Data_visualization

Abstract

Pollinator sharing can have negative consequences for species fitness with the arrival of foreign pollen. However, the costs of heterospecific pollen are not yet well understood. For this reason, we have conducted a glasshouse experiment where we try to understand how phylogenetic relatedness and the different traits of these species are involved in this process. We experimentally crossed 10 species belonging to three different families: Brassicaceae, Solanaceae and Convolvulaceae. Overall, more than 4000 crosses were done and seed set and pollen tubes were considered as proxy of effect. We found that for all species foreign pollen (50% or less) reduced seed set. Moreover, the seed set reduction is not dependent on the degree of relatedness of the pollen donor. However, the effect is governed by the degree of relatedness and the traits of the species recipient. Our results show that the outcome of heterospecific pollen deposition is determined in greater degree by the traits of the pollen recipient than the pollen donor and that certain traits such as compatibility system are crucial to understand the costs of heterospecific pollen.

Figure 1. Seed set of Solanaceae species with the different pollination treatments (N=10) A) Petunia, B) Capsicum, C) Tomato, D) Eggplant. In total 9 different crosses with 50% heterospecific pollen appear from left to right. The focal treatment with itself is shown in blank to maintain simmetry among pannels. Moreover, for each species are also shown in grey hand cross pollination, hand self pollination, control (bagged emasculated flowers) and flower control (bagged flowers).
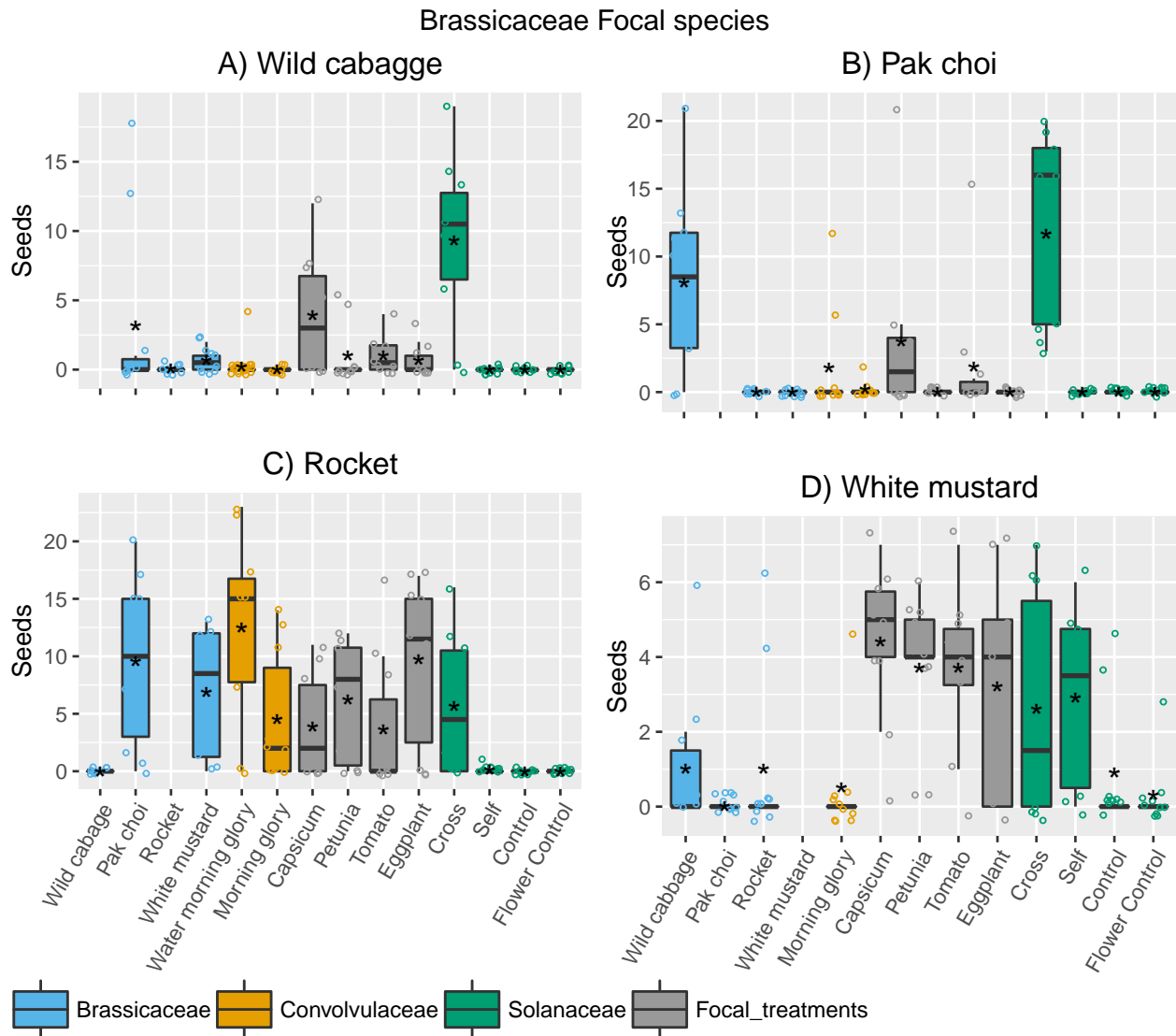
Figure 2. Seed set of Brassicaceae species with the different pollination treatments (N=10) A) Wild cabbage, B) Pak choi, C) Rocket, D) White mustard. In total 9 different crosses with 50% heterospecific pollen appear from left to right. The focal treatment with itself is shown in blank to maintain simmetry among pannels. Moreover, for each species are also shown in grey hand cross pollination, hand self pollination, control (bagged emasculated flowers) and flower control (bagged flowers).
