# Introduction to ceRNAmiRNAfun

ceRNAmiRNAfun Developers

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This document guides the user through all available functions of the ceRNAmiRNAfun package. ceRNAmiRNAfun aims to find out miRNA and ceRNA triplets based on both miRNA and gene expression data. Recent studies have shown that among the target genes of miRNAs, several algorithms have been developed to identify ceRNAs and their dynamic regulating systems. Most of the algorithms divide a miRNA into different groups based on its expression level and them perform the analysis accordingly. However, the expression level of a miRNA is actually a continuous variable instead of a discrete variable. To address this issue, we developed a new algorithm based on the circular binary algorithm. We got the correlation with each window, and the applied the circular binary algorithm to get the peaks from the miRNA expression level across samples.

#### Installation

ceRNAmiRNAfun is on Bioconductor and can be installed following standard installation procedure.

```
install.packages("BiocManager",repos = "http://cran.us.r-project.org")
BiocManager::install("ceRNAmiRNAfun")
system.file("exdata", "mirtest3.csv", package="ceRNAmiRNAfun")
```

To use,

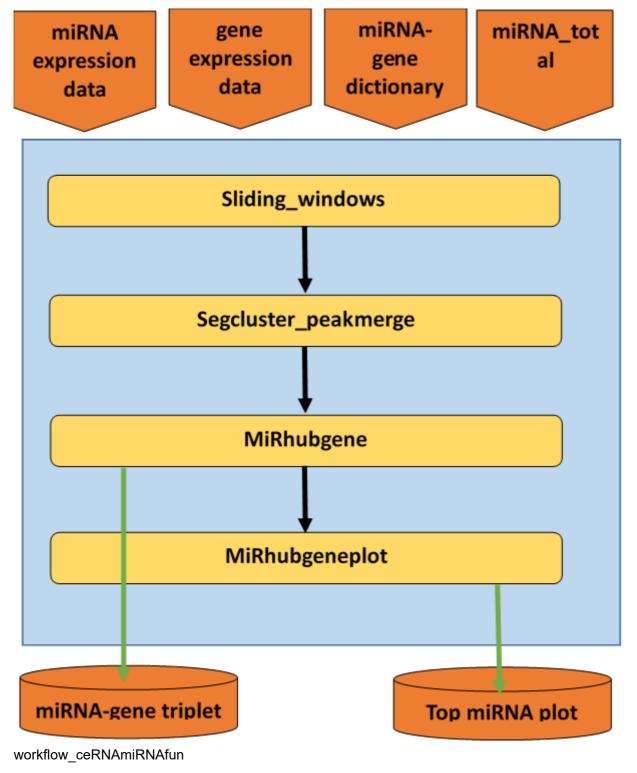
library(ceRNAmiRNAfun)

#### General Workflow

#### workflow steps:

Basically, there are four steps, corresponding to four R functions, to complete the whole analysis:

- 1.Sliding\_windows to get the correlation coefficients in each window.
- 2.Segcluster\_peakmerge to merge the continuous samples.
- 3.MiRhubgene to sort the data and then give the output table of miRNA and ceRNA triplets.
- 4.MiRhubgeneplot to plot the top miRNA.



#### **Data Source**

As shown in the workflow, not only samples of miRNA and gene expression data, but also the miRNA and corresponding genes library based on the same data resource for the analysis. The format of ceRNAmiRNAfun input data in expression is matrices, in dictionary is list. Data sources are platform- and technologyindependent. Therefore, expression data are all acceptable for ceRNAmiRNAfun. However, the raw miRNA and gene data have to be formatted into expression matrices and the related library should be necessary before using ceRNAmiRNAfun package.

### miRNA expression

Columns for samples. Rows for miRNAs

```
miRNA SampleA SampleB ...
                  0.5
          0.1
В
         -0.5
                   2.1
C
          0.4
                   0.3
```

# gene expression

Columns for samples. Rows for genes

```
GENE SmapleA
                SampleB ...
Α
         0.1
                   0.2
        -0.5
                   -0.3
C
         0.4
                   0.1
```

#### dictionary data

The first column of miRNA names, the second column of list form of candidate genes.

pairinformation	miRNA	genelist
Row1	Α	geneA1,geneA2
Row2	В	geneB1,geneB2
Row3	С	geneC1,geneC2

#### miRNA total data

The column of miRNA names, extracted from the first column of dictionary data.

```
miRNA
Row1
          Α
          В
Row2
           C
Row3
```

# **Usage Example**

Now, we show an example using internal data for ceRNAmiRNAfun workflow.

#### **Example Data Source**

To demonstrate the usage of the ceRNAmiRNAfun package, the package contains 475 paired miRNA and gene lung cancer samples. As for dictionary data, there are 137 paired miRNA and corresponding gene data in a list form, and miRNA total data has the first column of the dictionary data of the miRNA's names. Both of the miRNA and gene data comes from the TCGA lung adenocarcinoma dataset.

