

Covariation among reproductive traits in flowering plants shape their interactions with pollinators

Abstract

1. Globally, plants display enormous variation in life history strategies and trait combinations. However, evidence suggests that evolutionary and physiological constraints limit the number of plant ecological strategies. Although there have been recent advances in understanding correlations among plant traits, reproductive traits are rarely considered, despite their key role in shaping plant life-history strategies and interactions with pollinators.
2. Here, using a global dataset of 18 reproductive traits for 1,506 species, we investigate the reproductive spectrum of flowering plants to identify how it shapes interactions with pollinators.
3. We show that over 50% of all trait variation is explained by the first two reproductive axes, which represent the negative correlation between flower number and flower size, and the negative correlation between autonomous selfing and floral display size. In addition, these reproductive axes were associated with different numbers of interacting partners and visitation rates of the distinct pollinator guilds. However, reproductive axes explain a relatively small amount of variance in pollinator interactions highlighting the need to incorporate other factors along with reproductive traits to fully explain large scale patterns of plant-pollinator interactions.
4. Our study identifies the major reproductive trait correlations in flowering plants and their role in shaping plant-pollinator interactions at a macro-ecological scale.

These findings emphasize the importance of considering reproductive traits in the global spectrum of plant form and function, and the need to explore beyond floral morphological traits to broaden our understanding of plant-pollinator interactions.

Keywords: life-history strategies | plant reproductive traits | plant-pollinator interactions

1. Introduction

Flowering plants display an astonishing diversity of reproductive structures (Barrett, 2002; Schiestl & Johnson, 2013) that shape their interactions with animal pollinators (Dellinger, 2020; Fenster et al., 2004). The variety of reproductive strategies used by different plant species, from specialised reliance on a single pollinator, to full self-pollination, is incredibly rich. However, not all combinations of reproductive traits (i.e., morphological and physiological traits that determine a plant's ability to reproduce) are possible due to evolutionary and ecological constraints (Agrawal, 2020; Stearns, 1989). Despite recent advances in theoretical and empirical understanding of correlations between plant reproductive traits at large ecological scales (Friedman, 2020; Paterno et al., 2020; Roddy et al., 2021; Salguero-Gómez et al., 2016; Song et al., 2022), studies typically focus on just a few traits, often in isolation. Thus, to develop a comprehensive understanding of the plant reproductive spectrum of trait variation, as done recently for root (Laughlin et al., 2021), leaf (Wright et al., 2004) and wood (Chave et al., 2009) related traits, we require a multi-trait perspective with broad geographical coverage. Importantly, characterization of the plant reproductive spectrum could reveal key reproductive strategies among different species (Agrawal, 2020) and improve knowledge of associations between specific reproductive traits and pollinators (Roddy et al., 2021). With the recent availability of large trait databases (e.g., TRY Kattge et al., 2011; COMPADRE Salguero-Gómez et al., 2015), there has been increased research on plant ecolog-

ical strategies, which has produced identification of global patterns in plant form and function (Bruelheide et al., 2018; Carmona et al., 2021; Díaz et al., 2016; Salguero-Gómez et al., 2016). However, studies that look at multiple traits tend to overlook plant reproductive traits (E-Vojtkó et al., 2020; Roddy et al., 2021; Rüger et al., 2018), and focus on trait correlations in other plant organs such as morphological and physiological leaf traits (Donovan et al., 2011; Osnas et al., 2013; Shipley et al., 2006; Wright et al., 2004). Despite the lack of a holistic understanding that depicts reproductive trait covariation patterns, there are widely recognized associations between plant reproductive trait pairs such as the negative correlation between flower size and flower number (E-Vojtkó et al., 2022; Kettle et al., 2011; Sargent et al., 2007); the positive association between flower size and outcrossing rate (Goodwillie et al., 2010); or the positive correlation between outcrossing rate and lifespan (Barrett, 2003; Moeller et al., 2017; Munoz et al., 2016). Although identification of these trait correlations has allowed progress toward a conceptual framework that integrates different floral trait relationships (Roddy et al., 2021), we still lack empirical understanding of how these associations among floral and other reproductive traits (e.g., reproductive biology) shape plant life history strategies.

Trait-based approaches (Fenster et al., 2004; Rosas-Guerrero et al., 2014) and trait-matching analyses (Bartomeus et al., 2016; Stang et al., 2009) are powerful for exploring the underlying mechanisms driving plant-pollinator interactions. For example, plant traits can define species' network roles (e.g., specialists vs generalists Tur et al., 2013) and plant species that occupy reproductive trait space extremes are likely to be more specialised and display tighter trait-matching with pollinators (Coux et al., 2016; Junker et al., 2013). Indeed, the degree of morphological matching between plants and pollinators often determines whether or not interactions occur, and consequently influences the broader network structure (Ibanez, 2012; Stang et al., 2009). However, it is still unclear how specific reproductive traits (e.g., mating or compatibility system) influence plant-pollinator associations (Devaux et al., 2014; Sargent & Ackerly, 2008; Tur et al., 2013). For example, species with high selfing rates may evolve to invest less in floral rewards resulting in a lower number of pollinator visits (Devaux et al., 2014). Indeed,

both morphology and mating system can determine a species' functional role within pollination networks, and the combination of several traits increases power for predicting network structure (Eklöf et al., 2013). Because the species position within the multidimensional trait space can result in different interaction patterns or adaptations to pollinators (see Dehling et al., 2016 for a purely morphological approach), the use of the reproductive trait space to explore plant-pollinator interactions could help to identify species functional roles and associations of traits with specific pollinators (i.e., pollination syndromes).

Here, we explore the spectrum of reproductive traits in flowering plants, within a global set of plant-pollinator networks, to identify reproductive trait covariation and how this influences the network of interactions between plants and pollinators. First, we identify the major axes of reproductive trait variation and correlations among traits for plant species. Then, we investigate the associations between plant species' positions in reproductive trait space and the strength of their interactions with different pollinator guilds within networks. Finally, we determine how the main axes of reproductive trait variation influence plant species' functional roles within pollination networks, using a range of complementary interaction network metrics.

2. Methods

(a) Plant-pollinator network studies

We selected 28 studies from 18 different countries that constituted a total of 64 plant-pollinator networks (see table S1 and figure S1). These studies recorded plant-pollinator interactions in natural systems and were selected so that we had broad geographical representation across different ecological communities. The pollinators documented by these studies are primarily insect pollinators (97.8% of the total visits recorded) and the plants are characterised by a clear dominance of smaller life forms (i.e., 61% of herbs, 28% of shrubs and 11% of trees). Although these studies differ in sampling effort

and methodology, all studies provided information about plant-pollinator interactions (weighted and non-weighted), which we used to build a database of plant species that are likely to benefit, to some degree, from animal pollination. Many of these networks are freely available either as published studies (e.g., [Carvalho et al., 2014](#); [Fortuna et al., 2010](#); [Olesen et al., 2007](#)) or in online archives (e.g., [Mangal Poisot et al., 2016](#); The Web of Life [Fortuna et al., 2010](#)). In total, our network dataset constituted 60 weighted (number of visits) and four unweighted (interaction presence-absence) networks, each sampled at a unique location and year, as well as eight meta-networks where interactions were pooled across several locations and multiple years.

(b) Taxonomy of plants and pollinators

All species names, genera, families and orders were retrieved and standardised from the taxonomy data sources NCBI (<https://www.ncbi.nlm.nih.gov/taxonomy>) for plants and ITIS (<https://www.itis.gov/>) for pollinators, using the R package *taxize* ([Chamberlain et al., 2020](#)). We filled the ‘not found’ searches manually using <http://www.theplantlist.org/> and <http://www.mobot.org/> for plants and <http://www.catalogueoflife.org/> for pollinators.

(c) Plant traits

We selected a total of 18 quantitative and categorical functional traits that included reproductive traits and traits related with plant form and size (see table 1). Reproductive traits consisted of those related to floral and reproductive biology: (i) floral traits included those related to the size of floral organs (e.g., style length), floral display size (e.g., flower number) and floral rewards (e.g., pollen quantity), while (ii) reproductive biology traits represented the plant reproductive system (i.e., the breeding, mating and compatibility system). The remaining traits were related to plant size and form (i.e., plant height, lifespan and life form), which are commonly used to characterise the fast-slow continuum of plant trait variation ([Salguero-Gómez et al., 2016](#)). For each plant species, we undertook an extensive literature and online search for all traits across

Table 1. Quantitative and categorical traits used in this study.

| Traits | Numerical range/Categories |
|--------------------------------|--|
| Quantitative | |
| Plant height | 0.01-30m |
| Flower width | 0.50-205mm |
| Flower length | 0.20-195mm |
| Inflorescence width | 0.80-300mm |
| Style length | 0-138mm |
| Ovules per flower | 1-25000 |
| Flowers per plant | 1-1.8 × 10 ⁵ |
| Microliters of nectar | 0-160μl |
| Milligrams of nectar | 0-6.1mg |
| Pollen grains per flower | 13-2 × 10 ⁸ |
| Autonomous selfing (fruit set) | 0-100% |
| Categorical | |
| Lifepan | Short-lived and perennial |
| Life form | Herb, shrub and tree |
| Flower shape | Brush, campanulate, capitulum, open, papilionaceous and tube |
| Flower symmetry | Actinomorphic and zygomorphic |
| Nectar | Presence and absence |
| Compatibility system | Self-incompatible, partially self-compatible and self-compatible |
| Breeding system | Hermaphrodite, monoecious and dioecious |

a wide range of resources (plant databases, online floras, books, journals and images). From a matrix comprising a total of 27,108 possible cells (considering all traits and plant species: 18 columns × 1,506 species), we were able to fill 22,796 cells (84.1% of the dataset, see figure S2 for missing value information for each trait). An extended description of each trait and how it was obtained can be found in Appendix S1.