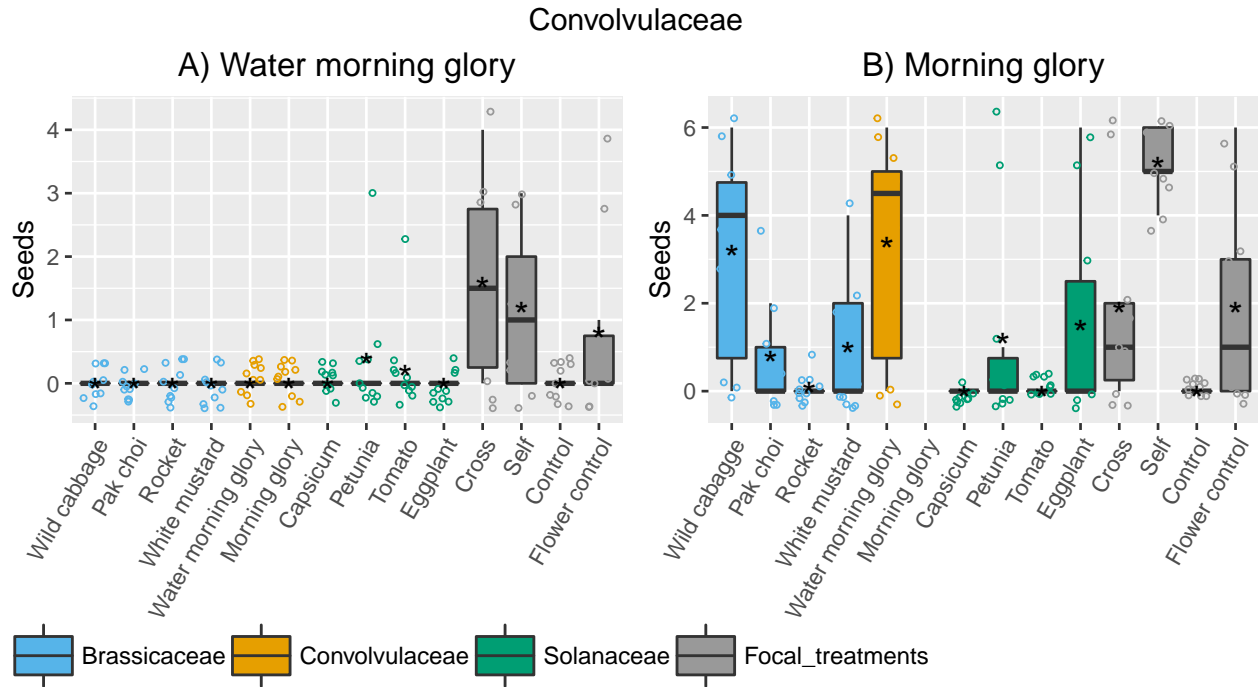
Figure 3. Seed set of Convolvulaceae species with the different pollination treatments (N=10) A) Water morning glory, B) Morning glory. In total 9 different crosses with 50% heterospecific pollen appear from left to right. The focal treatment with itself is shown in blank to maintain simmetry among pannels. Moreover, for each species are also shown in grey hand cross pollination, hand self pollination, control (bagged emasculated flowers) and flower control (bagged flowers).
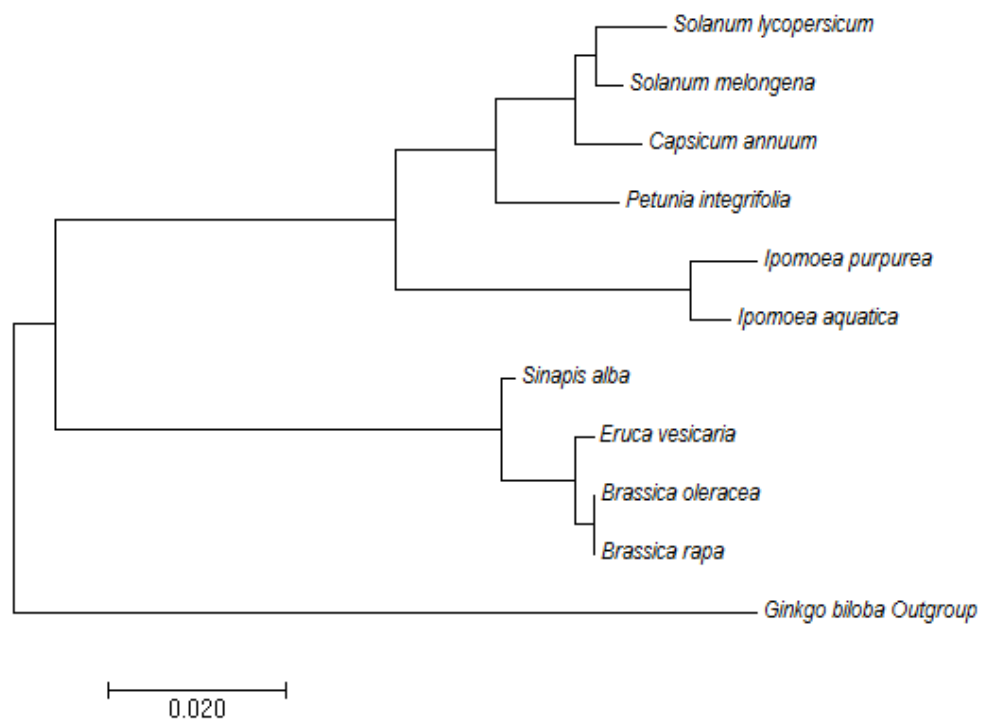
CHOLOROPLAST RBCL PHYLOGENY

Figure 4. Evolutionary distances of the ten species used in the experiment using maximum composite likelihood method.
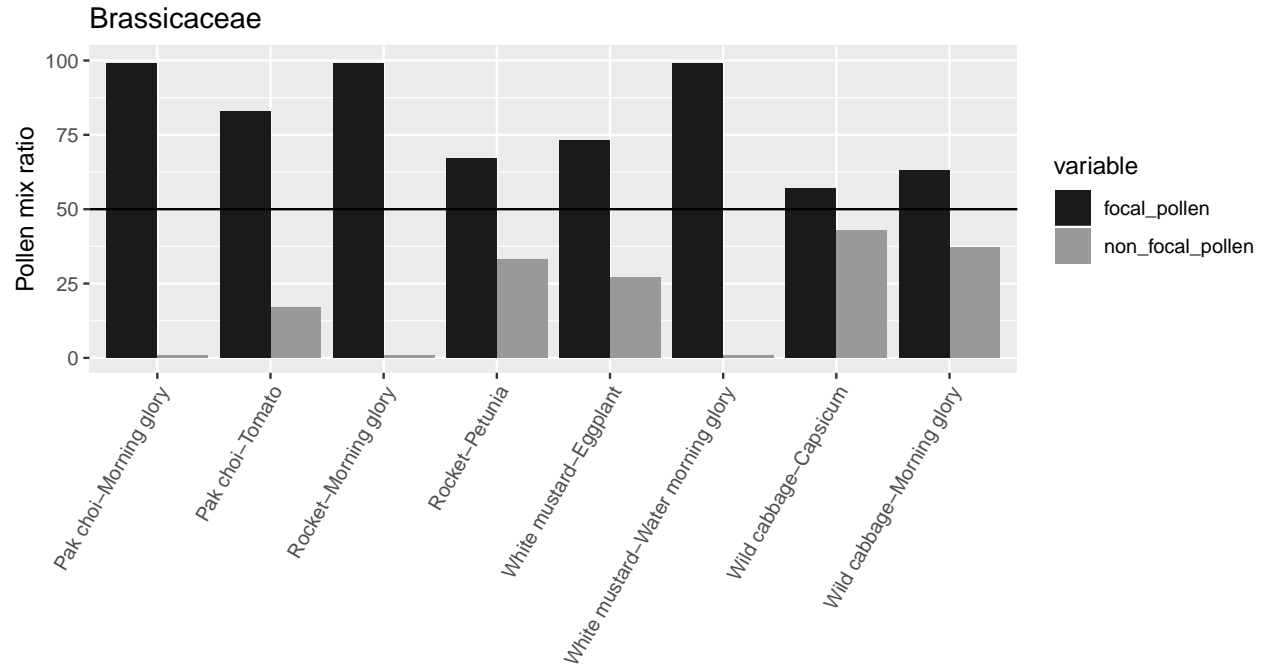
Brassicaceae

Figure 5. Percentage of pollen deposited on the stigma of a subset of species of the Brassicaceae family with pollen mixes with the other two families (N=3). The black line represents the theoretical 50% pollen percentage of our mixes. The sample that was in ethanol was dried in a heater (few days) and then the solution was filled with sodium hydroxide to degradate the tissue. Once the tissue was soft, all the solution plus the tissue was placed on a grid and the whole quantity of the microtube was counted.