### Format of Input Data

First of all, load the internal data and check the format.

```
data(mirna_sam)
data(gene_sam)
data(dictionary)
data(miRNA_total)
```

Basically, the format of input data should be the same as the internal data. For miRNA expression data should be like.

```
mirna_sam[1:3, 1:3]
```

As for gene expression data,

```
gene_sam[1:3, 1:3]
```

As for the dictionary data, a list form of miRNA and corresponding genes.

```
dictionary[1:3,]
```

And miRNA total

```
miRNA_total[1:3]
```

### Sliding windows

Set the size of the window, and then take the input such as dictionary, mirna sam, gene sam, miRNA total, and the window size w into the function

Realdata=Datalist\_sliding\_w(w=10,dictionary=dictionary,miRNA\_total=miRNA\_total,mirna\_sam=mirn a\_sam,gene\_sam=gene\_sam)

```
## Error in makeCluster(no_cores, outfile = ""): 沒有這個函數 "makeCluster"
```

The function will calculate correlation coefficients within each window, a sliding mover that contains putative ceRNA triplets composed of a miRNA and several genes. And the corresponding miRNA expression of each window was the average expression of samples within different windows. The output will be Realdata, a list of data frames with correlation coefficients of genes within each window for each corresponding miRNA average expression.

# Segcluster peakmerge

After getting correlation coefficients with sliding windows, we try to merge some samples within a cluster to decrease the noise of the all correlations. We have to put cor shreshold, dictionary, mirna sam, gene sam, miRNA\_total and Realdata from sliding\_windows into the function.

Result TCGA LUSC =segcluster peakmerge(cor shreshold peak=0.85,dictionary=dictioanry,miRNA to tal=miRNA\_total,mirna\_sam=mirna\_sam,gene\_sam=gene\_sam,Realdata=Realdata)

```
## Error in makeCluster(no_cores, outfile = ""): 沒有這個函數 "makeCluster"
```

The function will cluster noisy correlation into neighboring regions of distinct correlation levels. And then do the peak merging by considering low probability of multi-peaks occurring. Also, the segments with few samples detected by circular binary segmentation which are less than three and make no sense to the interactions would merge to a single peak. The output will be Result\_TCGA\_LUSC, a list format with miRNAs, candidate ceRNA-ceRNA pirs, peak locations, and the number of samples occuring ceRNA-ceRNA interactions.

### MiRhubgene

After the peak merge, we will sort the information of miRNA and ceRNA to find out triplets. We should put dictionary and Result TCGA LUSC from segcluster peakmerge into the function.

```
miRhubgeneoutput=miRhubgene(dictionary=dictionary,Result TCGA LUSC= Result TCGA LUSC)
```

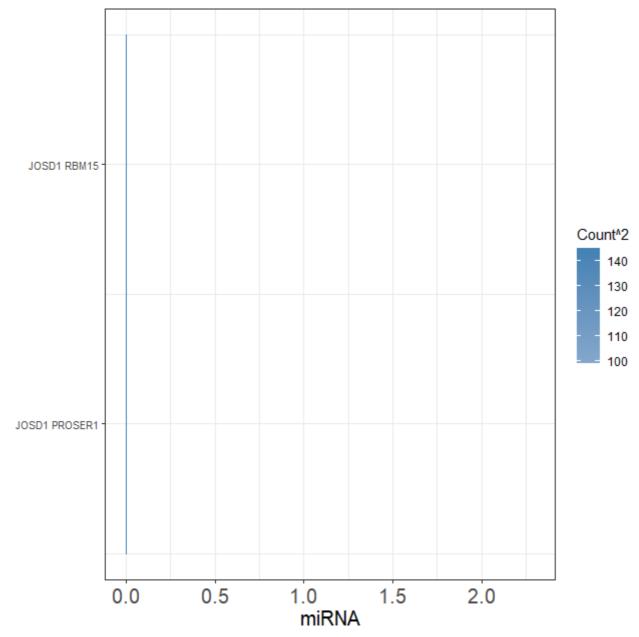
The output will be miRhubgeneoutput, a dataframe formats with miRNA names with the number of bridging ceRNA triplets and the corresponding genes with the number of ceRNA triplets.

# MiRhubgeneplot

Last, we do the plot of the top miRNA to see its expression situation. The input should be dictionary, mirna sam, gene sam, Result TCGA LUSC from segcluster peakmerge, miRhubgeneoutput from miRhubgene, and also the window size w and the number of samples N into the function.

plotp=miRhubgeneplot(dictionary=dictionary, Result TCGA LUSC=Result TCGA LUSC, mirna sam=mirna sam,miRhubgeneoutput=miRhubgeneoutput,w=10,N=475)

The function will plot the top miRNA, for visualizing the result of the triplets. The output will be plotp a plot format with miRNA expression in x-axis and the interaction ceRNA in y-axis.



plotp