Covariation among reproductive traits in flowering plants shape their interactions with pollinators

Abstract

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- 1. Globally, plants display enormous variation in life history strategies and trait combinations. However, evidence suggests that evolutionary and physiological constraints limit the number of plant ecological strategies. Although there have been recent advances in understanding correlations among plant traits, reproductive traits are rarely considered, despite their key role in shaping plant life-history strategies and interactions with pollinators.
 - 2. Here, using a global dataset of 18 reproductive traits for 1,506 species, we investigate the reproductive spectrum of flowering plants to identify how it shapes interactions with pollinators.
- 3. We show that over 50% of all trait variation is explained by the first two reproductive axes, which represent the negative correlation between flower number and flower size, and the negative correlation between autonomous selfing and floral display size. In addition, these reproductive axes were associated with different numbers of interacting partners and visitation rates of the distinct pollinator guilds. However, reproductive axes explain a relatively small amount of variance in pollinator interactions highlighting the need to incorporate other factors along with reproductive traits to fully explain large scale patterns of plant-pollinator interactions.
- 4. Our study identifies the major reproductive trait correlations in flowering plants and their role in shaping plant-pollinator interactions at a macro-ecological scale.

- These findings emphasize the importance of considering reproductive traits in the global spectrum of plant form and function, and the need to explore beyond floral morphological traits to broaden our understanding of plant-pollinator interactions.
- Keywords: life-history strategies | plant reproductive traits | plant-pollinator interactions

1. Introduction

Flowering plants display an astonishing diversity of reproductive structures (Barrett, 33 2002; Schiestl & Johnson, 2013) that shape their interactions with animal pollinators (Dellinger, 2020; Fenster et al., 2004). The variety of reproductive strategies used by 35 different plant species, from specialised reliance on a single pollinator, to full selfpollination, is incredibly rich. However, not all reproductive trait combinations in 37 plants are possible due to evolutionary and ecological constraints (Agrawal, 2020; Stearns, 1989). Despite recent advances in theoretical and empirical understanding of correlations between plant reproductive traits at large ecological scales (Friedman, 2020; Paterno et al., 2020; Roddy et al., 2021; Salguero-Gómez et al., 2016; Song et al., 41 2022), studies typically focus on just a few traits, often in isolation. Thus, to develop a comprehensive understanding of the plant reproductive spectrum of trait variation, as done recently for root (Laughlin et al., 2021), leaf (Wright et al., 2004) and wood (Chave et al., 2009) related traits, we require a multi-trait perspective with broad geographical coverage. Importantly, characterization of the plant reproductive spectrum could reveal key reproductive strategies among different species (Agrawal, 2020) and improve knowledge of associations between specific reproductive traits and pollinators (Roddy et al., 2021). With the recent availability of large trait databases (e.g., TRY Kattge et al., 2011; COM-

PADRE Salguero-Gómez et al., 2015), there has been increased research on plant ecolog-