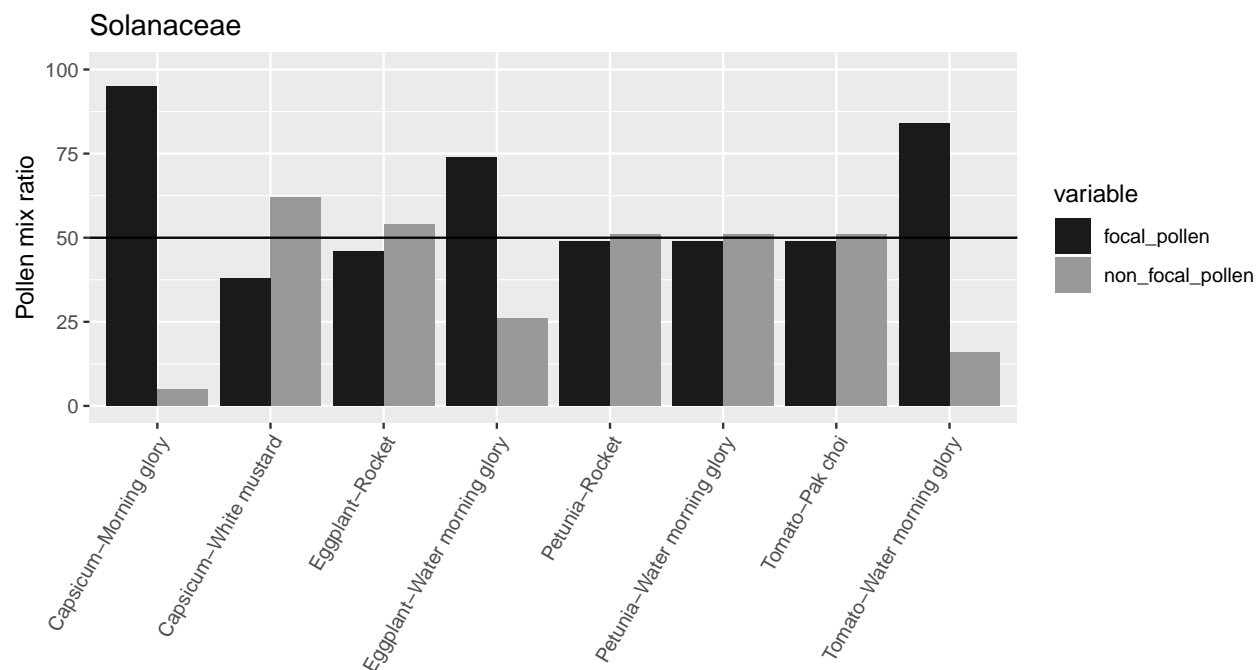
Figure 6. Percentage of pollen deposited on the stigma of a subset of species of the Solanaceae family with pollen mixes with the other two families (N=3). The black line represents the theoretical 50% pollen percentage of our mixes. The sample that was in ethanol was dried in a heater (few days) and then the solution was filled with sodium hydroxide to degradate the tissue. Once the tissue was soft, all the solution plus the tissue was placed on a grid and the whole quantity of the microtube was counted.
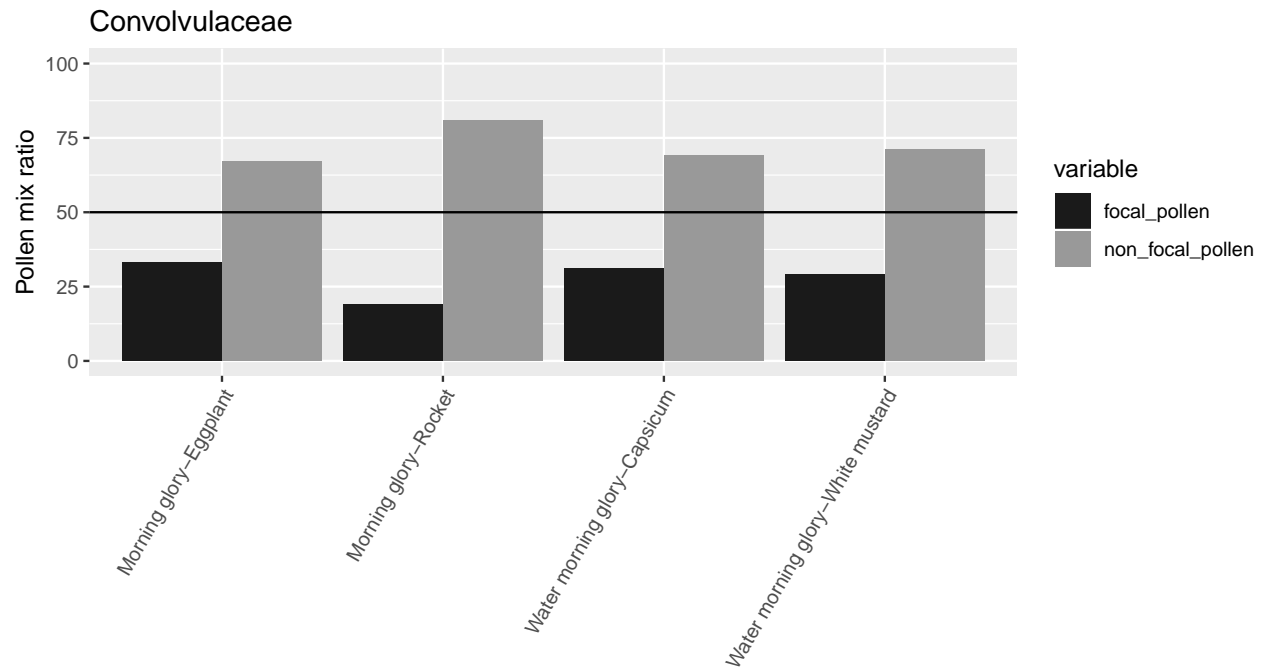
Figure 7. Percentage of pollen deposited on the stigma of a subset of species of the Convolvulaceae family with pollen mixes with the other two families (N=3). The black line represents the theoretical 50% pollen percentage of our mixes. The sample that was in ethanol was dried in a heater (few days) and then the solution was filled with sodium hydroxide to degradate the tissue. Once the tissue was soft, all the solution plus the tissue was placed on a grid and the whole quantity of the microtube was counted.
