ENCODE DREAM Challenge

 $John\ Reid$

Joining, by = c("TF", "cell")

##		TT	0011	anli+	+mus blind	anli+ blind
##	1	TF ARID3A	cell HepG2		true.blind FALSE	split.blind train
	2	ARIDSA		ladder	FALSE	ladder
	3	ATF2	GM12878	train	FALSE	train
	4	ATF2	H1-hESC	train	FALSE	train
	5	ATF2	MCF-7	train	FALSE	train
##	6	ATF2	K562	ladder		ladder.blind
##	7	ATF2	HepG2	submit	TRUE	submit
##	8	ATF3	HCT116	train	FALSE	train
##	9	ATF3	H1-hESC	train	FALSE	train
##	10	ATF3	HepG2	train	FALSE	train
##	11	ATF3	K562	train	FALSE	train
##	12	ATF3		ladder	FALSE	ladder
##	13	ATF7	GM12878	train	FALSE	train
##	14	ATF7	HepG2		FALSE	train
##	15	ATF7	K562	train	FALSE	train
##	16	ATF7		ladder		ladder.blind
##	17	CEBPB	A549	train	FALSE	train
##	18	CEBPB	H1-hESC HCT116	train	FALSE	train
## ##	19 20	CEBPB CEBPB	HeLa-S3		FALSE FALSE	train train
	21	CEBPB	HepG2		FALSE	train
	22	CEBPB	IMR-90	train	FALSE	train
	23	CEBPB	K562	train	FALSE	train
	24	CEBPB		ladder	FALSE	ladder
	25	CREB1	GM12878	train	FALSE	train
##	26	CREB1	H1-hESC	train	FALSE	train
##	27	CREB1	HepG2	train	FALSE	train
##	28	CREB1	K562	train	FALSE	train
##	29	CREB1	MCF-7	ladder	TRUE	ladder.blind
##	30	CTCF	A549	train	FALSE	train
##	31	CTCF	H1-hESC	train	FALSE	train
##	32	CTCF	HeLa-S3	train	FALSE	train
	33	CTCF	HepG2		FALSE	train
##	34	CTCF	IMR-90	train	FALSE	train
##	35	CTCF	K562	train	FALSE	train
	36	CTCF	MCF-7	train	FALSE	train
##		CTCF	GM12878		FALSE	ladder
## ##		CTCF		submit	TRUE TRUE	submit submit
##		E2F1	<pre>induced_pluripotent_stem_cell</pre>	train	FALSE	train
##		E2F1	HeLa-S3	train	FALSE	train
##		E2F1		submit	TRUE	submit
##		E2F6	A549	train	FALSE	train
##		E2F6	H1-hESC		FALSE	train
##		E2F6	HeLa-S3		FALSE	train
##		E2F6		ladder	FALSE	ladder

шш	47	ECD4	CM10070	*	FALSE	*
## ##		EGR1	GM12878			train
		EGR1	H1-hESC		FALSE	train
##		EGR1	HCT116		FALSE	train
##		EGR1	MCF-7		FALSE	train
##		EGR1		ladder	FALSE	ladder
##		EGR1		submit	TRUE	submit
##	53	EP300	GM12878	train	FALSE	train
##	54	EP300	H1-hESC	train	FALSE	train
##	55	EP300	HeLa-S3	train	FALSE	train
##	56	EP300	HepG2	train	FALSE	train
##	57	EP300	K562	train	FALSE	train
##	58	EP300	SK-N-SH	train	FALSE	train
##	59	EP300	MCF-7	ladder	FALSE	ladder
##	60	FOXA1	HepG2	train	FALSE	train
##	61	FOXA1	-	ladder		ladder.blind
##	62	FOXA1	liver	submit	TRUE	submit
##	63	FOXA2	HepG2		FALSE	train
##	64	FOXA2	-	submit	TRUE	submit
##		GABPA	GM12878	train	FALSE	train
	66	GABPA	H1-hESC		FALSE	train
##		GABPA	HeLa-S3		FALSE	train
	68	GABPA	HepG2		FALSE	train
##		GABPA	MCF-7		FALSE	train
	70	GABPA	SK-N-SH	train	FALSE	train
	71	GABPA		ladder	FALSE	ladder
	72	GABPA		submit	TRUE	submit
	73	GATAS	A549	train	FALSE	train
	74	GATAS	SK-N-SH		FALSE	train
	75	GATAS		ladder	FALSE	ladder
	76	HNF4A	HepG2	train	FALSE	train
	77	HNF4A	-	submit	TRUE	submit
##	78	JUND	HCT116	train	FALSE	
	79	JUND	HeLa-S3			train train
	80				FALSE	train
	81	JUND	HepG2 K562		FALSE	
##	82	JUND	MCF-7		FALSE FALSE	train
##		JUND				train
		JUND	SK-N-SH	train	FALSE	train
##		JUND	H1-hESC		FALSE	ladder
## ##		JUND		submit	TRUE	submit
		MAFK	GM12878	train	FALSE	train
##		MAFK	H1-hESC		FALSE	train
##		MAFK	HeLa-S3		FALSE	train
##		MAFK	HepG2		FALSE	train
##		MAFK	IMR-90		FALSE	train
##		MAFK		ladder	FALSE	ladder
##		MAFK		ladder	FALSE	ladder
##		MAX	A549	train	FALSE	train
##		MAX	GM12878	train	FALSE	train
##		MAX	H1-hESC	train	FALSE	train
##		MAX	HCT116	train	FALSE	train
##		MAX	HeLa-S3	train	FALSE	train
##		MAX	HepG2		FALSE	train
##		MAX	K562		FALSE	train
##	100	MAX	SK-N-SH	train	FALSE	train