ical strategies, which has produced identification of global patterns in plant form and function (Bruelheide et al., 2018; Carmona et al., 2021; Díaz et al., 2016; Salguero-Gómez et al., 2016). However, studies that look at multiple traits tend to overlook plant repro-54 ductive traits (E-Vojtkó et al., 2020; Roddy et al., 2021; Rüger et al., 2018), and focus on 55 trait correlations in other plant organs such as morphological and physiological leaf traits (Donovan et al., 2011; Osnas et al., 2013; Shipley et al., 2006; Wright et al., 2004). Despite the lack of a holistic understanding that depicts reproductive trait covariation 58 patterns, there are widely recognized associations between plant reproductive trait pairs such as the negative correlation between flower size and flower number (E-Vojtkó et al., 2022; Kettle et al., 2011; Sargent et al., 2007); the positive association between flower size and outcrossing rate (Goodwillie et al., 2010); or the positive correlation 62 between outcrossing rate and lifespan (Barrett, 2003; Moeller et al., 2017; Munoz et al., 2016). Although identification of these trait correlations has allowed progress toward a conceptual framework that integrates different floral trait relationships (Roddy et al., 2021), we still lack empirical understanding of how these associations among floral and other reproductive traits (e.g., reproductive biology) shape plant life history strategies. Trait-based approaches (Fenster et al., 2004; Rosas-Guerrero et al., 2014) and traitmatching analyses (Bartomeus et al., 2016; Stang et al., 2009) are powerful for exploring the underlying mechanisms driving plant-pollinator interactions. For example, plant traits can define species' network roles (e.g., specialists vs generalists Tur et al., 2013) 71 and plant species that occupy reproductive trait space extremes are likely to be more 72 specialised and display tighter trait-matching with pollinators (Coux et al., 2016; Junker 73 et al., 2013). Indeed, the degree of morphological matching between plants and pollinators often determines whether or not interactions occur, and consequently influences the broader network structure (Ibanez, 2012; Stang et al., 2009). However, it is still 76 unclear how specific reproductive traits (e.g., mating or compatibility system) influence 77 plant-pollinator associations (Devaux et al., 2014; Sargent & Ackerly, 2008; Tur et al., 2013). For example, species with high selfing rates may evolve to invest less in floral rewards resulting in a lower number of pollinator visits (Devaux et al., 2014). Indeed,

both morphology and mating system can determine a species' functional role within pollination networks, and the combination of several traits increases power for predicting network structure (Eklöf et al., 2013). Because the species position within the multidimensional trait space can result in different interaction patterns or adaptations to pollinators (see Dehling et al., 2016 for a purely morphological approach), the use of the reproductive trait space to explore plant-pollinator interactions could help to identify species functional roles and associations of traits with specific pollinators (i.e., pollination syndromes).

Here, we explore the spectrum of reproductive traits in flowering plants, within a global set of plant-pollinator networks, to identify reproductive trait covariation and how this influences the network of interactions between plants and pollinators. First, we identify the major axes of reproductive trait variation and correlations among traits for plant species. Then, we investigate the associations between plant species' positions in reproductive trait space and the strength of their interactions with different pollinator guilds within networks. Finally, we determine how the main axes of reproductive trait variation influence plant species' functional roles within pollination networks, using a range of complementary interaction network metrics.

2. Methods

99 (a) Plant-pollinator network studies

We selected 28 studies from 18 different countries that constituted a total of 64 plantpollinator networks (see table S1 and figure S1). These studies recorded plant-pollinator
interactions in natural systems and were selected so that we had broad geographical
representation across different ecological communities. The pollinators documented by
these studies are primarily insect pollinators (97.8% of the total visits recorded) and
the plants are characterised by a clear dominance of smaller life forms (i.e., 61% of
herbs, 28% of shrubs and 11% of trees). Although these studies differ in sampling effort

and methodology, all studies provided information about plant-pollinator interactions 107 (weighted and non-weighted), which we used to build a database of plant species 108 that are likely to benefit, to some degree, from animal pollination. Many of these 109 networks are freely available either as published studies (e.g., Carvalheiro et al., 2014; 110 Fortuna et al., 2010; Olesen et al., 2007) or in online archives (e.g., Mangal Poisot et al., 111 2016; The Web of Life Fortuna et al., 2010). In total, our network dataset constituted 112 60 weighted (number of visits) and four unweighted (interaction presence-absence) 113 networks, each sampled at a unique location and year, as well as eight meta-networks 114 where interactions were pooled across several locations and multiple years.

(b) Taxonomy of plants and pollinators

All species names, genera, families and orders were retrieved and standardised from the taxonomy data sources NCBI (https://www.ncbi.nlm.nih.gov/taxonomy) for plants and ITIS (https://www.itis.gov/) for pollinators, using the R package *taxize* (Chamberlain et al., 2020). We filled the 'not found' searches manually using http: //www.theplantlist.org/ and http://www.mobot.org/ for plants and http://www.ca talogueoflife.org/ for pollinators.