(d) Phylogenetic Distance

We calculated the phylogenetic distance between all plant species considered using the function *get_tree* from the package *rrees* (<https://github.com/daijiang/rrees>), which downloads phylogenetic distances from the extended R implementation of the Open Tree of Life (Jin & Qian, 2019; Smith & Brown, 2018).

(e) Data Imputation

To avoid excluding species with missing trait data, we imputed missing values with the help of the function *missForest* (Stekhoven & Bühlmann, 2012) which allows imputation of datasets with continuous and categorical variables. We accounted for the phylogenetic distance among species on the imputation process by including the eigenvectors from principal component analysis of the phylogenetic distance (PCoA), which has been shown to improve the performance of *missForest* (Penone et al., 2014). To extract the eigenvectors, we used the function *PVRdecomp* from the package *PVR* (Chamberlain et al., 2018) based on a previous conceptual framework that considers phylogenetic eigenvectors (Diniz-Filho et al., 2012). We conducted two different imputations, one for the full set of species (1,506 species: 5.79% of missing values), and excluding nectar and pollen traits because of the high percentage of missing values (figure S2), and a second one for the subset of species with data for pollen per flower and microliters of nectar (755 species: 8.01% of missing values). To corroborate that our imputation of missing values did not affect our results, we evaluated the reproductive spectrum (see section below) with and without imputed values and results for the two datasets were consistent (Fig S3 and Fig S4).

(f) Plant strategies analysis

We explored the association between the different quantitative plant traits using a phylogenetically informed Principal Component Analysis (pPCA). We did not include the quantitative variables of flower length and inflorescence width because they were highly and moderately correlated to flower width, respectively (Pearson's correlation = 0.72, $P < 0.01$ and Pearson's correlation = 0.36, $P < 0.01$), and thus we avoided overemphasise flower size on the spectrum of trait variation. Prior to the analyses, we excluded outliers and standardised the data. Specifically, due to the high sensitivity of dimensionality reduction to outliers (Legendre & Legendre, 2012; Serneels & Verdonck, 2008), we excluded values outside the 2.5th–97.5th percentile range, and thus our final

dataset had 1,236 species. Then, we log transformed all continuous variables to reduce the influence of extreme values and then z-transformed ($X=0$, $SD=1$) these variables so that they were within the same numerical range, as recommended for principal component analysis (Legendre & Legendre, 2012). Although qualitative traits were not included in the dimensionality reduction analysis, we also investigated their position in the trait space and the statistical association of the different qualitative traits with the main axes of trait variation using an ANOVA and Tukey test. Similarly, we also explored for each pollinator guild (see “Visitation patterns” subsection below) the position in the trait space of the plants that interacted with them. We performed the pPCA using the function *phyl.pca* from the package *phytools* (Revell, 2012) with the method lambda (λ) that calculates the phylogenetic correlation between 0 (phylogenetic independence) and 1 (shared evolutionary history) and we implemented the mode “covariance” because values for each variables were on the same scale following transformation (Abdi & Williams, 2010). Finally, we conducted an additional phylogenetic informed principal component analysis for the subset of species with pollen and nectar quantity data. For this, we included all quantitative traits included in the main pPCA, plus pollen grains and microlitres of nectar per flower.

(g) Phylogenetic signal

We calculated the phylogenetic signal of the different quantitative traits on the imputed dataset with the full set of species ($N = 1,506$) with the package *phytools* (Revell, 2012) and used Pagel’s λ as a measurement of the phylogenetic signal. However, for pollen and nectar traits, phylogenetic signal was calculated only on the subset of species that had quantitative information for these traits ($N = 755$).

(h) Network analyses

(i) Visitation patterns

We used Bayesian modelling (see details below) to investigate how the different guilds of pollinators interacted along the main axes of reproductive trait variation using qualitative and quantitative measures of plant-pollinator interactions. Specifically, interactions between plants and pollinators in binary networks were defined as the presence or absence of floral visitation, while in weighted networks, interactions were measured as the the number of visits from each insect species to different plant species. Although floral visitors are not always pollinators and the number of visits does not consider the pollination efficiency of each species (Ballantyne et al., 2015), the number of visits does provide valuable information about the relative contributions different floral visitors make to pollination (Vázquez et al., 2005, 2012). Therefore, despite the fact that we do not evaluate pollinator effectiveness we refer to the different floral visitors as pollinators.

We divided pollinators into six main taxonomical guilds that differ in life form, behaviour and thus likely play different ecological roles: (i) bees (Hymenoptera-Anthophila), (ii) non-bee-Hymenoptera (Hymenoptera-non-Anthophila), (iii) syrphids (Diptera-Syrphidae), (iv) non-syrphid-Diptera (Diptera-non-Syrphidae), (v) Lepidoptera, and (vi) Coleoptera. Note that we excluded non-insect pollinators from our analyses as they comprised only a 2.2% of the total visits recorded in our set of studies. Moreover, because bees had the greatest number of occurrences in networks (2,256 records) and the highest frequency of visits, we further broke this group down into the main bee families (Andrenidae, Apidae, Colletidae, Halictidae, and Megachilidae). In addition, we found that *Apis mellifera* was the pollinator with the largest proportion of occurrences in networks (7.55% of the total). This is consistent with previous research showing that *A. mellifera* was the most frequent pollinator in a similar dataset of 80 plant-pollinator networks in natural ecosystems (Hung et al., 2018). Hence, to control for the effect of *A. mellifera* on observed visitation patterns of bees, we conducted an

analogous analysis with presence-absence of the interaction and number of visits excluding *A. mellifera*. We found that *A. mellifera*, was partly driving some of the observed trends on PC1 (figure S5). However, we did not detect major differences on PC2 and PC3.

We used Bayesian generalised linear mixed models in the R package *brms* (Bürkner, 2017) to model the presence-absence of observed interactions and number of visits as a function of the main axes of plant trait variation (i.e., PC) and their interactions with pollinator guilds (i.e., PGs; e.g., number of visits \sim PC1 \times PGs + PC2 \times PGs + PC3 \times PGs). Because we were interested in possible differences in visitation patterns among pollinator guilds to plants with different reproductive strategies, we included interactions between the main axes of trait variation (PC1, PC2 and PC3) and the different pollinator guilds in the model. We added a nested random effect of networks within the study system to capture the variation in networks among studies and within networks. Moreover, we included the phylogenetic covariance matrix as a random factor due to the possible shared evolutionary histories of species and therefore lack of independence among them. We specified for the presence or absence of an interaction and number of visits with Bernoulli and a zero inflated negative binomial distributions, respectively. The models were run with 3,000 iterations with previous 1,000 warm up iterations and with non or very weakly informative priors from the *brm* function so the priors had negligible influence on the results (Bürkner, 2017). We set delta (Δ) to 0.99 to avoid divergent transitions and visualised the posterior predictive checks with the function *pp_check* using the *bayesplot* package (Gabry et al., 2019).

(ii) Plant species network roles

We also used Bayesian modelling to investigate how the main axes of trait variation determined different plant species' functional roles within pollination networks. For this, we selected complementary species-level network metrics, commonly applied in bipartite network studies (Dormann et al., 2008), with clear ecological interpretations and are relevant for defining species' functional roles in networks. Specifically, the

species-level metrics we included were: (i) sum of visits per plant species; (ii) normalised degree, calculated as the number of links per plant species divided by the total possible number of partners; and (iii) selectivity (d') (Blüthgen et al., 2006), which measures the deviation of an expected random choice of the available interaction partners and ranges between 0 (maximum generalisation) and 1 (maximum specialisation). Normalised degree and selectivity were calculated with the *specieslevel* function in the R package *bipartite* (Dormann et al., 2008). We modelled each plant species metric (sum of visits, normalised degree and plant selectivity) as a function of the three main axes of trait variation (plant species level metric \sim PC1 + PC2 + PC3). For the different response variables in each model (i.e., each plant species level metric), we used different distribution families (zero inflated negative binomial for the sum of visits, weibull for normalised degree, and zero-one inflated beta for selectivity). Finally, we used the same random factors, model settings and conducted the same posterior predictive checks for each model as detailed above in the ‘visitation patterns’ section.

Our analyses were conducted on the subset of 60 weighted networks (out of a total of 64 networks) sampled in a unique flowering season and site, which included 556 plant and 1,126 pollinator species. In total, our network dataset (excluding meta- and non-weighted networks) included 2,256 interactions for bees, 1,768 for non-syrphid-dipterans, 845 for syrphids, 437 for lepidopterans, 432 for coleopterans, and 362 for non-bee-hymenopterans. Sampling methods varied across studies but this was accounted for in analyses by including study identity as a random effect in our models. All analyses were conducted in R version 4.0.3.

