## Carnegie Mellon University

# Midterm Presentation for the Merck Capstone Project

#### **Protein Folding**

Joon Jung, Britney Wang, Fangzhou Yuan

# Agenda

Background information - Protein Folding, Tools available
Scholarly articles research
Installation of Protein Folding Tools on Bridges-2 PSC
Summary of "AlphaFold2 can predict single-mutation effects"

# What is Protein Folding?

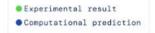
- Process where linear sequence of amino acids folds into structure
- Crucial for its functionality: Correct folding of protein is crucial for its functionality
  - Enzymes binding to substrates, signaling molecules to receptors
  - Neurodegenerative diseases : Alzheimer's, Parkinson's disease
- Important to understand and be able to predict correctly
  - Biological function, disease mechanism, drug design





T1037 / 6vr4 90.7 GDT (RNA polymerase domain)

T1049 / 6y4f 93.3 GDT (adhesin tip)



### Protein Prediction Tasks

- **Structure Prediction:** prediction of how the backbone and side chains fold in 3D space.
  - Primary, Secondary, Tertiary, Quaternary
- Prediction of Protein Disorder Regions: some regions of proteins (eg: RNA) are intrinsically disordered or highly flexible, which can be critical for their function.
- Antibody-antigen Structure Prediction
- Effects of Mutations Prediction: how changes in the amino acid sequence (due to mutations) can affect the overall structure and function of the protein.

# Why is it so hard?

- Complexity of Protein folding process: Highly complex process that involves numerous interatomic interactions
  - Hydrogen bonds, hydrophobic interactions, van der Waals force, etc
- Limited Experimental data: Collecting data through experiments are time-consuming and expensive.
- Conformational Flexibility: Proteins adopt into multiple conformations under different environmental conditions or with other molecules
- Computational Complexity: complexity of prediction increases exponentially as the sequence gets long and size of protein increases (degrees of freedom, local minima)

### Model Comparison (Research)

- AlphaFold2 (DeepMind): network-based model, relies on Multiple Sequence Alignments (MSAs)
  - Winner of CASP13 (2018), CASP14 (2020)
  - High accuracy, got a GDT score above 90
- RoseTTAFold (UW): inspired by AF2, but different similar NN architecture
  - RMSD score is comparable to AF2's
- ESMFold (Meta AI): large-scale language based model
  - Requires only a single input sequence
  - Faster prediction speed, but lower accuracy
  - Predict the structures of orphan proteins with higher accuracy than AF
- OmegaFold (Helixon): DL-based method that uses only single primary sequence and no MSAs
  - better suited for proteins that have low sequence coverage

### Model Evaluation Metrics

#### **Overall Structural Accuracy**

- GDT\_TS and GDT\_HA: overall shape and fold of a protein (Used in CASP)
- TM-score: overall topological similarity rather than specific atomic positions

#### **Local Structural Accuracy**

• **pIDDT:** confidence in the accuracy of local structural features

#### **Backbone Conformation**

RMSD: average distance between the atoms (usually the backbone atoms)

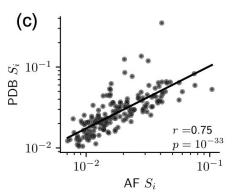
### Evaluation on Mutations Effects Prediction

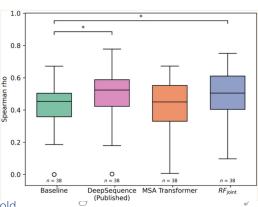
#### **Metrics**

- pLDDT insufficient to sort folding from nonfolding proteins
- **Effective Strain(ES)**: measure local deformation more robust measure of structural change upon mutation.
- Log odds ratio: how often the mutant (altered) amino acid appears at a specific position in a set of related proteins

#### **Model Results**

- AlphaFold2
- RoseTTAFold Joint (RFjoint )





# Installing Folding Models

### **Bridges-2 (thanks to CMU and PSC)**

- Storage (~1TB for our current database, up to 2.6TB for the full database)
- GPU

# Installing Folding Models

### AlphaFold2 (AlphaFold Non-Docker)

- AlphaFold release v2.3.1
- OpenMM patches
- Small genetic database (instead of the full database)
- Docker restrictions -> Singularity containers -> AlphaFold non-Docker
- Environment has been set up
- Errors in prediction phase
  - Reduced database/test sequence issues
  - o cudatoolkit and nvidia cuda driver version compatibility issues

# Installing Folding Models

### OmegaFold (OmegaFold v1.1.0)

- Fully operational
- A single .fasta file -> A list of .pdb files

### **ESMFold** (**ESM-2 v1.0.3**)

- Half way in environment setup
- Alternative implementations: ColabFold and ESM Metagenomic Atlas

## Next step

- AlphaFold
  - Debugging (possible source of error: cudatoolkit compatibility issues)
  - Running test sequences
- OmegaFold
  - Running test sequences
- ESMFold & RoseTTAFold2
  - Finishing the setup

## Paper summary

### "AlphaFold2 can predict single-mutation effects"

- Comparing AF predictions with curated set of proteins from PDB
  - "AF can detect the effect of mutation on structure by identifying local deformations between protein pairs differing by 1-3 mutations.
  - Recent evidence suggests that AF learns the energy functional underlying folding
- "AF can predict local structural change."
  - wild-type(WT 6BDD\_A) and single mutant (6BDE\_A) structures of H-NOX protein
  - Metrics used(local deformation): Effective strain (ES) per residue Si, and phenotype change (no consistent correlation with RMSD, pLDDT)
- "AF predicts structure, not folding"
  - They emphasize that AF is only trained to predict structures of stable proteins, and no claim whether protein will indeed fold into predicted structure

# Paper summary

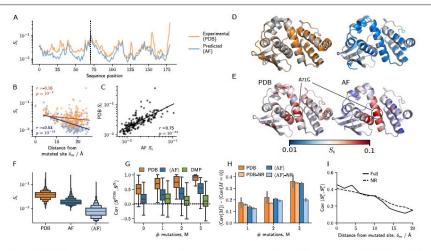


FIG. 1. A: Local deformation per residue measured by effective strain,  $S_i$ , between wild-type (WT) and mutant (A71G) H-NOX protein, for experimental (orange) and AF-predicted (blue) structures. Dotted line indicates the mutated residue. B:  $S_i$  vs distance from the nearest mutated site,  $\delta_m$ . C: Comparison of  $S_i$  obtained from experimental and predicted structures. D: Overlaid WT (grey, 68DD\_A) and mutant (colour, 68DE\_A), experimental (orange) and predicted (blue) structures. E: Wild type protein with residues coloured by  $S_i$ ; location of A71G mutation is shown. F: Distribution of  $S_i$  between matched pairs of structures with the same sequence (M = 0), for PDB, AF, and averaged AF ( $\langle AF \rangle$ ) structures. G: Distribution of correlation between PDB strain fields and equivalent fields from PDB, AF and DMPfold, shown for different numbers of mutations, M. H: Residual correlation that is due to mutations, shown for the full dataset and a non-redundant version (NR); whiskers show bootstrapped 95 % confidence intervals. I: Correlation between PDB and  $\langle AF \rangle$  strain fields,  $S_i^P$ , across all pairs P and residues i that are within a distance  $\delta_m$  from a mutated site, shown for the full dataset and a non-redundant version (NR).



#### **Questions?**

We want to thank CMU, PSC, and Merck.co team for the resources and time to allow us do this capstone project.