

#### **REVIEW**

# **REVISED** Recent Advances in Biopesticide Research and

# **Development with a Focus on Microbials**

[version 5; peer review: 3 approved, 1 approved with reservations]

Previously titled: Recent Advances in Biopesticide Research and Development: A Focus on Microbial: A

Review

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#### **Abstract**

Biopesticides are pest control products derived from natural sources such as microbes, macro-organisms (insects and pathogens), plant extracts, and certain minerals. Many biopesticides are considered environmentally safe and can complement or substitute conventional chemical pesticides. They can also be highly specific or broad spectrum with a unique mode of action controlling a wide range of pest species. Due to their target-specificity and low to no environmental residuality, biopesticides conform to the 3 pillars of Climate-Smart Agriculture, the Sustainable Development Goals, and, ultimately, the Paris Agreement. This review focuses largely on microbial biopesticides derived from fungi, bacteria, viruses, and nematodes. It discusses (i) the various microbial biopesticide formulations, (ii) the mode of microbial biopesticide action, (iii) the factors that affect the potential efficacy of biopesticides, (iv) challenges to the adoption of microbial biopesticides, and (v) the role



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of microbial biopesticides in Integrated Pest Management programs. Finally, advancements in application techniques, as well as future research directions and gaps, are highlighted.

#### **Keywords**

biocontrol, bioprotection, formulation, climate-smart agriculture, sustainable pest management

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## **REVISED** Amendments from Version 4

We have modified the paragraph: "The products are designed as wettable powders and can be used with various seed treatment fungicides (Table 1). Agrobactenum radiobacter containing strain K1026, a modified version of K84, was registered as a biopesticide in several countries, including Australia, the United States, Turkey, and New Zealand and is used explicitly as a microbial control agent to combat crown gall disease in plants (Kerr and Bullard, 2020), by competing with the pathogenic strain of Agrobacterium tumefaciens, due to its ability to produce an antibiotic called agrocin 84, while being genetically modified to prevent the transfer of this resistance to other bacteria; making it a safer and more targeted microbial agent (Penyalver et al. 2000). K1026 showed strong efficacy against agrocin 84-sensitive and resistant strains of A. tumefaciens (Vicedo et al. 1993)."

We have added the following references in the references

Kerr, A.; Bullard, G. Biocontrol of Crown Gall by *Rhizobium rhizogenes*: Challenges in Biopesticide Commercialisation. *Agronomy* 2020, 10, 1126. https://doi.org/10.3390/agronomy10081126

Penyalver R, Vicedo B & López MM. Use of the Genetically Engineered Agrobacterium Strain K1026 for Biological Control of Crown Gall. European Journal of Plant Pathology (2000). 106, 801-810 https://doi.org/10.1023/A:1008785813757

Vicedo B, Peñalver R, Asins MJ, López MM. Biological Control of *Agrobacterium tumefaciens*, Colonization, and pAgK84 Transfer with *Agrobacterium radiobacter* K84 and the Tra- Mutant Strain K1026. Appl Environ Microbiol 1993; 59 (1); 309-315: https://doi.org/10.1128/aem.59.1.309-315.1993.

Any further responses from the reviewers can be found at the end of the article

### 1. Introduction

Biopesticides are naturally derived pest control compounds that offer a relatively safer and target-specific complement to conventional chemical pesticides (Leo and Rathore 2015; Tijjani et al. 2016; Nuruzzaman et al. 2019; Temiz 2020; Wattimena and Latumahina 2021; Huang et al. 2022). Biopesticides include 1) biochemicals (plant extracts, semiochemicals, microbial extracts/fermentation products, insect growth regulators, compounds synthesized by other organisms, and inorganic compounds), 2) microbial biopesticides (bacteria, fungi, protozoa, viruses, oomycetes, yeast, and algae), 3) macrobials (insect predators, parasitoids, and entomopathogenic nematodes) and classical biocontrol agents (Copping and Leonard 2000; Chandler et al. 2008; Leahy et al. 2014; Liu et al. 2021; Ram and Singh 2021). Biopesticides play a crucial role in promoting climate-smart agriculture (CSA) by offering effective pest management solutions, improving crop yield and quality, and reducing pest resistance while minimizing negative impacts on human health and the environment (Szewczyk et al. 2006; Satish et al. 2017; Kumari et al. 2022). The growing need for biopesticides is partly a consequence of the decrease in the use of conventional pesticides due to farmers increasingly adopting climatesmart agricultural practices and reducing their carbon footprint (FAO 2019). This shift is driven by several advantages associated with biopesticides, including 1) their ability to slow down the development of pest resistance, 2) low to no toxicity to humans and the environment, hence safeguarding biodiversity, preserving soil health and enhancing food security, 3) their ability to complement chemical pesticides, 4) their low environmental fate, 5) host specificity, 6) biodegradability, 7) minimal risk of post-harvest contamination, 8) resilience against environmental stresses, and 9) compatibility with IPM practices (Deravel et al. 2014; Kalpana 2021). Biopesticides may, however, not be equally effective against all pests, and their efficacy can vary depending on the target species. Additionally, some pests may have natural resistance or tolerance to certain biopesticides (Isman 2006).

The excessive reliance on conventional chemical pesticides in agriculture poses a significant threat to environmental health and sustainability, as the agriculture sector contributes a third of the total amount of greenhouse gas emissions. Climate-smart agriculture (CSA) was developed as a framework to contribute to achieving the Sustainable Development Goals (SDGs) under an uncertain climate. Agricultural production systems need to tackle three challenges simultaneously, namely: 1) sustainably increasing agricultural productivity and incomes, 2) building resilience to the impacts of climate change by implementing biopesticide regimes and reducing conventional chemical pesticide use, and 3) contributing to climate change mitigation, where possible (FAO 2017).

The rising consumer demand for sustainably produced food, coupled with the growing resistance of pests to conventional chemical pesticides, the emergence of novel pest threats, and the impacts of climate change, underscores the increasing importance of biopesticides in the future of climate-smart agriculture systems (Chandler et al. 2008). To address this pressing issue, adopting environmentally safe and host-specific pest control options alongside IPM programs can create a winning combination for growers (Chang et al. 2003; Essiedu et al. 2020).

Microbial biopesticides are a broad range of pest control products obtained from microorganisms, including bacteria, viruses, fungi, protozoa, and entomopathogenic nematodes. Many microbial biopesticides can be cost-competitive with other pesticides and fungicides (Ruiu 2018). Microbial biopesticides are the most commonly used in pest control programs (Kvakkestad et al. 2020). Most entomopathogenic bacteria belong to the Bacillaceae family, but the most widely used commercial bacterial biopesticides are from the genus Bacillus, especially the species Bacillus thuringiensis (Bt) and Bacillus subtilis (Bs) (Alsaedi et al. 2017; Akutse et al. 2020). These are aerobic bacteria that produce insecticidal crystal proteins that are toxic to specific insect orders (Gill and Cowles 1992; Raymond et al. 2009). Over 200 Bt-based biopesticides have been registered (Desneux et al. 2022) and are effective against various lepidopterans, coleopterans, dipterans, and hemipterans. However, resistance to some Bt products has been noted. For instance, laboratory resistance to Bt kurstaki has been reported for diamondback moth larvae (Liao et al. 2019), tobacco budworm, and beet armyworm (Blanco et al. 2009; Muhammad et al. 2019). More recently, western corn rootworms have shown field resistance to Bt (Legwaila et al. 2015). Actinomycetes - filamentous, aerobic bacteria resembling fungi - are naturally found in soils and have also been developed into biopesticides (Pitterna et al. 2009). Some Actinomycete species can kill insects, including Tuta absoluta. Two species, Saccharopolyspora spinosa and Streptomyces avermitilis, are used to make bacterial biopesticides (Akutse et al. 2020). Spinosad, a microbial biopesticide made from the aerobic fermentation of S. spinosa contains secondary metabolites called spinosyns (Higa 1994).

Most entomopathogenic fungi belong to the two orders, *Entomophthorales* and *Hypocreales* (previously *Hypomycetes*) (Hajek and Leger 1994). The majority *of entomophthoralean* fungi are obligate parasites with limited host ranges and generate both sexual and asexual spores. Using epizootics, many of these species inhibit insects, the most prevalent genera being *Entomophaga*, *Entomophthora*, and *Zoophthora* (Goettel and Inglis 2000). *Beauveria spp.*, *Metarhizium*, *Isaria*, and *Lecanicillium* spp. are fungi that infect and kill insect pests through direct contact or by penetrating their cuticle, following which the spores germinate and multiply in the insect's body (de Faria and Wraight 2007; Kumari et al. 2022). Viruses from families of baculoviruses, cypoviruses, and densoviruses have been successfully used as viral biopesticides (Abd-Alla et al. 2020). For example, the *nucleopolyhedro* virus effectively infects and kills insect larvae (Cory 2000) and is used to control a variety of serious insect pests, especially moths (Gramkow et al. 2010). Insect viruses are considered environmentally safe due to their high selectivity and lack of non-target organisms species shifts. They also degrade naturally and do not persist in the environment (Abd-Alla et al. 2020). Despite their advantages, insect viruses face certain challenges as microbial biopesticide agents. These include their slow action, narrow host range, and potential for resistance development in insect hosts (Abd-Alla et al. 2020). Addressing these challenges through research and development efforts is crucial to fully realize the potential of insect viruses as sustainable pest control solutions.



**Figure 1. The 3 pillars of Climate-Smart Agriculture and the Sustainable Development Goals (FAO, 2019).** The visual mapping illustrates how the three pillars of Climate smart agriculture (CSA) interact with and impact each of the 17 Sustainable Development Goals (SDGs). The second part demonstrates how the five steps for implementing CSA intersect with and affect the 17 SDGs in terms of synergies and trade-offs.

Generally, microbial biopesticides can have shorter shelf lives compared to chemical pesticides. They may also be sensitive to environmental factors such as sunlight, temperature, and moisture (McManus 1989; Matos et al. 2020; Wattimena and Latumahina 2021). However, if stored according to the label instructions, they could last up to 3-6 months (Akutse et al. 2020). There is a potential for microbial biopesticides to be integrated into IPM strategies in agriculture without significantly compromising productivity and yield (Aroraet al. 2020). However, several limitations currently hinder the widespread adoption of microbial biopesticides. These include 1) the high cost of refined commercial products, 2) policy and regulatory issues, 3) challenges in meeting global market demand, 4) lack of standardized preparation methods and guidelines, 5) difficulties in determining appropriate dosages of active ingredients, 6) susceptibility to environmental factors, 7) limited stability, and 8) slow adoption by end-users. While research breakthroughs are expected to address these limitations in the coming years, farmers, particularly those in rural areas, can still benefit from using microbial solutions as a means of plant protection to enhance crop quality and ensure farms are productive and profitable (Stevenson et al. 2017). It is important to note that the maximum effectiveness of biopesticide applications, including microbial biopesticides, is achieved when they are integrated into an overall IPM approach (Quarles 2013) and aligned to the 3 pillars of climate-smart agriculture (FAO 2017) (Figure 1). Climate-smart agriculture (CSA) has a strong potential to contribute to achieving the 17 Sustainable Development Goals (SDGs). A critical precondition for realising this scope is identifying potential synergies and trade-offs between the three pillars of CSA and the five implementation steps of CSA and the SDGs. These provide entry points for targeted CSA planning to enhance synergies and reduce trade-offs. CSA and the SDGs have synergies and trade-offs at different levels for each of the three pillars of CSA, where some of the synergies & trade-offs overlap all three pillars of CSA for some of the SDGs, namely SDGs 2 and 6. The overlapping synergies & trade-offs for the implementation steps and SDGs are different, as only step 2 has overlapping trade-offs and synergies in SDGs 6, 6, 9, 13, and 15 (United Nations 2015).

## 2. Methods

To comprehensively gather information on biopesticides, we employed a systematic and rigorous approach. We utilized a wide range of electronic sources, including Google Scholar, PubMed, Scopus (Elsevier), Web of Science, Semantic Scholar, Academia, and other relevant websites, to conduct extensive literature searches. Our analysis of over 200 scientific papers and other relevant online resources enabled us to amass a comprehensive archive of pertinent literature. Only microbial (bacterial, fungal, viral, nematode, and protozoa) biopesticides and other types were excluded. Our primary focus was on recent advancements in the development and application of biopesticides within the framework of IPM within the climate-smart agriculture framework. The literature outcomes were categorized into the (i) classification of biopesticides, (ii) importance of biopesticides in sustainable agriculture, (iii) role of microbial biopesticides in integrated pest management, (iv) different types and formulations of microbial biopesticides, and (v) advancement and future perspectives of microbial biopesticides.

# 3. Concept and classification of biopesticides

Biopesticides are defined by the US Environmental Protection Agency (USEPA) as 'naturally occurring substances, microorganisms, and plant-produced substances that control pests' (EPA 2023). While the USEPA defines biopesticides as "pesticides derived from naturally occurring substances", this term is not universally recognized. According to the Southern African Development Community (SADC), the term biopesticide is generally applied to a substance derived from nature, such as a microorganism or botanical or semiochemical, that may be formulated and applied like a conventional chemical pesticide and that is normally used for short-term pest control (SADC 2019). The East African Community (EAC) also classified the term "biopesticides" as microbial, macrobial, botanical, and semiochemical biopesticides derived from or based on genetically modified organisms (GMOs) and pest control agents. The International Biocontrol Manufacturer's Association (IBMA) and the International Organization for Biological Control (IOBC) prefer the term "biocontrol agents" (BCAs) instead. IBMA classifies BCAs into four groups, namely macrobiotics (predators and parasites), microbial (bacteria, fungi, and viruses that kill or harm pests), natural products (plant extracts), and semiochemicals (chemical signals) (Guillon 2003). The European Environmental Agency define biopesticide as "a pesticide in which the active ingredient is a virus, fungus, bacteria, or a natural product derived from a plant source" (European Environment Agency 2023).

Biopesticides offer several advantages over conventional chemical pesticides, including lower toxicity to non-target organisms, reduced environmental impacts, and less likelihood of developing resistance to pests. Biopesticides effectively control insects, diseases, weeds, and nematodes, promote plant health, and enhance the productivity and profitability of farmers. Importantly, many of them are non-toxic to beneficial organisms and wildlife. Additionally, their biodegradability reduces pollution concerns associated with many chemical pesticides (Shilpi and Promila 2012; Essiedu et al. 2022). Due to their targeted action and effectiveness in small quantities, biopesticides are gaining wide application on plants and crops. They are playing an increasingly significant role in agriculture, promoting sustainable practices and a healthier environment (EPA 2023), thus complying with the climate-smart agriculture principles (FAO 2017), and, ultimately, the Paris Agreement (United Nations 2015), the Nagoya Protocol of Article 6 (Access to Genetic

Resources) and Article 7 (safeguards the rights of indigenous and local communities regarding their traditional knowledge and ensures they benefit from its use) (The Nagoya Protocol 2014).

#### 3.1 Mechanism of action and efficacy of biopesticides

How do they work? Biopesticides work through various mechanisms, such as 1) biochemical disruption,-where they interfere with the biochemical processes of pests, affecting their growth, development, or reproduction; 2) microbial activity-infecting or parasitizing pests; 3) predation and competition-acting as natural predators or competitors of pests to limit their population growth; and 4) repellency-deterring pests from attacking crops (Essiedu et al. 2022). They can play a crucial role in IPM strategies and contribute to sustainable agricultural practices by reducing the environmental impact and minimizing the risk of pesticide resistance (Essiedu et al. 2022), thereby complying with the 3 pillars of climate-smart agriculture and the synergies with the SDGs (FAO 2024) (Figure 1).

## 4. Importance of biopesticides in sustainable agriculture

Biopesticides are an important tool in sustainable agriculture, and their use will likely continue growing. The use of biopesticides has steadily increased at a Compound Annual Growth Rate (CAGR) of 11.0% between 2018 and 2022 (CAGR) between 2018 and 2022. The global biopesticides market reached a value of US\$ 8,123.8 million in 2023. This upward trend is expected to continue, with the market expanding at a 10.3% CAGR and reaching US\$ 21,827.6 million by 2033 (Persistence Market Research 2023). Food security is essential due to the world's growing population. Humans and global economies depend on a stable and reliable food supply. As the population grows, food security will become more threatened (FAO 2024). Sustainable farming and maximizing soil fertility require non-toxic and environmentally sustainable pesticides. Thus, the need for biopesticides will likely increase over time (Bharti and Ibrahim 2020). Sustainable farming depends on biopesticides, which are poised to advance significantly in the coming years (Copping et al. 2000; Alavanja 2009). Optimizing manufacturing processes to increase yields and reduce costs can make biopesticides more economically viable. By using biopesticides, farmers can reduce their reliance on chemical pesticides, protect human health and the environment, and produce safe and healthy food (Hezakiel et al. 2023; FAO 2024).

#### 5. Role of biopesticides in integrated pest management

Biopesticides play a vital role in IPM programs, which aim to control pests effectively whilst reducing the reliance on conventional chemical pesticides (Jozsef 2020). As the demand for more sustainable, environmentally safe, and sustainable agricultural practices grows, the use of biopesticides is becoming increasingly popular as part of IPM strategies. IPM aims to combine multiple pest control methods, including biopesticides, to minimize the use of chemical pesticides and maintain a balanced ecosystem in agricultural environments (Jozsef 2020). When used within IPM programs, biopesticides offer several advantages, including 1) targeted pest control, 2) diverse modes of action, 3) resistance management, 4) reduced negative environmental impact (Isman 2006), 5) sustainable crop production (Gurr et al. 2017), and 6) reduced risk of pesticide resistance in non-target organisms (Prasanna et al. 2018). Many biopesticides have the potential to contribute to economic growth and rural livelihood improvement, thus complying with the SDGs (Figure 1).

Additionally, biopesticides offer potential benefits in the face of climate change challenges, and they align with the growing consumer demand for environmentally friendly and sustainable agricultural practices (FAO 2017; Akutse et al. 2020). Besides, biopesticides 1) generate fewer greenhouse gasses (GHG) than chemical pesticides, 2) improve nutrient availability for plants, 3) increase soil fertility, and 4) offer a more environmentally safe approach to pest control in agriculture, which can be useful in the fight against climate change.

#### 5.1 Biopesticide categories

Biopesticide is a generic term generally applied to a substance derived from nature, such as a botanical or semiochemical, that may be formulated and applied like a conventional chemical pesticide and that is normally used for short-term pest control (USEPA 2020). Naturally derived or laboratory-made chemicals with structures and functions similar to naturally occurring ones are known as biochemical pesticides. They differ from traditional pesticides in their origin (source) and how they control or kill pests (O'Brien and Jones 2009). Microbial pesticides are natural pest control agents harnessing the power of microscopic living organisms (Clemson 2007). These "living bullets" can be either spores or active organisms, often specifically chosen for their pathogenic nature towards targeted pests. Common examples include biofungicides (*Trichoderma*, *Pseudomonas*, *Bacillus*, *Colletotrichum*), bioherbicides (*Phytophthora*, *Cylindrobasidium*, *Colletotrichum acutatum*), and bioinsecticides (*Bt*) (MacGregor 2006; Gupta 2010). Microbial biopesticides can originate from naturally occurring or genetically modified bacteria, fungi, algae, viruses, or protozoans. They subdue pests through a diverse range of strategies (toxic metabolites, disease-causing, competitive exclusion, and other modes of action), depending on the specific microbe (Clemson 2007).

Microbial biopesticides offer diverse delivery methods to effectively reach crop targets. These methods include live organisms, dead organisms, and spores (O'Brien and Jones 2009). Globally, microbials account for 41% of all BCAs used worldwide, macrobiotics comprise 33% of the global market, and other natural products represent the remaining 26% (Guillon 2003). Based on their origin or active ingredients, they can be classified as bacterial-based biopesticides, fungal biopesticides, viral biopesticides, nematodes, and protozoan-based biopesticides.

#### 5.2 Bacterial-based biopesticides

Among all microbial pesticides, bacterial biopesticides are the most widely used. These versatile microbes attack pests in multiple ways, primarily targeting insects. While mostly used against various orders of insect pests in agriculture, bacterial biopesticides can also curb the growth of plant-harming bacteria and fungi (O'Brien and Jones 2009; Jouzani and Valijanian 2017). Bacterial pathogens need direct contact with their target to be effective, often requiring ingestion. They secrete endotoxins, protein-based toxins specific to the targeted pest that wreak havoc on their digestive system (Schallmey and Singh 2004). Bacillus thuringiensis (Bt) dominates the biopesticide market, accounting for a whopping 90% share in the USA alone (Chattopadhyay and Bhatnagar 2004). Bacillus thuringiensis is the most common species used commercially to control a diverse range of insect pest species, including lepidopterans, hemipterans, and coleopterans in agriculture, forestry, and even medicine since its discovery in 1901 (Mazid 2011). It is a sporeforming bacterium with several strains that produce diverse insecticidal crystal proteins, often targeting different pests (Table 1). The crystal proteins are toxic and, once ingested, damage the gut tissues, leading to paralysis of the gut. The infected insects stop feeding and eventually die from gut impairment and starvation. Bt var. kurstaki specifically targets caterpillars, while the genes encoding Cry proteins have been transferred into crops like cotton, hence reducing reliance on chemical pesticides (Mazid 2011). Commercial Bt-based products are available in various forms, such as powders containing a combination of dried spore and crystal toxins or formulations in liquid suspensions (Boyetchko et al. 2020). Bacillus thuringiensis's high target specificity and environmental safety make it an ideal complement to traditional chemical pesticides for insect pest control (Roy and Moktan 2007; Kumar 2012). Figure 2 illustrates the effects of Bacillus thuringiensis on insect larvae. Other entomopathogenic bacteria of interest include Chromobacterium subtsugae, Brevibacillus laterosporus, Lysinibacillus sphaericus, Paenibacillus popilliae, Serratia marcescens, and Yersinia entomophagy (Gangwar et al. 2021).

Table 1. Microbial biopesticide categories, mode of action, and their use in pest management.

Fungal-based biopesticides	Application type and mode of action	Use	Remark
Beauveria bassiana	It infects the host insect through the cuticle, colonizes its body, and ultimately causes death (Baldiviezo et al. 2020). This fungus is known for its rapid multiplication and the production of various toxins that result in exogenous infections (Nakahara et al. 2009; Naqqash et al. 2016).	Widely used fungal biopesticides are effective against numerous insect pests (Naqqash et al. 2016).	It is among the most effective biocontrol fungi (Naqqash et al. 2016).
Metarhizium anisopliae	It can infect and kill a variety of insect pests, and this product is suitable for use in non-food areas such as ornamental greenhouses, nurseries, residential and institutional lawns, and landscape perimeters. However, it should not be used in areas where there is a risk of water contamination (Naqqash et al. 2016; Montalva et al. 2016). It can also be used for the control of mosquitoes in the public health domain (Vivekanandhan et al. 2022)	This product targets a variety of pests, including ticks and beetles, as well as aphids, mealybugs, fruit flies, root weevils, grasshoppers, whiteflies, gnats, and thrips. etc. (Montalva et al. 2016), and mosquitos (Vivekanandhan et al. 2022). <i>M rileyi</i> controls lepidopterans.	It can be applied at terrestrial non-food sites and as indoor residual spraying for mosquitoes.