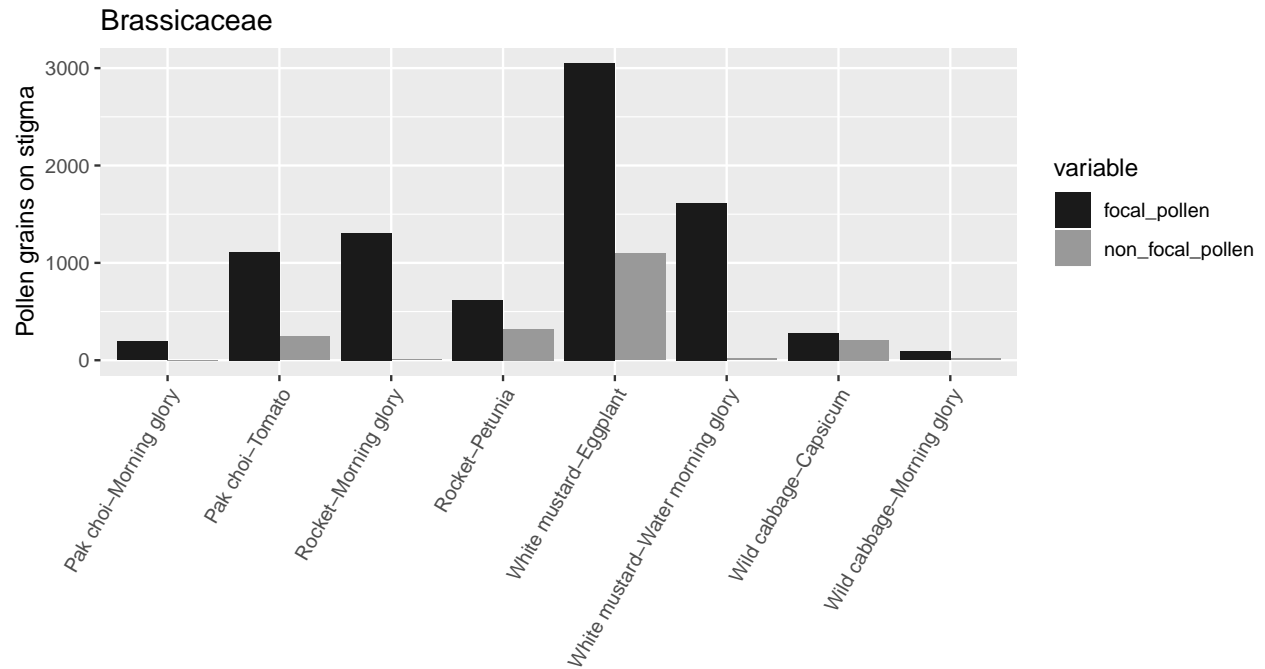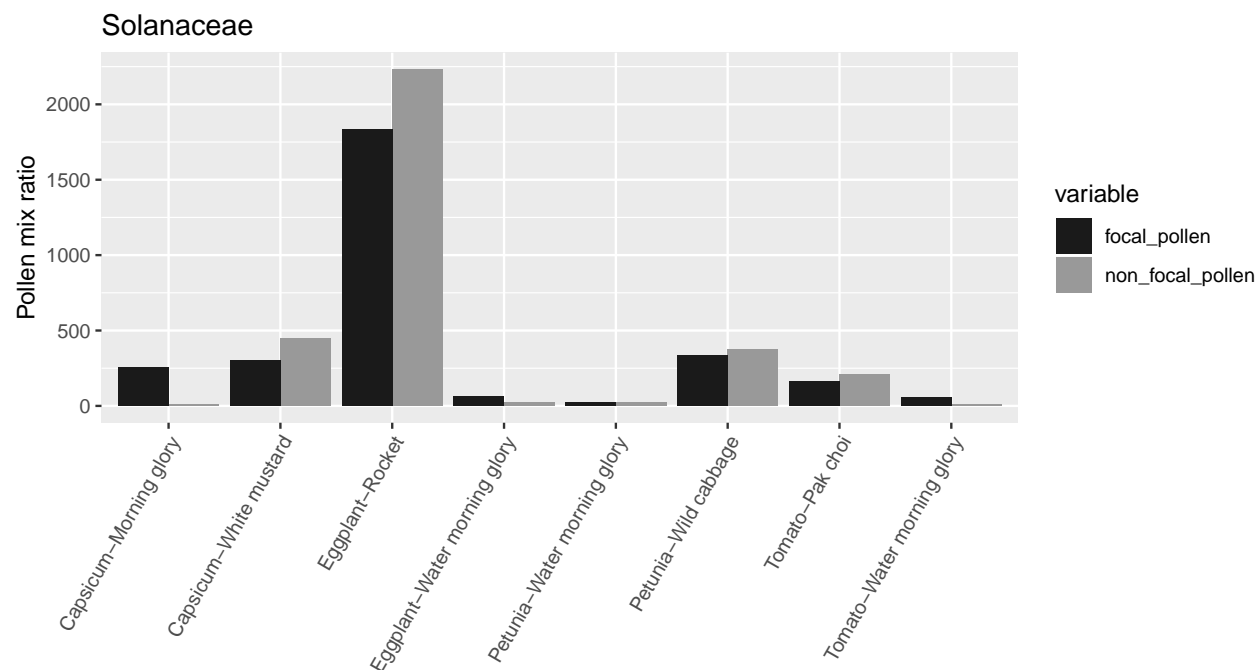
TOTAL POLLEN ON STIGMA -NEXT 3 PAGES-

Figure 8. Total pollen deposited on the stigma of a subset of species of the Brassicaceae family with pollen mixes with the other two families (N=3). The sample that was in ethanol was dried in a heater (few days) and then the solution was filled with sodium hydroxide to degradate the tissue. Once the tissue was soft, all the solution plus the tissue was placed on a grid and the whole quantity of the microtube was counted.