3. Results

(a) Plant strategies analysis

Our phylogenetically informed principal component analysis captured 51.8% and 70.97% of the reproductive trait variation in the first two and three axes, respectively

(figure 1 and figure S6). The first principal component (PC1; the flower number - flower size axis) represented 26.72% of the trait variation and indicated a negative correlation between flower number and flower size. The main contributing traits to PC1 were plant height, flower number, ovule number and flower size (loadings $> |0.5|$; table S3) but style length also contributed moderately (loading = -0.33). One end of this axis comprised species with high investment in flower number and plant height but small flower size, short style length and low ovule number. For instance, *Cornus florida* occurred at this end of the spectrum, which, on average, has approximately 10,000 flowers, is 7.5 m high, has flowers that are 3 mm wide, styles 3.5 mm long and two ovules per flower. The other end of this spectrum had species that were short in height, with few large flowers that had long styles and many ovules. For example, *Petunia axillaris* on average, is 0.5 m high and has approximately 10 flowers per plant that are over 50 mm wide with 25 mm long styles and over 200 ovules. The second principal component (PC2; the autonomous selfing - floral display axis) encompassed 25.05% of the trait variation and represented variation from low to high autonomous selfing (i.e., high to low pollinator dependence). The main driver of trait variation on PC2 was autonomous selfing (loading = 0.85) but all other traits (except ovule number) made moderate contributions (loadings ranging between 0.27 to 0.4; table S3). Generally speaking, species with low autonomous selfing had larger and many flowers with long styles, and were tall. *Zuccagnia punctata* occurred at this extreme of the spectrum, which is a self-incompatible species, completely dependent on animal pollination for seed production, producing approximately 1,500 flowers per plant with an average height of 3 m and 20 mm long styles. In contrast, species with high autonomous selfing tended to have fewer and smaller flowers with shorter styles, and were shorter in height. *Veronica peregrina* occurred at this end of the spectrum, and is a self-compatible plant that requires little or no pollination by animals, it has approximately 20 flowers per plant, is 0.2 m high and has 0.25 mm long styles. The third principal component (PC3; the style length - autonomous selfing axis) explained a considerable amount of trait variability (19.17%) and represented a positive correlation between style length (loading

305 = -0.66) and autonomous selfing (loading = -0.51), where species with short styles
306 had low autonomous selfing and species with long styles high autonomous selfing.
307 The remaining traits, apart from ovule number, were all positively correlated with style
308 length and autonomous selfing (loadings from -0.23 to -0.46; table S3). In addition, the
309 pPCA with the subset of species that we had nectar and pollen quantity data for showed
310 that nectar quantity (microlitres of nectar per flower) and pollen grains per flower were
311 positively associated with floral display size. Floral rewards were particularly strongly
312 correlated with flower size but weakly correlated with flower number (figure S4). The
313 first two principal components for this pPCA explained similar variance (45.52%) and
314 similar associations of traits, despite some variability in the loadings (table S4).

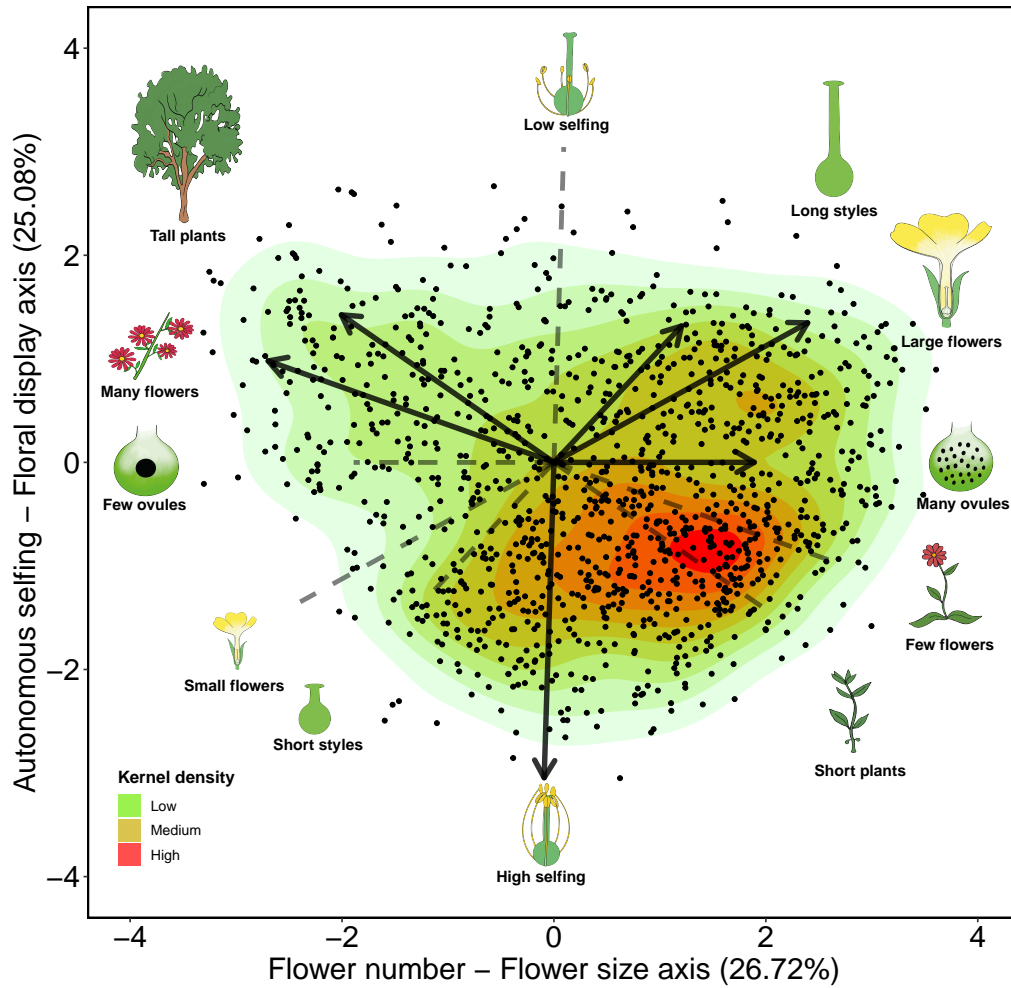


Figure 1. Phylogenetically informed principal component analysis (pPCA) of 1,236 plant species from 28 plant-pollinator network studies. The solid arrows indicate the direction of the different quantitative traits (flower number, plant height, style length, flower size, ovule number and level of autonomous selfing) across the two main axes of trait variation. The length of the arrows indicate the weight of the variables on each principal component and the dashed lines show the opposed direction of trait variation. The icons at both ends of arrows and dashed lines illustrate the extreme form of the trait continuum.

315 As expected, due to individual trait correlations between qualitative and quantitative
 316 traits (e.g., life form and plant height), we found that most categorical traits were
 317 clustered on the trait space and statistically associated with the first two axes of trait
 318 variation (figure s2 and table s2). However, the position of plants connected to each of
 319 the pollinator guilds was scattered without clear aggregation patterns on the trait space
 320 (figure s7; but see “Visitation patterns” section below for detailed results from quantita-
 321 tive analyses). Flower symmetry, which was only associated with PC2 (Sum of squares

322 = 8.51, F-value = 14.72, $P < 0.01$), and nectar provision, which was independent of PC1
 323 and PC2 (PC1: Sum of squares = 0.37, F-value = 0.29 , $P = 0.59$; PC2: Sum of squares =
 324 0.83, F-value = 1.43, $P = 0.23$) showed a lack of statistical association. In addition, we
 325 found statistical differences between the different categorical trait levels within the trait
 326 space (figure S8). Regarding self-compatibility, we found larger differences on PC2. For
 327 instance, species with unisexual flowers that were self-incompatible, were statistically
 328 differentiated from species with partial or full self-compatibility on this axis (figure S8a
 329 and figure S8b; Tukey test $P < 0.05$). Life forms differed statistically across both axes of
 330 trait variation and followed a gradient of larger life forms (trees and shrubs) with lower
 331 autonomous selfing to smaller ones (herbs) with higher autonomous selfing (figure
 332 S8c and figure S8d; Tukey test $P < 0.05$). Consequently, lifespan also followed this
 333 gradient but perennial and short lived species only differed statistically on PC2 (figure
 334 S8e and figure S8f; Tukey test $P < 0.05$). Species with unisexual flowers (monoecious
 335 and dioecious) were clustered on both extremes of the first two principal components
 336 and had both the lowest autonomous selfing levels and the highest number of flowers
 337 (figure S8g and figure S8h; Tukey test $P < 0.05$). Moreover, we found that campanulate
 338 and capitulum flower shapes were differentiated from tube, papilionaceous, open and
 339 brush shapes in the trait space; the former had larger flowers and low autonomous
 340 selfing levels, while the latter had more flowers and high autonomous selfing (figure S8i
 341 and figure S8j; Tukey test $P < 0.05$). Finally, in terms of flower symmetry, zygomorphic
 342 flowers were associated with high levels of autonomous selfing, whereas actinomorphic
 343 flowers were more dependent on pollinators (figure S8k and figure S8l; Tukey test $P <$
 344 0.05).