 Table 1. Continued

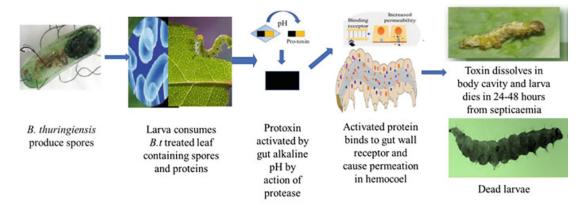
Fungal-based biopesticides	Application type and mode of action	Use	Remark
Verticillium lecanii also known as Lecanicillium lecanii	This product includes fragments of <i>Verticillium lecanii</i> fungi, along with spores. When these spores encounter the outer layer of a targeted pest insect, they germinate, penetrate their cuticle, and proliferate within their bodies. This eventually leads to the insect's demise, as it becomes drained of nutrients (Lee et al. 2006).	Used for controlling aphids, whiteflies, and other sucking insects from fruits, vegetable crops, etc. (Lee et al. 2006; Feng et al. 2000). Lecanicillium lecanii targets whiteflies, leafminers, aphids, and scale insects (Ruiu 2018).	This agro-input is harmless, environmentally safe, and cost-effective.
Paecilomyces lilacinus		Controls plant parasitic nematodes (Akutse et al. 2020).	
Conidiobolus thromboides Acari	Hemiptera, Thysanoptera		
Isaria fumosorosea		I. fumosorosea targets soft- bodied insects such as whiteflies (Akutse et al. 2020).	
Trichoderma sp.	Trichoderma spp. is a type of fungicide that is highly effective in combating soilborne diseases like root rot. It is especially beneficial for dryland crops such as groundnut, black gram, green gram, and chickpea, which are prone to these diseases (Ghayur 2000).	They are known for their antagonistic interactions with various plant pathogens, including fungi, bacteria, and nematodes (Ghayur 2000). <i>Trichoderma asperellum</i> against soilborne pests.	The use of <i>Trichoderma</i> spp. As a biopesticide is promising to control plant diseases and insect pests. (Howell 2003).
Phytophthora palmwora MWV	Liquid bioherbicide formulation	used to control stranglervinrie weed.	(Kachhawa 2017)
Colletotrichum gIloeosponoides	A dry powder bioherbicide	Targeted to control Northern joint vetch	(Kachhawa 2017)
Colletotrichum acutatum	A liquid bioherbicide formulation	Targeted to control <i>Hakea</i> sericea and <i>H. gibbosa</i>	it is used in agriculture, forestry, and conservation applications to control these alien and invasive species (Muir 2023).
Bacterial-based pe	sticide		
Bacillus thuringiensis (Bt)	The most widely used biopesticide worldwide is Bt. It primarily targets lepidopterous pests, which are known to cause grave damage. Some examples are American bollworms in cotton and stem borers in rice. It releases toxins that harm the midgut of the pest when the larvae consume it, leading to its demise (Ghayur 2000).	Lepidopteran pests, such as stem borers in rice.	It is a highly specific, safe, and effective organism for insect control (Roh et al. 2007).
B. subtilis	Produce a variety of antimicrobial compounds, such as antibiotics, lipopeptides (e.g., surfactin, iturin), and cyclic peptides (e.g., bacillomycin), which inhibit the growth and activity of plant pathogens (Djaenuddin et al. 2020).	This product targets pests such as wilts, crown rot, root rot, and other seedborne diseases that are caused by fungi such as Fusarium, Aspergillus, Pythium, and Rhizoctonia (Djaenuddin et al. 2020).	It is safe for the environment and humans.

 Table 1. Continued

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Fungal-based biopesticides	Application type and mode of action	Use	Remark		
Btvar kurstakz	This product is used as a bioinsecticide	It is used to target Lepidopteran spp.	No side effects to non- target organisms.		
Bt israelensis	controls gnats fly	It is used to control gnats flies	(Ruiu 2018)		
Bt tenebrionsis	Beauveria bassiana	shows activity against Coleopteran adults and larvae	(Rajaput et al. 2019)		
Bt. Vnar. sun dlego Bt var. tenebnoms	Both are bioinsecticides effective against targeted pests.	Developed by Ecogen and Thermo Trilogy company to control Coleopteran pests.	No phytotoxicity effects		
Pseudomonas fluorescens-	Pseudomonas species are significant bacteria in agriculture. Studies have demonstrated that they can enhance plant growth and shield them from pathogens and herbivores (Daniel 2020).	It can colonize the rhizosphere and produce secondary metabolites that suppress soil-borne pathogens (Glick 2012). These bacteria also play a role in phytoremediation and are part of the essential microbiome of numerous plants (Adesemoye and Kloepper 2009; Germaine et al. 2009).	No toxicity effects to non-target organisms.		
Streptomyces species	Filamentous bacteria that produce various antifungal compounds. <i>Streptomyces strains</i> possess valuable applications in agriculture due to their ability to biologically control phytopathogens, particularly fungi that are harmful to plants (Law et al. 2017).	They have been utilized as biocontrol agents against plant pathogenic fungi, such as <i>Fusarium</i> and <i>Phytophthora</i> species (Gonzalez-Franco 2009; Ara et al. 2014).	Streptomyces strains display antibacterial, antifungal (Ara et al. 2014), and immunosuppressive tendencies.		
Viral-based biopes	ticides				
Cydia pomonella granulovirus- (CpGV)	They are commonly used for the control of the codling moth ( <i>Cydia pomonella</i> ), a major pest of apple and pear orchards. <i>CpGV</i> is a safe, effective, and environmentally friendly biocontrol agent for codling moth (EPPO 2019).	The CpGV virus particles infect and kill the codling moths' larvae, reducing pest populations (EPPO 2019).	It is a valuable tool for sustainable agriculture.		
Baculoviruses	Viruses are aimed at specific targets and are capable of infecting and eliminating various significant plant pests. Known as baculovirus insecticides. Nucleopolyhedroviruses (NPVs) control Helicoverpa armigera, Helicoverpa spp. Spodoptera exigua, S litura. Granuloviruses (GVs) control Plutella xylostella, Thaumatotibia leucotreta, Pieris rapae, Cydia pomonella	These viruses have been proven to be highly effective against pests such as lepidopterous in cotton, rice, and vegetables (Chang et al. 2003; Ruiu 2018; Kumar et al. 2021).	They are insect-specific viruses widely used as biopesticides.		
Cylindrobasidium laeve	A liquid bioherbicide formulation	Targeted for the control of Acacia mearnsii and A. decurrans	it is used in agriculture, forestry, and conservation applications to control these alien and invasive species (Muir 2023).		

Table 1. Continued

Fungal-based biopesticides	Application type and mode of action	Use	Remark		
Nematode based biopesticides					
Steinernema spp. and Heterorhabditis spp.		are used on different host crops to control pests such as leafminers, thrips, weevils, beetles, cutworms, scarab grubs, lepidopterans, and fungus gnats.	(Koppenhöfer et al. 2020; Tarasco et al. 2023; Rae et al. 2023).		
Phasmarhabditis hermaphrodita		used against several slug and snail species.	(Rae et al. 2023)		
H. marelatus		It is used to control white grub black vine weevils.	os (scarabs), cutworms, and		
H. bacteriophora	White grubs (scarabs), cutworms, black vine weevils, flea beetles, corn rootworms, citrus root weevils ( <i>Diaprepes</i> spp.) (Kachhawa 2017)				
H. megidis	Used to control weevils				
S. glaseri	Used to control white grubs (scarabs, especially Japanese beetle, <i>Popillia</i> sp.), banana root borer (Kachhawa 2017)				



Mode of action of B.t

Figure 2. Effects of Bacillus thuringiensis (Bt gene and Cry protein) on insect larvae (Singh et al. 2019).

Recombinant DNA technology has been used to develop several new Bt-based products. MVP<sup>TM</sup> and M-Trackr<sup>TM</sup>, for example, were created using Mycogen Corporation's CellCap<sup>®</sup> encapsulation method. This procedure entails removing a gene from *Bt* that encodes the delta-endotoxin protein, incorporating it onto a plasmid, and inserting it into a *Pseudomonas fluorescens* strain (Pitterna et al. 2009; Legwaila et al. 2015). Before being killed by heat and chemical treatment, the recombinant cells were cultured in aerobic culture and induced to express delta-endotoxin. The dead bacterial cells in the aqueous formulation acted as microcapsules, protecting the fragile *Bt* toxin from destruction in the environment.

Pathogenic bacteria have also been considered for biological weed control. However, one of the challenges to using phytopathogenic bacteria for biological weed control is the necessity of free water for dispersal and wounds or natural holes for bacterial penetration into the weed plant (Zidack et al. 1992; Johnson et al. 1996). Previously, researchers have studied *Xanthomonas campestns pv. Poae* for the control of annual bluegrass (*Pea annua* L.) and discovered that cutting or mowing turfgrass allows bacteria to enter the plant (Imaizumi et al. 1997). Furthermore, bacteria administered at a rate of log CFU/mL at high water levels (400 mL/m²) reduced disease severity in annual bluegrass by more than 90%. Silwet L-77 (0.2%), an organosilicon surfactant, increased bacterial penetration and entrance into plant stomata and hydathodes (Johnson et al. 1996; Boyetchko et al. 2020). A low surface tension of 30 dynes/cm or less is necessary to convey liquid into a leaf's stomata. Silwet decreases the surface tension of water to 20 dynes/cm. Compared to plants sprayed with

bacteria without the surfactant, the application of *Pseudomonas syringae* pv. *Tagetes* with this surfactant facilitated the penetration and entry of bacteria into stomata and hydathodes, significantly increasing disease severity and incidence (Johnson et al. 1996).

Bacterial biopesticides are environmentally safe and precise alternatives to traditional pest control methods. They offer several advantages, including safety for humans, wildlife, and beneficial insects. These biopesticides can be used in conjunction with other pest control strategies, including some chemical methods. Additionally, they leave no harmful residues, don't harm pollinators and other valuable organisms, and can be applied close to harvest time. The additional benefit is that they can self-propagate, extending their effectiveness beyond the initial application and into subsequent growing seasons (Meenatchi and Aditi 2021). Bacterial biopesticides, while effective, come with a few disadvantages. For instance, they only target specific species or groups of insects, leaving other pests to survive and continue causing damage. This means that additional biopesticides are needed to complement the treatment when dealing with different pests. Hence, they should be applied alongside other IPM strategies. Finally, the effectiveness of microbial insecticides can be influenced by factors like ultraviolet radiation and heat, so it's important to apply them strictly according to the label (Meenatchi and Aditi 2021).

## 5.3 Fungal biopesticides

Mycopesticides include naturally occurring fungi and fungi cell components. These fungi attach to their targets through sticky spores. These spores then germinate and form microscopic tubes that pierce the insect's skin, releasing potent hydrolytic enzyme mixtures along with toxins (Langner and Göhre 2016). This internal invasion disrupts the insect's physiology, leading to its eventual death. The fungi then grow outward from the dead insect, producing new spores to continue the cycle, hence perpetuating natural pest control (Pineda et al. 2007; Gabarty et al. 2014). The role of hydrolytic enzymes, especially chitinases, in the killing process, and the possible use of chitin synthesis inhibitors are prime research areas. Popular commercially available mycoinsecticides are often derived from species like *Beauveria bassiana*, *Metarhizium anisopliae*, *Isaria fumosorosea*, *Hirsutella thompsonii*, *Phytophthora palmivora*, *Alternaria cassia*, and *Lecanicillium spp*. (Table 1). Notably, *Beauveria bassiana and Metarhizium anisopliae* are two common ascomycetes known for their broad effectiveness against pests like aphids, beetles, grasshoppers, and caterpillars. These fungi are typically applied as conidia (spores) or mycelium, which then sporulate after application, ensuring sustained pest control (Lacey et al. 2015). *Cylindrobasilium laeve* for the control of *Acacia mearnsii* (black wattle) and *Colletotrichum acutatum* for the control of *Hakea sericea* (silky hakea) are some of the available mycoherbicides for integrated alien and invasive species management.

Fungal biopesticides hold immense promise as environmentally safe options against a diverse range of insect and mite pests. They possess several desirable traits for biocontrol: they selectively target pests, leaving beneficial insects like bees and predators unharmed. Additionally, they generally pose no threat to the growth or development of other beneficial organisms like earthworms and springtails. This makes them ideal candidates for IPM and sustainable agricultural practices, promoting biodiversity and protecting the environment (Goettel et al. 2008; Kim and Goettel 2011; Koike et al. 2011). Several fungi have been successfully mass-produced and formulated for commercial application as mycoinsecticides, paving the way for their widespread use in pest control (Chandler et al. 2008).

The remarkable potential of entomopathogenic fungi is actively being harnessed for use in IPM programs. Mass production techniques and ecological approaches are constantly refined to optimize their effectiveness and ensure seamless integration into sustainable agricultural practices (Chandler et al. 2011; Lacey et al. 2015). Furthermore, promising results have been achieved by combining fungal biopesticides with traditional insecticides, often leading to significantly boosted pest mortality rates. For example, studies have shown that combining *B. bassiana* with low-dose insecticides enhances potato beetle control, while its synergy with neem oil proves effective against tobacco thrips eggs and nymphs (Chandler et al. 2011; Lacey et al. 2015; Sarwar 2015).

Fungi-based biopesticides boast several advantages over other biocontrol methods. Their broader host range allows them to tackle a diverse range of pests across fields, greenhouses, storage facilities, and soil, unlike bacteria and viruses with limited targets (Amruta et al. 2019). Commercially important fungi like *Beauveria, Metarhizium, Lecanicillium*, and *Isaria* are surprisingly easy to mass produce, requiring significantly less substrate compared to alternative methods. These fungi are highly productive, delivering powerful, targeted pest control while reducing the impact on the environment. They pose no risk to beneficial organisms, readily biodegrade, and integrate seamlessly into IPM programs. Additionally, their persistence in the environment offers long-lasting protection against pest outbreaks, making them a potentially cost-effective solution for sustainable agriculture (Naveenkumar et al. 2017; Rajaput et al. 2019; Sufyan et al. 2020; Upamanya and Bhattacharyya 2020).

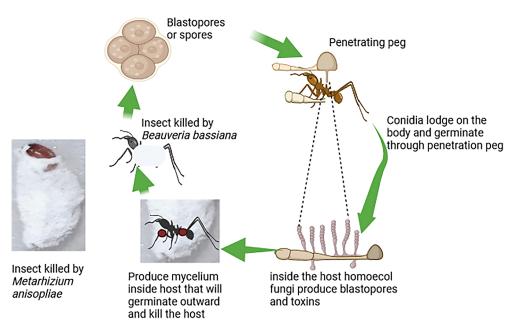


Figure 3. Mode of action of fungi-based biopesticides.

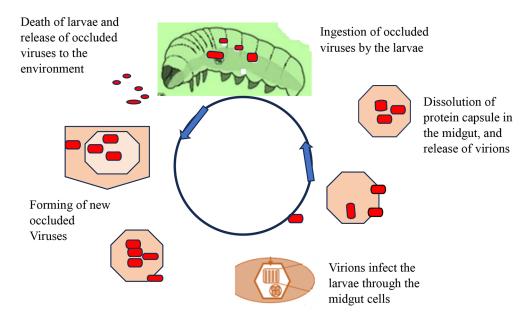
Fungi-based biopesticides, however, face some environmental hurdles. Optimal spore germination and penetration into insect cuticles often require humidity levels exceeding 80%, depending on the specific fungal species. Temperature fluctuations and exposure to UV radiation can significantly impact their survival. Germinating conidia, essential for active pest control, are often delicate and susceptible to environmental damage. Conversely, the production of durable resting spores for long-term persistence can be expensive compared to other microbial methods like bacteria-based biopesticides (Sabaratnam 2002; Kolombet et al. 2008; Kordali et al. 2008; Li et al. 2014). Figure 3 illustrates the mode of action of fungi-based biopesticides, highlighting the steps by which these biopesticides attack target insect pests (Figure 3).

#### 5.4 Viral biopesticides

Baculoviruses (BVs) are tiny, naturally occurring viruses that have emerged as powerful allies in the fight against insect pests. They belong to a group known as entomopathogenic viruses, specifically targeting and infecting insects and other arthropods. Unlike harmful conventional chemical pesticides, baculoviruses pose no threat to humans, wildlife, or beneficial insects, making them an environmentally safe option for pest control (Cory 2000; Jackson and Crawford 2005; Ramle et al. 2005; Moscardi et al. 2011), and are considered as the most commercial viral biopesticides (Granados 1986; Moore and King 1987). Baculoviruses involve circular supercoiled double-stranded DNA genomes, which range from 80 to 180 kbp (Rohrmann 2013). Within the *Baculoviridae* family, scientists have identified three main subgroups: nuclear polyhedrosis viruses (NPVs), cytoplasmic polyhedrosis viruses (CPVs), and granulosis viruses (GVs). These subgroups differ in the structure and number of their protective protein coats, called occlusion bodies (Moscardi et al. 2011). These bodies allow the virus to survive outside the host, waiting for its next target. NPVs and CPVs form polyhedral bodies with numerous virus particles, while GVs have smaller, granular bodies containing just one particle. Notably, each subgroup employs a unique method to uncoat and establish infection within their host: NPVs in the nucleus, CPVs in the cytoplasm, and GVs within the nuclear pore complex (Cory 2000; Moscardi et al. 2011).

Baculoviruses offer highly specific targeting, primarily focused on lepidopteran larvae and hymenopterans (butterfly and moth) pests that damage crops like cotton, rice, and vegetables. One example is *Heliothis zea nucleopolyhedrosis* virus (HzNPV), the first commercially successful broad-spectrum viral insecticide. Interestingly, HzNPV demonstrates versatility, effectively controlling pests across various crops like soybeans, sorghum, maize, tomatoes, and beans. While other entomopathogenic viruses exist, such as tetraviruses and cypoviruses, their use in crop protection remains limited compared to the widespread adoption of baculoviruses (Sarwar 2015). A comprehensive summary of viral biopesticides and their target pests is presented by (Usta 2013; Singh et al. 2019; Meenatchi and Aditi 2021).

Figure 4 shows how viral replication and infection take place and attack their target host. After infecting a target cell, viral replication in the nucleus or cytoplasm unfolds in three distinct phases: early (0-6 hours), second (6-24 hours), and very



**Figure 4. Mode of action of viral biopesticides.** The virions enter the midgut cell nucleus, at which point the virus replicates within the nuclei of susceptible tissue cells, and tissue susceptibility varies greatly between viruses, with some NPVs being capable of infecting almost all tissue types and most GVs being tissue-specific replications. In the terminal stage of infection, the insect liquefies and thus releases polyhedral, which can infect other insects upon ingestion. A single caterpillar at its death may contain over 109 occlusion bodies from an initial dose of 1000. Under optimal conditions, target pests may be killed in 3-7 days, but when the condition is not suitable, death may be caused in 3-4 weeks (Kalamkoff 2004).

late (24-72 hours). During the late phase, the virus forms protective protein coats called occlusion bodies (Obs) or virions. These tiny packages containing multiple virus particles can lead to natural outbreaks (enzootics) that drastically reduce pest populations. However, sunlight poses a challenge: occlusion bodies can rapidly lose potency when exposed to ultraviolet radiation (UV) between 280-320 nm (Killick 1990). Interestingly, studies show that plastic greenhouses can help mitigate this issue by filtering out over 90% of harmful UV-B rays, boosting infection rates in larvae (Lasa et al. 2007).

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Several studies have been conducted to explore ways to improve the effectiveness of viral biopesticides. Certain formulations, like stilbene, can enhance susceptibility to NPV infection by disrupting the insect's digestive barrier (Okuno et al. 2003) or inducing cell death in the midgut (Dougherty et al. 2006). Additionally, researchers are investigating genetically engineered viruses. For example, vAcTaITX-1 and vAcDTX9.2, derived from specific spiders, show promise as commercial biopesticides against lepidopteran pests (Hughes et al. 1997). While viral biopesticides offer numerous advantages over conventional chemical pesticides, including specificity and environmental safety, they face certain challenges for large-scale adoption. Producing them efficiently, particularly recombinant viruses, can be costly and labor-intensive, requiring specialized equipment and lengthy procedures (Gupta 2010; Lacey et al. 2015). Despite these hurdles, organizations like IPM centers and state agricultural departments are actively supporting the development and small-scale production of these promising biopesticides.

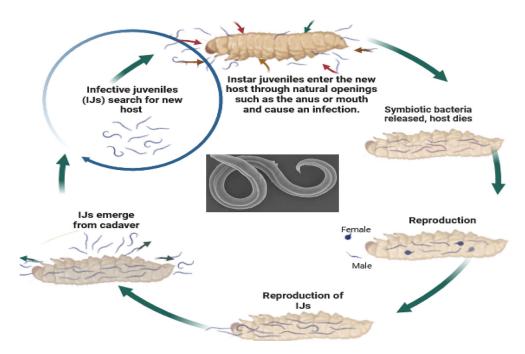
What is the main advantage of viral biopesticides? Viral biopesticides offer a safer alternative, protecting human health, wildlife, and even beneficial insects. These targeted pathogens would not trigger resistance in pests, making them a lasting solution. Moreover, they seamlessly work with other pest control methods for a comprehensive defense (Singh et al. 2019; Meenatchi and Aditi 2021). While viral biopesticides offer enticing, environmentally safe alternatives to chemical pesticides, they come with limitations. Unlike the broad spectrum of some conventional chemical insecticides,

their use often remains targeted to specific pest species. Additionally, successful pest control can be a slow process, as infections may take a significant amount of time to become lethal and can be easily deactivated by high temperatures and ultraviolet radiation. Understanding these vulnerabilities is crucial for choosing the right pest control strategy (Singh et al. 2019; Meenatchi and Aditi 2021) for the targeted pest and bioclimatic zone.