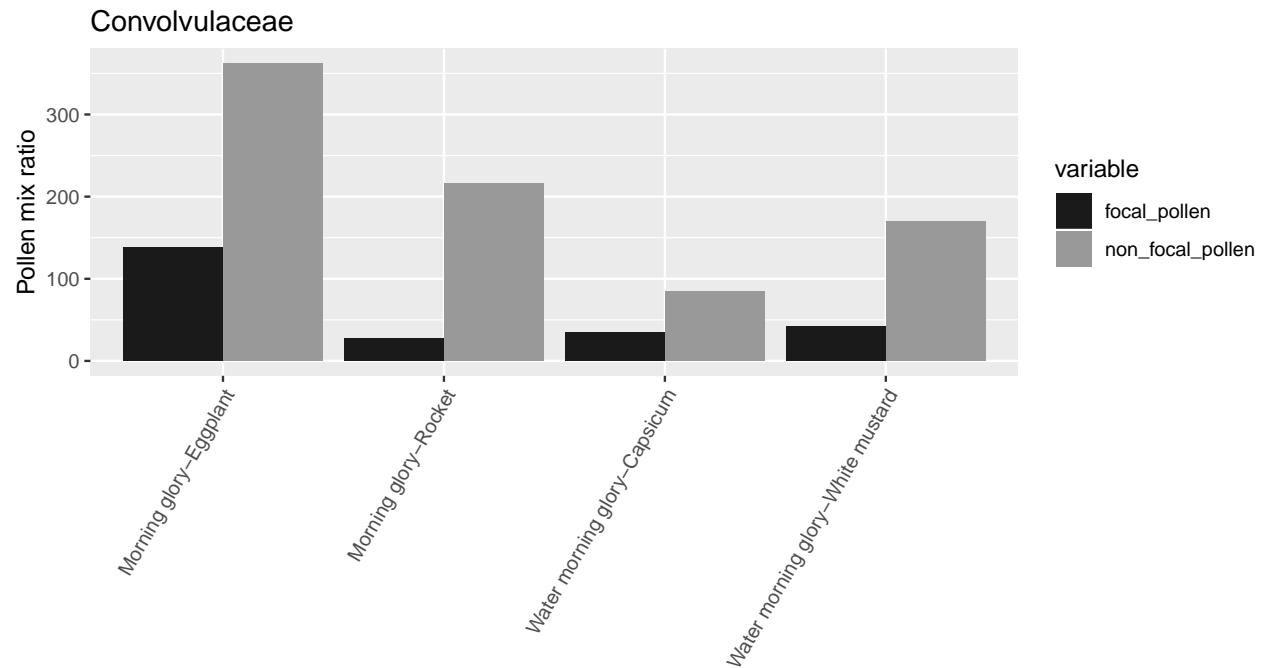
Figure 9. Total pollen deposited on the stigma of a subset of species of the Solanaceae family with pollen mixes with the other two families (N=3). The sample that was in ethanol was dried in a heater (few days) and then the solution was filled with sodium hydroxide to degradate the tissue. Once the tissue was soft, all the solution plus the tissue was placed on a grid and the whole quantity of the microtube was counted.

Convolvulaceae

Figure 10. Total pollen deposited on the stigma of a subset of species of the Convolvulaceae family with pollen mixes with the other two families (N=3). The sample that was in ethanol was dried in a heater (few days) and then the solution was filled with sodium hydroxide to degradate the tissue. Once the tissue was soft, all the solution plus the tissue was placed on a grid and the whole quantity of the microtube was counted.
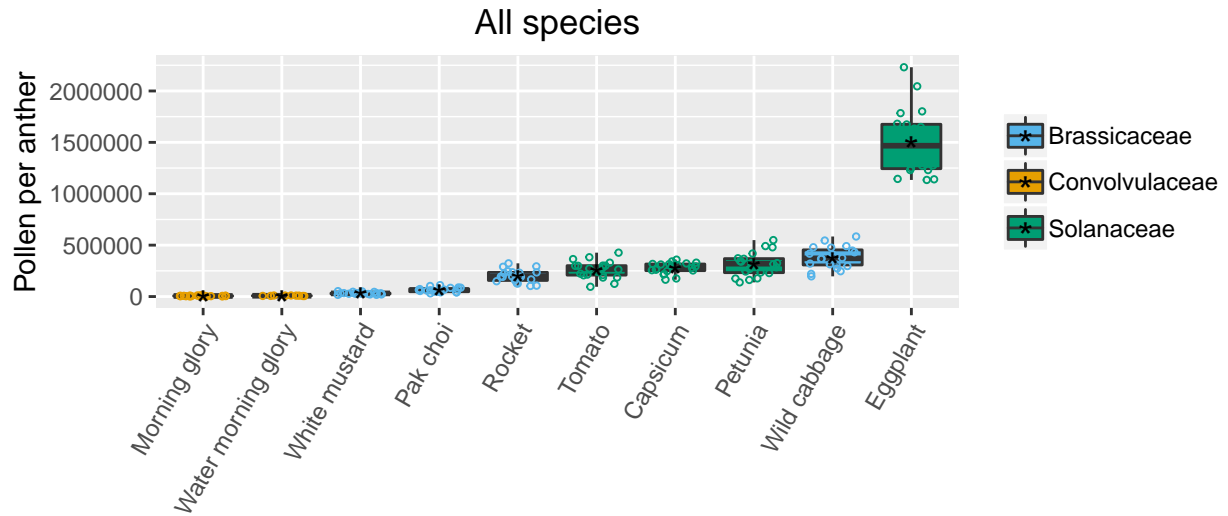
All species

Figure 11. Pollen per anther of the ten species used in our artificial community belonging to three different families, Brassicaceae, Solanaceae and Convolvulaceae (N=20, I.aquatica N=10). Species are coloured by family. Pollen was counted with an hemocytometer.