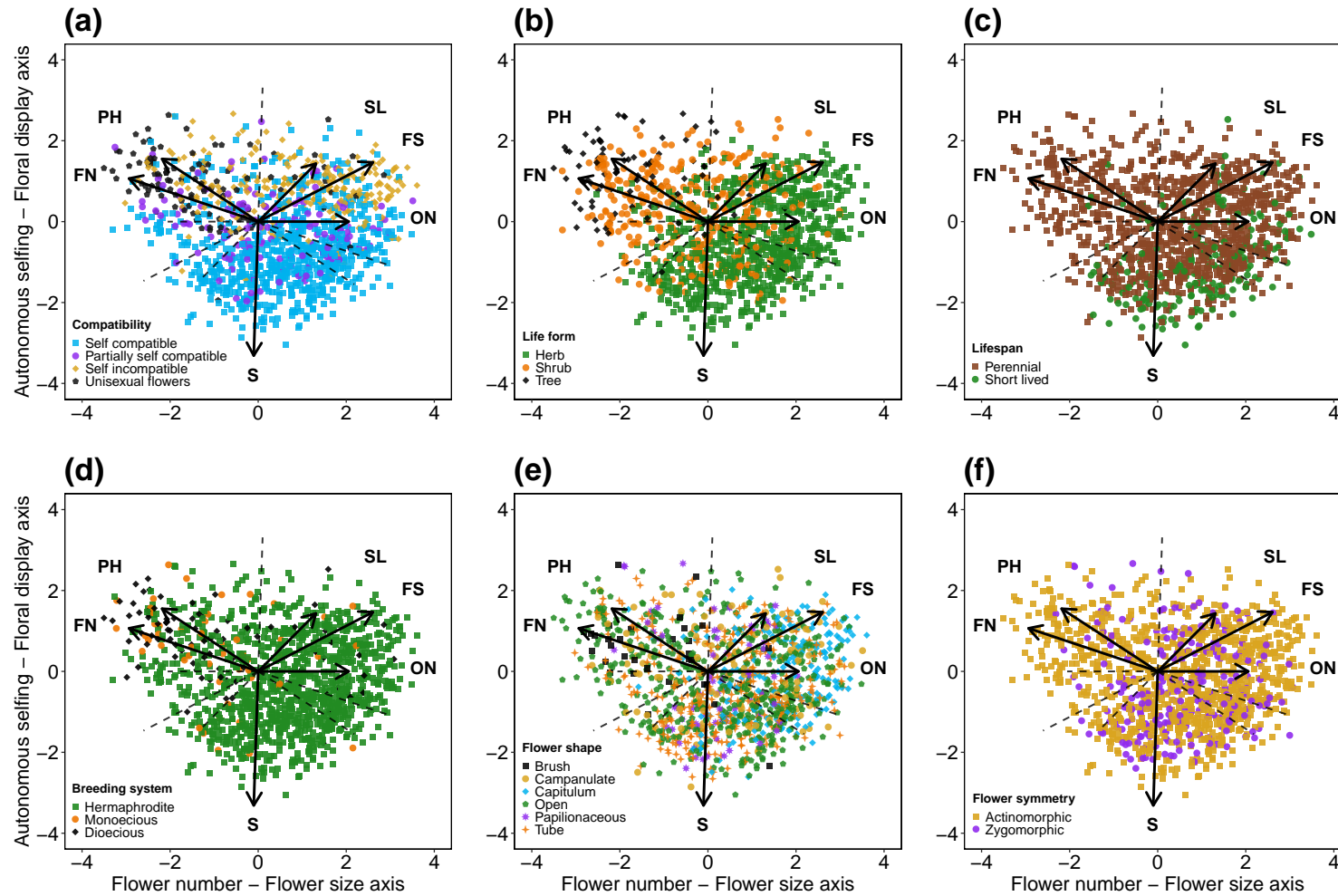


Figure 2. Location of the different qualitative traits in the trait space. The figure panel is composed of the traits that showed statistical association with the first two axes of trait variation: compatibility system (a), life form (b), lifespan (c), breeding system (d), flower shape (e) and flower symmetry (f). The solid arrows indicate the direction of variation of the different quantitative traits shown in figure 1: flower number (FN), plant height (PH), style length (SL), flower size (FS), ovule number (ON) and the level of autonomous selfing (S).

(b) Phylogenetic signal

We found a strong phylogenetic signal ($P < 0.01$) for most quantitative traits (table S5). The traits that showed the highest phylogenetic signal were ovule number ($\lambda = 1$), pollen grains per flower ($\lambda = 1$) and plant height ($\lambda = 0.96$), followed by flower length ($\lambda = 0.75$), flower width ($\lambda = 0.73$) and number of flowers per plant ($\lambda = 0.69$). The traits that had a moderate phylogenetic signal included inflorescence width ($\lambda = 0.57$), style length ($\lambda = 0.49$) and autonomous selfing ($\lambda = 0.34$). Finally, microliters of nectar per flower showed the lowest phylogenetic signal of all the traits we measured ($\lambda = 0.14$).

(c) Visitation patterns

The main axes of trait variation (PC1, PC2 and PC3) partly explained the presence-absence of interaction partners (conditional $R^2 = 0.26$; marginal $R^2 = 0.20$) but little of the total number of visits (conditional $R^2 = 0.31$; marginal $R^2 = 0.06$). However, we found relevant differences across the different pollinator guilds for both the presence-absence of interactions and number of visits (figure 3). On the flower number - flower size axis, we found that plants with many small flowers tended to have interaction partners that were Coleoptera, non-bee-Hymenoptera and Diptera (figure 3a), whereas plants with few large flowers mostly had interaction partners that were bees and Lepidoptera. We found similar trends for visitation rates on the flower number - flower size axis, but bees and syrphids visited plant species with many small flowers more frequently (figure 3d). On the autonomous selfing - floral display axis, we found that all plant species with lower autonomous selfing had many interaction partners and received many visits from all pollinator guilds (figure 3b and figure 3e). Finally, on the style length - autonomous selfing axis, plant species with short styles and low autonomous selfing had more interaction partners for all guilds, except bees which interacted more frequently with plant species with long styles and high autonomous selfing (figure 3c). Plants with long styles and high autonomous selfing interacted more frequently with Lepidoptera and non-bee-Hymenoptera (figure 3f).

372 The additional models for the most represented bee families, for both presence-absence
373 of interactions (marginal $R^2 = 0.29$; conditional $R^2 = 0.19$) and the number of visits
374 (marginal $R^2 = 0.30$; conditional $R^2 = 0.03$), showed that the Apidae family was the main
375 driver of the observed patterns. The contrast between interaction presence/absence
376 and number of visits for bees on PC1 (figure 3a and figure 3d) was driven by the
377 Andrenidae family, which had more interaction partners but made fewer visits to plant
378 species with few large flowers (figure S9).

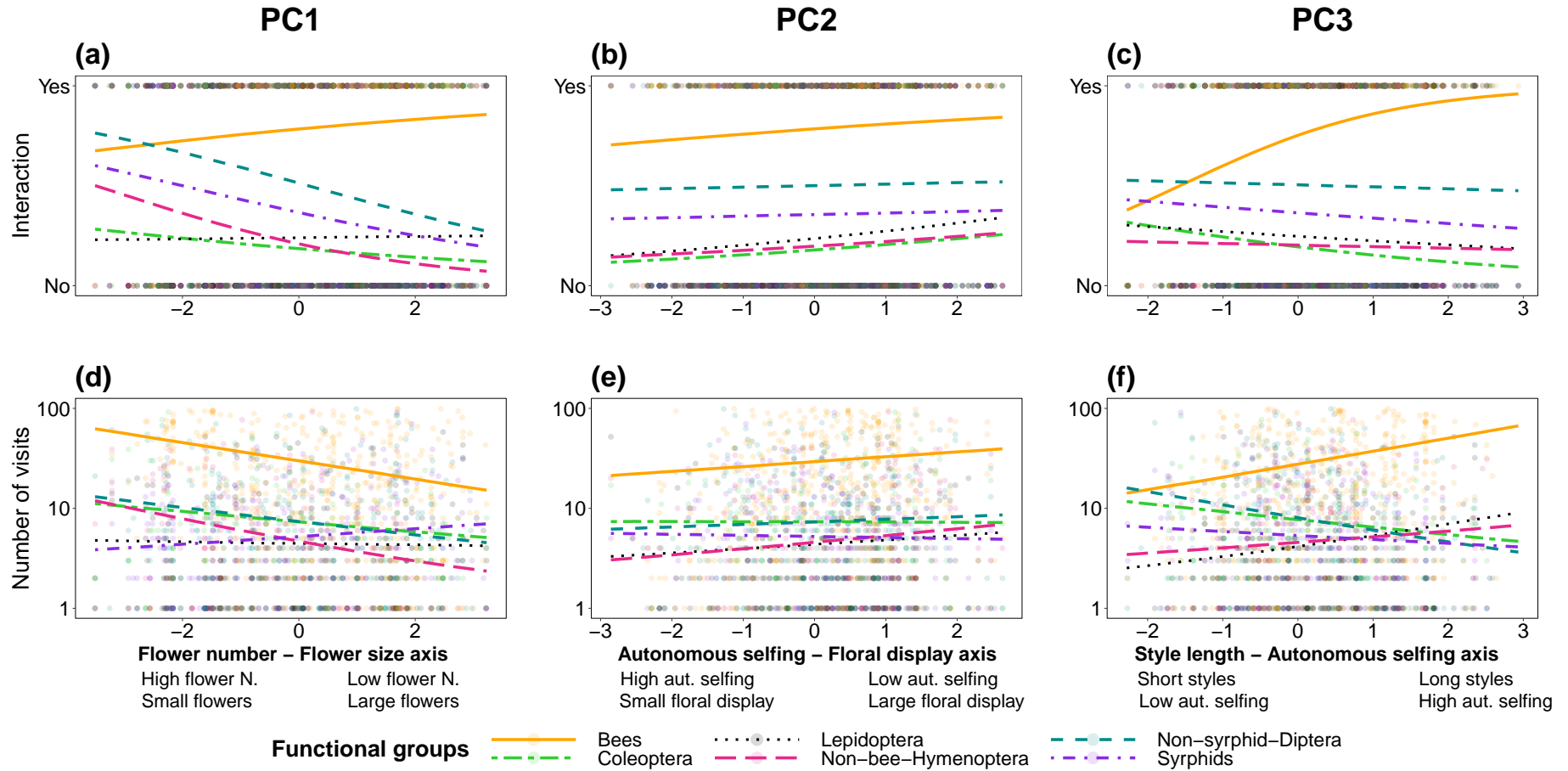


Figure 3. Fitted posterior estimates of the presence-absence of interaction (a, b and c) and number of visits (d, e and f) of the different pollinator guilds in relation to the main axes of plant reproductive trait variation (PC1, PC2 and PC3). PC1 represents the flower number - flower size axis, PC2 represents the autonomous selfing - floral display axis and PC3 represents the style length - autonomous selfing axis. For visualization purposes due to large differences between number of visits from bees and the rest of the pollinator guilds, the number of visits was log-transformed (Y-axis of lower panel).