# 5.5 Nematodes as biopesticides

Entomopathogenic nematodes (EPNs) are classed as microbial pesticides, even though they are multicellular. EPNs are tiny soil-dwelling organisms that live in the water films around soil particles, fit nicely into IPM programs because they are considered nontoxic to humans, relatively specific to their target pests, and can be applied with standard pesticide equipment (Shapiro-Ilan et al. 2006). These beneficial parasites help maintain the balance in the ecosystem by targeting specific pests without harming other organisms (Kachhawa 2017). Two main families of EPNs, *Steinernematidae*, and *Heterorhabditidae*, have been enlisted in the fight against pests (Bhat and Chaubey 2020). Species like *S. carpocapsae*, *S. thermophilum*, *H. bacteriophora*, and *H. indica* are commercially available and actively deployed in pest control programs in India (Meenatchi and Aditi 2021). Commercial biopesticides based on EPNs contain non-feeding, third-stage infective juveniles (IJs) as the active ingredients. *Heterorhabditis* and *Steinernema* are mutualistically associated with bacteria of the genera *Photorhabdus* and *Xenorhabdus*, respectively (Ferreira 2014).

So, how do these tiny organisms take down their targets? EPNs, in their infective juvenile stage, barely millimeters long, use their keen senses to locate insects through carbon dioxide emissions, vibrations, and other chemical cues. They then wriggle their way into the insect's body through natural openings like the mouth, anus, or breathing holes. Once inside, the nematodes release their symbiotic bacterial companions, setting off a natural pest control cascade. These bacteria produce toxins that quickly kill the insect within a week. The EPNs then feast on the insect's liquefied remains, multiplying within the host to create new generations of tiny warriors. Finally, once their feast is over, the next generation of infective juveniles emerges, ready to seek out their next victim and continue the cycle (Figure 5). Under ideal conditions, the impact of EPNs becomes visible within 5-7 days. Look for browning or tanning of insects infected by *Steinernematidae*, while those taken down by *Heterorhabditidae* turn a distinctive red (Gupta 2010; Koul 2011; Ruiu 2018).



**Figure 5. Mode of action of Entomopathogenic nematodes.** The juvenile stage releases cells of their symbiotic bacteria from their intestines into the hemocoel. The bacteria multiply in the insect hemolymph, and the infected host usually dies within 24 to 48 hours. After the death of the host, nematodes continue to feed on the host tissue, mature, and reproduce (Bedding 1982). The progeny nematodes develop through four juvenile stages to the adult. Depending on the available resources, one or more generations may occur within the host cadaver, and a large number of infective juveniles are eventually released into the environment to infect other hosts and continue their life cycle (Bedding 1982). Nematodes enter the body cavity of insects and release their symbiotic bacteria into the host's intestine.

Advantages of EPNs: EPNs offer a double win: safety and effectiveness. These organisms pose no threats to human health, plants, or animals, requiring no protective gear or waiting periods. Moreover, they leave no harmful residues, making them a sustainable choice for healthy crops and clean environments. Their target list reads like a bug buffet: cranberry girdlers, root weevils, webworms, and even wood borers tremble at their approach. However, these tiny heroes have their preferences. Moist soil around roots, protection from harsh UV, and short stints with other pest control methods are necessary for them to thrive, ensuring their targeted pest control doesn't disrupt the natural ecosystem (Ruiu 2018; Singh et al. 2019; Meenatchi and Aditi 2021).

The disadvantages of EPNs: EPNs are highly susceptible to adverse weather conditions. Harsh UV rays and sizzling temperatures can quickly deactivate these tiny heroes, making their effectiveness dependent on fickle environmental conditions. Compared to their chemical counterparts, EPNs come with a hefty price tag. This, along with their short shelf life, makes them a less financially enticing option for some applications. EPNs can't reach aerial pests, limiting their range of action. Additionally, relying solely on EPNs can disrupt the natural predator-prey balance, requiring repeated applications to maintain pest control. Furthermore, EPNs thrive in moisture. Pre- and post-irrigation, along with application during cooler hours, are essential for their success (Meenatchi and Aditi 2021).

#### 5.6 Protozoan biopesticides

Protozoans, also known as microsporidians, are intracellular parasites that can only survive by living inside other cells. They are found almost everywhere and can attack certain insects, like lepidopteran and orthopteran, making them useful in IPMs. Examples include *Nosema* sp. and *Vairimorpha* sp. However, despite their pest-specific nature and slow-acting properties, the use of protozoa as biopesticides is not as effective as other organisms, such as bacteria, viruses, and fungi. They can cause chronic and debilitating effects on their targets, but their success rate is lower compared to other biopesticides (Meenatchi and Aditi 2021). Protozoans have a specific mode of action. Microsporidia infects the European corn borer, *Ostrinia nubilalis*, by being eaten by insects. The spores germinate in the midgut region, and the sporoplasm is then injected into the midgut cells. The spores then spread to different tissues and organs, multiply, and cause tissue breakdown and *septicaemia* (Senthil-Nathan 2015).

#### 6. Natural enemies of pests as biopesticides

Two powerful allies in this environmentally safe pest control approach are parasitoids and predators.

**Parasitoids**: These insects specialize in laying their eggs on or inside other insects. Their young hatch and feast on the host's body from the inside out, eventually leading to the host's demise. Examples include wasps that parasitize caterpillars and flies that target beetle larvae.

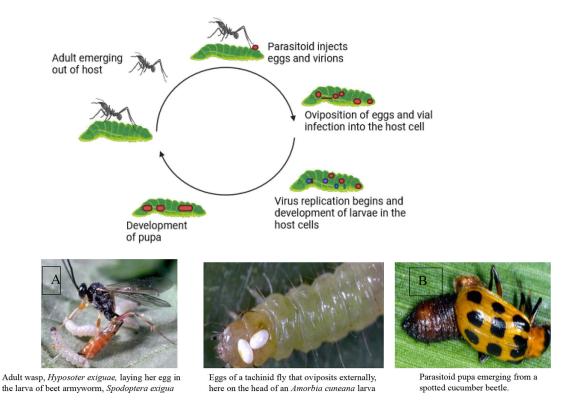
Parasitoids have a shorter life cycle than predators and can multiply at a faster rate. Therefore, they can be more effective in controlling insects. However, their presence may not be noticeable unless you examine samples of the insects to see if any adult parasitoids have emerged (Yelitza et al. 2020).

**Predators**: These insects are fierce hunters who actively seek out and devour their prey. They're often larger than their victims and use their speed, agility, and sharp senses to track them down. Ladybugs munch on aphids, dragonflies swoop on mosquitoes, and ground beetles patrol the soil for tasty treats. Introducing these natural predators into your environment creates a pest-fighting force on patrol. Figure 6 illustrates the mode of action of natural enemies in pest control, specifically focusing on A) the mode of action of parasitoids and B) natural predators, including lady beetles (*Coccinellidae*), wasps, and fungus gnats.

Meenatchi and Aditi (2021) and Gautam (2008) have summarized several promising biopesticides that are effective against various crop pests. These specialized assassins are laser-guided for specific pests, efficiently hunting them even at low levels. They curb reproduction, keep infestations below damage thresholds, and play well with other pest-management tactics. Additionally, they help disrupt pest feeding and lessen the damage to your plants. They are environmentally safe pest control solutions (Gautam 2008).

# 7. Biopesticide formulations

Biopesticide formulations, like conventional pesticide formulations, involve preparing a product from an active ingredient (biologically active metabolite or microbe) by adding various functional (active) and inert substances. This creates a product suitable for application to the target weed, pathogen, or insect using existing equipment. The active ingredient is combined with a carrier material and various additives to improve the biopesticide's lifespan and efficacy (Grewal and Peters 2005). Several factors significantly influence the economic viability of biopesticide products. These include the impact on the target pest, market size, pest spectrum, field performance consistency, production costs,



**Figure 6. Natural enemies' mode of action.** A) Mode of action of parasitoids, and B) Natural predators (the insects are Lady beetles, (Coccinellidae), wasps, and fungus gnats, respectively).

and technological challenges (fermentation, formulation, and delivery systems), all of which pose potential hurdles to commercialization. Optimizing product formulations can significantly improve field performance consistency. This is crucial for the successful adoption and economic viability of many biopesticides (Glass 1993). However, slow progress in research on formulation and delivery systems remains a major bottleneck for biopesticide development (Lumsden et al. 1995).

The formulation is arguably the most crucial step in the development of biopesticides and is the cornerstone of product development (Leggett et al. 2016). Ideally, a user-friendly formulation must meet several important criteria, including preserving and expressing the pesticidal properties of the microorganism, extending the shelf life to at least six months and preferably up to two years under ambient conditions, and enabling application using existing equipment (Pusey 1994). Moreover, enhanced formulations can improve the efficacy of biopesticides by boosting their dispersion, attachment, and persistence at the target site (Droby et al. 2009). Stickers improve adherence to foliage, enhancing persistence (Schisler et al. 2004). Formulation stabilizes the biopesticide throughout production, distribution, storage, handling, and application to ensure persistence and activity at the target site. Cognizance must be given to ensure that the co-formulants used in biopesticide formulations are not chemical pesticide derivatives, are environmentally safe, and do not impact human health.

#### 7.1 Biopesticide formulation requirements and challenges

The effectiveness of a biopesticide hinges on the biological activity of its active microorganism or metabolite. In its raw state, this active ingredient requires formulation to enable its practical handling, storage, and application (Harald 2019). A crucial distinction between conventional (chemical) pesticide formulation and biopesticide formulation lies in the biopesticide's living nature. Biopesticides are sensitive to storage conditions and environmental factors, necessitating careful formulation to preserve their biological viability (Harald 2019). Formulating biopesticides faces several challenges, including ensuring market viability, ease of production and application, product stability during storage and transport, and long-term viability and efficacy of the biopesticide (Lumsden et al. 1995). Contributing factors to their limited commercial success include production difficulties, sensitivity to environmental stressors, and lack of appropriate formulation (Powell and Jutsum 1993). Another challenge is the slime layer produced by gram-positive bacteria like *Bacillus spp.* during endospore formation. This slime can clog sprayer filters and nozzles, hindering application. While

removing the slime through methods like centrifugation is possible, it may also remove valuable components contributing to the biopesticide's effect (Boyette et al. 1996).

The goal of formulation is to improve product stability, bioactivity, delivery, and integration into pest management systems. Successful formulations also prioritize user convenience, compatibility with equipment and practices, and effectiveness at practical rates. Environmental factors like temperature, moisture, and UV exposure significantly influence efficacy, particularly for foliar-applied biopesticides (Alan 1988). Soil properties, moisture, temperature regimes, and microbial competition impact soil-applied biopesticides. All these factors need careful consideration during formulation development.

# 7.2 Types of biopesticide formulations

#### 7.2.1 Formulation of bacteria biopesticide

Several recent studies highlight the growing use of bacteria-based biofungicide formulations as a safe and effective complement to conventional chemical pesticides (Cook 1993; Lumsden et al. 1995; Elad 1995). *Bacillus*-based products are already boosting crop yields in various countries, from China to the United States (Zhang et al. 1996). These wettable powders, compatible with existing fungicides, target diseases caused by *Rhizoctonia* and *Fusarium* in a range of crops like cotton, legumes, vegetables, and ornamentals.

Mass production of these biofungicide bacteria formulations typically involves deep-tank liquid fermentation. However, in some cases, methods like semisolid or solid-state fermentation might be more suitable. Key factors influencing both bacterial growth and their beneficial metabolite production include the medium's nutrient composition and optimal growth conditions (Paau 1988). Cost-effectiveness and readily available components are prioritized. The final formulation, usually involving additional ingredients after fermentation, comes in various forms, like solids, slurries, powders, or granules. Long-term viability during transport and storage (at least 4 months) is crucial (Paau 1988). The formulation significantly impacts the biofungicide's performance by enhancing bacterial survival after application. This involves creating a protective environment that maximizes the bacteria's potential for successful colonization and disease control. Choosing the right formulation can help ensure consistent results in the field.

The products are designed as wettable powders and can be used with various seed treatment fungicides (Table 1). Agrobactenum radiobacter containing strain K1026, a modified version of K84, was registered as a biopesticide in several countries, including Australia, the United States, Turkey, and New Zealand and is used explicitly as a microbial control agent to combat crown gall disease in plants (Kerr and Bullard 2020), by competing with the pathogenic strain of Agrobacterium tumefaciens, due to its ability to produce an antibiotic called agrocin 84, while being genetically modified to prevent the transfer of this resistance to other bacteria; making it a safer and more targeted microbial agent (Penyalver et al. 2000). K1026 showed strong efficacy against both agrocin 84-sensitive and resistant strains of A. tumefaciens (Vicedo et al. 1993). Streptomyces griseoviridis (Mycostop), formulated as a wettable powder, tackles damping-off and root rot in vegetables and ornamentals caused by Fusarium, Phomopsis, and Pythium. It can be applied dry to seeds or as a liquid drench and is compatible with chemical pesticides (Muir 2023). Three Burkholderia assian strains (Blue Circle, Deny, and Intercept) come in liquid and fight a wider range of enemies. These bacterial warriors target fungi like Fusarium, Phytophthora, and Pythium alongside nematodes, including Globodera rostochensis, Heterodera glycines, and Hoplolaimus columbus (Kumari et al. 2022).

Most commercial bioinsecticide formulations today harness the power of *Bacillus thuringiensis* (*Bt*), a friendly, spore-forming bacterium. This gram-positive warrior wields a powerful weapon: delta-endotoxin proteins that wreak havoc on the mid-guts of susceptible insect larvae (Cannon 1993) (Table 1). *Bt* proteins are highly specific, only affecting a few species in the Lepidoptera (butterflies and moths), Coleoptera (beetles), and Diptera (flies) families. *Bt* biopesticides come in an array of formulations, from concentrated liquids to handy dusts, catering to different application needs.

### 7.2.2 Formulation of fungal biopesticides

Fungicide formulations primarily include *filamentous fungi* (e.g. *Gliocladium virens* and *Trichoderma harzzanum*), yeast-like fungi (e.g. *Pseudozymajlocculosa* (Bélanger 1997) and *Tllletiopsis pallescens* (Urquhart and Punja 1997). These biofungicides are used to control root-infecting pathogens (e.g. *Pythlum, Rhizoctonia*), and foliar fungal pathogens (e.g. powdery mildew (Bélanger 1997; Urquhart and Punja 1997; Punja 1997) and *Botrytzs* (Elad 1995). Environmental conditions, such as temperature and moisture, can impact the growth and survival of fungal biofungicides. Various formulations have been developed for the application of spore inocula, including granules, pellets, dust, and wettable

powders. These formulations can be applied directly or suspended in water or oil, as is the case with *Cylindobasidium leave* (Muir 2023).

Granular formulations not only protect against desiccation but also offer a food base for the fungus. Meanwhile, powders can be easily sprayed and can provide coverage for large areas. An example of this is the large-scale aerial application of sawdust covered in *Colletotrichum acutatum* spores for the control of *Hakea sericea* (Jacobson and Azevedo 2023). It is also possible to treat seeds with liquids or dust for the application of these biopesticides. Moreover, spore formulations in inverted emulsions have been tested for yeasts, such as *Tilletiopsis* (Urquhart and Punja 1997).

The use of alginate prill has been successfully developed to create a granular formulation of *Gliocladium virens* (Soil Gard), which helps control root-infecting fungi in potting media (Lumsden et al. 1995). Similarly, *Trichoderma* powder or dust formulations with pyrophyllite clay (Pyrax) have also been proven to be effective (Jayaraj and Radhakrishnan 2006). To produce the necessary biomass, appropriate nutrient substrates are used in large-scale deep tank fermenters. The resulting product can either be used wet or dried before formulation (Lumsden et al. 1995). Most of the factors that affect product development for fungi are like those for bacteria.

Entomopathogenic fungi are a fascinating group of soil-dwelling microorganisms that play a crucial role in biological control. These fungi are capable of infecting and killing insects through cuticle penetration. They don't need to be ingested by the insect hosts; instead, they directly invade through the insect's outer covering. This unique mode of action allows them to control a wide range of insect pests, including sucking insects. However, they cannot effectively combat viruses and bacteria similarly (Shishupala 2022). Entomopathogenic fungi are a type of fungi that can target and control pests without harming beneficial organisms such as pollinators. They are an environmentally safe alternative to traditional chemical pesticides for managing agricultural pests. Some of the most well-researched fungal species with bioinsecticidal properties include Akanthomyces muscarius, Beauveria bassiana, Cordyceps fumosorosea, Purpureocillium lilacinum, Verticillium, and the Metarhizium anisopliae species complex (Shishupala 2022). A recent study by Luo et al. (2022) isolated a new entomopathogenic fungus from an adult cadaver of D. citri. The fungus was identified as Cordyceps fumosorosea based on its morphology and ITS sequence analysis. The researchers named this specific isolated C. fumosorosea SCAU-CFDC01. The study assessed the pathogenicity of the strain against D. citri nymphs and adults in both laboratory and greenhouse settings.

Various fungi can be used to control different types of insects. *Verticillium lecanil* can control aphids, *Beauveria bassiana* can control whiteflies, locusts, and beetles, *Metarhizium anisopliae* and *M. flavoviride* can control mosquitos and locusts respectively, while *Lagenidium gigantem* can control mosquito larvae (Lacey and Goettel 1995; Bateman 1997; FAO 2017; Savita 2019). These fungi can be applied to insects in different ways, such as wettable powders, emulsions, specks of dust, baits, or traps, or even added to the soil (Auld 1992; Feng et al. 1994; Goettel et al. 1995; Inglis et al. 1996) or used in indoor residual spraying. It is important to formulate these fungi to protect against environmental conditions such as moisture, temperature, UV damage, and desiccation. For example, *B. bassiana* conidia on foliage can be damaged by UV-B radiation, which is part of the sunlight spectrum (Caudwell and Gatehouse 1996a; Daoust and Pereirn 1986). In field conditions, these entomopathogens can be applied at ultralow volumes (ULV) to increase their effectiveness and protect against UV damage (McGUIRE and Shasha 1992; Moore et al. 1993; Inglis et al. 1996). To improve survival and shelf-life, sunlight blockers (such as clay) or UV-B absorbing compounds (such as Tinopal) can be added to inoculum formulations or starch encapsulation (Pereira and Roberts 1991; McGuire and Shasha 1992; Caudwell and Gatehouse 1996b).

#### 7.2.3 Formulation of mycoherbicides

Environmental factors, such as temperature and moisture, can greatly impact the effectiveness of Mycoherbicides. The moisture needed for the disease to develop is often determined by the amount of dew that occurs. "DeVine" is the first registered Mycoherbicide, which is a liquid formulation consisting of chlamydospores of *Phytophthora palmivora* (Burnett et al. 1974; TeBeest and Templeton 1985). This product has a limited shelf life of only 6 weeks when refrigerated due to its instability. Another Mycoherbicide called "Collego" (*Colletotrichum gloeosporioides f.sp. aeschynomene*) is formulated as dried spores and is available in a wettable powder (Boyette 1994). The commercialized formulation of *Colletotrichum acutatum* is formulated as dried spores that are mixed with water when ready to apply, and Stumpout is *Cylindrobasidium laeve* formulated as dried spores in an iron carrier that is mixed with standard cooking oil when ready to apply (Muir 2023).

There are several ways to improve the effectiveness of mycoherbicides, which are used to control weeds (Boyette et al. 1996). One way is to use adjuvants and other additives that can help spores grow, make the pathogen more stable, change

the environment, or make it possible to use bioherbicides on more plants (Boyette 1994). For example, a fungus called *Colletotrichum truncatum* can kill a weed called hemp sesbania, but it needs a lot of moisture to work (Boyette et al. 1993). Adding unrefined corn oil to the biocontrol agent can help it work better, so less moisture is needed. This makes it easier to use the bioherbicide and reduces the amount of spray needed (Boyette 1994).

Surfactants are ingredients used in formulations to help wet plants by reducing surface tension and improving the dispersal of fungal spores in spray droplets. Some surfactants used include Tween 20 with *Fusarlum lateritum*, nonoxynol with *Alternaria macrospora* and *A. assia*, and *sorbitol* with *Colletotrichum coccodes* (Boyette 1994). However, evaluating the inhibitory or stimulatory effects on spore-germination and infection is important before selecting appropriate surfactants and ensuring that the surfactants do not contain pesticide derivatives. Water-in-oil invert emulsions have been used with foliar fungal biopesticides to provide a favorable environment for germination and infection (Connick et al. 1990; Daigle et al. 1990; Auld and Morin 1993; Boyette et al. 1996). Using inverted emulsions (Boyette et al. 1993) has been found significantly to improve the efficacy of *C. truncatum* and *Alternaria assia*. However, inverted emulsions are very viscous and may demonstrate phytotoxicity in some target plants (Walker and Boyette 1985; Amsellem et al. 1990; Goettel et al. 1995). Connick et al. (1990) developed an inverted emulsion with improved water-retention properties that was less viscous. Additionally, vegetable oils can enhance the efficacy of mycoherbicides, such as *Colletotrichum orbiculare*, for the control of spiny cocklebur (Auld 1992) and *Collototrichum acutatum* for the control of *Hakea sericea* and *H. gibbose*. No phytotoxicity and improvements in the spread of the inverted emulsion were observed.

Solid-based formulations of mycoherbicides have been developed for weeds that are infected at or below the soil surface, which is more suitable for preemergence mycoherbicides (Daigle and Connick 1990; Boyette et al. 1996). These formulations can provide a food base for the pathogen, act as a buffer in extreme environmental conditions, and retain inoculum that may not be easily washed away. One such formulation, known as "PESTA," uses a wheat-gluten matrix consisting of liquid inoculum, semolina wheat flour, and kaolin. This formulation includes fungal agents like *C. truncatum*, *A. crassa*, and *Fusarium lateritium* and can be applied as preemergent and soil-incorporated treatments (Boyette et al. 1984). The shelf life of the product can be improved by manipulating the water activity and sucrose content. Other solid substrates used to formulate mycoherbicides include bran, wheat kernels, cornmeal/sand, vermiculite (Auld and Morin 1993; Boyette et al. 1996), and sawdust (Muir 2023). For example, cornmeal/cornmeal sand was used to formulate mycelium, micro- and macroconidia, and chlamydospores of *Fusarium solani f.sp. cucurbitae* for the control of Texas gourd. This pre-emergent granular formulation provided 96% control of the weed (Boyette et al. 1984).

# 7.2.4 Formulation of viruses

Baculoviruses have been studied as a potential solution for controlling insect pests that belong to the Lepidoptera, Hymenoptera, and Coleoptera families (Gory and Bishop 1995). These viruses have several advantages, such as high specificity, not harming beneficial insects, and persisting in the environment, which makes long-term control of insect pests possible without damaging the environment. Nuclear polyhedrosis viruses (NPVs) and granulosis viruses (GVs) are examples of baculoviruses. However, there are some limitations to using these biopesticides, such as their slow speed of biological activity, low stability under UV light, and difficulties in production (Powell and Jutsum 1993).