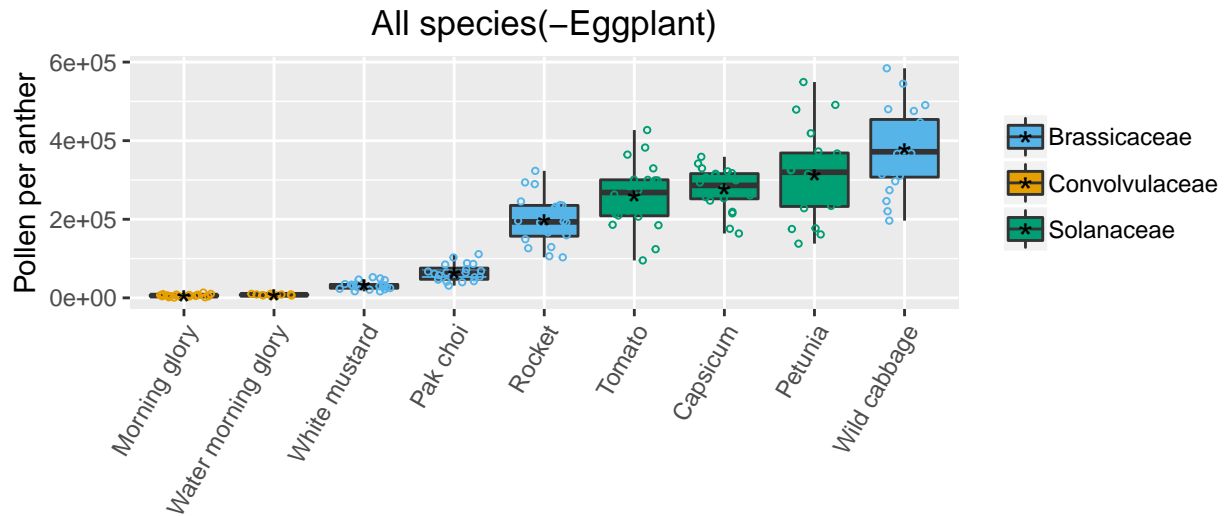
Figure 12. Pollen per anther of the species used in our artificial community belonging to three different families, Brassicaceae, Solanaceae and Convolvulaceae (N=20, I.aquatica N=10). Eggplant excluded in order to improve data visualization. Species are coloured by family. Pollen was counted with an hemocytometer.
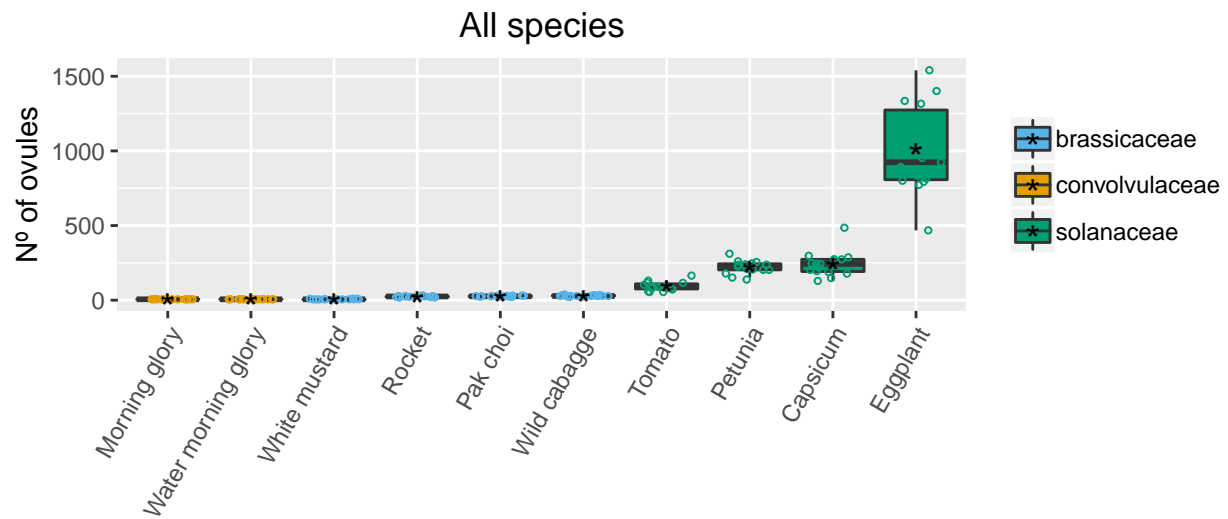
Figure 13. Number of ovules of the ten species (N=15). Species are coloured by family.
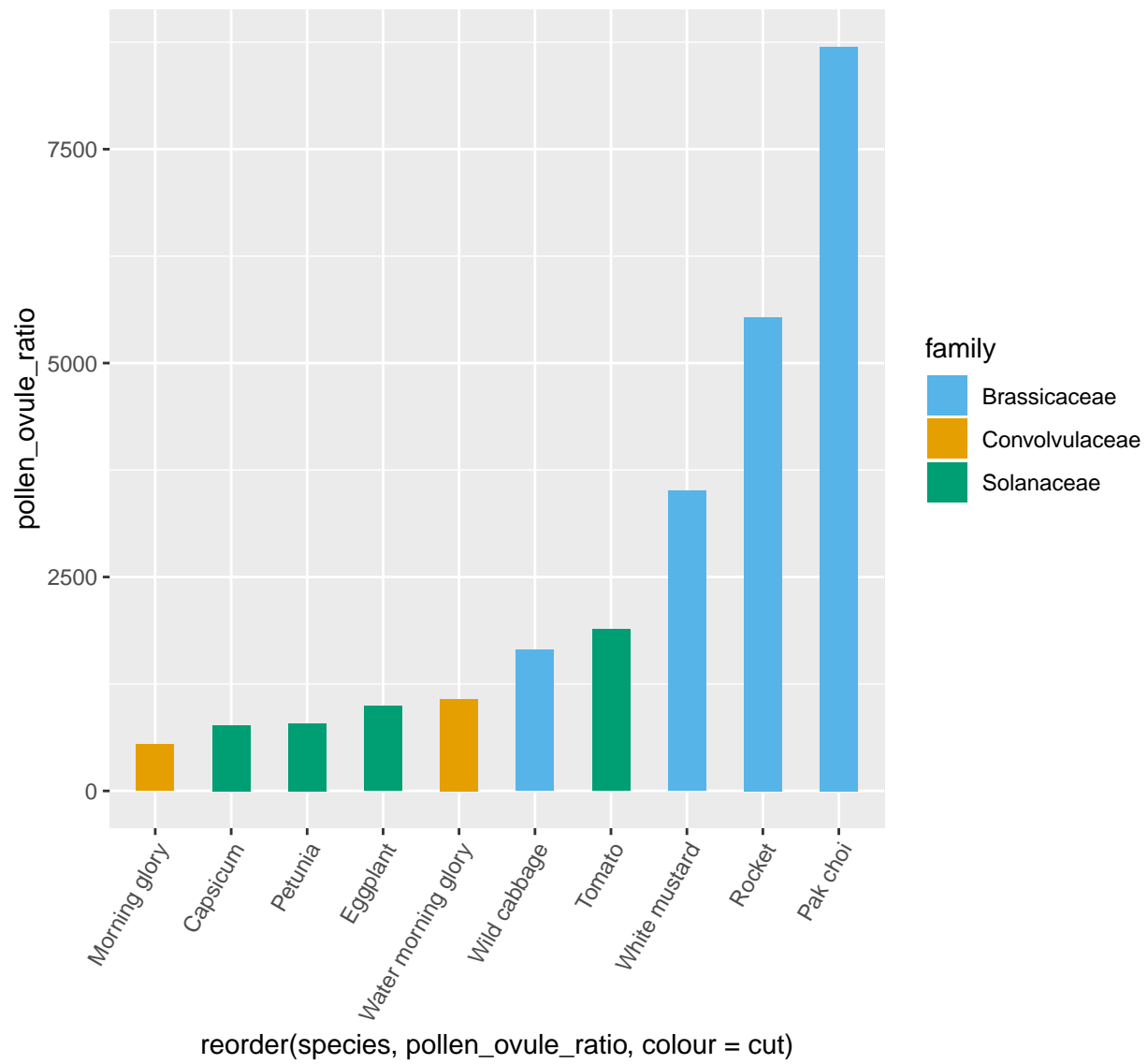
Figure 14. Pollen ovule ratio for all the species. Total pollen per flower was estimated by multiplying by the total number of anthers of the flower. Species are coloured by family.
