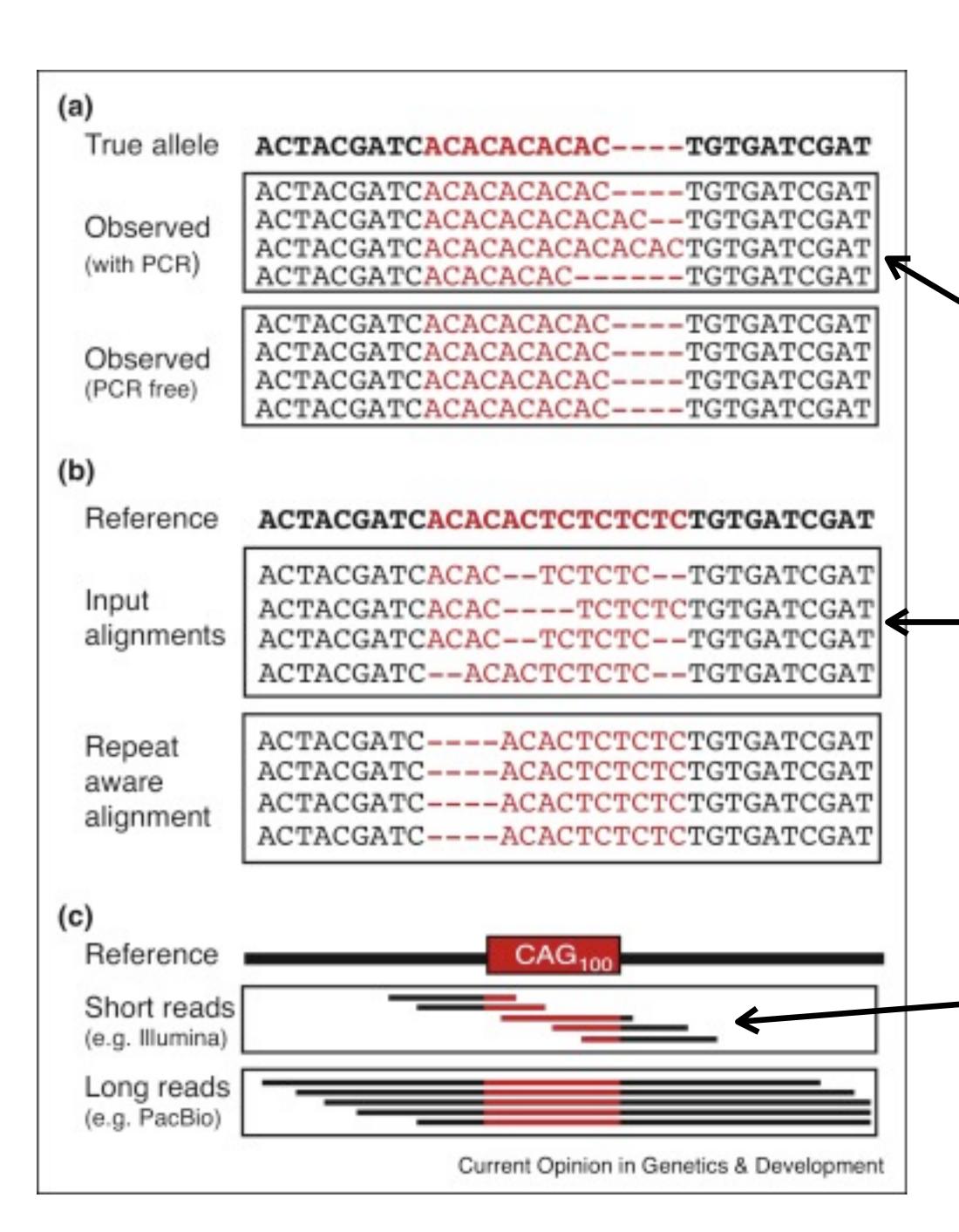


Classic use case: paternity testing



Functional consequences of STR mutations





Why STRs are challenging to genotype

PCR induces mistakes during sequencing (addressed by PCR-free methods)

Not all alignment algorithms are 'repeat aware'

Short reads do not always span longer STRs, which makes determining their length very hard

Gymrek, Melissa. 'A Genomic View of Short Tandem Repeats'. *Current Opinion in Genetics and Development* 44 (1 June 2017): 9–16. https://doi.org/10.1016/j.gde.2017.01.012.