The stability of the baculoviruses is often a function of viability, but it is not a significant problem for small-scale field trials as the viruses can be collected from macerated larvae, mixed with water, and stored for short periods through refrigeration. However, these systems are not suitable for large-scale production and application (Gory and Bishop 1995). The formulation of these viruses is an important aspect of product development, but researchers have not given it as much attention as bacteria and fungi.

Baculoviruses are usually available in the form of concentrated wettable powders. The biocontrol product called "Elcar" for corn earworm (*Helicoverpa zea*) NPV can be spray-dried or air-dried after being diluted with an inert carrier. The gypsy moth (*Lymantria dispar* L.) NPV is freeze-dried either with carbohydrates or by acetone precipitation (Gory and Bishop 1995). The virus can be inactivated by UV radiation, especially wavelengths of 290-320 nm. UV protectants like reflections or absorbers can be added to formulations to stabilize baculoviruses. Several dyes, such as lissamine green, acridine yellow, alkali blue, and mercurochrome, have been used as UV protectants, especially to absorb UV-A irradiation (Shapiro 1995). Optical brighteners (fluorescent brighteners), which are commonly used in soaps, detergents, and fabric softeners, also absorb UV light. They have been shown to significantly reduce the photodegradation of NPVs and enhance viral activity (Shapiro 1992; Dougherty et al. 1995; Shapiro 1995). Care must be taken to ensure that these optical brighteners do not adversely affect the environment due to residues and residuality.

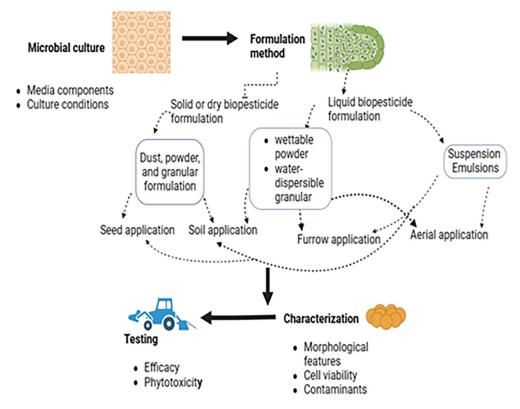


Figure 7. General workflow of the procedures involved in the development of microbial biocontrol formulation. Straight line arrows indicate process (steps), and broken lines indicate categories or classifications.

There are two types of biopesticide formulations: liquid and dry, which differ based on their physical state (Figure 7). Dry formulations (dust powders, granules, seed dressers, wettable powders, and wettable) dispersible powder. They are produced using various technologies, including spray drying, freeze drying, or air drying, with or without a fluidized bed (Li 2002). They typically require binders, dispersants, and wetting agents for optimal efficacy and application. Liquid formulations can be water-based, oil-based, polymer-based, or combinations. Water-based formulations, like suspension concentrates, suspo-emulsions, capsule suspensions, and ultra-low volume liquids, require additional inert ingredients for stability and performance (Boyetchko et al. 1999). These include stabilizers, stickers, surfactants, coloring agents, antifreeze compounds, and additional nutrients. Care must be taken to ensure that these inert ingredients do not adversely affect the environment due to residues and residuality and do not contain pesticide derivatives.

### 8. Advances in biopesticide and delivery technologies enhance the efficacy of biological products

Advances in biopesticide application techniques have been driven by the need for more efficient and targeted pest control methods while minimizing environmental impacts and fate. These advancements aim to improve the delivery, efficacy, and performance of biopesticides. Several improved spray technologies and precision targeting methods have been developed, including controlled droplet application, variable rate technology, intelligent spray systems, air-assisted sprayers, robotics, and automation technologies, automated targeting systems (Wang et al. 2019), thermal fogging, and aerial application. The improved spray technologies play an important role in delivering droplets of consistent size and velocity, improving spray deposition, reducing non-target drift, optimizing chemical pesticide usage, reducing costs, and minimizing environmental impacts and fate. Additionally, some of these technologies utilize sensors and algorithms to detect and target pests in real-time, enabling precise application of biopesticides only where needed. Other techniques improve the effectiveness of biopesticides by reaching target areas that are otherwise difficult to treat, and some spray techniques improve coverage and reduce the amount of biopesticide needed.

## 8.1 Seed treatments and soil applications

Seed treatments and soil applications involve various methods for applying biopesticides to protect seeds and enhance crop establishment. Seed coatings and treatments apply biopesticides onto the seed surface, providing early protection and improving crop establishment (Khan et al. 2020). Incorporating biopesticides into soil amendments can improve soil health, control soil-borne pests, and enhance plant growth, offering sustainable pest management and environmental

benefits. Other methods include soil drenching, which involves applying biopesticides as a liquid solution directly to the soil around the plant base. This technique targets pests in the soil, such as nematodes and soil-borne pathogens. Biopesticide-based soil fumigation utilizes the controlled release of compounds to control pests and diseases in the soil, providing an alternative to chemical fumigation but with reduced environmental impacts. Rhizosphere application focuses on applying biopesticides directly to the root zone of plants. This technique targets pests that interact with plant roots, such as soil-dwelling insects and nematodes. These seed treatment and soil application methods offer effective pest management while minimizing environmental impacts and fate.

## 8.2 Factors affecting the efficacy of biopesticides

Various challenges and limitations, including environmental conditions such as temperature, humidity, and sunlight, can influence by the efficacy of biopesticides during pest control. Some biopesticides may require specific temperatures or moisture levels for optimal performance. Additionally, biopesticides often exhibit narrow target specificity that is effective against specific pests but not others. This limitation reduces their broad-spectrum efficacy, and it may be necessary to use multiple biopesticides to target different pests (Gurr et al. 2017). Furthermore, the formulation and storage conditions of biopesticides can affect their stability and shelf life. Factors such as pH, temperature, and exposure to light can degrade the effectiveness of biopesticides over time (Riga et al. 2017).

It's important to note that these challenges and limitations can vary depending on the specific biopesticide and pest control scenario. By considering and addressing these factors, it is possible to optimize the efficacy of biopesticides in pest control practices. The efficacy of biopesticides can be influenced by the application methods used. Factors such as spray volume, droplet size, and coverage play a crucial role in the distribution and effectiveness of the biopesticide on the target pests (Chandler et al. 2011). Ensuring that the application method is properly calibrated is important to achieve adequate coverage and contact with the pests (International Symposium on Food Safety and Control).

Biopesticides may have a shorter persistence compared to chemical pesticides, which may require more frequent applications. Their residual activity on plants or the environment can be limited, necessitating careful timing and repeated treatments. Proper timing of application is essential to target pests during vulnerable stages of their life cycle thus, it is critical to fully understand the pest's lifecycle before implementing a biopesticide programme. In addition to the challenges mentioned earlier, several other factors can limit the efficiency of biopesticides. These include regulatory approval, which can be a complex and time-consuming process. Limited availability and accessibility of biopesticides can pose challenges for farmers (Biondi et al. 2021). And their compatibility with integrated pest management can be a complex issue. Lack of standardization and quality control protocols may affect the consistent performance of biopesticides. Furthermore, biopesticides can be more expensive compared to chemical pesticides, affecting their

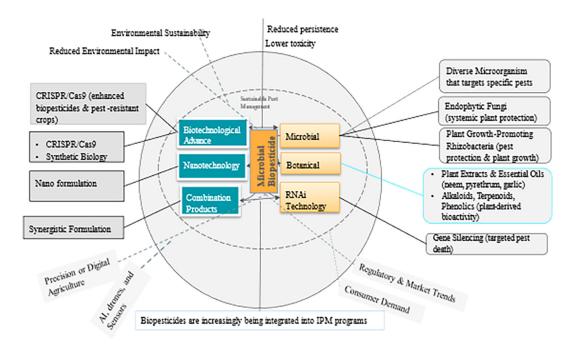


Figure 8. Summary of the emerging trends in biopesticide development.

cost-effectiveness. The efficacy of biopesticides can vary depending on factors such as pest population density, pest development stage, and resistance mechanisms. Pests can develop resistance to specific biopesticides over time, which can reduce their effectiveness. Therefore, it is important to monitor pest populations and implement IPM strategies that incorporate the use of different biopesticides and other pest control measures to mitigate the development of resistance.

#### 9. Future perspectives and research directions

Emerging trends in biopesticide development are driven by the need for sustainable agricultural practices and the increasing demand for ecologically safe pest control methods. Some of the key trends are summarized in Figure 8. Future perspectives and research directions in the field of microbial biopesticides including key research directions include Genetic engineering to improve microbial agents, synergistic interactions with other pest control methods, advanced formulation and delivery techniques, resistance management strategies, environmental impact assessments, integration into sustainable agriculture, and public awareness and adoption are essential for advancing their development and application. The future of microbial biopesticides looks promising, with ongoing research focusing on enhancing their efficacy, specificity, and environmental safety (Karlsson et al. 2020). Microbiopesticides are gaining significant attention in modern agriculture due to their environmentally safe nature and reduced impact on human health. Researchers and companies are actively involved in developing new biopesticide formulations and strategies to enhance their efficacy. Some emerging trends in biopesticide development include microbial biopesticides, plant extracts and essential oils, RNA-based biopesticides, nanotechnology in biopesticides, and the combination of different biopesticides for improved pest control (Karlsson et al. 2020). As the demand for sustainable and ecologically safe pest management strategies has increased, biopesticides have made tremendous strides in recent years. One of the most significant discoveries is the use of microbial biopesticides, such as bacteria, fungi, and viruses, with qualities enhanced through genetic modification or natural selection. By combining CRISPR/Cas9 and RNAi, researchers can develop microbial biopesticides with improved specificity, efficacy, and environmental safety. These technologies offer exciting possibilities for sustainable agriculture. Figure 8 provides a comprehensive summary of the emerging trends in biopesticide development and the integration of biopesticides with other integrated pest management (IPM) practices and pest control methods.

These trends indicate a growing focus on developing innovative, effective, and sustainable pest control solutions that align with environmental conservation and agricultural productivity goals.

#### 10. Conclusion

The adoption of biopesticides is an increasingly popular trend in agriculture due to various environmental, health, and economic benefits. Biopesticides consist of naturally occurring substances, including insects, nematodes, microorganisms, plants, and semiochemicals, along with their by-products, offering a sustainable alternative to traditional chemical pesticides. Despite the benefits, several challenges can hinder the widespread adoption of biopesticides. Novel microbial strain creation with improved features, such as greater efficacy and larger target spectra, should be prioritized in future research and the use of biopesticides. To maximize their efficiency and reduce the possibility of resistance, biopesticides' modes of action must be understood. Along with other tactics, using biopesticides in complete pest management programs can improve overall pest control. The stability, adhesion, and targeted distribution of biopesticides will be improved through improvements in formulation and delivery techniques. Risk assessment and regulatory frameworks must be created to guarantee the safety and effectiveness of biopesticides. To encourage knowledge and acceptance of biopesticides, efforts should be made to educate and raise public awareness. Collaboration and knowledge sharing among researchers, stakeholders, and regulatory agencies will facilitate progress in the field. By focusing on these recommendations, biopesticides can contribute to sustainable and environmentally safe pest management practices in agriculture.

## **Authors contribution**

Conceptualization: K.T.M.; Original draft preparation: K.T.M. and L.M.; Writing: K.T.M., D. M.; and D.M.; Editing, Reviewing, and formatting: D.N.: and Reviewed drafts of the paper: J.G.; Language editing and preparation of tables and/or figures: K.T.M. and D.N.; Final approval of the review to be published.

All authors have read and agreed to the published version of the manuscript.

## Data availability statement

No data are associated with this article.

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#### References

Abd-Alla AMM, Meki IK, Demirbas-Uzel G: Insect Viruses as Biocontrol Agents: Challenges and Opportunities. El-Wakeil N, Saleh M, Abuhashim M, editors. Cottage Industry of Biocontrol Agents and Their Applications. Cham: Springer; 2020; pp. 277–295

Adesemoye AO, Kloepper JW: Plant-microbes interactions in enhanced fertilizer-use efficiency. Appl. Microbiol. Biotechnol. 2009; 85: 1-12. PubMed Abstract | Publisher Full Text

Akutse KS, Subramanian S, Maniania N, et al.: Entomopathogenic fungus isolates for adult *Tuta absoluta* (Lepidoptera: Gelechiidae) management and their compatibility with Tuta pheromone. J. Appl. Entomol. 2020; 144: 777-787.

Alan SP: Formulations useful in applying beneficial microorganisms to seeds. Trends Biotechnol. 1988; 6: 276-2

**Publisher Full Text** 

Alavanja MCR: Introduction: Pesticides Use and Exposure, Extensive Worldwide. J. Rev. Environ. Heal. 2009; 24: 303–309.

PubMed Abstract | Publisher Full Text | Free Full Text

Alsaedi G. Ashouri A. Talaei-Hassanloui R: Evaluation of Bacillus thuringiensis to control Tuta absoluta (Meyrick) (Lepidoptera: Gelechiidae) under laboratory conditions. Agric. Sci. 2017; 8: 559-591

Amruta N. Kumar MKP. Kandikattu HK. et al.: Bio-priming of rice seeds with novel bacterial strains, for management of seedborne Magnaporthe oryzae L. Plant Physiol. Rep. 2019; 24: 507-520. **Publisher Full Text** 

Amsellem Z, Sharon A, Gressel J, et al.: Complete abolition of high inoculum threshold of two mycoherbicides (Alternaria cassiae and A. crassa) when applied in invert emulsion. Phytopathology. 1990; 80: 925-929.

**Publisher Full Text** 

Ara I, Bukhari NA, Aref N, et al.: Antiviral activities of streptomycetes against tobacco mosaic virus (TMV) in Datura plant: evaluation of different organic compounds in their metabolites. Afr. J. Biotechnol. 2014; **11**: 2130-2138.

**Publisher Full Text** 

Arora NK, Mishra J, Dutta V: Biopesticides in India: technology and sustainability linkages. 3 Biotech. 2020; 10: 210–212. PubMed Abstract | Publisher Full Text | Free Full Text

Auld BA, Morin L: Constraints in the development of bioherbicides. Weed Technol. 1993; 9: 638-652.

**Publisher Full Text** 

Auld BA: Mass production, formulation, and application of fungi as biocontrol agents, m Biological Control of Locusts and Grasshoppers. Wallingford, UK: CAB International; 1992.

Baldiviezo LV, Pedrini N, Santana M, et al.: Isolation of Beauveria bassiana from the chagas disease vector Triatoma infestans in the gran chaco region of argentina: assessment of gene expression during host-

pathogen interaction. J. Fungi. 2020; 6: 219. PubMed Abstract | Publisher Full Text | Free Full Text

Bateman R: The development of a mycoinsecticide for the control of locusts and grasshoppers. *Outlook Agric.* 1997; **26**: 13–18.

Bedding MA: Penetration of insect cuticle by infective juveniles of Heterorhabditis spp. (Heterorhabditidae: Nematoda). Nematologica. 1982; 28: 354–359.

**Publisher Full Text** 

Bélanger RR: Challenges and prospects for integrated control of powdery mildews in the greenhouse. Can. J. Plant Pathol. Rev. Can. Phytopathol. 1997; 19: 310-314.

**Publisher Full Text** 

Bharti V, Ibrahim S: Biopesticides: Production, Formulation and Application Systems. Int. J. Curr. Microbiol. App. Sci. 2020; 9: 3931-3946.

Bhat AH, Chaubey AT: **Global distribution of entomopathogenic nematodes**, **Steinernema** and **Heterorhabditis**. *Egypt J. Biol. Pest. Control.* 

Biondi A, Guedes RNC, Wan FH, et al.: Pest Management: From Chemical Control to Sustainable Integrated Pest Management. Sustainable Pest Management. Academic Press; 2021.

Blanco CA, Andow DA, Abel CA, et al.: Bacillus thuringiensis Cry1Ac resistance frequency in tobacco budworm (Lepidoptera: Noctuidae). J. Econ. Entomol. 2009; 102: 381–387.
PubMed Abstract | Publisher Full Text

Boyetchko S, Pedersen E, Punja RM: Formulations of biopesticides. Hall FR, Menn JJ, editors. *Biopesticides: use and delivery. Methods in biotechnology.* Vol 5. Totowa: Humana Press; 1999; pp. 487–508

Boyetchko S, Eric Pedersen ZP, Reddy M: Formulations of Biopesticides. Methods in Biotechnology, vol. 5: Biopesticides: Use and Delivery. Hall FR, Menn JJ, editors. Totowa, NJ: Humana Press; 2020.

Boyette CD, Qmmby PC, Caesar AJ, et al.: Adjuvants, formulations, and spraying systems for improvement of mycoherbrcides. Weed Technol. 1996; **10**: 637-644.

**Publisher Full Text** 

Boyette CD, Templeton GE, Oliver LR: Texas gourd (Cucurbita texana) control with Fusarium solani f. sp. cucurbitae. Weed Sci. 1984; 32: 649-655

**Publisher Full Text** 

Boyette CD, Quimby PC, Bryson CT, et al.: Biological control of hemp sesbania (Sesbania exaltata) under field conditions with Colletotrichum truncatum formulated in an invert emulsion. Weed Sci. 1993; **41**: 497-500. **Publisher Full Text** 

Boyette CD: Unrefined corn oil improves the mycoherbicide activity of *Colletotrichum truncatum* for hemp sesbania (*Sesbanza exaltata*). Control Weed Technol. 1994; 8: 526-529.

Burnett HC, Tucker DPH, Ridings WH: **Phytophthora root and stem rot of milkweed vine**. *Plant Dis. Rep.* 1974; **58**: 355–357.

Cannon RJ: Prospects and progress for Bacillus thuringiensis-based pesticides. Pestic. Sci. 1993; 37: 331-335

**Publisher Full Text** 

Caudwell RW, Gatehouse AG: Laboratory and field trials of bait formulations of the fungal pathogen, Metarhizium flavoviride, against a tropical grasshopper and locust. Biocontrol Sci. Tech. 1996a; 6:

Caudwell RW, Gatehouse AG: Formulation of grasshopper and locust entomopathogens in baits using starch extrusion technology. Crop Prot. 1996b; 15: 33-37.

**Publisher Full Text** 

Chandler D, Bailey AS, Tatchell GM, et al.: The development, regulation and use of biopesticides for integrated pest management. Philos T R Soc. B Biol. Sci. 2011; 366: 1987-1998.

Chandler D, Davidson G, Grant WP, et al.: Microbial biopesticides for integrated crop management: an assessment of environmental and regulatory sustainability. *Trends Food Sci. Technol.* 2008; **19**: 275–283.

Chang JH, Choi JY, Jin BR, et al.: An improved baculovirus insecticide producing occlusion bodies that contain Bacillus thuringiensis insect toxin. J. Invertebr. Pathol. 2003; 84: 30-37.

PubMed Abstract | Publisher Full Text

Chattopadhyay A, Bhatnagar NBBR: **Bacterial insecticidal toxins.** *Crit. Rev. Microbiol.* 2004; **30**: 33–54.

**Publisher Full Text** 

Clemson HGIC: Organic pesticides and biopesticides, Clemson extension, home and garden information center. 2007; 23

Connick Jr, Lewis JA, Quimby PC: Formulation of biocontrol agents for use in plant pathology. New directions in biological control, alternatives for suppressing agricultural pests and diseases. Proceedings of a UCLA colloquium held at Frisco, Colorado, January 20-27, 1989. Wiley-Liss. 1990.

Cook RJ: Making greater use of introduced microorganisms for biological control of plant pathogens. Annu. Rev. Phytopathol. 1993; 31:

PubMed Abstract | Publisher Full Text

Copping, Leonard G, Julius JM: Biopesticides: a review of their action, applications, and efficacy. Pest Manag. Sci. 2000; 56: 651-676

Cory JS: Assessing the risks of releasing genetically modified virus insecticides: progress to date. Crop Prot. 2000; 19: 779-785. **Publisher Full Text** 

Daigle DJ, Connick WJ, Quimby PC, et al.: Invert emulsions carrier and water source for the mycoherbicide Alternaria castle. Weed Technol. 1990; 4(327-33): 1.

Daigle DJ, Connick WJ: Formulation and application technology for microbial weed control. Microbes and Microbial Products as Herbic (Hoagland, R. E, ed.), ACS Symposium series 439, American Chemical Society, Washington, DC. 1990.

Daniel RT: Effect of the Bioherbicide Pseudomonas fluorescens D7 on Downy Brome (Bromus tectorum). Rangel. Ecol. Manag. 2020; 73: 753-755.

**Publisher Full Text** 

Daoust RA, Pereirn RM: Stability of entomopathogenic fungi Beauveria bassiana and Metarhizium anisopliae on beetle-attracting tubers and cowpea foliage in Brazil. Environ. Entomol. 1986; 15: 1237–1243. **Publisher Full Text** 

de Faria MR, Wraight SP: Mycoinsecticides and mycoacaricides: a comprehensive list with worldwide coverage and international classification of formulation types. *Biol. Control.* 2007; **43**: 237–256.

Deravel J, Krier F, Jacques P: **Biopesticides**, a **complementary and alternative approach to the use of agrochemicals**. **A review**. *Biotechnol. Agron. Soc. Environ.* 2014; **18**: 220–232.

Desneux N, Han P, Mansour R, et al.: Integrated pest management of *Tuta absoluta*: practical implementations across different world regions. J. Pest. Sci. 2022; 2004: 1–23.

#### **Publisher Full Text**

Djaenuddin N, Suriani, Muis A: **Effectiveness of Bacillus subtilis TM4 biopesticide formulation as biocontrol agent against maydis leaf blight disease on corn.** International Conference on Sustainable Cereals and Crops Production Systems in the Tropics 23-25 September 2019, Makassar City, Indonesia. p 484 012096. 2020.

Dougherty EM, Guthrte K, Shapuo M: In Vitro effects of fluorescent brightener on the efficacy of occluston body dissolution and polyhedral derived virions. *Bloi Control.* 1995; **5**: 383–388. Publisher Full Text

Dougherty EM, Narang N, Loeb M, et al.: Fluorescent brightener inhibits apoptosis in baculovirus-infected gypsy moth larval midgut cells in vitro. *Biocontrol Sci. Tech.* 2006; **16**: 157–168.

#### **Publisher Full Text**

Droby S, Wisniewski M, Macarisin WC: **Twenty years of postharvest biocontrol research: is it time for a new paradigm?** *Postharvest Biol. Technol.* 2009; **52**: 137–145.

#### **Publisher Full Text**

Elad Y: Practical approaches for biocontrol implementation. Novel approaches to integrated pest management; 1995; 323–338.

EPA: Ingredients Used in Pesticide Products: Pesticides. What Are Biopesticides? 2023.