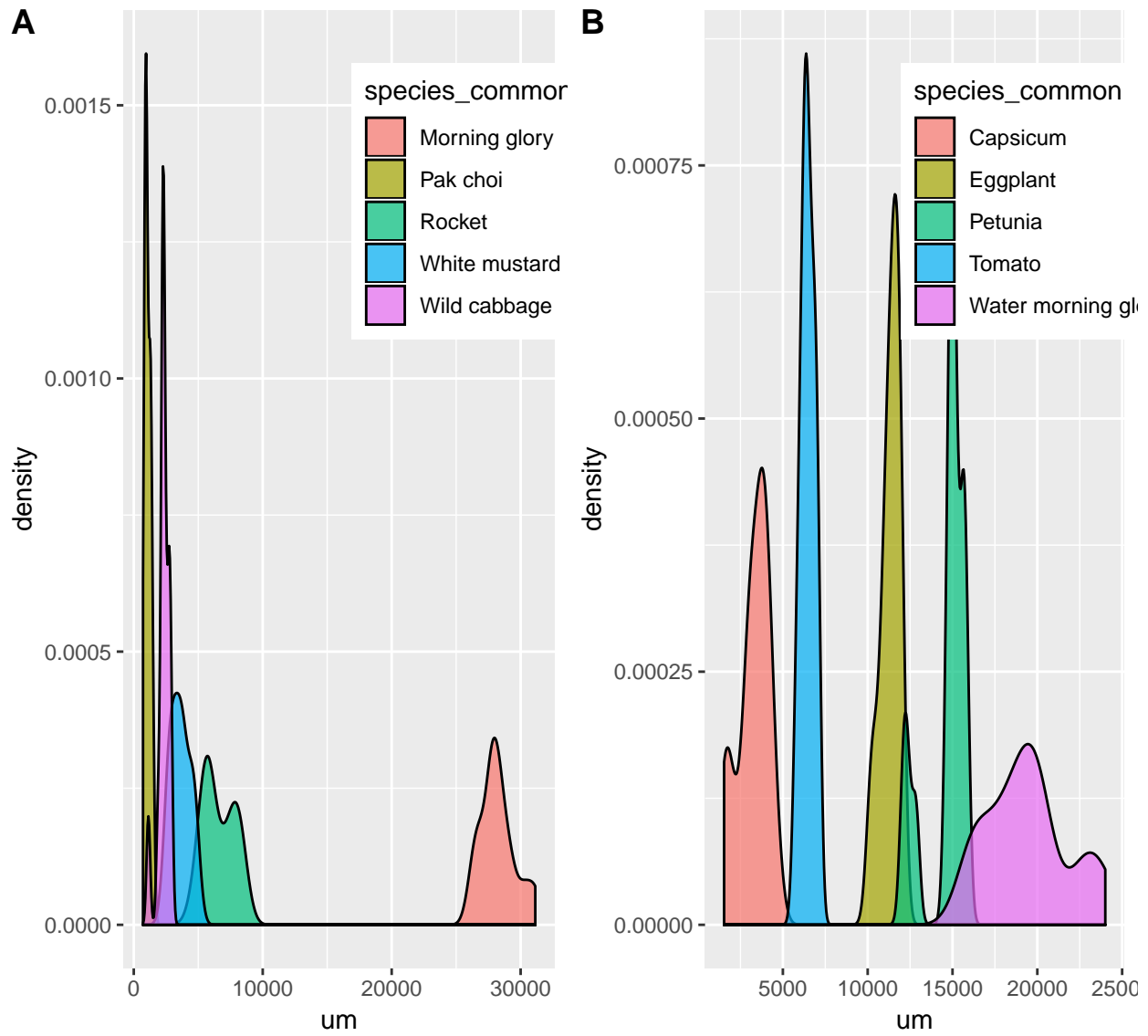
Figure 15. Density plot of the stigmatic area. The measurements were conducted with a stereo microscope. Species are divided in two pannels of 5 species each (4 Brassicaceae + 1 Convolvulaceae and 4 Solanaceae + 1 Convolvulaceae). The x axis shows the area in square micrometers.