##	101	MAX	MCF-7	ladder	FALSE	ladder
	102	MAX		submit	TRUE	submit
	103	MYC	A549	train	FALSE	train
	104	MYC	HeLa-S3	train	FALSE	train
##	105	MYC	K562	train	FALSE	train
##	106	MYC	MCF-7	train	FALSE	train
##	107	MYC	HepG2	ladder	FALSE	ladder
##	108	NANOG	H1-hESC	train	FALSE	train
##	109	NANOG	<pre>induced_pluripotent_stem_cell</pre>	submit	TRUE	submit
##	110	REST	H1-hESC	train	FALSE	train
##	111	REST	HeLa-S3	train	FALSE	train
##	112	REST	HepG2	train	FALSE	train
##	113	REST	MCF-7	train	FALSE	train
##	114	REST	Panc1	train	FALSE	train
##	115	REST	SK-N-SH	train	FALSE	train
##	116	REST	K562	${\tt ladder}$	FALSE	ladder
##	117	REST	liver	${\tt submit}$	TRUE	submit
##	118	RFX5	GM12878	train	FALSE	train
##	119	RFX5	HeLa-S3	train	FALSE	train
##	120	RFX5	MCF-7	train	FALSE	train
##	121	RFX5	SK-N-SH	train	FALSE	train
##	122	RFX5	HepG2	ladder	FALSE	ladder
##	123	SPI1	GM12878	train	FALSE	train
##	124	SPI1	K562	ladder	FALSE	ladder
##	125	SRF	GM12878	train	FALSE	train
##	126	SRF	H1-hESC	train	FALSE	train
##	127	SRF	HCT116	train	FALSE	train
##	128	SRF	HepG2	train	FALSE	train
##	129	SRF	K562	train	FALSE	train
##	130	SRF	MCF-7	ladder	FALSE	ladder
##	131	STAT3	HeLa-S3	train	FALSE	train
##	132	STAT3	GM12878	ladder	FALSE	ladder
##	133	TAF1	GM12878	train	FALSE	train
	134	TAF1	H1-hESC	train	FALSE	train
	135	TAF1	HeLa-S3		FALSE	train
	136	TAF1	K562		FALSE	train
	137	TAF1	SK-N-SH	train	FALSE	train
	138	TAF1	HepG2	ladder	FALSE	ladder
	139	TAF1		submit	TRUE	submit
	140	TCF12	GM12878		FALSE	
	141	TCF12	H1-hESC		FALSE	train
	142	TCF12		train	FALSE	train
	143	TCF12	SK-N-SH		FALSE	train
	144	TCF12		ladder		ladder.blind
		TCF7L2		train	FALSE	train
		TCF7L2	HeLa-S3		FALSE	train
		TCF7L2	Panc1		FALSE	train
		TCF7L2		ladder	FALSE	ladder
	149	TEAD4	A549		FALSE	train
	150	TEAD4	H1-hESC		FALSE	train
	151	TEAD4	HCT116		FALSE	train
	152	TEAD4	=	train	FALSE	train
	153	TEAD4	K562		FALSE	train
##	154	TEAD4	SK-N-SH	train	FALSE	train