123 (c) Plant traits

We selected a total of 18 quantitative and categorical functional traits that included reproductive traits and traits related with plant form and size (see table 1). Reproductive traits consisted of those related to floral and reproductive biology: (i) floral traits included those related to the size of floral organs (e.g., style length), floral display size (e.g., flower number) and floral rewards (e.g., pollen quantity), while (ii) reproductive biology traits represented the plant reproductive system (i.e., the breeding, mating and compatibility system). The remaining traits were related to plant size and form (i.e., plant height, lifespan and life form), which are commonly used to characterise the fast-slow continuum of plant trait variation (Salguero-Gómez et al., 2016). For each plant species, we undertook an extensive literature and online search for all traits across

Table 1. Quantitative and categorical traits used in this study.

Traits	Numerical range/Categories
Quantitative	
Plant height	0.01-30m
Flower width	0.50-205mm
Flower length	0.20-195mm
Inflorescence width	0.80-300mm
Style length	0-138mm
Ovules per flower	1-25000
Flowers per plant	$1-1.8 \times 10^5$
Microliters of nectar	$0-160\mu$ l
Milligrams of nectar	0-6.1mg
Pollen grains per flower	$13-2 \times 10^8$
Autonomous selfing (fruit set)	0-100%
Categorical	
Lifepan	Short-lived and perennial
Life form	Herb, shrub and tree
Flower shape	Brush, campanulate, capitulum,
	open, papilionaceous and tube
Flower symmetry	Actinomorphic and zygomorphic
Nectar	Presence and absence
Compatibility system	Self-incompatible, partially self-compabtile and self-compatible
Breeding system	Hermaphrodite, monoecious and dioecious

¹³⁴ a wide range of resources (plant databases, online floras, books, journals and images).

From a matrix comprising a total of 27,108 possible cells (considering all traits and

plant species: 18 columns \times 1,506 species), we were able to fill 22,796 cells (84.1% of

the dataset, see figure S2 for missing value information for each trait). An extended

description of each trait and how it was obtained can be found in Appendix S1.

139 (d) Phylogenetic Distance

¹⁴⁰ We calculated the phylogenetic distance between all plant species considered using the

function *get_tree* from the package *rtrees* (https://github.com/daijiang/rtrees), which

downloads phylogenetic distances from the extended R implementation of the Open

Tree of Life (Jin & Qian, 2019; Smith & Brown, 2018).

(e) Data Imputation

To avoid excluding species with trait missing values, we imputed trait missing values 145 with the help of the function *missForest* (Stekhoven & Bühlmann, 2012) which allows 146 imputation of datasets with continuous and categorical variables. We accounted for 147 the phylogenetic distance among species on the imputation process by including the 148 eigenvectors from principal component analysis of the phylogenetic distance (PCoA), which has been shown to improve the performance of *missForest* (Penone et al., 2014). 150 To extract the eigenvectors, we used the function *PVRdecomp* from the package *PVR* (Chamberlain et al., 2018) based on a previous conceptual framework that considers 152 phylogenetic eigenvectors (Diniz-Filho et al., 2012). We conducted two different imputations, one for the full set of species (1,506 species: 5.79% of missing values), and 154 excluding nectar and pollen traits because of the high percentage of missing values 155 (figure S2), and a second one for the subset of species with data for pollen per flower 156 and microliters of nectar (755 species: 8.01% of missing values). To corroborate that our 157 imputation of missing values did not affect our results, we evaluated the reproductive 158 spectrum (see section below) with and without imputed values and results for the two 159 datasets were consistent (Fig S3 and Fig S4). 160