(d) Plant species network roles

Variance within plant species-level network metrics was poorly explained by the three main axes of trait variation (figure S10; number of visits ~ PCs, conditional $R^2 = 0.11$, marginal $R^2 = 0.02$; normalised degree ~ PCs, conditional $R^2 = 0.24$, marginal $R^2 = 0.02$; and, selectivity ~ PCs, conditional $R^2 = 0.37$, marginal $R^2 = 0.03$). Overall, the most notable trends were found on the flower number - flower size axis (PC1) and the style length - autonomous selfing axis (PC3) for number of visits and selectivity. On PC1, the number of visits per flower was higher for plant species with more flowers but was lower for plant species with larger flowers (figure S10a). However, selectivity (d') was higher for plant species with large flowers but lower for plant species with small flowers (figure S10g). On PC3, the number of visits was lower for plants with shorter styles and lower autonomous selfing but higher for plant species with longer styles and higher autonomous selfing (figure S10c). Again, selectivity showed the opposite trend to the number of visits (figure S10i).

4. Discussion

We found strong covariation among traits within the flowering plant reproductive spectrum, which shapes interactions with pollinators at a global scale. Despite the enormous variability in plant reproductive structures, the first two and three axes captured over 50% and 70% of trait variation, respectively, and these axes encapsulated correlations among flower number and flower size, autonomous selfing and floral display, and style length and autonomous selfing. Although explained trait variation on the first two axes was less than previous studies of morphological and physiological traits (Carmona et al., 2021; Díaz et al., 2016), our results align with the two largest and most recent studies of plant life strategies that included reproductive, form and size related traits (E-Vojtkó et al., 2022; Salguero-Gómez et al., 2016). The different plant reproductive axes that we identified were associated with different numbers of interacting partners and visitation rates of the distinct pollinator guilds. However, the

unexplained variability found highlights the need to account for other local factors, such as species relative abundances (Bartomeus et al., 2016), along with reproductive traits, to better explain plant-pollinator associations.

Our study provides strong evidence for several widely discussed trait associations with the help of a great diversity of phylogenetic lineages (170 families and over 1,200 species). Specifically, by using a multitrait approach, we are able to link the reproductive spectrum of plant trait variation with the previously described “fast-slow continuum” in plant (Salguero-Gómez et al., 2016) and animal (Healy et al., 2019) life-history strategies. This fast-slow continuum was illustrated by the associations between reproductive traits and plant height, life form and lifespan traits. For instance, although the negative correlation between flower number and flower size has received substantial attention (Kettle et al., 2011; Sargent et al., 2007; Worley et al., 2000), it has only been investigated across a few hundred Angiosperm species and generally lacking a multitrait perspective. We found support for this trait association and show that other reproductive traits (e.g., style length or ovule number) covary along this flower number - flower size axis. Our study also complements previous work linking plant breeding systems and life forms (Friedman, 2020; Petit & Hampe, 2006) by showing that most species with unisexual flowers also have woody life forms and many flowers. Moreover, we identify the previously documented positive association between outcrossing rate and floral display size (Goodwillie et al., 2010), where species with low autonomous selfing levels also allocate large resource quantities to attract pollinators.

Although we did not include floral rewards in our main analysis due to insufficient data, we did find that plant species with large floral displays and, in particular, large flowers, tended to have higher pollen and nectar quantity. Thus, we demonstrate that globally, reproductive trait covariation in flowering plants likely imposes constraints on the broader spectrum of plant trait variation.

Overall, the main plant reproductive axes were insufficient to fully capture plant-pollinator associations. However, these axes helped explain, to some extent, interaction

patterns among and within pollinator guilds, indicating that they partly mediate these associations across large spatial scales. For example, we found that all pollinator guilds interacted more frequently with species with large floral displays and low ability to self pollinate, which aligns with previous findings (Hegland & Totland, 2005; Kaiser-Bunbury et al., 2014; Lázaro et al., 2013) and predictions from evolutionary life-history theory (Stebbins, 1970). These predictions suggest that larger investment in floral display and rewards to attract pollinators occurs when dependence on pollinators is high. Although pollinators play a major role in shaping plant reproductive strategies, these different strategies are also constrained by the cost of reproductive organs and trait correlations (Galen, 1999). For instance, smaller flowers tend to receive fewer visits than larger flowers, but small flowers are inexpensive to produce (e.g., have few ovules). Therefore, low per-flower visitation can be compensated by overall plant-level visitation through producing many small flowers that develop into numerous small fruits (Kettle et al., 2011). Our results also indicate that bees interact more with plants that have higher levels of autonomous selfing and large styles, which can help with a more efficient pollen transfer and prevent pollen competition (Ashman & Arceogómez, 2013; Lanuza et al., 2021; Tong & Huang, 2016). Similarly, the specialised floral structures of zygomorphic flowers promote precise pollen transfer (Gong & Huang, 2009; Sargent, 2004), although we found that they were associated with high levels of autogamy. The annual herb *Lathyrus clymenum* is a clear example of this pollinator specialization - selfing strategy that may get “the best of both worlds” (Goodwillie & Weber, 2018; Lloyd & Schoen, 1992). It has showy zygomorphic flowers with relatively long styles and few ovules per flower that are likely fertilised with a single legitimate floral visit but can avoid reproductive failure in the absence of pollinators through autonomous self pollination. Our findings are consistent with the hypothesis that selfing is not rare within specialised pollination systems (Fenster & Marten-Rodriguez, 2007) and emphasise the need to consider different reproductive assurance mechanisms (e.g., early versus delayed selfing; Goodwillie & Weber, 2018) to better understand plant reproductive trait correlations and plant-pollinator interactions.

463 Because flowering plants can adapt to their animal pollinators to optimise pollen trans-
 464 fer, the correlation between distinct suites of plant reproductive traits and pollinator
 465 guilds could indicate the existence of pollination syndromes ([Dellinger, 2020](#); [Faegri](#)
 466 [& Van Der Pijl, 2013](#); [Fenster et al., 2004](#); [Rosas-Guerrero et al., 2014](#)). However, our
 467 work shows a weak association between the different plant reproductive strategies and
 468 pollinator guilds, which supports previous findings from a similar pollinator guild
 469 classification and spatial scale ([Ollerton et al., 2009](#)). Although our findings contrast
 470 with a vast amount of literature on pollinator syndromes dating back over half a cen-
 471 tury ([Faegri & Van Der Pijl, 2013](#); [Fenster et al., 2004](#)), the ecological scale of our study
 472 differs from the majority of floral syndrome studies that rarely consider plant-pollinator
 473 associations beyond the community scale (ca. 3% of studies; [Dellinger, 2020](#)). We
 474 argue that the predictive power of floral syndromes gets blurred when increasing the
 475 number of species and scale, reinforcing the view that strong specialisation may be the
 476 exception rather than the norm in plant-pollinator interactions ([Jordano, 1987](#); [Waser](#)
 477 [et al., 1996](#)). Further, there are several factors that might have influenced our findings,
 478 (e.g., the suite of reproductive traits and type of pollinator guilds we included). For
 479 instance, including other key reproductive traits, such as floral scent or flower colour,
 480 could help to improve predictions of plant-pollinator interactions. In addition, our use
 481 of broad taxonomic pollinator guild groupings may have obscured finer scale patterns
 482 that occur at lower taxonomic ranks (e.g., genus or species level). The fact that our
 483 study only accounts for insect pollinators may have also influenced the observed asso-
 484 ciations between floral traits and pollinator guilds in comparison with other studies, as
 485 some other guilds (e.g., hummingbirds or bats) generally display strong trait-matching
 486 ([Lagomarsino et al., 2017](#); [Wilson et al., 2004](#)). Regardless, it is worth noting that local
 487 factors (e.g., species relative abundances, phenologies or intraspecific variation) likely
 488 contribute to the variability in interactions between the plant reproductive spectrum
 489 and pollinators ([Arroyo-Correa et al., 2023](#); [Bartomeus et al., 2016](#); [Encinas-Viso et](#)
 490 [al., 2012](#); [Vázquez et al., 2007](#)). We suggest that while plant reproductive traits do
 491 influence which pollinators they interact with, at a macroecological scale, pollination

syndromes possibly take the form of a multifaceted, context dependent continuum (e.g., ornithophilous versus entomophilous pollination of red flowers; [León-Osper & Narbona, 2022](#)).