##	155	TEAD4	MCF-7	ladder	FALSE	ladder
##	156	YY1	GM12878	train	FALSE	train
##	157	YY1	H1-hESC	train	FALSE	train
##	158	YY1	HCT116	train	FALSE	train
##	159	YY1	HepG2	train	FALSE	train
##	160	YY1	SK-N-SH	train	FALSE	train
##	161	YY1	K562	ladder	FALSE	ladder
##	162	ZNF143	GM12878	train	FALSE	train
##	163	ZNF143	H1-hESC	train	FALSE	train
##	164	ZNF143	HeLa-S3	train	FALSE	train
##	165	ZNF143	HepG2	train	FALSE	train
##	166	ZNF143	K562	ladder	FALSE	ladder

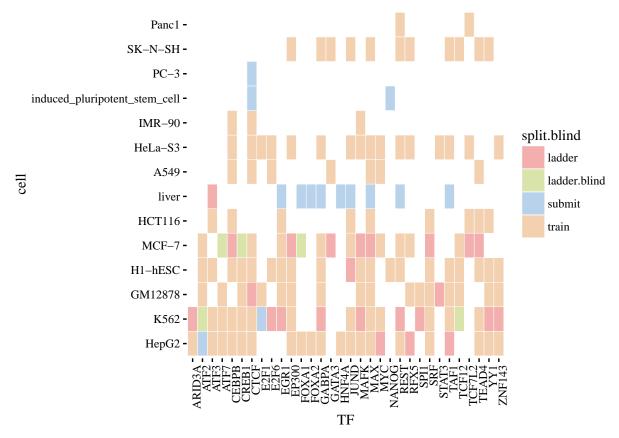
The ENCODE-DREAM challenge

The challenge is to predict cell-type specific binding of transcription factors (TFs) using four types of data:

- DNA sequence: a reference human genomic sequence
- In-vitro DNA shape: the physical shape of the genome in an in-vitro system
- DNase-seq: how open and accessible regions of the genome are
- RNA-seq: levels of gene expression

TF/cell-type combinations

TFs have different binding profiles in different cell types. 32 TFs and 14 cell types are represented in the data although not all combinations are present.



There are essentially three (two!?) prediction tasks:

- Held-out chromosomes: For each TF/cell-type combination, the ChIP-seq data will not be available for 3 chromosomes. Methods will be assessed by their predictive performance on these chromosomes.
- Across cell-type: Methods will be assessed by their performance on cell-types for which no training data has been made available (PC-3, induced pluripotent stem cells, liver).
- Within cell-type: Methods will be assessed on

Data

TF binding

TF binding is measured using the ChIP-seq protocol and converted to a binary score for sliding windows of 200bp. The windows slide by 50bp. For each 200bp location, binding is defined as bound (B), unbound (U) or ambiguous (A). In addition the challenge provides more detailed information from the ChIP-seq experiments including conservative and relaxed estimates of peaks and fold-control signals showing how enriched the ChIP-seq experiment was over a control background experiment. It is not clear how useful the extra information will be as it will obviously not be available on the held-out data.

DNA sequence

The human reference genome is over 3 billion base pairs long. Each base is represented as a character: adenine (A); cytosine (C); guanine (G) and thymine (T). The challenge is restricted to chromosomes 1-22 and chromosome X. All data in the challenge are defined with respect to release GRCh37/hg19. TFs tend to prefer to bind specific sequences. These preferences are summarised in binding motifs but these are not

known for all TFs. The only external data that is allowed to be used in this challenge are libraries of TF-DNA binding motifs. Obviously this data is not cell-type specific.

DNase-seq

Information on chromatin accessibility on a per-cell-type basis will be available in four formats:

- · conservative peaks
- relaxed peaks
- filtered BAM alignment files
- fold-enrichment signal coverage tracks

The first two will be much easier to use as they summarise the last two.

DNA shape

Participants are encouraged to use the DNAshapeR to calculate DNA shape features across the genome. This information is not cell-type specific but has been shown to be predictive of TF-DNA binding. Note that DNAshapeR requires a version of R > 3.3.

Gene expression

Gene expression is regulated by TF binding and so should be useful indirectly to infer binding. The major difficulties are that - It is known which locations on the genome regulate which genes. Commonly TF binding sites regulate the closest gene but this is not always the case. - Several TFs can combine in an unknown way to regulate a gene. - There are other mechanisms of gene regulation that will confound the relationship.

Transcription factors

Motifs

• Hard to find motif for TAF1. Is TBP a good motif to use?