EPPO: Carpovirusine (CpGV). European and Mediterranean Plant Protection

Essiedu JA, Adepoju FO, Ivantsova MN: **Benefits and limitations in using biopesticides: A review.** In *AIP Conference Proceedings.* AIP Publishing; 2020 Dec 9: Vol. 2313. No. 1.

Essiedu JA, Adepoju FO, Ivantsova MN: **Benefits and limitations in using biopesticides**: **A review**. *Proceedings of the VII International Young Researchers' Conference—Physics, Technology, Innovations (PTI-2020). Ekaterinburg, Russia, pp 080002, 18–22 May.* 2022.

European Environment Agency: How pesticides impact human health and ecosystems in Europe EEA Briefings 06 (2023). 2023.

FAO: Climate-smart agriculture Sustainable Development Goals: In: Mapping interlinkages, synergies, and trade-off s and guidelines for integrated implementation. Rome, Italy: Food and Agriculture Organization of The United Nations; 2019; 12.

FAO: Climate- Smart Agriculture Source Book. Second ed. Rome Italy: Food and Agriculture Organization of The United Nations; 2017.

FAO: The 3 pillars of Climate-Smart Agriculture and the Sustainable Development Goals Synergies & Trade-offs. 2024.

Feng MG, Poprawski TJ, Khachatourians GG: **Production, formulation** and application of the entomopathogenic fungus *Beauveria bassiana* for insect control: current status. *Biocontrol Sci. Tech.* 1994; 4: 3–34.

Feng KC, Liu BL, Tzeng YM: *Verticillium lecanii* spore production in solidstate and liquid-state fermentations. *Bioprocess Eng.* 2000; **23**: 25–29. Publisher Full Text

Ferreira MA: Xenorhabdus and Photorhabdus, bacterial symbionts of the entomopathogenic nematodes Steinernema and Heterorhabditis and their in vitro liquid mass culture: a review. *Afr. Entomol.* 2014; **22**:

#### Publisher Full Text

Gabarty A, Salem HM, Fouda MA, et al.: Pathogencity induced by the entomopathogenic fungi Beauveria bassiana and Metarhizium anisopliae in Agrotisipsilon (Hufn.). J. Radiat. Res. Appl. Sci. 2014; 7: 95–100.

#### **Publisher Full Text**

Gangwar P, Trivedi M, Tiwari RK: **Entomopathogenic Bacteria**. In: Omkar, editors. *Microbial Approaches for Insect Pest Management*. Singapore: Springer; 2021. **Publisher Full Text** 

Gautam RD: Biological pest suppression. Westville Pub. House; 2008.

Germaine KJ, Keogh E, Ryan D, et al.: **Bacterial endophyte-mediated naphthalene phytoprotection and phytoremediation**. *FEMS Microbiol*. *Lett*. 2009; **296**: 226–234.

PubMed Abstract | Publisher Full Text

Ghayur A: A Study of Biopesticides and Biofertilisers in Haryana. India: 2000.

Gill SS, Cowles PP: **Mode of action of** *Bacillus thuringiensis* **endotoxins**. *Annu. Rev. Entomol.* 1992; **37**: 615–634.

Publisher Full Text

Glass DJ: Formulation of microbial herbicides to Improve performance mthe field. Proceedings of 8th EWRS Symposium "Quantitative Approaches to Weed and Herbicide Research and Their Practical Application. Braunschwerg, Germany. 1993; pp. 219–225.

Glick BR: Plant growth-promoting bacteria: mechanisms and applications. *Scientifica (Cairo)*. 2012; 963401.

#### **Publisher Full Text**

Goettel MS, Johnson DL, Inglis GD: **The role of fungi in the control of grasshoppers**. *J. Bot.* 1995; **73**: 71–75. **Publisher Full Text** 

Goettel MS, Inglis GDWS: **Fungi**.Lacey LA, Kaya HK, editors. *Field Manual of Techniques in Invertebrate Pathology.* 2nd Edition. Dordrecht: Springer; 2000; pp. 255–281.

#### **Publisher Full Text**

1986.

Goettel MS, Koike M, Kim JJ, et al.: Potential of Lecanicillium spp. for the management of insects, nematodes, and plant diseases. J. Invertebr. Pathol. 2008: 98: 256–261.

#### PubMed Abstract | Publisher Full Text

Gonzalez-Franco CR: Actinomycetes as biological control agents of phytopathogenic fungi. *Tecnociencia Chihuahua*. 2009; **3**: 64–73.

Gory JS, Bishop DH: *Use of baculoviruses as biological insecticides.* Humana Press: 1995

Gramkow AW, Perecmanis S, Sousa RLBNE, et al.: Insecticidal activity of two proteases against *Spodoptera frugiperda* larvae infected with recombinant baculoviruses. *J. Virol.* 2010; **7**: 143.

Granados FB: The Biology of Baculoviruses. Boca Raton, FL, USA: CRC Press;

Grewal PS, Peters A: Formulation and quality. In: Grewal, P.S., Ehlers, R.U. and Shapiro-Ilan, D.I. ©CAB International 2017 − for Tarique Hassan Askary 282 T.H. Askary and M. Jamal Ahmad (eds). In: Nematodes as Biocontrol Agents. CAB Internationa. CAB International, Wallingford, UK. 2005; pp. 79–90.

Guillon ML: **Regulation of biological control agents in Europe.** *International symposium on biopesticides for developing countries.* Turrialba: CATIE; 2003; pp. 143–147

Gupta SDA: **Biopesticides: an ecofriendly approach for pest control.** *J. Biopest.* 2010; **3**: 186–188.

Gurr GM, Wratten SD, Landis DA, et al.: **Habitat management to suppress pest populations: Progress and prospects**. Annu. Rev. Entomol. 2017; **62**: 91–109

#### PubMed Abstract | Publisher Full Text

Hajek AE, Leger LR: **Interactions between fungal pathogens and insect hosts**. *Annu. Rev. Entomol.* 1994; **39**: 293–322. **Publisher Full Text** 

Harald BT: Pesticides & Biopesticides: Formulation & Mode of Action. 2019

Hezakiel HE, Thampi M, Rebello S, et al.: Biopesticides: a Green Approach Towards Agricultural Pests. Appl. Biochem. Biotechnol. 2023; 1–30.
Publisher Full Text

Higa PJ: Beneficial and effective microorganisms for sustainable agriculture and environment. Atami, Japan: International Nature Farming Research Center; 1994; 16.

Howell CR: Mechanisms Employed by Trichoderma Species in the Biological Control of Plant Diseases: The History and Evolution of Current Concepts. *Plant Dis.* 2003; **87**: 4–10.

# PubMed Abstract | Publisher Full Text

Huang Y, Li Z, Luo X, et al.: Biopesticides extension and rice farmers' adoption behavior: a survey from Rural Hubei Province, China. Environ. Sci. Pollut. Res. 2022; 29: 51744–51757.

# PubMed Abstract | Publisher Full Text

Hughes PR, Wood HA, Breen JP, et al.: Enhanced bioactivity of recombinant baculoviruses expressing insect-specific spider toxins in lepidopteran crop pests. J. Invertebr. Pathol. 1997; 69: 112–118.

PubMed Abstract | Publisher Full Text

Imaizumi S, Nishino T, Miyabe K, et al.: Biological Control of Annual Bluegrass (Poa annual.) with a Japanese Isolate of Xanthomonas campestrispv. poae (JT-P482). Biol. Control. 1997; 8: 7–14. Publisher Full Text

Inglis GD, Johnson DL, Goettel MS: **Effect of bait substrate and formulation on infection of grasshopper nymphs by Beauveria bassiana**. *Biocontrol Sci. Tech.* 1996; **6**: 35–50.

#### **Publisher Full Text**

Isman MB: Botanical insecticides, deterrents, and repellents in modern agriculture and an increasingly regulated world. *Annu. Rev. Entomol.* 2006; **51**: 45–66.

# PubMed Abstract | Publisher Full Text

Jackson TA, Crawford GT: Oryctes virus—time for a new look at a useful biocontrol agent. J. Invertebr. Pathol. 2005; 89: 91–94.
PubMed Abstract | Publisher Full Text

Jacobson TKBGD, Azevedo JC: **Invasiveness, Monitoring and Control of** *Hakea sericea*: **A Systematic Review.** *Plants.* 2023; **12**: 751.

PubMed Abstract | Publisher Full Text | Free Full Text

Jayaraj J, Radhakrishnan VR: **Development of formulations of** *Trichoderma harzianum* **strain M1 for control of damping-off of tomato caused by** *Pythium aphanidermatum***.** *Arch. Phytopathol. Plant Protect.* **2006: <b>39**: 1–8.

#### **Publisher Full Text**

Johnson DR, Wyse DL, Jones KJ: **Controlling weeds with phytopathogenic bacteria**. *Weed Technol*. 1996; **10**: 621–624. **Publisher Full Text** 

Jouzani GS, Valijanian SR: **Bacillus thuringiensis: a successful insecticide** with new environmental features and tidings. *Appl. Microbiol. Biotechnol.* 2017; **101**: 2691–2711.

PubMed Abstract | Publisher Full Text

Jozsef Kiss MDD: **Implementing biopesticides as part of an integrated pest management (IPM) programme.** *Biopesticides for sustainable agriculture*. France: Académie d'Agriculture de France; 2020.

Kachhawa D: Microorganisms as a biopesticides. J. Entomol. Zool. Stud. 2017; 5: 468–473.

Kalamkoff JWV: *Baculovirus*. Dunedin, New Zealand: University of Otago; 2004

2004. Kalpana AK: **A review of biopesticides and their plant phytochemicals** 

Karlsson, Green K, Stenberg LÅ: Making sense of Integrated Pest Management (IPM) in the light of evolution. *Evol. Appl.* 2020; **13**: 1791–1805.

PubMed Abstract | Publisher Full Text | Free Full Text

information. Ann. Rom. Soc. Cell Biol. 2021; 3576-3588

Kerr A, Bullard G: **Biocontrol of Crown Gall by Rhizobium rhizogenes: Challenges in Biopesticide Commercialisation.** *Agronomy.* 2020; **10**: 1126.

**Publisher Full Text** 

Khan AA, Park S, Ali S, et al.: Seed coating: A technique for delivering biopesticides for sustainable pest management. Advances in Seed Priming. Springer; 2020.

Killick HJ: Influence of droplet size, solar ultraviolet light and protectants, and other factors on the efficacy of baculovirus sprays against *Panolis flammea* (Schiff.) (Lepidoptera: Noctuidae). *Crop Prot.* 1990; 9: 21–28.

**Publisher Full Text** 

Kim JJ, Goettel MS: Evaluation of *Lecanicillium longisporum*, Vertalec against the cotton aphid, *Aphis gossypii*, and cucumber powdery mildew, *Sphaerotheca fuliginea* in a greenhouse environment. *Crop Part* 2011: 29: 540-544

Koike M, Shinya R, Aiuchi D, et al.: Future biological control for soybean cyst nematode. El-Shemy HA, editor. Soybean physiology and biochemistry. Croatia: Intech Open Access; 2011; pp. 193–208

Kolombet LV, Zhigletsova SK, Kosareva NI, et al.: Development of an extended shelf-life, liquid formulation of the biofungicide Trichoderma asperellum. World J. Microbiol. Biotechnol. 2008; 24: 122-121

Publisher Full Text

Koppenhöfer AM, Shapiro-Ilan DI, Hiltpold I: **Entomopathogenic Nematodes in Sustainable Food Production.** *Front. Sustain. Food Syst.* 2020; **4**: 125.

Publisher Full Text

Kordali S, Cakir A, Ozer H, et al.: Antifungal, phytotoxic and insecticidal properties of essential oil isolated from Turkish Origanum acutidens and its three components, carvacrol, thymol and p-cymene. Bioresour. Technol. 2008: 99: 8788–8795.

**Publisher Full Text** 

Koul O: Microbial biopesticides: opportunities and challenges. *CAB Rev.* 2011; **6**: 1–26.

Kumar J, Ramlal A, Mallick D, et al.: An overview of some biopesticides and their importance in plant protection for commercial acceptance. *Plants.* 2021; **10**: 1185.

PubMed Abstract | Publisher Full Text | Free Full Text

Kumar S: **Biopesticides:** a need for food and environmental safety. *J Biofert Biopest.* 2012; **03**: 1–3.

Publisher Full Text

Kumari I, Hussain R, Sharma S, et al.: Microbial biopesticides for sustainable agricultural practices. Biopesticides. Elsevier; 2022; pp. 301–317.

Kvakkestad V, Sundbye A, Gwynn KI: **Authorization of microbial plant protection products in the Scandinavian countries: A comparative analysis.** *Environ. Sci. Policy.* 2020; **106**: 115–124.

Publisher Full Text

Lacey LA, Goettel MS: Current developments in microbial control of insect pests and prospects for the early 21st century. *Entomophaga*. 1995; 40: 3–27.

**Publisher Full Text** 

Lacey LA, Grzywacz D, Shapiro-Ilan DI, et al.: Insect pathogens as biological control agents: back to the future. J. Invertebr. Pathol. 2015; 132: 1–41.

PubMed Abstract | Publisher Full Text

Langner T, Göhre G: Fungal chitinases: function, regulation, and potential roles in plant/pathogen interactions. *Curr. Genet.* 2016; **62**: 243–254.

**PubMed Abstract | Publisher Full Text** 

Lasa R, Ruiz-Portero C, Alcázar MD, et al.: Efficacy of optical brightener formulations of *Spodoptera exigua* multiple nucleopolyhedrovirus (SeMNPV) as a biological insecticide in greenhouses in southern Spain. *Biol. Control.* 2007; 40: 89–96.

Law JW, Ser HL, Khan TM, et al.: The Potential of Streptomyces as Biocontrol Agents against the Rice Blast Fungus, Magnaporthe oryzae (Pyricularia oryzae). Front. Microbiol. 2017; 8: 3.

PubMed Abstract | Publisher Full Text | Free Full Text

Leahy J, Mendelsohn M, Kough J, et al.: Biopesticide oversight and registration at the US Environmental Protection Agency. Biopestic. State Art Futur. Oppor. 2014; 3–18.

**Publisher Full Text** 

Lee JY, Kang SW, Yoon C: *Verticillium lecanii* Spore Formulation Using UV Protectant and Wetting Agent and the Biocontrol of Cotton Aphids. *Biotechnol. Lett.* 2006; **28**: 1041–1045.

PubMed Abstract | Publisher Full Text

Leggett MJP, Setlow SA, Sattar JYM: **Assessing the activity of microbicides against bacterial spores: knowledge and pitfalls.** *Appl. Microbiol.* 2016; **120**: 1174–1180.

PubMed Abstract | Publisher Full Text

Legwaila MM, Munthali DC, Kwerepe BC: Efficacy of Bacillus thuringiensis (var. kurstaki) qAgainst Diamondback Moth (*Plutella xylostella* L.) Eggs and Larvae on Cabbage Under Semi-Controlled Greenhouse Conditions. *Int. J. Insect. Sci.* 2015; 7: 39–45.
PubMed Abstract | Publisher Full Text | Free Full Text

Leo ML, Rathore HS: Biopesticides Handbook. 1st ed. Natu: CRC Press; 2015.
Li BOTS: Effects of trehalose on stress tolerance and biocontrol

efficacy of *Cryptococcus laurentii*. *J. Appl. Microbiol*. 2002; **100**: 854–861. Li H. Wei I. Pan SY. *et al.*: Antifungal. phytotoxic and toxic metabolites

Li H, Wei J, Pan SY, et al.: Antifungal, phytotoxic and toxic metabolites produced by *Penicillium purpurogenum*. Nat. Prod. Res. 2014; **28**: 2358–2361.

PubMed Abstract | Publisher Full Text

Liao J, Xue Y, Xiao G, et al.: Inheritance and fitness costs of resistance to Bacillus thuringiensis toxin Cry2Ad in laboratory strains of the diamondback moth, Plutella xylostella (L.). Sci. Reports. 2019; 9: 6113. PubMed Abstract | Publisher Full Text | Free Full Text

Liu X, Cao A, Yan D, et al.: **Overview of mechanisms and uses of biopesticides.** Int. J. Pest Manag. 2021; **67**: 65–72.

**Publisher Full Text** 

Lumsden RD, Lewis JA, Fravel DR: Formulation and delivery of biocontrol agents for use against soilborne plant pathogens. In Hall FR, Barry JW, editors. Biorational Pest Control Agents Formulation and Delivery, ACS Symposium Series 595. Washington, DC. 1995; p. 166182.

Luo Y, Wu S, He X, et al.: Identification of a Cordyceps fumosorosea Fungus Isolate and Its Pathogenicity against Asian Citrus Psyllid, Diaphorina citri (Hemiptera). (Liviidae). Insects. 2022; 13: 374. PubMed Abstract | Publisher Full Text | Free Full Text

MacGregor JT: **Genetic toxicity assessment of microbial pesticides: needs and recommended approaches.** *Intern. Assoc. Env. Mutagen. Soc.* 2006; **1**: 17.

Matos MP, da Silva AM, El-Din NS, et al.: Biopesticides: An overview on the recent developments and perspectives. Bioorg. Med. Chem. 2020; 28: 115255.

**Publisher Full Text** 

Mazid KJ: A review on the use of biopesticides in insect pest management. *Int. J. Sci. Adv. Technol.* 2011; **1**: 169–178.

McGuire MR, Shasha BS: **Adherent starch granules for encapsulation of insect control agents.** *J. Econ. Entomol.* 1992; **85**: 1425–1433. **Publisher Full Text** 

McManus ML: Biopesticides: an overview. 1989.

Meenatchi R, Aditi N: Biopesticides for pest management. Sustainable Bioeconomy: Pathways to Sustainable Development Goals. 2021.

Montalva C, Rocha LFN, Fernandes ÉKK, et al.: Conidiobolus macrosporus (Entomophthorales), a mosquito pathogen in Central Brazil.

J. Invertebr. Pathol. 2016; 139: 102–108.

PubMed Abstract | Publisher Full Text

Moore D, Bridge PD, Higgins PM, et al.: Ultra-violet radiation damage to *Metarhizium flavoviride* conidia and the protection given by vegetable and mineral oils and chemical sunscreens. *Ann. Appl. Biol.* 1993; 122: 605–616.

**Publisher Full Text** 

Moore NF, King LAPR: Viruses of insects. Insect. Sci. Appl. 1987; 3: 275–289.

Moscardi F, de Souza ML, de Castro MEB, et al.: Baculovirus pesticides: **present state and future perspectives.** Ahmad I, Ahmad F, Pichtel J, editors. *Microbes and microbial technology.* New York: Springer; 2011;

Muhammad IS, Muhammad A, Mansoor UHMA: Efficacy of Cry1Ac toxin from *Bacillus thuringiensis* against the beet armyworm, *Spodoptera* exigua (Hübner) (Lepidoptera: Noctuidae). Egypt J. Biol. Pest. Control.

#### **Publisher Full Text**

Muir D: *Environmental Programmes Pesticide policy*. Department of Forestry, Fisheries and the Environment; 2023; vol. **2023**.

Nakahara Y, Shimura S, Ueno C, et al.: Purification and characterization of silkworm hemocytes by flow cytometry. Dev. Comp. Immunol. 2009; **33**: 439-448.

#### PubMed Abstract | Publisher Full Text

Naqqash MN, Gökçe A, Bakhsh A, et al.: Insecticide resistance and its molecular basis in urban insect pests. Parasitol. Res. 2016; 115: 1363-1373

#### PubMed Abstract | Publisher Full Text

Naveenkumar R, Muthukumar A, Sangeetha GMR: **Developing eco-**friendly biofungicide for the management of major seed borne diseases of rice and assessing their physical stability and storage life. C. R. Biol. 2017; 340: 214-225.

#### PubMed Abstract | Publisher Full Text

Nuruzzaman M, Liu Y, Rahman MM, et al.: Nanobiopesticides: Composition and preparation methods. Nano-biopesticides today and future perspectives. Elsevier; 2019; Academic Press; pp. 69-131. .

O'Brien SF, Iones II: Green Chemistry and Sustainable Agriculture The Role of Biopesticides. 2009.

Okuno S, Takatsuka J, Nakai M, et al.: Viral-enhancing activity of various stilbene-derived brighteners for a *Spodoptera litura* (Lepidoptera: Noctuidae) nucleopolyhedrovirus. *Biol. Control.* 2003; **26**: 146–152.

Paau AS: Formulations useful in applying beneficial microorganisms to seeds. Trends Biotechnol. 1988: 6: 276-279.

#### Publisher Full Text

Penyalver R, Vicedo B, López MM: Use of the Genetically Engineered Agrobacterium Strain K1026 for Biological Control of Crown Gall. Eur. J. Plant Pathol. 2000; 106: 801–810.

#### **Publisher Full Text**

Pereira RM, Roberts DW: Alginate and cornstarch mycelial formulations of entomopathogenic fungi. Beguveria bassiana and Metarhizium anisopliae. J. Econ. Entomol. 1991; 84: 1657–1661.

Persistance Market Research: **Biopesticides, derived from natural** sources like plants, animals, bacteria, and certain minerals, offer a safe and sustainable alternative to synthetic pesticides. These ecofriendly solutions include natural pest-controlling substances (biochemicals), be. 2023; 1-6.

Pineda S, Alatorre R, Schneider MA: Pathogenicity of two entomopathogenic fungi on Trialeurodes vaporariorum and field evaluation of a Paecilomyces fumosoroseus isolate. Southwest. Entomol. 2007: 32: 43-52.

Pitterna T, Cassayre J, Hüter OF, et al.: New ventures in the chemistry of avermectins. *Bioorg. Med. Chem.* 2009; **17**: 4085–4095. PubMed Abstract | Publisher Full Text

Powell KA, Jutsum AR: Technical and commercial aspects of biocontrol products Pesticide Science. Pestic. Sci. 1993; 32: 315-321.

Prasanna BM, Huesing JE, Eddy R: Biopesticides in integrated pest management: Advantages and limitations. Integrated Pest Management. Springer; 2018.

Punja ZK: Comparative efficacy of bacteria, fungi, and yeasts as biological control agents for diseases of vegetative crops. Can. J. Plant Pathol. 1997; 19: 315-323.

## **Publisher Full Text**

Pusey PL: Enhancement of biocontrol agents for postharvest diseases and their integration with other control strategies. Wilson CL, Wisniewski MD, editors. Biological control of postharvest diseases - theory and practice. Boca Raton: CRC Press; 1994; pp. 77-88

Quarles W: New biopesticides for IPM and organic production. IPM Pr. 2013; 33: 1-20.

Rae R, Sheehy L, McDonald-Howard K: **Thirty years of slug control using the parasitic nematode** *Phasmarhabditis hermaphrodita* and beyond. Pest Manag. Sci. 2023; **79**: 3408–3424.