Our study provides a comprehensive assessment of reproductive trait covariation patterns in flowering plants, using a global dataset of traits for plant species within plant-pollinator networks. We show the importance of pollinators in mediating reproductive trait correlations and identify the need to consider broader plant reproductive strategies, rather than just flower morphology, to understand patterns of plant-pollinator interactions. Further, we provide empirical evidence of reproductive trait associations that complement trait covariation studies for other plant organs, highlighting the need to include reproductive traits in the global spectrum of plant form and function. Future work incorporating underrepresented areas of the world and additional relevant reproductive traits (e.g., flower life span and colour) from a broader range of life forms will be valuable for enhancing our understanding of the plant reproductive spectrum and associated plant-pollinator interactions.

References

- Abdi, H., & Williams, L. J. (2010). Principal component analysis. *WIREs Comp. Stats.*, 2(4), 433–459. <https://doi.org/10.1002/wics.101>
- Agrawal, A. A. (2020). A scale-dependent framework for trade-offs, syndromes, and specialization in organismal biology. *Ecology*, 101(2), e02924. <https://doi.org/10.1002/ecy.2924>
- Arroyo-Correa, B., Jordano, P., & Bartomeus, I. (2023). Intraspecific variation in species interactions promotes the feasibility of mutualistic assemblages. *Ecology Letters*. <https://doi.org/10.1111/ele.14163>
- Ashman, T.-L., & Arceo-Gómez, G. (2013). Toward a predictive understanding of the fitness costs of heterospecific pollen receipt and its importance in co-flowering communities. *American Journal of Botany*, 100(6), 1061–1070. <https://doi.org/10.3732/ajb.1200496>
- Ballantyne, G., Baldock, K. C. R., & Willmer, P. G. (2015). Constructing more informative plant–pollinator networks: Visitation and pollen deposition networks in a heathland plant community. *Proc. Royal Soc. B*, 282(1814), 20151130. <https://doi.org/10.1098/rspb.2015.1130>
- Barrett, S. C. H. (2002). The evolution of plant sexual diversity. *Nat. Rev. Genet.*, 3(4), 274–284. <https://doi.org/10.1038/nrg776>
- Barrett, S. C. H. (2003). Mating strategies in flowering plants: The outcrossing-selfing paradigm and beyond. *Philos. Trans. R. Soc. Lond. B Biol. Sci.*, 358(1434), 991–1004. <https://doi.org/10.1098/rstb.2003.1301>
- Bartomeus, I., Gravel, D., Tylianakis, J. M., Aizen, M. A., Dickie, I. A., & Bernard-Verdier, M. (2016). A common framework for identifying linkage rules across different types of interactions. *Funct. Ecol.*, 30(12), 1894–1903. <https://doi.org/10.1111/1365-2435.12666>
- Blüthgen, N., Menzel, F., & Blüthgen, N. (2006). Measuring specialization in species interaction networks. *BMC Ecol.*, 6(1), 9. <https://doi.org/10.1186/1472-6785-6-9>

- Bruehlheide, H., Dengler, J., Purschke, O., Lenoir, J., Jiménez-Alfaro, B., Hennekens, S. M., Botta-Dukát, Z., Chytrý, M., Field, R., Jansen, F., et al. (2018). Global trait–environment relationships of plant communities. *Nat. Ecol. Evol.*, 2(12), 1906–1917. <https://doi.org/10.1038/s41559-018-0699-8>
- Bürkner, P.-C. (2017). Brms: An R package for Bayesian multilevel models using Stan. *J. Stat. Softw.*, 80(1), 1–28. <https://doi.org/10.18637/jss.v080.i01>
- Carmona, C. P., Tamme, R., Pärtel, M., Bello, F. de, Brosse, S., Capdevila, P., González-M, R., González-Suárez, M., Salguero-Gómez, R., Vásquez-Valderrama, M., & Toussaint, A. (2021). Erosion of global functional diversity across the tree of life. *Sci. Adv.*, 7(13), eabf2675. <https://doi.org/10.1126/sciadv.abf2675>
- Carvalho, L. G., Biesmeijer, J. C., Benadi, G., Fründ, J., Stang, M., Bartomeus, I., Kaiser-Bunbury, C. N., Baude, M., Gomes, S. I. F., Merckx, V., Baldock, K. C. R., Bennett, A. T. D., Boada, R., Bommarco, R., Cartar, R., Chacoff, N., Dänhardt, J., Dicks, L. V., Dormann, C. F., ... Kunin, W. E. (2014). The potential for indirect effects between co-flowering plants via shared pollinators depends on resource abundance, accessibility and relatedness. *Ecol. Lett.*, 17(11), 1389–1399. <https://doi.org/10.1111/ele.12342>
- Chamberlain, S., Szoecs, E., Foster, Z., Arendsee, Z., Boettiger, C., Ram, K., Bartomeus, I., Baumgartner, J., O'Donnell, J., Oksanen, J., Tzovaras, B. G., Marchand, P., Tran, V., Salmon, M., Li, G., & Grenié, M. (2018). Package “PVR.” *R package version 0.3*. At <https://CRAN.r-project.org/package=PVR>.
- Chamberlain, S., Szoecs, E., Foster, Z., Arendsee, Z., Boettiger, C., Ram, K., Bartomeus, I., Baumgartner, J., O'Donnell, J., Oksanen, J., Tzovaras, B. G., Marchand, P., Tran, V., Salmon, M., Li, G., & Grenié, M. (2020). *Taxize: Taxonomic information from around the web*. *R package version 0.9.99*. At <https://CRAN.r-project.org/package=taxize>.
- Chave, J., Coomes, D., Jansen, S., Lewis, S. L., Swenson, N. G., & Zanne, A. E. (2009). Towards a worldwide wood economics spectrum. *Ecol. Lett.*, 12(4), 351–366. <https://doi.org/10.1111/j.1461-0248.2009.01285.x>
- Coux, C., Rader, R., Bartomeus, I., & Tylianakis, J. M. (2016). Linking species functional roles to their network roles. *Ecol. Lett.*, 19(7), 762–770. <https://doi.org/10.1111/ele>

- Dehling, D. M., Jordano, P., Schaefer, H. M., Böhning-Gaese, K., & Schleuning, M. (2016). Morphology predicts species' functional roles and their degree of specialization in plant–frugivore interactions. *Proc. Royal Soc. B*, 283(1823), 20152444. <https://doi.org/10.1098/rspb.2015.2444>
- Dellinger, A. S. (2020). Pollination syndromes in the 21st century: Where do we stand and where may we go? *New Phytol.*, 228(4), 1193–1213. <https://doi.org/10.1111/nph.16793>
- Devaux, C., Lepers, C., & Porcher, E. (2014). Constraints imposed by pollinator behaviour on the ecology and evolution of plant mating systems. *J. Evol. Biol.*, 27(7), 1413–1430. <https://doi.org/10.1111/jeb.12380>
- Díaz, S., Kattge, J., Cornelissen, J. H. C., Wright, I. J., Lavorel, S., Dray, S., Reu, B., Kleyer, M., Wirth, C., Colin Prentice, I., Garnier, E., Bönisch, G., Westoby, M., Poorter, H., Reich, P. B., Moles, A. T., Dickie, J., Gillison, A. N., Zanne, A. E., ... Gorné, L. D. (2016). The global spectrum of plant form and function. *Nature*, 529(7585), 167–171. <https://doi.org/10.1038/nature16489>
- Diniz-Filho, J. A. F., Bini, L. M., Rangel, T. F., Morales-Castilla, I., Olalla-Tárraga, M. Á., Rodríguez, M. Á., & Hawkins, B. A. (2012). On the selection of phylogenetic eigenvectors for ecological analyses. *Ecography*, 35(3), 239–249. <https://doi.org/10.1111/j.1600-0587.2011.06949.x>
- Donovan, L. A., Maherali, H., Caruso, C. M., Huber, H., & Kroon, H. de. (2011). The evolution of the worldwide leaf economics spectrum. *Trends in Ecology & Evolution*, 26(2), 88–95. <https://doi.org/10.1016/j.tree.2010.11.011>
- Dormann, C. F., Gruber, B., & Fründ, J. (2008). Introducing the bipartite package: Analysing ecological networks. *R News*, 8/2.
- Eklöf, A., Jacob, U., Kopp, J., Bosch, J., Castro-Urgal, R., Chacoff, N. P., Dalsgaard, B., Sassi, C. de, Galetti, M., Guimaraes, P. R., et al. (2013). The dimensionality of ecological networks. *Ecol. Lett.*, 16(5), 577–583. <https://doi.org/10.1111/ele.12081>
- Encinas-Viso, F., Revilla, T. A., & Etienne, R. S. (2012). Phenology drives mutualistic

network structure and diversity. *Ecol. Lett.*, 15(3), 198–208. <https://doi.org/10.1111/j.1461-0248.2011.01726.x>

E-Vojtkó, A., Bello, F. de, Durka, W., Kühn, I., & Götzenberger, L. (2020). The neglected importance of floral traits in trait-based plant community assembly. *J. Veg. Sci.*, 31(4), 529–539. <https://doi.org/10.1111/jvs.12877>

E-Vojtkó, A., Junker, R. R., Bello, F. de, & Götzenberger, L. (2022). Floral and reproductive traits are an independent dimension within the plant economic spectrum of temperate central europe. *New Phytologist*. <https://doi.org/10.1111/nph.18386>

Faegri, K., & Van Der Pijl, L. (2013). *Principles of pollination ecology* (3rd edn). Elsevier.