We need special tools to genotype STRs!

e.g. Table 1 in https://doi.org/10.1016/j.gde.2017.01.012

• We will use GangSTR today: https://github.com/gymreklab/GangSTR



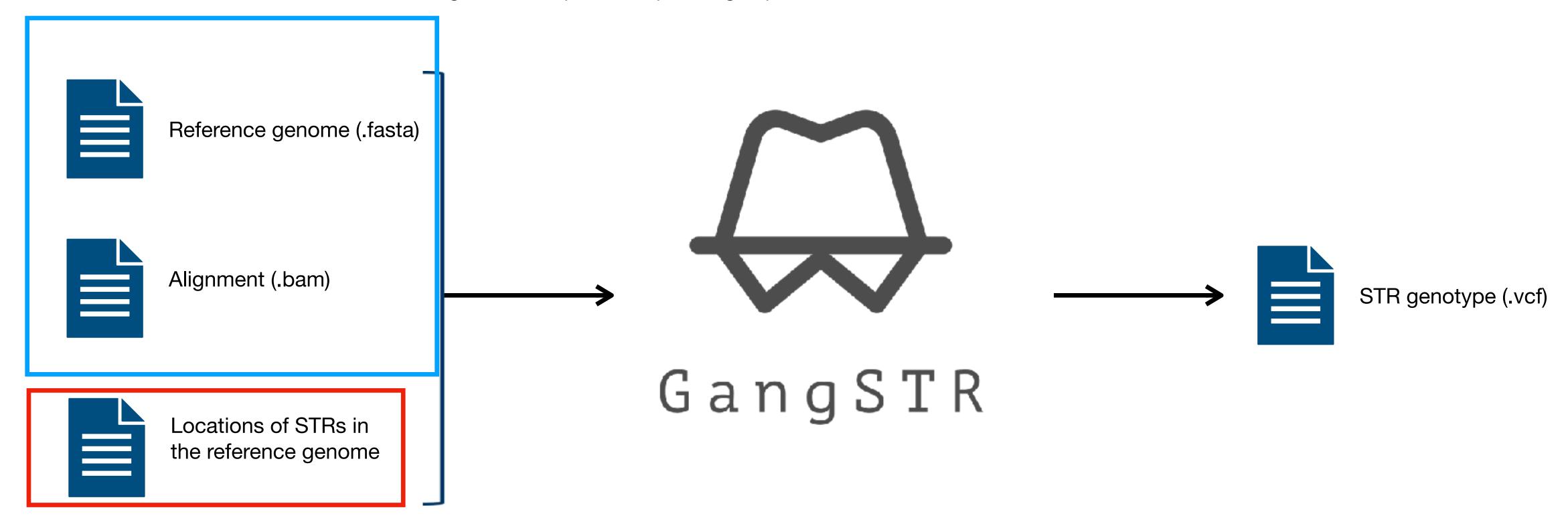
GangSTR

GangSTR intput/output

These files we have!

Reference: available online

Alignment: output of sequencing experiment



This file we need to generate ourselves!

Detecting (short) tandem repeats in genomic sequence

- Computationally expensive for large sequences (especially imperfect repeats)
- Many different algorithms exists, each with different strengths/weaknesses

...GCTACGTACCTACCTACCTACCTAA...

TR unit alignment

ACGTACCT

AC-TACCT

ACCTACCT

Tandem repeat annotation library (TRAL)

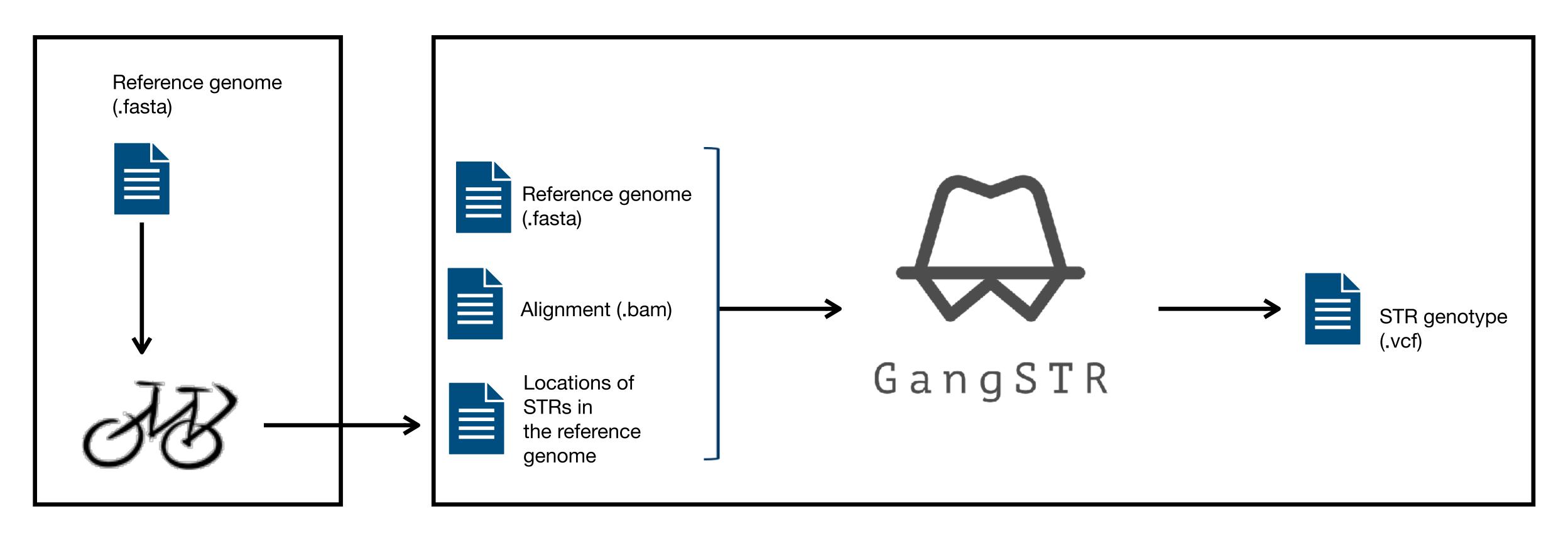
- A python library aimed at addressing challenges in repeat detection:
 - Allows for the running of several repeat detection algorithms on the input sequence
 - Calculates scores for each detected repeat for filtering
 - Can make the set of detected repeats non-redundant