#### PubMed Abstract | Publisher Full Text

Rajaput J, Rao MSL, Hegde RV, et al.: Biopriming: a novel seed treatment options to manage the seed-borne fungal infection of tomato. J. Pharmacogn. Phytochem. 2019; 8: 659-661.

Ram K, Singh R: Efficacy of different fungicides and biopesticides for the management of lentil wilt (Fusarium oxysporum f. sp. lentis):

Management of lentil wilt. J. AgriSearch. 2021; 8: 55-58.

Ramle M. Wahid MB. Norman K. et al.: The incidence and use of Orvctes virus for control of rhinoceros beetle in oil palm plantations in Malaysia. J. Invertebr. Pathol. 2005; 89: 85-90.

#### **PubMed Abstract | Publisher Full Text**

Raymond B, Johnston PR, Wright DJ, et al.: A mid-gut microbiota is not required for the pathogenicity of Bacillus thuringiensis to diamondback moth larvae. Environ. Microbiol. 2009; 11: 2556-2563.

#### PubMed Abstract | Publisher Full Text

Riga E, Lenteren JCV, Medrzycki P, et al.: Challenges and Opportunities for Biopesticides to Debut in Conventional Agricultural Systems. Integrated Pest Management: Experiences with Implementation, Global

Roh JY, ChoiJY LMS, Jin BR, et al.: Bacillus thuringiensis as a specific, safe, and effective tool for insect pest control. J. Microbiol. Biotechnol. 2007; **17**: 547–559.

#### **PubMed Abstract**

Rohrmann GF: Baculovirus Molecular Biology. Third ed. Bethesda (MD): National Center for Biotechnology Information (US); 2013

Roy A, Moktan SP: Characteristics of Bacillus cereus isolates from legume-based Indian fermented foods. Food Control. 2007; 18: 1555-1564

#### **Publisher Full Text**

Ruiu L: Microbial biopesticides in agroecosystems. Agron. 2018; 8: 235. 8235

#### **Publisher Full Text**

Sabaratnam TJ: Formulation of a Streptomyces biocontrol agent for the suppression of Rhizoctonia damping-off in tomato transplants. Biol. Control. 2002; 23: 245-253.

#### **Publisher Full Text**

SADC Secretariat: SADC (Southern African Development Community) Guidelines for Pesticides Management and Risk Reduction. 2019.

Sarwar M: Biopesticides: an effective and environmentally friendly insect-pests inhibitor line of action. Int. J. Eng. Adv. Res. Tech. 2015; 1:

Satish G, Ashokrao DM, Arun SK: Microbial degradation of pesticide: a review. Afr. J. Microbiol. Res. 2017; 11: 992-1012.

Savita SA: Fungi as Biological Control Agents. Giri B, Prasad R, Wu QS, et al., editors. Biofertilizers for Sustainable Agriculture and Environment Soil Biology. Vol. 55. Cham: Springer; 2019.

Schallmey M, Singh WO: Developments in the use of Bacillus species for industrial production. Can. J. Microbiol. 2004; **50**: 1–17.

#### **Publisher Full Text**

Schisler DA, Slininger PJ, Behle JM: Formulation of Bacillus spp. for biological control of plant diseases. Phytopathology. 2004; 94

#### **Publisher Full Text**

Senthil-Nathan S: A Review of biopesticides and their mode of action **against insect pests.** *Environmental sustainability.* New Delhi: Springer; 2015; pp. 49–63.

Shapiro-Ilan DI, Gough DH, Piggott SJ, et al.: Application technology and environmental considerations for use of entomopathogenic nematodes in biological control. *Biol. Control.* 2006; **38**: 124–133.

Shapiro M: Radiation protection and activity enhancement of viruses. 1995.

Shapiro M: Use of optical brighteners as radiation protectants for gypsy moth (Lepidoptera: Lymantriidae) nuclear polyhedrosis virus. J. Econ. Entomol. 1992; 85: 1682-1686.

Shilpi S, Promila M: Biopestcides: Types and Applications. Int J Adv Pharm Biol Chem. 2012; 1: 508-515.

Shishupala S: Biocontrol Potential of Fungi for Pest and Pathogen Management. Rajpal VR, Singh I, Navi SS, editors. Fungal diversity, ecology and control management. Fungal Biology. Singapore: Springer;

Singh A, Bhardwaj R, Singh IK: Biocontrol Agents: Potential of Biopesticides for Integrated Pest Management. Giri B, et al., editors Biofertilizers for Sustainable Agriculture and Environment, Soil Biology. 2019;

Stevenson PC, Isman MB, Belmain SR: Pesticidal plants in Africa: a global vision of new biological control products from local uses. Ind. Crop. Prod. 2017; 110: 2-9. **Publisher Full Text** 

Stockwell VO, Stack JP: Using Pseudomonas spp. for integrated biological control. Phytopathology. 2007; 97: 244-249. PubMed Abstract | Publisher Full Text

Sufyan M, Tahir MI, Haq MIU, et al.: Effect of seed bio priming with rhizobacteria against root associated pathogenic fungi in chickpea. Pak. J. Phytopathol. 2020; 32: 89-96.

Szewczyk B, Hoyos-Carvajal L, Paluszek M, et al.: Baculoviruses—reemerging biopesticides. Biotechnol. Adv. 2006; 24: 143-160. PubMed Abstract | Publisher Full Text

Tarasco E, Fanelli E, Salvemini C, et al.: Entomopathogenic nematodes and their symbiotic bacteria: from genes to field uses. Front. Insect. Sci. 2023; **3**: 1195254.

PubMed Abstract | Publisher Full Text | Free Full Text

TeBeest DO, Templeton GE: Mycoherbicides: Progress in the Biologic. Plant Dis. 1985; **69**: 7.

Temiz Ö: Biopesticide emamectin benzoate in the liver of male mice: evaluation of oxidative toxicity with stress protein, DNA oxidation, and apoptosis biomarkers. Environ. Sci. Pollut. Res. 2020; 27:

PubMed Abstract | Publisher Full Text

The Nagoya Protocol: The Nagoya Protocol. 2014.

Tijjani A, Bashir KA, Mohammed I, et al.: **Biopesticides for pests control:** A review. J. Biopestic Agric. 2016; **3**: 6–13.

Upamanya GK, Bhattacharyya DP: Consortia of entomo-pathogenic fungi and bio-control agents improve the agro-ecological conditions for brinjal cultivation of Assam. 3 Biotech. 2020; 10: 450. PubMed Abstract | Publisher Full Text | Free Full Text

United Nations: The Paris Agreement. 2015.

Reference Source

Urquhart E, Punja ZK: Epiphytic growth and survival of Tilletiopsis pallescens, a potential biological control agent of Sphaerotheca fuliginea, on cucumber leawes. Can. J. Bot. 1997; 75: 892-901. **Publisher Full Text** 

USEPA: What are Biopesticides? 2020.

Usta C: Microorganisms in biological pest control—a review (bacterial toxin application and effect of environmental factors). Curr. Prog. Biol. Res. 2013; 13: 287-317. **Publisher Full Text** 

Vicedo B, Peñalver R, Asins MJ, et al.: Biological Control of Agrobacterium tumefaciens, Colonization, and pAgK84 Transfer with Agrobacterium radiobacter K84 and the Tra Mutant Strain K1026. Appl. Environ. Microbiol. 1993: 59(1): 309-315.

PubMed Abstract | Publisher Full Text | Free Full Text

Vivekanandhan P, Swathy K, Murugan AC: Insecticidal Efficacy of Metarhizium anisopliae Derived Chemical Constituents against Disease-Vector Mosquitoes. J. Fungi. 2022; 8: 300. PubMed Abstract | Publisher Full Text | Free Full Text

Walker HL, Boyette CD: Biocontrol of sicklepod (Cassia obtusifolia) in soybeans (Glycine max) with Alternaria cassiae. Weed Sci. 1985; 33: 212–215.

Wang J, Wu W, Qin Y, et al.: Automated targeting systems for pesticide application in agriculture: A review. Comput. Electron. Agric. 2019; 156:

Wattimena CMA, Latumahina FS: Effectiveness of botanical biopesticides with different concentrations of termite mortality. J. Belantara. 2021; 4: 66-74. Publisher Full Text

Yelitza C, Mundstock S, Sampaio M, et al.: Use of Parasitoids as a Biocontrol Agent in the Neotropical Region: Challenges and Potential. IntechOpen; 2020. Hortic Crop.

**Publisher Full Text** 

Zhang SA, Xu WM, Yan Z, et al.: Research and commercialization of yield-increasing bacteria (YIB) in China. Tang WHRCAR, editors. Advances in Biological Control of Plant Diseases. Beijing: China Agricultural University Press; 1996; pp. 47-53.

Zidack NK, Backman PA, Shaw JJ: **Promotion of bacterial infection of** leaves by an organosilicone surfactant: implications for biological weed control. Biol. Control. 1992; 2: 111-117.

**Publisher Full Text** 

# **Open Peer Review**

**Current Peer Review Status:** 







Version 4

Reviewer Report 19 February 2025

https://doi.org/10.5256/f1000research.177450.r365526

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# ? Alessandro Mattedi 🗓

- <sup>1</sup> University of L'Aquila, Coppito, L'Aquila, Italy
- <sup>2</sup> University of L'Aquila, Coppito, L'Aquila, Italy

Thank you for your suggestions. We have included the following references.

- Abdallah, Dorra Ben, Slim Tounsi, Houda Gharsallah, Adnane Hammami, and Olfa Frikha-Gargouri. "Lipopeptides from Bacillus amyloliquefaciens strain 32a as promising biocontrol compounds against the plant pathogen Agrobacterium tumefaciens." *Environmental Science and Pollution Research* 25 (2018): 36518-36529.
- Brown, Pamela JB, Jeff H. Chang, and Clay Fuqua. "Agrobacterium tumefaciens: a transformative agent for fundamental insights into host-microbe interactions, genome biology, chemical signaling, and cell biology." *Journal of Bacteriology* 205, no. 4 (2023): e00005-23.
- Mnif, Inès, and Dhouha Ghribi. "Potential of bacterial derived biopesticides in pest management." Crop Protection 77 (2015): 52-64."

Reply: The cited papers do not talk about the biopesticide properties of *Agrobacterium tumefaciens* that the manuscript mentions, and you did not explain in what they consist. Please verify whether they are the correct references.

"14) "*Agrobactenum tumefaciens* is available commercially in Australia, the United States, and New Zealand"

What about other countries?

Please also give examples of commercial biopesticide products containing A. tumefaciens.

Response: Thank you for your suggestions There are no recent A. tumefaciens products"

Reply: If there are no products available, then how it is possible that it is "available commercially" in the mentioned countries as a biopesticide? Please clarify this section.

"15) I also recommend the possibility to mention Agrobacterium radiobacter and how it is sometimes confused.

• Response: Thank you for your suggestions and comments. A strain of *A. radiobacter* known as K84 is used as a biopesticide to control crown gall disease (Stockwell et al. 1993).

Stockwell, V. O., L. W. Moore, and J. E. Loper. "Fate of Agrobacterium radiobacter K84 in the environment." *Applied and Environmental Microbiology* 59, no. 7 (1993): 2112-2120."

Reply: Perhaps due to an oversight, in the manuscript you added the wrong reference (Stockwell & Stack, 2007), plus you did not mention how the species is sometimes confused. Please check.

Competing Interests: No competing interests were disclosed.

**Reviewer Expertise:** Environmental microbiology

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard, however I have significant reservations, as outlined above.

# Version 3

Reviewer Report 11 January 2025

https://doi.org/10.5256/f1000research.174749.r351244

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# 🚶 Alessandro Mattedi 🗓

- <sup>1</sup> University of L'Aquila, Coppito, L'Aquila, Italy
- <sup>2</sup> University of L'Aquila, Coppito, L'Aquila, Italy
- <sup>3</sup> University of L'Aquila, Coppito, L'Aquila, Italy

Thanks to the authors for writing a long review about this topic. I have some minor corrections and suggestions about the manuscript. Please forgive me in advance if my language is not clear enough or heavy to read:

- 1) The review in general could benefit from a more synthetic language. Often, paragraphs can be shortened while conveying the same amount of information. Occasionally, it also happens that the same concepts are repeated, for example when illustrating the several advantages of biopesticides or how they align with several different target strategies. There are other examples that could benefit from reading again the text and summarizing repetitions. I suggest the authors to consider this possibility, as it can help to promote their ideas in a more effective and concise way.
- 2) "However, resistance to some *Bt* products has been noted." I recommend to add a brief mention on how and why this might have happened.

3) It could be worthy to note that recently some concerns have arisen about the possibility that Bt toxins could be involved in a fraction of foodborne outbreaks, although there are still relevant gaps in knowledge that needs to be filled and we cannot yet draw definitive conclusions. Still, the reader might be interested in pursuing further studying and investigation of the topic. A reference, Ref (1)

Not strictly necessary, depending on the goals.

- 4) There are other entomopathogenic bacteria of interest such as Lysinibacillus sphaericus, Brevibacillus laterosporus, Serratia marcescens, Yersinia entomophaga etc.. The topic is much larger and has quite interesting considerations regarding the feasibility of applications, their effectiveness, and the large-scale limitations. For example: Ref (2)
- 5) "Besides, biopesticides 1) generate fewer greenhouse gasses (GHG) than chemical pesticides, 2) improve nutrient availability for plants, 3) increase soil fertility, and 4) offer a more environmentally safe approach to pest control in agriculture, which can be useful in the fight against climate change."

Please provide references for these points.

Please elaborate points 2 and 3: if you are stating that biopesticides per se are also biostimulants, this requires a better explanation.

Please expand point 4 or condensate it with point 1.

6) "Microbial biopesticides offer diverse delivery methods to effectively reach crop targets. These methods include live organisms, dead organisms, and spores"

Please describe these methods here for a better understanding of the scientific mechanisms.

7) "Additionally, they leave no harmful residues, don't harm pollinators and other valuable organisms, and can be applied close to harvest time."

This is actually a slight approximation. Recent research show that there could be some negative effects after all, and that there are knowledge gaps regarding pollinators. It is always a matter of balancing risks and benefits. While the low impact status of biopesticides is not changed, it is still worth to remember peculiarities and potential limits. For reference: Ref (3)

8) "Bacterial biopesticides, while effective, come with a few disadvantages. For instance, they only target specific species or groups of insects, leaving other pests to survive and continue causing damage."

This is conceptually a bit clashing with the previous statement that specificity is an advantage, in order to avoid non-target insects. It is better to reformulate in a clearer way that the specificity gives that advantages but requires complementarity of treatment when dealing with different pests.

9) The review doesn't adhere to the same structure within its sections. For example, section 5.5 gives two explicit separated paragraphs titled "advantages" and "disadvantages" of EPNs, while other sections discuss them as part of the flow in the text. I suggest to uniform the style for better clarity and readibility.

- 10) Section 6 is a bit superficial and doesn't mention the need to assess that alloctonous predator can be safely controlled i.e. they cannot become invasive. Since the review focuses on microbials, it doesn't need to go in detail about macrobials, but better accuracy is required.
- 11) Some phrases like "reduce chomping on your precious plants", "This gram-positive warrior wields a powerful weapon: delta-endotoxin proteins that wreak havoc on the mid-guts of susceptible insect larvae" or "These technologies offer exciting possibilities for sustainable agriculture" sound quite odd and out of place for a technical review. Are they referencing something else? I suggest to rephrase anyway.
- 12) "Mass production of these biofungicide bacteria formulations typically involves deep-tank liquid fermentation. However, in some cases, methods like semisolid or solid-state fermentation might be more suitable."

Please explain, summarizing what these processes are and detailing why one could be preferred in certain conditions.

- 13) Please explain what are the biopesticide properties of Agrobacterium tumefaciens and give recent detailed references for specific sentences.
- 14) "Agrobactenum tumefaciens is available commercially in Australia, the United States, and New Zealand"

What about other countries?

Please also give examples of biopesticide commercial products containing A. tumefaciens.

- 15) I also recommend the possibility to mention Agrobacterium radiobacter and how it is sometimes confused.
- 16) "Entomopathogenic fungi are a type of fungi that can target and control pests without harming beneficial organisms such as pollinators."

This is not necessarily true and there is still the need to verify the safety of formulations against pollinators. For example: Ref (4)

17) Abstract: "Biopesticides are pest control products derived from natural sources such as microbes, macro-organisms (insects and pathogens), plant extracts, and certain minerals."

Reprised in the conclusions: "Biopesticides, derived from natural materials like animals, plants, bacteria, and certain minerals"

This is a direct copy and paste from the EPA definition, which is credited in other sections: https://www.epa.gov/ingredients-used-pesticide-products/what-are-biopesticides

I recommend to avoid such exact transpositions.

18) The manuscript shifts in detail. Even within the same section, some parts are more accurate

while others are more superficial. Examples are not always fully comprehensive. There are parts which are not fully clarified, and sometimes a specific example could induce to think that it is broadly applicable to other cases. I recommend to rework in order to convey the information in a clearer way.

- 19) The article aims to explain recent advances, but it is for the most part an introductory presentation and summary of the known concepts in the field. I suggest to mention more in detail recent publications to discuss specifically what they add to the established literature. One example: Baculoviruses are introduced with references going back to the 90s, did they have recent innovations?
- 20) The review mentions CRISPR/Cas9 as a tool for the development of microbial biopesticides. The brief mention doesn't explain why. I suggest to slightly expand.
- 21) There are images of low quality, for example figure 8. Please create new images of higher quality.
- 21) Do the photos of figure 6 have authors and copyright?

#### References

- 1. Biggel M, Jessberger N, Kovac J, Johler S: Recent paradigm shifts in the perception of the role of Bacillus thuringiensis in foodborne disease. *Food Microbiol*. 2022; **105**: 104025 PubMed Abstract | Publisher Full Text
- 2. Ruiu L: Insect Pathogenic Bacteria in Integrated Pest Management. *Insects*. 2015; **6** (2): 352-67 PubMed Abstract | Publisher Full Text
- 3. Cappa F, Baracchi D, Cervo R: Biopesticides and insect pollinators: Detrimental effects, outdated guidelines, and future directions. *Sci Total Environ*. 2022; **837**: 155714 PubMed Abstract | Publisher Full Text
- 4. Leite MOG, Alves DA, Lecocq A, Malaquias JB, et al.: Laboratory Risk Assessment of Three Entomopathogenic Fungi Used for Pest Control toward Social Bee Pollinators. *Microorganisms*. 2022; **10** (9). PubMed Abstract | Publisher Full Text

# Is the topic of the review discussed comprehensively in the context of the current literature?

Partly

Are all factual statements correct and adequately supported by citations?

Partly

Is the review written in accessible language?

Partly

Are the conclusions drawn appropriate in the context of the current research literature? Partly

Competing Interests: No competing interests were disclosed.

# **Reviewer Expertise:** Environmental microbiology

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard, however I have significant reservations, as outlined above.

Author Response 23 Jan 2025

# **Kahsay Tadesse Mawcha**

Dear Reviewer,

We appreciate your time and appreciate your comments and suggestions. We will address them in the main text of the manuscript.

**Competing Interests:** There is no computing interest.

Author Response 23 Jan 2025

# **Kahsay Tadesse Mawcha**

# Authors' response to reviewers' comments

We thank the reviewer for their time and expertise in evaluating our review paper.

Thanks to the authors for writing a long review about this topic. I have some minor corrections and suggestions about the manuscript. Please forgive me in advance if my language is not clear enough or heavy to read:

- 1. The review in general could benefit from a more synthetic language. Often, paragraphs can be shortened while conveying the same amount of information. Occasionally, it also happens that the same concepts are repeated, for example when illustrating the several advantages of biopesticides or how they align with several different target strategies. There are other examples that could benefit from reading again the text and summarizing repetitions. I suggest the authors to consider this possibility, as it can help to promote their ideas in a more effective and concise way.
  - Authors Response: Thank you for your comment, and we made a few changes to the text.
- 2. "However, resistance to some *Bt* products has been noted.". I recommend adding a brief mention of how and why this might have happened.
  - Response: Thank you for your question. For the "How?" through natural selection.
     Insect pests that are susceptible to Bt toxins die, and those resistant can survive and reproduce.
- 3. It could be worthy to note that recently some concerns have arisen about the possibility that Bt toxins could be involved in a fraction of foodborne outbreaks, although there are still relevant gaps in knowledge that needs to be filled and we cannot yet draw definitive conclusions. Still, the reader might be interested in pursuing further studying and investigation of the topic. A reference, Ref (1)

Not strictly necessary, depending on the goals.

- Response: Implementing several strategies, including cultivating refuge areas of non-Bt crops, alternating Bt crops with non-Bt varieties, and employing different Bt toxins that operate via various mechanisms, may reduce the threat of resistance. However, the effectiveness of these strategies can vary depending on the specific insect species and the environmental conditions.
- 4. There are other entomopathogenic bacteria of interest such as Lysinibacillus sphaericus, Brevibacillus laterosporus, Serratia marcescens, Yersinia entomophaga etc.. The topic is much larger and has quite interesting considerations regarding the feasibility of applications, their effectiveness, and the large-scale limitations. For example: Ref (2)
  - Response: Thanks for your comments. We have included the above bacterial biopesticides in the main text. Other entomopathogenic bacteria of interest include Chromobacterium subtsugae, Brevibacillus laterosporus, Lysinibacillus sphaericus, Paenibacillus popilliae, Serratia marcescens, and Yersinia entomophagy (Gangwar et al. 2021).

Gangwar, P., Trivedi, M., Tiwari, R.K. (2021). Entomopathogenic Bacteria. In: Omkar (eds) Microbial Approaches for Insect Pest Management. Springer, Singapore. https://doi.org/10.1007/978-981-16-3595-3\_2

5) "Besides, biopesticides 1) generate fewer greenhouse gasses (GHG) than chemical pesticides, 2) improve nutrient availability for plants, 3) increase soil fertility, and 4) offer a more environmentally safe approach to pest control in agriculture, which can be useful in the fight against climate change."

Please provide references for these points.

 Response: We have included several references from FAO 2017; 2020; Climate-smart, and the Paris Agreement. We have also added Fenibo et al. (2022)

Fenibo EO, Ijoma GN, Nurmahomed W, Matambo T. The Potential and Green Chemistry Attributes of Biopesticides for Sustainable Agriculture. *Sustainability*. 2022; 14(21):14417. https://doi.org/10.3390/su142114417

5) Please elaborate points 2 and 3: if you are stating that biopesticides per se are also biostimulants, this requires a better explanation.

Please expand point 4 or condensate it with point 1.