Fenster, C. B., Armbruster, W. S., Wilson, P., Dudash, M. R., & Thomson, J. D. (2004). Pollination Syndromes and Floral Specialization. *Annu. Rev. Ecol. Evol. Syst.*, 35(1), 375–403. <https://doi.org/10.1146/annurev.ecolsys.34.011802.132347>

Fenster, C. B., & Marten-Rodriguez, S. (2007). Reproductive assurance and the evolution of pollination specialization. *International Journal of Plant Sciences*, 168(2), 215–228. <https://doi.org/10.1086/509647>

Fortuna, M. A., Stouffer, D. B., Olesen, J. M., Jordano, P., Mouillot, D., Krasnov, B. R., Poulin, R., & Bascompte, J. (2010). Nestedness versus modularity in ecological networks: Two sides of the same coin? *J. Anim. Ecol.*, 79(4), 811–817. <https://doi.org/10.1111/j.1365-2656.2010.01688.x>

Friedman, J. (2020). The evolution of annual and perennial plant life histories: Ecological correlates and genetic mechanisms. *Annu. Rev. Ecol. Evol. Syst.*, 51, 461–481. <https://doi.org/10.1146/annurev-ecolsys-110218-024638>

Gabry, J., Simpson, D., Vehtari, A., Betancourt, M., & Gelman, A. (2019). Visualization in Bayesian workflow. *J. R. Stat. Soc. Ser. A Stat. Soc.*, 182(2), 389–402. <https://doi.org/10.1111/rssa.12378>

Galen, C. (1999). Why do flowers vary? The functional ecology of variation in flower size and form within natural plant populations. *Bioscience*, 49(8), 631–640. <https://doi.org/10.2307/1313439>

Gong, Y.-B., & Huang, S.-Q. (2009). Floral symmetry: Pollinator-mediated stabilizing

selection on flower size in bilateral species. *Proceedings of the Royal Society B: Biological Sciences*, 276(1675), 4013–4020. <https://doi.org/10.1098/rspb.2009.1254>

Goodwillie, C., Sargent, R. D., Eckert, C. G., Elle, E., Geber, M. A., Johnston, M. O., Kalisz, S., Moeller, D. A., Ree, R. H., Vallejo-Marin, M., & Winn, A. A. (2010). Correlated evolution of mating system and floral display traits in flowering plants and its implications for the distribution of mating system variation. *New Phytol.*, 185(1), 311–321. <https://doi.org/10.1111/j.1469-8137.2009.03043.x>

Goodwillie, C., & Weber, J. J. (2018). The best of both worlds? A review of delayed selfing in flowering plants. *American Journal of Botany*, 105(4), 641–655. <https://doi.org/10.1002/ajb2.1045>

Healy, K., Ezard, T. H. G., Jones, O. R., Salguero-Gómez, R., & Buckley, Y. M. (2019). Animal life history is shaped by the pace of life and the distribution of age-specific mortality and reproduction. *Nat. Ecol. Evol.*, 3(8), 1217–1224. <https://doi.org/10.1038/s41559-019-0938-7>

Hegland, S. J., & Totland, Ø. (2005). Relationships between species' floral traits and pollinator visitation in a temperate grassland. *Oecologia*, 145(4), 586–594. <https://doi.org/10.1007/s00442-005-0165-6>

Hung, K.-L. J., Kingston, J. M., Albrecht, M., Holway, D. A., & Kohn, J. R. (2018). The worldwide importance of honey bees as pollinators in natural habitats. *Proc. Royal Soc. B*, 285(1870), 20172140. <https://doi.org/10.1098/rspb.2017.2140>

Ibanez, S. (2012). Optimizing size thresholds in a plant-pollinator interaction web: Towards a mechanistic understanding of ecological networks. *Oecologia*, 170(1), 233–242. <https://doi.org/10.1007/s00442-012-2290-3>

Jin, Y., & Qian, H. (2019). VPhyloMaker: An R package that can generate very large phylogenies for vascular plants. *Ecography*, 42(8), 1353–1359. <https://doi.org/10.1111/ecog.04434>

Jordano, P. (1987). Patterns of mutualistic interactions in pollination and seed dispersal: Connectance, dependence asymmetries, and coevolution. *The American Naturalist*, 129(5), 657–677. <https://doi.org/10.1086/284665>

- Junker, R. R., Blüthgen, N., Brehm, T., Binkenstein, J., Paulus, J., Schaefer, H. M., & Stang, M. (2013). Specialization on traits as basis for the niche-breadth of flower visitors and as structuring mechanism of ecological networks. *Funct. Ecol.*, 27(2), 329–341. <https://doi.org/10.1111/1365-2435.12005>
- Kaiser-Bunbury, C. N., Vázquez, D. P., Stang, M., & Ghazoul, J. (2014). Determinants of the microstructure of plant–pollinator networks. *Ecology*, 95(12), 3314–3324. <https://doi.org/10.1890/14-0024.1>
- Kattge, J., Díaz, S., Lavorel, S., Prentice, I. C., Leadley, P., Bönsch, G., Garnier, E., Westoby, M., Reich, P. B., Wright, I. J., Cornelissen, J. H. C., Violle, C., Harrison, S. P., Bodegom, P. M. V., Reichstein, M., Enquist, B. J., Soudzilovskaia, N. A., Ackerly, D. D., Anand, M., ... Wirth, C. (2011). TRY – a global database of plant traits. *Glob. Chang. Biol.*, 17(9), 2905–2935. <https://doi.org/10.1111/j.1365-2486.2011.02451.x>
- Kettle, C. J., Maycock, C. R., Ghazoul, J., Hollingsworth, P. M., Khoo, E., Sukri, R. S. H., & Burslem, D. F. R. P. (2011). Ecological Implications of a Flower Size/Number Trade-Off in Tropical Forest Trees. *PLoS One*, 6(2), e16111. <https://doi.org/10.1371/journal.pone.0016111>
- Lagomarsino, L. P., Forrestel, E. J., Muchhala, N., & Davis, C. C. (2017). Repeated evolution of vertebrate pollination syndromes in a recently diverged andean plant clade. *Evolution*, 71(8), 1970–1985.
- Lanuza, J. B., Bartomeus, I., Ashman, T.-L., Bible, G., & Rader, R. (2021). Recipient and donor characteristics govern the hierarchical structure of heterospecific pollen competition networks. *Journal of Ecology*, 109(6), 2329–2341. <https://doi.org/10.1111/1365-2745.13640>
- Laughlin, D. C., Mommer, L., Sabatini, F. M., Bruelheide, H., Kuyper, T. W., McCormack, M. L., Bergmann, J., Freschet, G. T., Guerrero-Ramírez, N. R., Iversen, C. M., et al. (2021). Root traits explain plant species distributions along climatic gradients yet challenge the nature of ecological trade-offs. *Nat. Ecol. Evol.*, 1–12. <https://doi.org/10.1038/s41559-021-01471-7>
- Lázaro, A., Jakobsson, A., & Totland, Ø. (2013). How do pollinator visitation rate and

seed set relate to species' floral traits and community context? *Oecologia*, 173(3), 881–893. <https://doi.org/10.1007/s00442-013-2652-5>

Legendre, P., & Legendre, L. (2012). *Numerical ecology* (2nd edn). Elsevier, Amsterdam.

León-Osper, M., & Narbona, E. (2022). Unravelling the mystery of red flowers in the mediterranean basin: How to be conspicuous in a place dominated by hymenopteran pollinators. *Functional Ecology*, 36(11), 2774–2790. <https://doi.org/10.1111/1365-2435.14166>

Lloyd, D. G., & Schoen, D. J. (1992). Self-and cross-fertilization in plants. I. Functional dimensions. *International Journal of Plant Sciences*, 153(3, Part 1), 358–369. <https://doi.org/10.1086/297040>

Moeller, D. A., Runquist, R. D. B., Moe, A. M., Geber, M. A., Goodwillie, C., Cheptou, P.-O., Eckert, C. G., Elle, E., Johnston, M. O., Kalisz, S., Ree, R. H., Sargent, R. D., Vallejo-Marin, M., & Winn, A. A. (2017). Global biogeography of mating system variation in seed plants. *Ecol. Lett.*, 20(3), 375–384. <https://doi.org/10.1111/ele.12738>

Munoz, F., Violle, C., & Cheptou, P.-O. (2016). CSR ecological strategies and plant mating systems: Outcrossing increases with competitiveness but stress-tolerance is related to mixed mating. *Oikos*, 125(9), 1296–1303. <https://doi.org/10.1111/oik.02328>

Olesen, J. M., Bascompte, J., Dupont, Y. L., & Jordano, P. (2007). The modularity of pollination networks. *PNAS*, 104(50), 19891–19896. <https://doi.org/10.1073/pnas.0706375104>

Ollerton, J., Alarcón, R., Waser, N. M., Price, M. V., Watts, S., Cranmer, L., Hingston, A., Peter, C. I., & Rotenberry, J. (2009). A global test of the pollination syndrome hypothesis. *Annals of Botany*, 103(9), 1471–1480.