161 (f) Plant strategies analysis

We explored the association between the different quantitative plant traits using a 162 phylogenetically informed Principal Component Analysis (pPCA). We did not include 163 the quantitative variables of flower length and inflorescence width because they were 164 highly and moderately correlated to flower width, respectively (Pearson's correlation 165 = 0.72, P < 0.01 and Pearson's correlation = 0.36, P < 0.01), and thus we avoided 166 overemphasise flower size on the spectrum of trait variation. Prior to the analyses, we 167 excluded outliers and standardised the data. Specifically, due to the high sensitivity of 168 dimensionality reduction to outliers (Legendre & Legendre, 2012; Serneels & Verdonck, 2008), we excluded values outside the 2.5th–97.5th percentile range, and thus our final

dataset had 1,236 species. Then, we log transformed all continuous variables to reduce 171 the influence of extreme values and then z-transformed (X=0, SD=1) these variables 172 so that they were within the same numerical range, as recommended for principal 173 component analysis (Legendre & Legendre, 2012). Although qualitative traits were not 174 included in the dimensionality reduction analysis, we also investigated their position 175 in the trait space and the statistical association of the different qualitative traits with the 176 main axes of trait variation using an ANOVA and Tukey test. Similarly, we also explored 177 for each pollinator guild (see "Visitation patterns" subsection below) the position in the 178 trait space of the plants that interacted with them. We performed the pPCA using the 179 function *phyl.pca* from the package *phytools* (Revell, 2012) with the method lambda (λ) 180 that calculates the phylogenetic correlation between 0 (phylogenetic independence) and 181 1 (shared evolutionary history) and we implemented the mode "covariance" because values for each variables were on the same scale following transformation (Abdi & 183 Williams, 2010). Finally, we conducted an additional phylogenetic informed principal 184 component analysis for the subset of species with pollen and nectar quantity data. For 185 this, we included all quantitative traits included in the main pPCA, plus pollen grains 186 and microlitres of nectar per flower. 187

188 (g) Phylogenetic signal

We calculated the phylogenetic signal of the different quantitative traits on the imputed dataset with the full set of species (N = 1,506) with the package *phytools* (Revell, 2012) and used Pagel's λ as a measurement of the phylogenetic signal. However, for pollen and nectar traits, phylogenetic signal was calculated only on the subset of species that had quantitative information for these traits (N = 755).

194 (h) Network analyses

195 (i) Visitation patterns

We used Bayesian modelling (see details below) to investigate how the different guilds 196 of pollinators interacted along the main axes of reproductive trait variation using qualitative and quantitative measures of plant-pollinator interactions. Specifically, 198 interactions between plants and pollinators in binary networks were defined as the presence or absence of floral visitation, while in weighted networks, interactions were 200 measured as the the number of visits from each insect species to different plant species. Although floral visitors are not always pollinators and the number of visits does not 202 consider the pollination efficiency of each species (Ballantyne et al., 2015), the number of visits does provide valuable information about the relative contributions different 204 floral visitors make to pollination (Vázquez et al., 2005, 2012). Therefore, despite the fact that we do not evaluate pollinator effectiveness we refer to the different floral 206 visitors as pollinators. 207

We divided pollinators into six main taxonomical guilds that differ in life form, 208 behaviour and thus likely play different ecological roles: (i) bees (Hymenoptera-209 Anthophila), (ii) non-bee-Hymenoptera (Hymenoptera-non-Anthophila), (iii) syrphids (Diptera-Syrphidae), (iv) non-syrphid-Diptera (Diptera-non-Syrphidae), (v) Lepi-211 doptera, and (vi) Coleoptera. Note that we excluded non-insect pollinators from our 212 analyses as they comprised only a 2.2% of the total visits recorded in our set of studies. 213 Moreover, because bees had the greatest number of occurrences in networks (2,256 214 records) and the highest frequency of visits, we further broke this group down into the 215 main bee families (Andrenidae, Apidae, Colletidae, Halictidae, and Megachilidae). In 216 addition, we found that *Apis mellifera* was the pollinator with the largest proportion of 217 occurrences in networks (7.55% of the total). This is consistent with previous research 218 showing that A. mellifera was the most frequent pollinator in a similar dataset of 80 219 plant-pollinator networks in natural ecosystems (Hung et al., 2018). Hence, to control 220 for the effect of *A. mellifera* on observed visitation patterns of bees, we conducted an

analogous analysis with presence-absence of the interaction and number of visits excluding *A. mellifera*. We found that *A. mellifera*, was partly driving some of the observed trends on PC1 (figure S5). However, we did not detect major differences on PC2 and PC3.