- **Response**: Thank you for your suggestion. However, the two points are clear, and we believe there is no need to elaborate.
- 6) "Microbial biopesticides offer diverse delivery methods to reach crop targets effectively. These methods include live organisms, dead organisms, and spores"

Please describe these methods here for a better understanding of the scientific mechanisms.

• **Response**: Thank you for your suggestion. We have addressed and incorporated your comment.

7) "Additionally, they leave no harmful residues, don't harm pollinators and other valuable organisms, and can be applied close to harvest time."

This is actually a slight approximation. Recent research show that there could be some negative effects after all, and that there are knowledge gaps regarding pollinators. It is always a matter of balancing risks and benefits. While the low impact status of biopesticides is not changed, it is still worth to remember peculiarities and potential limits. For reference: Ref (3)

- **Response**: Many Thanks for your suggestion
- 8) "Bacterial biopesticides, while effective, come with a few disadvantages. For instance, they only target specific species or groups of insects, leaving other pests to survive and continue causing damage."

This is conceptually a bit clashing with the previous statement that specificity is an advantage, in order to avoid non-target insects. It is better to reformulate in a clearer way that the specificity gives that advantages but requires complementarity of treatment when dealing with different pests.

- **Response**: Thank you for your suggestions and comments. We have addressed them in the main text of the manuscript.
- 9) The review doesn't adhere to the same structure within its sections. For example, section 5.5 gives two explicit separated paragraphs titled "advantages" and "disadvantages" of EPNs, while other sections discuss them as part of the flow in the text. I suggest to uniform the style for better clarity and readibility.
  - Response: Thank you for your suggestions and comments. We have mentioned the
    advantages and disadvantages of the other types of microbial pesticides. We did this
    purposefully, as they are different in type.
- 10) Section 6 is a bit superficial and doesn't mention the need to assess that alloctonous predator can be safely controlled i.e. they cannot become invasive. Since the review focuses on microbials, it doesn't need to go in detail about macrobials, but better accuracy is required.
  - Response: Thank you for your suggestions. Macrobial agents (living organisms, such as insects, mites, or nematodes, that are used to control pests) and microbial agents (microorganisms, such as bacteria, fungi, or viruses, that are used to control pests) are both used in pest control. Still, they differ significantly in their nature and mode of action. While both macrobial and microbial agents are biological control methods, they differ in their composition and how they target pests. Macrobial agents are living organisms that directly attack the pest, whereas microbial agents are microorganisms that can infect, kill, or produce toxins harmful to the pest. So, the focus of this review is to discuss microbial biopesticides.

- 11) Some phrases like "reduce chomping on your precious plants", "This gram-positive warrior wields a powerful weapon: delta-endotoxin proteins that wreak havoc on the midguts of susceptible insect larvae" or "These technologies offer exciting possibilities for sustainable agriculture" sound quite odd and out of place for a technical review. Are they referencing something else? I suggest to rephrase anyway.
  - Response: Thank you for your suggestions. We have rephrased it in the main text of the manuscript as "Meenatchi and Aditi (2021) and Gautam (2008) have summarized several promising biopesticides that are effective against various crop pests. These specialized assassins are laser-guided for specific pests, efficiently hunting them even at low levels. They curb reproduction, keep infestations below damage thresholds, and play well with other pest-management tactics. Additionally, they help disrupt pest feeding and lessen the damage to your plants. They are environmentally safe pest control solutions "
- 12) "Mass production of these biofungicide bacteria formulations typically involves deeptank liquid fermentation. However, in some cases, methods like semisolid or solid-state fermentation might be more suitable."

Please explain, summarizing what these processes are and detailing why one could be preferred in certain conditions.

- Response: Thank you for your suggestions. The mass production of biofungicide bacteria formulations typically involves deep-tank liquid fermentation for several reasons: high yield, controlled environment, scalability, automation and Costeffectiveness. If we explain it more than the present, it may need more pages, and this could be beyond the page limit of the review.
- 13) Please explain what are the biopesticide properties of Agrobacterium tumefaciens and give recent detailed references for specific sentences.

**Response**: Thank you for your suggestions. We have included the following references.

- Abdallah, Dorra Ben, Slim Tounsi, Houda Gharsallah, Adnane Hammami, and Olfa Frikha-Gargouri. "Lipopeptides from Bacillus amyloliquefaciens strain 32a as promising biocontrol compounds against the plant pathogen Agrobacterium tumefaciens." *Environmental Science and Pollution Research* 25 (2018): 36518-36529.
- Brown, Pamela JB, Jeff H. Chang, and Clay Fuqua. "Agrobacterium tumefaciens: a transformative agent for fundamental insights into host-microbe interactions, genome biology, chemical signaling, and cell biology." *Journal of Bacteriology* 205, no. 4 (2023): e00005-23.
- Mnif, Inès, and Dhouha Ghribi. "Potential of bacterial derived biopesticides in pest management." *Crop Protection* 77 (2015): 52-64.
- 14) "Agrobactenum tumefaciens is available commercially in Australia, the United States, and New Zealand"

What about other countries?

Please also give examples of commercial biopesticide products containing A. tumefaciens.

- Response: Thank you for your suggestions There are no recent A. tumefaciens products
- 15) I also recommend the possibility to mention Agrobacterium radiobacter and how it is sometimes confused.
  - Response: Thank you for your suggestions and comments. A strain of A. radiobacter known as K84 is used as a biopesticide to control crown gall disease (Stockwell et al. 1993).

Stockwell, V. O., L. W. Moore, and J. E. Loper. "Fate of Agrobacterium radiobacter K84 in the environment." *Applied and Environmental Microbiology* 59, no. 7 (1993): 2112-2120.

16) "Entomopathogenic fungi are a type of fungi that can target and control pests without harming beneficial organisms such as pollinators."

This is not necessarily true and there is still the need to verify the safety of formulations against pollinators. For example: Ref (4)

- Response: Thank you for your suggestions and comments. The statement "Entomopathogenic fungi are a type of fungi that can target and control pests without harming beneficial organisms" is clear. Without harming beneficial organisms, such organisms cause minimal or no damage to beneficial organisms.
- 17) Abstract: "Biopesticides are pest control products derived from natural sources such as microbes, macro-organisms (insects and pathogens), plant extracts, and certain minerals."

Reprised in the conclusions: "Biopesticides, derived from natural materials like animals, plants, bacteria, and certain minerals"

This is a direct copy and paste from the EPA definition, which is credited in other sections: https://www.epa.gov/ingredients-used-pesticide-products/what-are-biopesticides

I recommend to avoid such exact transpositions.

- Response: Thank you for your suggestions and comments. We have addressed the
  comment in the main text of the manuscript "Biopesticides consist of naturally
  occurring substances, including insects, nematodes, microorganisms, plants, and
  semiochemicals, along with their by-products. They offer a sustainable alternative to
  traditional chemical pesticides.
- 18) The manuscript shifts in detail. Even within the same section, some parts are more accurate while others are more superficial. Examples are not always fully comprehensive. There are parts which are not fully clarified, and sometimes a specific example could induce to think that it is broadly applicable to other cases. I recommend to rework in order to convey the information in a clearer way.
  - Response: Thank you for your suggestions and comments, and we have made changes to the main text. However, this comment is very general one.
- 19) The article aims to explain recent advances, but it is for the most part an introductory

presentation and summary of the known concepts in the field. I suggest to mention more in detail recent publications to discuss specifically what they add to the established literature. One example: Baculoviruses are introduced with references going back to the 90s, did they have recent innovations?

# Response: Thank you for your suggestions and comments. We have added the following recent references to the list.

- o Das, Sumistha, Arunava Goswami, and Nitai Debnath. "Application of baculoviruses as biopesticides and the possibilities of nanoparticle mediated delivery." In Nanobiopesticides today and future perspectives, pp. 261-280. Academic Press, 2019.
- o Gelaye, Yohannes, and Belete Negash. "The role of baculoviruses in controlling insect pests: A review." Cogent Food & Agriculture 9, no. 1 (2023): 2254139.
- Ruiu, Luca. "Microbial biopesticides in agroecosystems." *Agronomy* 8, no. 11 (2018):
- Wilson, Kenneth, David Grzywacz, Igor Curcic, Freya Scoates, Karen Harper, Annabel Rice, Nigel Paul, and Aoife Dillon. "A novel formulation technology for baculoviruses protects biopesticide from degradation by ultraviolet radiation." Scientific Reports 10, no. 1 (2020): 13301.
- 20) The review mentions CRISPR/Cas9 as a tool for the development of microbial biopesticides. The brief mention doesn't explain why. I suggest to slightly expand.
  - Response: Thank you for your suggestions and comments. However, the current content of the paragraph about CRISPR/Cas9 is sufficient, and we believe that adding more information on CRISPR/Cas9 may become overwhelming.
- 21) There are images of low quality, for example figure 8. Please create new images of higher quality.
  - Response: Thank you for your suggestions and comments
- 21) Do the photos of figure 6 have authors and copyright?
  - Response: Thank you for your suggestions and comments. Yes. It is our own work.

**Competing Interests:** No competing interests were disclosed.

Reviewer Report 30 December 2024

https://doi.org/10.5256/f1000research.174749.r348089

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Sonia Sethi 🗓



#### Written properly

Recent advances in biopesticide research highlight microbial agents like bacteria, fungi, and viruses as eco-friendly alternatives to chemical pesticides. Innovations focus on enhancing efficacy, formulation stability, and scalability for agricultural use. Genetic engineering, nanotechnology, and integrated pest management strategies are driving improvements, addressing resistance issues, and promoting sustainable crop protection globally.

# Is the topic of the review discussed comprehensively in the context of the current literature?

Yes

Are all factual statements correct and adequately supported by citations?

Is the review written in accessible language?

Are the conclusions drawn appropriate in the context of the current research literature?

**Competing Interests:** No competing interests were disclosed.

Reviewer Expertise: Agriculture microbiology

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

Reviewer Report 27 December 2024

https://doi.org/10.5256/f1000research.174749.r346594

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# Mehari Desta Hawku 🗓



- <sup>1</sup> Crop Sciences, University of Rwanda College of Agriculture Animal Sciences and Veterinary Medicine (Ringgold ID: 183043), Busogo, Musanze, Rwanda
- <sup>2</sup> Crop Sciences, University of Rwanda College of Agriculture Animal Sciences and Veterinary Medicine (Ringgold ID: 183043), Busogo, Musanze, Rwanda
- <sup>3</sup> Crop Sciences, University of Rwanda College of Agriculture Animal Sciences and Veterinary

<sup>&</sup>lt;sup>1</sup> Dr. B. Lal Institute of Biotechnology, Malviya Nagar, Jaipur, India

<sup>&</sup>lt;sup>2</sup> Dr. B. Lal Institute of Biotechnology, Malviya Nagar, Jaipur, India

<sup>&</sup>lt;sup>3</sup> Dr. B. Lal Institute of Biotechnology, Malviya Nagar, Jaipur, India

Medicine (Ringgold ID: 183043), Busogo, Musanze, Rwanda

No further comment.

Is the topic of the review discussed comprehensively in the context of the current

Yes

Are all factual statements correct and adequately supported by citations?

Yes

Is the review written in accessible language?

Yes

Are the conclusions drawn appropriate in the context of the current research literature? Yes

**Competing Interests:** No competing interests were disclosed.

Reviewer Expertise: My research interests encompass a broad spectrum of plant pathology including: Identification and molecular characterization of resistance or susceptible genes in plantmicrobe interactions; Understanding the intricate mechanisms governinginteractions between plants and microbes; Examining the signaling pathways involved in activating plant defense responses against pathogens; Studying the molecular components that enable pathogens to invade and manipulate plant cells; Understanding the mechanisms of abiotic stress tolerance in plants

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

# Version 2

Reviewer Report 06 November 2024

https://doi.org/10.5256/f1000research.173837.r335979

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# Yordanys Ramos 🗓



<sup>1</sup> Instituto de Investigaciones Agropecuarias y Forestales, Universidad Michoacana de San Nicolas de Hidalgo, Morelia, Michoacán, Mexico

2

Instituto de Investigaciones Agropecuarias y Forestales, Universidad Michoacana de San Nicolas de Hidalgo, Morelia, Michoacán, Mexico

<sup>3</sup> Instituto de Investigaciones Agropecuarias y Forestales, Universidad Michoacana de San Nicolas de Hidalgo, Morelia, Michoacán, Mexico

<sup>4</sup> Instituto de Investigaciones Agropecuarias y Forestales, Universidad Michoacana de San Nicolas de Hidalgo, Morelia, Michoacán, Mexico

The authors have made the relevant modifications to the manuscript and have explained in detail certain aspects that helped improve the understanding of the paper.

# Is the topic of the review discussed comprehensively in the context of the current literature?

Yes

Are all factual statements correct and adequately supported by citations?

Yes

Is the review written in accessible language?

Yes

Are the conclusions drawn appropriate in the context of the current research literature? Yes

Competing Interests: No competing interests were disclosed.

Reviewer Expertise: Biological Control, Integrated Pest Management, Entomology, Microbiology

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

Reviewer Report 04 November 2024

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#### Mehari Desta Hawku 🕛



<sup>1</sup> Crop Sciences, University of Rwanda College of Agriculture Animal Sciences and Veterinary Medicine (Ringgold ID: 183043), Busogo, Musanze, Rwanda

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No further comments.

Is the topic of the review discussed comprehensively in the context of the current literature?

Yes

Are all factual statements correct and adequately supported by citations?

Yes

Is the review written in accessible language?

Yes

Are the conclusions drawn appropriate in the context of the current research literature? Yes

Competing Interests: No competing interests were disclosed.

**Reviewer Expertise:** My research interests encompass a broad spectrum of plant pathology including:Identification and molecular characterization of resistance or susceptible genes in plant-microbe interactions;Understanding the intricate mechanisms governinginteractions between plants and microbes;Examining the signaling pathways involved in activating plant defense responses against pathogens;Studying the molecular components that enable pathogens to invade and manipulate plant cells;Understanding the mechanisms of abiotic stress tolerance in plants

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

Author Response 04 Nov 2024

#### **Kahsay Tadesse Mawcha**

We really appreciate your time and expertise. Many thanks for approving our article.

Competing Interests: No competing interests were disclosed.

Version 1

Reviewer Report 16 October 2024

https://doi.org/10.5256/f1000research.169409.r326009

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# ? Mehari Desta Hawku 🗓

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The manuscript investigates the rising significance of microbial biopesticides as eco-friendly alternatives to traditional pesticides. It emphasizes the potential of utilizing natural organisms, such as bacteria, fungi, viruses, nematodes and protozoans, for effective pest control while reducing environmental harm. The authors provide a thorough examination of recent advancements and methodologies that enhance the effectiveness of these biopesticides. Furthermore, the review underscores the importance of integrating these biopesticides into comprehensive integrated pest management strategies. However, to offer a more comprehensive review, the following aspects should be addressed:

- 1. While the topic is broad, the literature focuses only on the role of microbial biopesticides in controlling insect pests, overlooking their role in disease and weed control
- 2. The role of recent cutting-edge technologies, such as CRISPR/Cas9 and RNA interference (RNAi), was not thoroughly discussed.
- 3. Regulatory frameworks and commercialization aspects, were not addressed.

#### **Minor comments:**

#### **Section 1: Introduction**

- 1. The introduction section lacks a comprehensive summary that clearly informs readers about the review's aims, the specific points the literature addresses, as well as the knowledge gaps and suggested future research directions. It is essential to include a concise overview outlining the main themes covered in the review. Additionally, highlighting the role of emerging technologies, such as CRISPR/Cas9 and RNA interference (RNAi) in biopesticide development, along with the potential for integrating biopesticides into sustainable agricultural practices, will provide clearer context for the reader. This summary should also emphasize the importance of interdisciplinary collaboration among researchers, policymakers, practitioners, and manufacturers to enhance the efficacy and acceptance of biopesticides in agricultural systems.
- 2. The image in Figure 1 is blurry; please consider replacing it with a clearer image and remove the word "legends". From the description of Figure 1, it appears that you have missed one image. Therefore, you can either remove the sentence, "The second part demonstrates how the five steps for implementing CSA intersect with and affect the 17 SDGs in terms of synergies and trade-offs," or consider incorporating the missing image.

#### **Section 2: Methods**

The methodology section is missing some key points, as outlined below:

- 1. It is important to clarify the specific research questions guiding the review
- 2. It's unclear how studies were selected for review. Please include specific inclusion and exclusion criteria used for selecting papers
- 3. The method does not specify the keywords or phrases used during the search process.
- 4. There's no description of how potential biases in selecting literature were mitigated, nor is there any indication of measures taken to avoid confirmation bias.
- 5. Mentioning how the quality of the papers was assessed would add rigor to the methodology
- 6. It would be helpful to specify the types of studies reviewed or considered for inclusion
- 7. Please specify the time frame of the literature reviewed
- 8. If there was any geographical focus, it would be beneficial to mention it.

# Section 3: Concept and classification of biopesticides

- 1. The second paragraph in this section is not related to the topic. Instead, this concept should be discussed under section 4.
- 2. The idea in the third paragraph should also be addressed on a separate topic as Mechanism of action and efficacy of microbial biopesticides

# Section 4: Importance of biopesticides in sustainable agriculture

- 1. The sentence 'The use of biopesticides has steadily increased by a steady 11.0% Compound Annual Growth Rate (CAGR) between 2018 and 2022.' under section 4 should be rephrased as 'The use of biopesticides has steadily increased at a Compound Annual Growth Rate (CAGR) of 11.0% between 2018 and 2022.' to avoid the redundancy of the word 'steadily' and 'steady'
- 2. Under this section the following points should be discussed in details; Environmental Safety, Reduced Chemical Dependency, Promoting Biodiversity, Compatibility with Organic Farming, and Sustainability and Resilience.

# Section 5: Role of biopesticides in integrated pest management

- 1. The different microbial biopesticides should be discussed under subsection 5.1 as a subsubsection not as a subsection.
- 2. In table 1, the role of microbial biopesticides in the IPM program is not mentioned within the table; therefore, please consider modifying the table's title.
- 3. In Table 1, I suggest adding a fifth column to list the references
- 4. In Table 1, could you clarify what is meant by 'application type'? If it refers to the method of delivery, this is not clearly specified. In addition, the mode of action is missing in several instances under the column 'Application Type and Mode of Action'. For some microbial biopesticides, important details are absent within the table. On Page 9, under Table 1, please replace 'fungal-based biopesticides' with 'viral-based biopesticides.' Overall, Table 1 lacks consistency and would benefit from clearer definitions and more complete information.

#### **Subsection 5.2: Bacterial based biopesticides**

- 1. After using the full name *Bacillus thuringiensis* (Bt), you should continue using its abbreviated form (Bt) throughout the text.
- 2. The last paragraph under sub-section 5.2 should be supported by relevant literature.
- 3. The image (particularly the text) in Figure 2 is unclear; please consider replacing it with a

clearer version. Additionally, remove the word 'legend' as well as the phrase 'The mode of action of Bt gene and Cry protein,' since this information is already specified in the figure itself

# **Subsection 5.3: Fungal biopesticides**

A citation is needed for Figure 3. Such figures are typically self-explanatory and do not require legends, so please remove the note stating, 'Legends are indicated in the figures. No additional legends.'

#### **Subsection 5.4: Viral biopesticides**

- 1. Please replace 'Baculovirus family' with Baculoviridae and do not italicize baculovirus
- 2. Would you please reconsider the first sentence of the third paragraph in subsection 5.4 to maintain clarity?
- 3. In Figure 4 (Page 12), please remove the term 'legends,' as it is unnecessary to mention since it is evident that you are describing the figure. Additionally, please ensure you provide proper citations for this figure, as you are referencing two sources: Singh (2019) and Kalamkoff (2004).
- 4. On page 13, the same legend is repeated; please consider removing it.

#### **Subsection 5.5: Nematodes as biopesticides**

1. Please add a citation after the caption of Figure 5 and remove the citation from the description. Figure 5 is somewhat blurry; please use a clearer image and consider removing the word 'legends'.

In **subsection 5.6**, please consider not italicizing the word 'septicaemia'

# Section 6: Natural enemies of pests as biopesticides

- 1. Since the primary focus of this review is on microbial biopesticides, and predators and parasitoids are classified as macrobials, I suggest removing this section.
- 2. The image (particularly the text) in Figure 6 is not clear; please consider replacing it.
- 3. Please add a citation to Figure 6.

Please add a citation to Figure 7 and consider using a clear image

#### **Subsection 8.2: Factors affecting efficacy of biopesticides**

- 1. Under this subsection, the first sentence should be modified as 'Various challenges and limitations, including environmental conditions such as temperature, humidity, and sunlight, can influence the efficacy of biopesticides during pest control'
- 2. In the second paragraph, please check the citation related to '(International Symposium on Food Safety and Control).'
- 3. The image in Figure 8 is blurry; please consider using a clearer and more visually appealing image. I recommend using photo editing software, such as Photoshop, to enhance the quality. Additionally, please remove the phrase 'own work' from the caption. Finally, ensure that the figure is either self-explanatory or includes a legend where necessary.

# Section 9. Future perspectives and research directions

Please remove subsection 9.1 from this section, as there are no other subsections following it. Additionally, avoid using a single sentence as a paragraph; instead, merge it with the preceding paragraph. In this section on future perspectives and research directions, it would be valuable to discuss in more detail the role of the cutting-edge technologies, such as CRISPR/Cas9 and RNA interference (RNAi) in pest control.

#### Section 10. Conclusion

The conclusion effectively encapsulates the key findings regarding the benefits and challenges of biopesticides. However, the sentence stating that "novel microbial strain creation with improved features, such as greater efficacy and larger target spectra, should be prioritized in future research and the use of biopesticides" may be more appropriate in a future perspectives section. This sentence emphasizes the need for ongoing research and innovation, which aligns better with a section dedicated to future research directions.

# Is the topic of the review discussed comprehensively in the context of the current literature?

Yes

Are all factual statements correct and adequately supported by citations?

Yes

Is the review written in accessible language?

Yes

Are the conclusions drawn appropriate in the context of the current research literature?

**Competing Interests:** No competing interests were disclosed.

**Reviewer Expertise:** My research interests encompass a broad spectrum of plant pathology including:Identification and molecular characterization of resistance or susceptible genes in plant-microbe interactions;Understanding the intricate mechanisms governinginteractions between plants and microbes;Examining the signaling pathways involved in activating plant defense responses against pathogens;Studying the molecular components that enable pathogens to invade and manipulate plant cells;Understanding the mechanisms of abiotic stress tolerance in plants

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard, however I have significant reservations, as outlined above.

Author Response 21 Oct 2024

#### **Kahsay Tadesse Mawcha**

Dear Chief Editor, Dear Reviewer,

We have carefully considered all of the reviewers' comments and suggestions. To address these, we have made substantial revisions to the manuscript. Below, we provide a detailed response to each specific point raised.