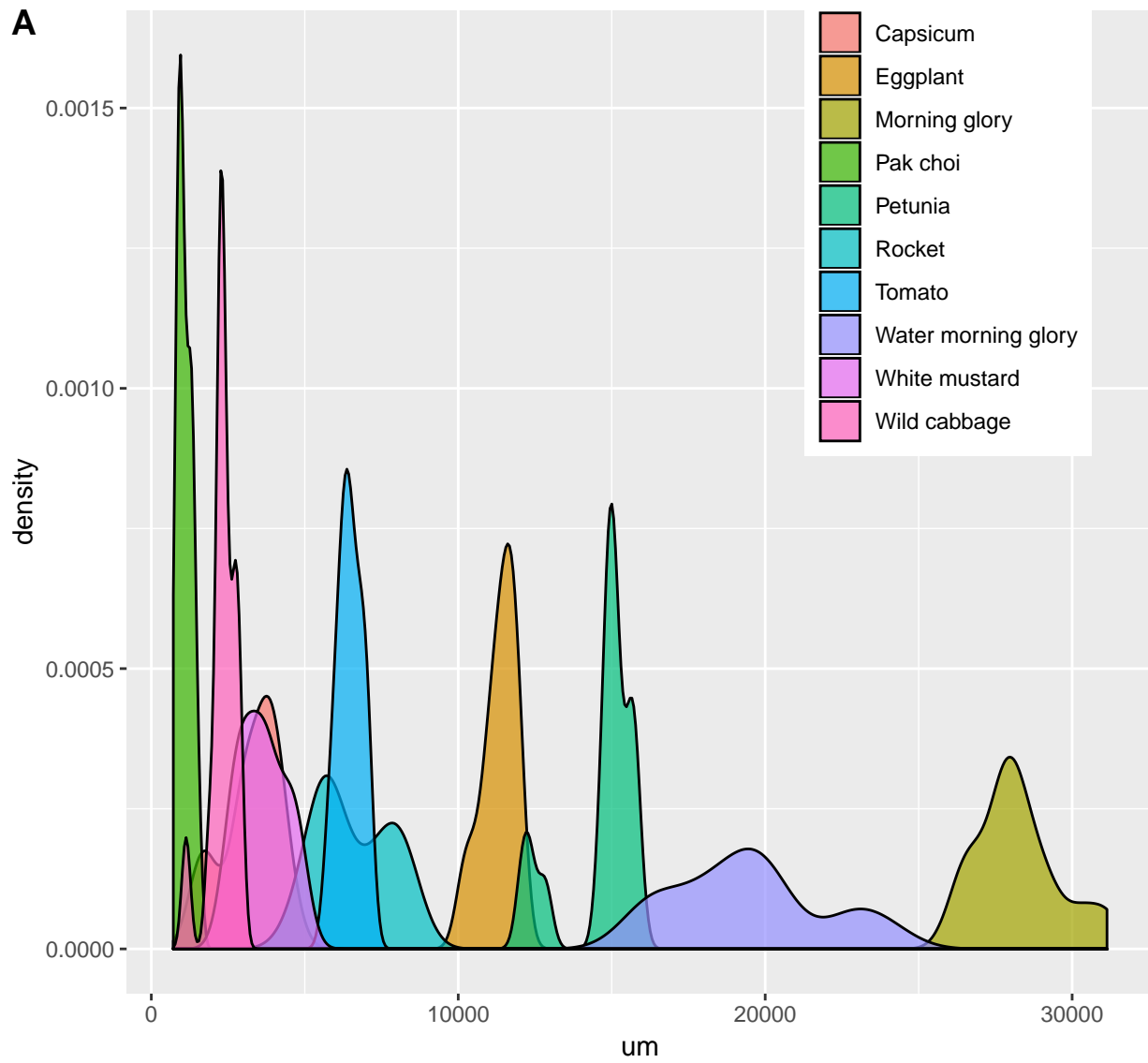
Figure 16. Density plot of the stigmatic area for all the species. The measurements were conducted with a stereo microscope. The x axis shows the area in square micrometers.
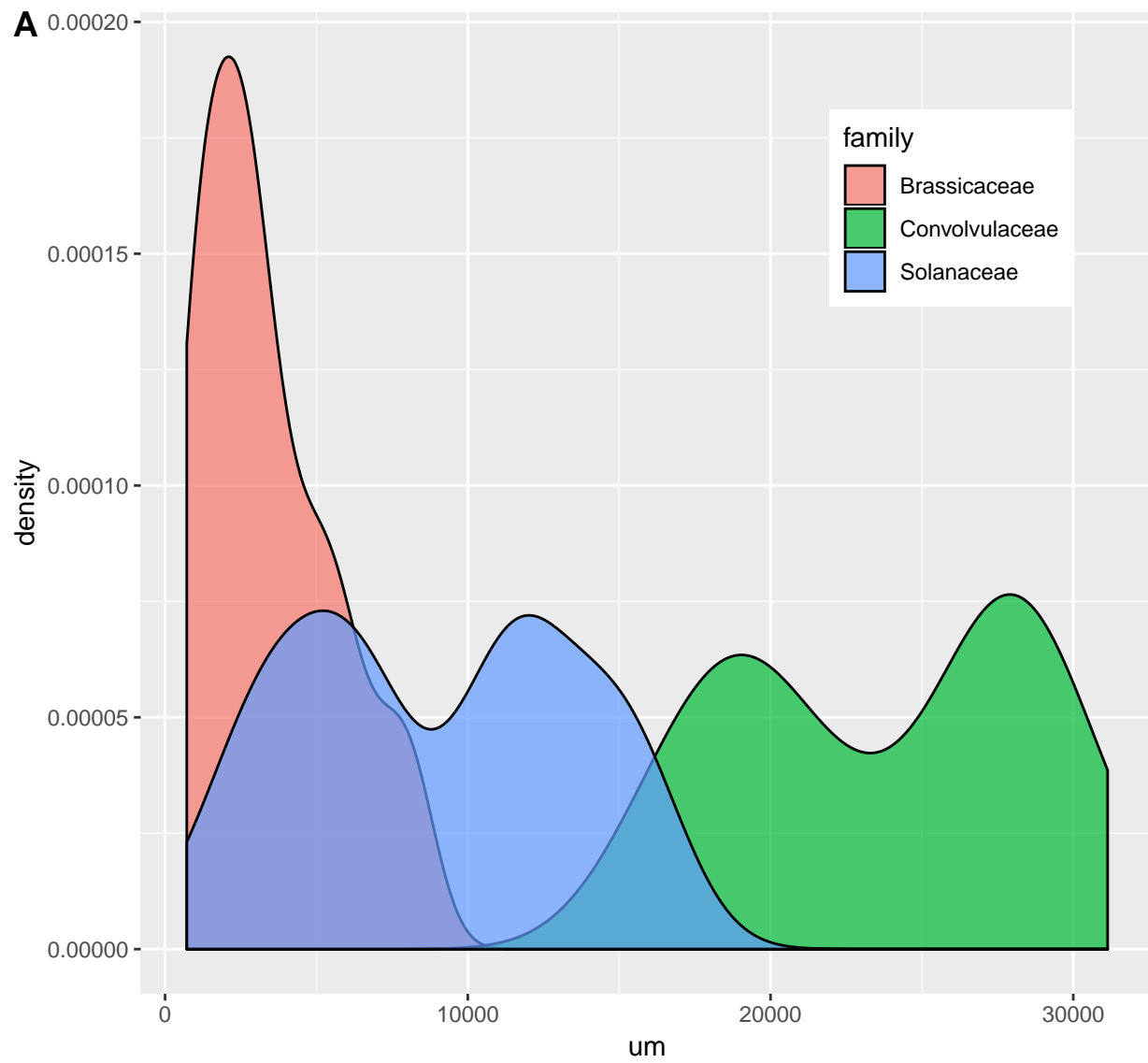
Figure 17. Density plot of the stigmatic area for the 10 species grouped by the three families: Brassicaceae, Solanaceae and Convolvulaceae. The measurements were conducted with a stereo microscope. The x axis shows the area in square micrometers.