Osnas, J. L., Lichstein, J. W., Reich, P. B., & Pacala, S. W. (2013). Global leaf trait relationships: Mass, area, and the leaf economics spectrum. *Science*, 340(6133), 741–744. <https://doi.org/10.1126/science.1231574>

Paterno, G. B., Silveira, C. L., Kollmann, J., Westoby, M., & Fonseca, C. R. (2020). The maleness of larger angiosperm flowers. *Proc. Natl. Acad. Sci.*, 117(20), 10921–10926.

<https://doi.org/10.1073/pnas.1910631117>

- Penone, C., Davidson, A. D., Shoemaker, K. T., Marco, M. D., Rondinini, C., Brooks, T. M., Young, B. E., Graham, C. H., & Costa, G. C. (2014). Imputation of missing data in life-history trait datasets: Which approach performs the best? *Methods Ecol. Evol.*, 5(9), 961–970. <https://doi.org/10.1111/2041-210X.12232>
- Petit, R. J., & Hampe, A. (2006). Some evolutionary consequences of being a tree. *Annu. Rev. Ecol. Evol. Syst.*, 37, 187–214. <https://doi.org/10.1146/annurev.ecolsys.37.091305.110215>
- Poisot, T., Baiser, B., Dunne, J. A., Kéfi, S., Massol, F., Mouquet, N., Romanuk, T. N., Stouffer, D. B., Wood, S. A., & Gravel, D. (2016). Mangal – making ecological network analysis simple. *Ecography*, 39(4), 384–390. <https://doi.org/10.1111/ecog.00976>
- Revell, L. J. (2012). Phytools: An R package for phylogenetic comparative biology (and other things). *Methods Ecol. Evol.*, 3(2), 217–223. <https://doi.org/10.1111/j.2041-210X.2011.00169.x>
- Roddy, A. B., Martínez-Perez, C., Teixido, A. L., Cornelissen, T. G., Olson, M. E., Oliveira, R. S., & Silveira, F. A. O. (2021). Towards the flower economics spectrum. *New Phytol.*, 229(2), 665–672. <https://doi.org/10.1111/nph.16823>
- Rosas-Guerrero, V., Aguilar, R., Martén-Rodríguez, S., Ashworth, L., Lopezaraiza-Mikel, M., Bastida, J. M., & Quesada, M. (2014). A quantitative review of pollination syndromes: Do floral traits predict effective pollinators? *Ecol. Lett.*, 17(3), 388–400. <https://doi.org/10.1111/ele.12224>
- Rüger, N., Comita, L. S., Condit, R., Purves, D., Rosenbaum, B., Visser, M. D., Wright, S. J., & Wirth, C. (2018). Beyond the fast–slow continuum: Demographic dimensions structuring a tropical tree community. *Ecol. Lett.*, 21(7), 1075–1084. <https://doi.org/10.1111/ele.12974>
- Salguero-Gómez, R., Jones, O. R., Archer, C. R., Buckley, Y. M., Che-Castaldo, J., Caswell, H., Hodgson, D., Scheuerlein, A., Conde, D. A., Brinks, E., Buhr, H. de, Farack, C., Gottschalk, F., Hartmann, A., Henning, A., Hoppe, G., Römer, G., Runge, J., Ruoff, T., ... Vaupel, J. W. (2015). The compadre Plant Matrix Database: An

open online repository for plant demography. *J. Ecol.*, 103(1), 202–218. <https://doi.org/10.1111/1365-2745.12334>

Salguero-Gómez, R., Jones, O. R., Jongejans, E., Blomberg, S. P., Hodgson, D. J., Mbeau-Ache, C., Zuidema, P. A., de Kroon, H., & Buckley, Y. M. (2016). Fast-slow continuum and reproductive strategies structure plant life-history variation worldwide. *Proc. Natl. Acad. Sci. U.S.A.*, 113(1), 230–235. <https://doi.org/10.1073/pnas.1506215112>

Sargent, R. D. (2004). Floral symmetry affects speciation rates in angiosperms. *Proceedings of the Royal Society of London. Series B: Biological Sciences*, 271(1539), 603–608. <https://doi.org/10.1098/rspb.2003.2644>

Sargent, R. D., & Ackerly, D. D. (2008). Plant–pollinator interactions and the assembly of plant communities. *Trends Ecol. Evol.*, 23(3), 123–130. <https://doi.org/10.1016/j.tree.2007.11.003>

Sargent, R. D., Goodwillie, C., Kalisz, S., & Ree, R. H. (2007). Phylogenetic evidence for a flower size and number trade-off. *Am. J. Bot.*, 94(12), 2059–2062. <https://doi.org/10.3732/ajb.94.12.2059>

Schiestl, F. P., & Johnson, S. D. (2013). Pollinator-mediated evolution of floral signals. *Trends Ecol. Evol.*, 28(5), 307–315. <https://doi.org/10.1016/j.tree.2013.01.019>

Serneels, S., & Verdonck, T. (2008). Principal component analysis for data containing outliers and missing elements. *Comput. Stat. Data Anal.*, 52(3), 1712–1727. <https://doi.org/10.1016/j.csda.2007.05.024>

Shipley, B., Lechowicz, M. J., Wright, I., & Reich, P. B. (2006). Fundamental trade-offs generating the worldwide leaf economics spectrum. *Ecology*, 87(3), 535–541. <https://doi.org/10.1890/05-1051>

Smith, S. A., & Brown, J. W. (2018). Constructing a broadly inclusive seed plant phylogeny. *Am. J. Bot.*, 105(3), 302–314. <https://doi.org/10.1002/ajb2.1019>

Song, B., Sun, L., Barrett, S. C., Moles, A. T., Luo, Y.-H., Armbruster, W. S., Gao, Y.-Q., Zhang, S., Zhang, Z.-Q., & Sun, H. (2022). Global analysis of floral longevity reveals latitudinal gradients and biotic and abiotic correlates. *New Phytologist*. <https://doi.org/10.1111/nph.18271>

- Stang, M., Klinkhamer, P. G. L., Waser, N. M., Stang, I., & van der Meijden, E. (2009). Size-specific interaction patterns and size matching in a plant–pollinator interaction web. *Ann. Bot.*, 103(9), 1459–1469. <https://doi.org/10.1093/aob/mcp027>
- Stearns, S. C. (1989). Trade-offs in life-history evolution. *Funct. Ecol.*, 3(3), 259–268. <https://doi.org/10.2307/2389364>
- Stebbins, G. L. (1970). Adaptive radiation of reproductive characteristics in angiosperms, i: Pollination mechanisms. *Annual Review of Ecology and Systematics*, 307–326. <https://doi.org/10.1146/annurev.es.01.110170.001515>
- Stekhoven, D. J., & Bühlmann, P. (2012). MissForest—non-parametric missing value imputation for mixed-type data. *Bioinformatics*, 28(1), 112–118. <https://doi.org/10.1093/bioinformatics/btr597>
- Tong, Z.-Y., & Huang, S.-Q. (2016). Pre-and post-pollination interaction between six co-flowering pedicularis species via heterospecific pollen transfer. *New Phytologist*, 211(4), 1452–1461. <https://doi.org/10.1111/nph.14005>
- Tur, C., Castro-Urgal, R., & Traveset, A. (2013). Linking Plant Specialization to Dependence in Interactions for Seed Set in Pollination Networks. *PLoS One*, 8(10), e78294. <https://doi.org/10.1371/journal.pone.0078294>
- Vázquez, D. P., Lomáscolo, S. B., Maldonado, M. B., Chacoff, N. P., Dorado, J., Stevani, E. L., & Vitale, N. L. (2012). The strength of plant–pollinator interactions. *Ecology*, 93(4), 719–725. <https://doi.org/10.1890/11-1356.1>
- Vázquez, D. P., Melián, C. J., Williams, N. M., Blüthgen, N., Krasnov, B. R., & Poulin, R. (2007). Species abundance and asymmetric interaction strength in ecological networks. *Oikos*, 116(7), 1120–1127. <https://doi.org/10.1111/j.0030-1299.2007.15828.x>
- Vázquez, D. P., Morris, W. F., & Jordano, P. (2005). Interaction frequency as a surrogate for the total effect of animal mutualists on plants. *Ecol. Lett.*, 8(10), 1088–1094. <https://doi.org/10.1111/j.1461-0248.2005.00810.x>
- Waser, N. M., Chittka, L., Price, M. V., Williams, N. M., & Ollerton, J. (1996). Generalization in pollination systems, and why it matters. *Ecology*, 77(4), 1043–1060.

<https://doi.org/10.2307/2265575>

Wilson, P., Castellanos, M. C., Hogue, J. N., Thomson, J. D., & Armbruster, W. S. (2004).

A multivariate search for pollination syndromes among penstemons. *Oikos*, 104(2),

345–361. <https://doi.org/10.1111/j.0030-1299.2004.12819.x>

Worley, A. C., Baker, A. M., Thompson, J. D., & Barrett, S. C. (2000). Floral display

in narcissus: Variation in flower size and number at the species, population, and

individual levels. *Int. J. Plant Sci.*, 161(1), 69–79. <https://doi.org/10.1086/314225>

Wright, I. J., Reich, P. B., Westoby, M., Ackerly, D. D., Baruch, Z., Bongers, F., Cavender-

Bares, J., Chapin, T., Cornelissen, J. H. C., Diemer, M., Flexas, J., Garnier, E., Groom,

P. K., Gulias, J., Hikosaka, K., Lamont, B. B., Lee, T., Lee, W., Lusk, C., ... Villar,

R. (2004). The worldwide leaf economics spectrum. *Nature*, 428(6985), 821–827.

<https://doi.org/10.1038/nature02403>