We believe that these revisions have significantly strengthened the manuscript and

improved its clarity, comprehensiveness, and overall quality. We are confident that the revised manuscript now meets the high standards of your journal.

Thank you again for your valuable feedback.

Sincerely,

#### **Reviewer 2**

#### **Comments and responses**

The manuscript investigates the rising significance of microbial biopesticides as eco-friendly alternatives to traditional pesticides. It emphasizes the potential of utilizing natural organisms, such as bacteria, fungi, viruses, nematodes and protozoans, for effective pest control while reducing environmental harm. The authors provide a thorough examination of recent advancements and methodologies that enhance the effectiveness of these biopesticides. Furthermore, the review underscores the importance of integrating these biopesticides into comprehensive integrated pest management strategies. However, to offer a more comprehensive review, the following aspects should be addressed:

- 1. While the topic is broad, the literature focuses only on the role of microbial biopesticides in controlling insect pests, overlooking their role in disease and weed control
- We accept the reviewer's comment. To address his comment, we have modified the title of the review to focus on Microbial biopesticides and recent advancements and methodologies
- 1. The role of recent cutting-edge technologies, such as CRISPR/Cas9 and RNA interference (RNAi), was not thoroughly discussed
- As the focus of the current review is microbial biopesticides, CRISPR/Cas9 and RNA interference (RNAi) have been discussed in detail under the emerging trends in biopesticides. We believe that there is no need to discuss them as a subheading.
- 1. Regulatory frameworks and commercialization aspects were not addressed.
- The review focuses on summarizing previous and recent studies about the
  effectiveness of biopesticides, particularly microbial products. Hence, regulatory
  frameworks and commercialization aspects are beyond the scope of the review.
  However, we have discussed a few points under the conclusion part about the
  importance of Regulatory frameworks and the need for harmonization of biopesticide
  guidelines as a future work.

We have also modified the title "Recent Advances in Biopesticide Research and Development with A Focus on Microbials"

Minor comments:

#### **Section 1: Introduction**

1. The introduction section lacks a comprehensive summary that clearly informs readers about the review's aims, the specific points the literature addresses, as well as the knowledge gaps and suggested future research directions. It is essential to include a

concise overview outlining the main themes covered in the review. Additionally, highlighting the role of emerging technologies, such as CRISPR/Cas9 and RNA interference (RNAi) in **biopesticide development**, **along with the potential for integrating biopesticides into sustainable agricultural practices**, will provide a clearer context for the reader. This summary should also emphasize the importance of interdisciplinary collaboration among researchers, policymakers, practitioners, and manufacturers to enhance the efficacy and acceptance of biopesticides in agricultural systems.

- We appreciate the suggestions given by the reviewer. However, most of the comments are out of the scope of the review content. Some of the comments, such as the role of emerging technologies, such as CRISPR/Cas9 and RNA interference (RNAi), and the potential for integrating biopesticides into sustainable agricultural practices, are highlighted in the conclusion part.
- 1. The image in Figure 1 is blurry; please consider replacing it with a clearer image and remove the word "legends". From the description of Figure 1, it appears that you have missed one image. Therefore, you can either remove the sentence, "The second part demonstrates how the five steps for implementing CSA intersect with and affect the 17 SDGs in terms of synergies and trade-offs," or consider incorporating the missing image.
- Within the figure, sections show the different linkages with climate-smart agriculture and how they are linked to the 17 SDGs. Hence, there is no missed image in this section. The figure provides information that is interlinked with each other. Climate-smart agriculture (CSA) has a strong potential to contribute to the achievement of the 17 Sustainable Development Goals (SDGs). A critical precondition for realising this scope is identifying potential synergies and trade-offs between the 3 pillars of CSA and the 5 implementation steps of CSA and the SDGs. These provide entry points for targeted CSA planning so as to enhance synergies and reduce trade-offs. CSA and the SDGs have synergies and trade-offs at different levels for each of the 3 pillars of CSA where some of the synergies & trade-offs overlap all 3 pillars of CSA for some of the SDGs, namely SDGs 2 and 6. The overlapping synergies & trade-offs for the implementation steps and SDGs are different, as only step 2 has overlapping trade-offs and synergies in SDG's, 6,9,13, and 15.
- We have also improved the paragraph and made some changes to it.

#### **Section 2: Methods**

The methodology section is missing some key points, as outlined below:

- 1. It is important to clarify the specific research questions guiding the review
- What are the current trends and advancements in the development and application of microbial biopesticides?
- What are the key challenges and limitations associated with the use of microbial biopesticides in agricultural settings?
- What are the most promising microbial agents currently being developed for use as biopesticides? Are the most important guiding questions
- 1. It's unclear how studies were selected for review. Please include specific inclusion and exclusion criteria used for selecting papers

- Only microbial (bacterial, fungal, viral, nematode, and protozoa) biopesticides were included and other types were excluded
- 1. The method does not specify the keywords or phrases used during the search process.
- Using various keywords, including biocontrol, formulations, bioprotection, climate-smart agriculture, and sustainable pest management, 197 relevant studies were found. We have already included the following phrases: The literature outcomes were categorised into the (i) classification of biopesticides, (ii) importance of biopesticides in sustainable agriculture, (iii) role of microbial biopesticides in integrated pest management, (iv) different types and formulations of microbial biopesticides, and (v) advancement and future perspectives of microbial biopesticides.
- 1. There's no description of how potential biases in selecting literature were mitigated, nor is there any indication of measures taken to avoid confirmation bias.
- To avoid biases in selecting literature, we used articles that are published in peerreviewed journals, which means available on the following search engines, Google Scholar, PubMed, Scopus (Elsevier), Web of Science, Semantic Scholar, Academia
- 1. Mentioning how the quality of the papers was assessed would add rigor to the methodology
- Most of our sources are published in peer-reviewed journals.
- It would be helpful to specify the types of studies reviewed or considered for inclusion
- We have included original research articles and reviews published in peer-reviewed journals and other relevant resources
- 1. Please specify the time frame of the literature reviewed
- Mostly, we used recently published articles (2018-2024) and very few; we have also included > 5 references, which are from 2003>.
- 1. If there were any geographical focus, it would be beneficial to mention it.
- The review is an overview of the current research and developments in microbial biopesticides. So, there are no specific geographic limitations.

# Section 3: Concept and classification of biopesticides

- 1. The second paragraph in this section is unrelated to the topic. Instead, this concept should be discussed under section 4.
- Section 3 deals with the classification/definition of biopesticides (general) by various organizations and associations. So, it is better to be as it is.
- 1. The idea in the third paragraph should also be addressed on a separate topic as Mechanism of action and efficacy of microbial biopesticides
- We accept the suggestion however, as we have discussed the mode of action and efficacy of each microbial product, this subtitle may cause a repetition.

#### Section 4: Importance of biopesticides in sustainable agriculture

- 1. The sentence 'The use of biopesticides has steadily increased by a steady 11.0% Compound Annual Growth Rate (CAGR) between 2018 and 2022.' under section 4 should be rephrased as 'The use of biopesticides has steadily increased at a Compound Annual Growth Rate (CAGR) of 11.0% between 2018 and 2022.' to avoid the redundancy of the word 'steadily' and 'steady'
- Accepted and addressed.
- 1. Under this section, the following points should be discussed in detail: Environmental

- Safety, Reduced Chemical Dependency, Promoting Biodiversity, Compatibility with Organic Farming, and Sustainability and Resilience.
- We are happy to discuss them in detail however, we may go beyond the scope of the topic!

# Section 5: Role of biopesticides in integrated pest management

- 1. The different microbial biopesticides should be discussed under subsection 5.1 as a sub-subsection, not as a subsection.
- Comment Accepted. However, we discuss the various types of biopesticides, including Biochemical, Botanical, and pheromone-based; the world limit and the number of pages may exceed the limit. So, it is better to focus on the title-i.e. microbial-based biopesticides!
- 1. In Table 1, the role of microbial biopesticides in the IPM program is not mentioned within the table; therefore, please consider modifying the table's title.
- Comment accepted, and Table 1. has been modified as "Microbial biopesticide categories, mode of action, and their use in pest management.
- 1. In Table 1, I suggest adding a fifth column to list the references
- We have included the references in line with each column.
- 1. In Table 1, could you clarify what is meant by 'application type'? If it refers to the method of delivery, this is not clearly specified. In addition, the mode of action is missing in several instances under the column 'Application Type and Mode of Action'. For some microbial biopesticides, important details are absent within the table.
- The table is self-explanatory and the title is also modified as suggested. The "application type" is NOT referring to the method. It is referring to "use". The remark column is for additional information.
- 1. On Page 9, under Table 1, please replace 'fungal-based biopesticides' with 'viral-based biopesticides.' Overall, Table 1 lacks consistency and would benefit from clearer definitions and more complete information.
- Accepted, but as the title of the table is formatted by the journal editor, it will be corrected during the production.

# **Subsection 5.2: Bacterial-based biopesticides**

- 1. After using the full name *Bacillus thuringiensis* (Bt), you should continue using its abbreviated form (Bt) throughout the text.
- Yes, but when we start a new line, we normally use the full name, that is, species and genus names.
- 1. The last paragraph under sub-section 5.2 should be supported by relevant literature.
- All paragraphs are cited. It is not clear!
- 1. The image (particularly the text) in Figure 2 is unclear; please consider replacing it with a clearer version. Additionally, remove the word 'legend' as well as the phrase 'The mode of action of Bt gene and Cry protein,' since this information is already specified in the figure itself
- Okay. Accepted, and the legend is deleted.

# **Subsection 5.3: Fungal biopesticides**

1. A citation is needed for Figure 3. Such figures are typically self-explanatory and do not require legends, so please remove the note stating, 'Legends are indicated in the figures. No additional legends.'

Okay. Accepted. The figure is our own work, and there is no need to include citations. **Subsection 5.4: Viral biopesticides** 

- 1. Please replace 'Baculovirus family' with Baculoviridae and do not italicise baculovirus
- Done!
- 1. Would you please reconsider the first sentence of the third paragraph in subsection 5.4 to maintain clarity?
- Accepted and modified as "Figure 4. shows how viral replication and infection take place and attack their target host".
- 1. In Figure 4 (Page 12), please remove the term 'legends,' as it is unnecessary to mention since it is evident that you are describing the figure. Additionally, please ensure you provide proper citations for this figure, as you are referencing two sources: Singh (2019) and Kalamkoff (2004).
- Accepted and the "Figure 4. Mode of action of viral biopesticides (Singh et al. 2019)."
   It was deleted in the text.
- 1. On page 13, the same legend is repeated; please consider removing it.
- Okay. Done!

### Subsection 5.5: Nematodes as biopesticides

- 1. Please add a citation after the caption of Figure 5 and remove the citation from the description. Figure 5 is somewhat blurry; please use a clearer image and consider removing the word 'legends'.
- Accepted. We have deleted the term "legend". However, the Figure is our own work, and there is no need to put citations.
- 1. In **subsection 5.6**, please consider not italicizing the word 'septicaemia'
- Accepted and done!

# Section 6: Natural enemies of pests as biopesticides

- 1. Since the primary focus of this review is on microbial biopesticides, and predators and parasitoids are classified as macrobials, I suggest removing this section.
- Sometimes, the definition of biopesticides may include parasitoids and predators, and their inclusion in this review enhances the broader reader of the journal.
- 1. The image (particularly the text) in Figure 6 is unclear; please consider replacing it.
- 2. Please add a citation to Figure 6.
- The figure is our own work and no need to include a citation.
- 1. Please add a citation to Figure 7 and consider using a clear image
- The figure is our own work and no need to include a citation.

#### Subsection 8.2: Factors affecting efficacy of biopesticides

- 1. Under this subsection, the first sentence should be modified as 'Various challenges and limitations, including environmental conditions such as temperature, humidity, and sunlight, can influence the efficacy of biopesticides during pest control'
- Accepted and done!
- 1. In the second paragraph, please check the citation related to '(International Symposium on Food Safety and Control).'
- o Comment accepted. We have added the full information and hyperlinked the details.

# https://conferences.iaea.org/event/351/

- 1. The image in Figure 8 is blurry; please consider using a clearer and more visually appealing image. I recommend using photo editing software, such as Photoshop, to enhance the quality. Additionally, please remove the phrase 'own work' from the caption. Finally, ensure that the figure is either self-explanatory or includes a legend where necessary.
- Accepted and done!

# Section 9. Future perspectives and research directions

1. Emerging trends in biopesticide development

Please remove subsection 9.1 from this section, as there are no other subsections following it. Additionally, avoid using a single sentence as a paragraph; instead, merge it with the preceding paragraph. In this section on future perspectives and research directions, it would be valuable to discuss in more detail the role of cutting-edge technologies, such as CRISPR/Cas9 and RNA interference (RNAi), in pest control.

# Accepted and addressed!

By combining CRISPR/Cas9 and RNAi, researchers can develop microbial biopesticides with enhanced specificity, efficacy, and environmental safety. These technologies offer exciting possibilities for the future of pest control and sustainable agriculture.

Additionally, we have added "Future perspectives and research directions in the field of microbial biopesticides, including key research directions include Genetic engineering to improve microbial agents, synergistic interactions with other pest control methods, advanced formulation and delivery techniques, resistance management strategies, environmental impact assessments, integration into sustainable agriculture, and public awareness and adoption are essential for advancing their development and application. The future of microbial biopesticides looks promising, with ongoing research focusing on enhancing their efficacy, specificity, and environmental safety".

### **Section 10. Conclusion**

- 1. The conclusion effectively encapsulates the key findings regarding the benefits and challenges of biopesticides. However, the sentence stating that "novel microbial strain creation with improved features, such as greater efficacy and larger target spectra, should be prioritized in future research and the use of biopesticides" may be more appropriate in a future perspectives section. This sentence emphasizes the need for ongoing research and innovation, which aligns better with a section dedicated to future research directions.
- Yes but the conclusion part also needs to show the future perspectives in the area, so we keep this section as it is.

Many thanks for the time you took to improve the paper. Sincerely,

Kahsay Tadesse

**Competing Interests:** No competing interests were disclosed.

Reviewer Report 04 October 2024

# https://doi.org/10.5256/f1000research.169409.r326005

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# 👔 Yordanys Ramos 🗓

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- <sup>4</sup> Instituto de Investigaciones Agropecuarias y Forestales, Universidad Michoacana de San Nicolas de Hidalgo, Morelia, Michoacán, Mexico
- <sup>5</sup> Instituto de Investigaciones Agropecuarias y Forestales, Universidad Michoacana de San Nicolas de Hidalgo, Morelia, Michoacán, Mexico

Introduction section, Page 3, last paragraph. Change *Beauveria bassiana* to *Beauveria spp.* since there are many more entomopathogenic species within the *Beauveria* genus, not just *Beauveria bassiana*. It would be more accurate to use the genus (*Beauveria spp.*), just as was done with *Metarhizium*, *Isaria*, and *Verticillium spp*.

In that same paragraph, the authors should change *Verticillium* to *Lecanicillium* according to the current classification of that fungal genus.

Page 5. Concepts and classifications of biopesticides. The authors should provide a citation for the concept of biopesticide provided by SADC, just as they did for the concept provided by USEPA. Likewise, they should include a citation for the concept provided by The European Environmental Agency.

Page 5: Please provide a citation for the Paris Agreement and the Nagoya Protocol. The authors can find both documents at these links:

https://unfccc.int/sites/default/files/english\_paris\_agreement.pdf https://www.cbd.int/abs/doc/protocol/nagoya-protocol-en.pdf

Page 6: Biopesticide Categories. The authors do not need to repeat the concept of biopesticide again, as this makes the review repetitive and less engaging for the reader, especially considering that a section was already dedicated to detailing the various existing concepts of this term.

Page 11: Figure 3, Mode of Action of Entomopathogenic Fungi. The cycle refers to the infection of *Beauveria bassiana*. Therefore, it would not be prudent to include a photo of an insect killed by *Metarhizium anisopliae* outside of the cycle. Additionally, it is quite confusing that the spore colors are shown as white in both cases. This is accurate for *Beauveria bassiana*, but when *Metarhizium* 

anisopliae causes mycosis in an insect, its spores are olive green.

In the section on entomopathogenic fungi as bioinsecticides, the authors should briefly mention that entomopathogenic fungi also have the ability to act as endophytes in plants and crops, and that by producing their secondary metabolites, they protect the plants from insect herbivory. This primarily includes fungi like *Beauveria bassiana* and *Metarhizium anisopliae*. This is also a bioinsecticidal activity of these microorganisms.

Page 13: What are the optimal conditions for the baculoviruses to cause rapid death within 3 to 7 days? This was left ambiguous, so it needs to be clarified.

Page 17: the numbering of the sections has been lost; it goes from 7.2.2 to 7.3.2. In any case, it should be 7.3. Please review.

# Is the topic of the review discussed comprehensively in the context of the current literature?

Partly

Are all factual statements correct and adequately supported by citations? Yes

Is the review written in accessible language?

Yes

Are the conclusions drawn appropriate in the context of the current research literature? Yes

**Competing Interests:** No competing interests were disclosed.

Reviewer Expertise: Biological Control, Integrated Pest Management, Entomology, Microbiology

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard, however I have significant reservations, as outlined above.

Author Response 11 Oct 2024

#### **Kahsay Tadesse Mawcha**

Thank you very much for the careful review of our manuscript entitled "Recent Advances in Biopesticide Research and Development: A Focus on Microbial: A Review. We appreciate your time and insightful comments on our manuscript.

We have addressed all the comments and suggestions forwarded from the reviewer. We made significant changes and addressed each point by point as follows:

Regrads,

Kahsay

#### Introduction section,

Page 3, last paragraph. Change Beauveria bassiana to Beauveria spp. since there are many more entomopathogenic species within the Beauveria genus, not just Beauveria bassiana. It would be more accurate to use the genus (Beauveria spp.), just as was done with Metarhizium, Isaria, and Verticillium spp. In that same paragraph, the authors should change Verticillium to Lecanicillium according to the current classification of that fungal genus.

= We want to mention the specific Beauveria bassiana. However, we agreed with the reviewer's suggestion and will improve the text to "Metarhizium, Isaria, and Lecanicillium spp."

Page 5. Concepts and classifications of biopesticides. The authors should provide a citation for the idea of biopesticide provided by SADC, just as they did for the concept provided by USEPA. Likewise, they should include a citation for the idea provided by The European Environmental Agency.

European Environment Agency, 2023. How pesticides impact human health and ecosystems in Europe EEA Briefings 06 (2023)

https://www.sciencedirect.com/science/article/pii/S0269749124005505#bib27

= For the SADC, we provided a link to the reference. We can also give the reference as ....(SADC 2019)

SADC Secretariat 2019. SADC (Southern African Development Community) Guidelines for Pesticides Management and Risk Reduction. http://faolex.fao.org/docs/pdf/sad212166.pdf

Page 5: Please provide a citation for the Paris Agreement and the Nagoya Protocol. The authors can find both documents at these links:

https://unfccc.int/sites/default/files/english\_paris\_agreement.pdf https://www.cbd.int/abs/doc/protocol/nagoya-protocol-en.pdf

<u>= We appreciate the suggestions. We include the citations for "</u>The Paris Agreement (United Nations 2015) which is available on the following link.

https://unfccc.int/sites/default/files/english\_paris\_agreement.pdf

<u>For</u> the Nagoya Protocol 2014. "The Nagoya Protocol on Access to Genetic Resources and The Fair and Equitable Sharing of Benefits Arising from Their Utilization to the Convention on Biological Diversity" <a href="https://www.cbd.int/abs/doc/protocol/nagoya-protocol-en.pdf">https://www.cbd.int/abs/doc/protocol/nagoya-protocol-en.pdf</a> Generally, both references can be used in the main text but we may not include them in the reference list as they are not peer-reviewed documents.

Page 6: Biopesticide Categories. The authors do not need to repeat the concept of biopesticide again, as this makes the review repetitive and less engaging for the reader, especially considering that a section was already dedicated to detailing the various existing concepts of this term.

= It is not a repetition. The first one is to show the concepts depending on EPA, EAC, The European Environmental Agency, the International Biocontrol Manufacturer's Association (IBMA), the International Organization for Biological Control (IOBC), and SADC. The purpose of discussing on page 6 is to demonstrate the different categories

#### of microbial biopesticides NOT all classifications of Biopesticides.

Page 11: Figure 3, Mode of Action of Entomopathogenic Fungi. The cycle refers to the infection of *Beauveria bassiana*. Therefore, it would not be prudent to include a photo of an insect killed by *Metarhizium anisopliae* outside of the cycle. Additionally, it is quite confusing that the spore colors are shown as white in both cases. This is accurate for *Beauveria bassiana*, but when *Metarhizium anisopliae* causes mycosis in an insect, its spores are olive green.

= The color of the spore doesn't matter when it comes to the dead insect, as it doesn't refer to spores when it's killed. The picture aims to illustrate that when microbial biopesticides are used, the target pest being killed is part of the cycle. This is just how it works, and the spores may produce different colors. The purpose of the picture is to demonstrate the mode of action of the entomopathogenic fungi.

In the section on entomopathogenic fungi as bioinsecticides, the authors should briefly mention that entomopathogenic fungi also have the ability to act as endophytes in plants and crops, and that by producing their secondary metabolites, they protect the plants from insect herbivory. This primarily includes fungi like *Beauveria bassiana* and *Metarhizium anisopliae*. This is also a bioinsecticidal activity of these microorganisms.

= We have mentioned Beauveria bassiana and Metarhizium anisopliae throughout the text. To avoid repetitions in the review, it is not necessary to include everything in this text. But, if we don't exceed the page number, we can include the text suggested by the reviewer.

Page 13: What are the optimal conditions for the baculoviruses to cause rapid death within 3 to 7 days? This was left ambiguous, so it needs to be clarified.

=The optimal conditions include optimum temperature and ultraviolet radiation. These have already been mentioned <u>under the advantages of viral biopesticides</u>. The common limitations of all viral products.

Page 17: the numbering of the sections has been lost; it goes from 7.2.2 to 7.3.2. In any case, it should be 7.3. Please review.

=Yes. Accepted. The subtitle "7.3.2 Formulation of fungal biopesticides" should be "7.2.3 Formulation of fungal biopesticides"

Many thanks for your time and consideration Sincerely, Kahsay Tadesse

Competing Interests: No

# Comments on this article

Version 1

Reader Comment 19 Sep 2024

**Moussa kassim ICGEB**, idm, International Centre for Genetic Engineering and Biotechnology, cap town, South Africa

Congratulations to the authors

**Competing Interests:** No competing interests were disclosed.

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