Summary

- Short tandem repeats can mutate rapidly, which can have functional consequences
- Repeat genotyping: determining how long repeats are in a given sequencing sample (e.g. using GangSTR)
- Repeat detection: determining where repeats are located and how long they are in the reference sequence (e.g. using TRAL)

Workflow for today



Morning

Afternoon

Task 1

STR background reading

Gymrek, Melissa. 'A Genomic View of Short Tandem Repeats'. *Current Opinion in Genetics and Development* 44 (1 June 2017): 9–16. https://doi.org/10.1016/j.gde.2017.01.012.

- Read the following sections of 'A genomic view of short tandem repeats':
 - Abstract + Introduction
 - Genotyping STRs from high-throughput sequencing data

- Afterwards, you should be able to answer the following questions:
 - Why is STR variation relevant to health and disease?
 - What are some of the challenges in analysing STRs from NGS data?

Task 2 Introduction to TRAL

Schaper, Elke, Alexander Korsunsky, Jūlija Pečerska, Antonio Messina, Riccardo Murri, Heinz Stockinger, Stefan Zoller, Ioannis Xenarios, and Maria Anisimova. 'TRAL: Tandem Repeat Annotation Library'. *Bioinformatics* 31, no. 18 (15 September 2015): 3051–53. https://doi.org/10.1093/bioinformatics/btv306.

• Read 'TRAL: tandem repeat annotation library'

- Afterwards, you should be able to answer the following questions:
 - Why should you use multiple tandem repeat detection algorithms to look for repeats in biological sequence?
 - What different functionalities does TRAL provide?

Task 3

STR detection in the APC gene

- Set up the 'BIO392_STRs' conda environment and activate it (see appendix 1)
- Start a jupyter server and open 'scripts/BIO392_TRAL.ipynb' (see appendix 2)
- Follow along with the steps in BIO392_TRAL.ipynb

Appendix 1

Setting up the BIO392_STRs conda env

1: Make sure you are in the right directory

2: type 'ls + ENTER' you should see this file

```
r∈sults
                       environment.yaml
                                                                    scripts
lmaxverbiest@clt-mob-n-2725/2022-09-30 % condd env create _f environment.yaml
Collecting package metadata (repodata.jsor): corc
Solving environment: done
==> WARNING: A newer version of condu exists. <==
  current version: 4.10.3
                                                                     3: type 'conda env create -f environment.yaml + ENTER'
 latest version: 22.9.0
                                                                           will create our environment (might take a while)
Please update conda by running
   $ conda update -n base -c defaults corce
Downloading and Extracting Packages
ppenjpeg-2.5.0
                     528 KB
ca-centificates-2022
                     150 KB
                      1.3 MB
libiconv-1.17
```

. . .

```
Installing collected packages: contigobj, plopython, tral successfully installed tropython—1.79 contigobj—5.0.6 tral=2.0

done

# # To activate this environment, use

# # conda activate E10392_STRs

# To deactivate an active environment, use

# # conda deactivate

# conda deactivate

# conda deactivate

# maxverblest@clt-mob-n=2720 2022-09=30 % conda activate B10392 STRs

(310392 STRs) maxverblest@clt-mob-n=2723 2022-09=30 %
```

Appendix 2

Starting a jupyter server

1: Make sure you are in the right directory

2: type 'jupyter notebook + ENTER'

```
(BIO392_STRs) maxverbiest@clt-mob-n-2720 2022-09-30 % jupyter notebook
[I 13:55:35.924 NotebookApp] Serving notebooks from local directory: /Users/maxverbiest/PhD/projects/BIO392/2022-09-30
[I 13:55:35.924 NotebookApp] Jupyter Notebook 6.3.0 is running at:
[I 13:55:35.924 NotebookApp] http://localhost:8888/
[I 13:55:35.924 NotebookApp] http://localhost:8888/
[I 13:55:35.924 NotebookApp] Use Control-to stop this server and shut down all kernels (twice to skip confirmation).
/Users/maxverbiest/miniconda3/envs/BIO392_STRs/lib/python3.6/json/encoder.py:257: UserWarning: date_default is deprecated since jupyter_client 7.0.0. Use j
upyter_client.jsonutil.json_default.
return _iterencode(o, 0)
```

3: The jupyter interface should open up in your browser If not, you can copy this URL and go there yourself