We used Bayesian generalised linear mixed models in the R package brms (Bürkner, 226 2017) to model the presence-absence of observed interactions and number of visits 227 as a function of the main axes of plant trait variation (i.e., PC) and their interactions 228 with pollinator guilds (i.e., PGs; e.g., number of visits ~ PC1 x PGs + PC2 x PGs + 229 PC3 x PGs). Because we were interested in possible differences in visitation patterns 230 among pollinator guilds to plants with different reproductive strategies, we included 231 interactions between the main axes of trait variation (PC1, PC2 and PC3) and the different pollinator guilds in the model. We added a nested random effect of networks 233 within the study system to capture the variation in networks among studies and within 234 networks. Moreover, we included the phylogenetic covariance matrix as a random 235 factor due to the possible shared evolutionary histories of species and therefore lack of independence among them. We specified for the presence or absence of an interaction 237 and number of visits with Bernoulli and a zero inflated negative binomial distributions, respectively. The models were run with 3,000 iterations with previous 1,000 warm up 239 iterations and with non or very weakly informative priors from the brm function so the priors had negligible influence on the results (Bürkner, 2017). We set delta (Δ) to 0.99 241 to avoid divergent transitions and visualised the posterior predictive checks with the 242 function *pp_check* using the *bayesplot* package (Gabry et al., 2019). 243

(ii) Plant species network roles

We also used Bayesian modelling to investigate how the main axes of trait variation determined different plant species' functional roles within pollination networks. For this, we selected complementary species-level network metrics, commonly applied in bipartite network studies (Dormann et al., 2008), with clear ecological interpretations and are relevant for defining species' functional roles in networks. Specifically, the

species-level metrics we included were: (i) sum of visits per plant species; (ii) nor-250 malised degree, calculated as the number of links per plant species divided by the 251 total possible number of partners; and (iii) selectivity (d') (Blüthgen et al., 2006), which 252 measures the deviation of an expected random choice of the available interaction part-253 ners and ranges between 0 (maximum generalisation) and 1 (maximum specialisation). 254 Normalised degree and selectivity were calculated with the *specieslevel* function in the 255 R package bipartite (Dormann et al., 2008). We modelled each plant species metric 256 (sum of visits, normalised degree and plant selectivity) as a function of the three main 257 axes of trait variation (plant species level metric ~ PC1 + PC2 + PC3). For the different 258 response variables in each model (i.e., each plant species level metric), we used different 259 distribution families (zero inflated negative binomial for the sum of visits, weibull for 260 normalised degree, and zero-one inflated beta for selectivity). Finally, we used the same random factors, model settings and conducted the same posterior predictive checks for 262 each model as detailed above in the 'visitation patterns' section.

Our analyses were conducted on the subset of 60 weighted networks (out of a total of 64 networks) sampled in a unique flowering season and site, which included 556 plant and 1,126 pollinator species. In total, our network dataset (excluding meta- and non-weighted networks) included 2,256 interactions for bees, 1,768 for non-syrphid-dipterans, 845 for syrphids, 437 for lepidopterans, 432 for coleopterans, and 362 for non-bee-hymenopterans. Sampling methods varied across studies but this was accounted for in analyses by including study identity as a random effect in our models. All analyses were conducted in R version 4.0.3.

3. Results

273 (a) Plant strategies analysis

Our phylogenetically informed principal component analysis captured 51.8% and 70.97% of the reproductive trait variation in the first two and three axes, respectively

(figure 1 and figure S6). The first principal component (PC1; the flower number - flower size axis) represented 26.72% of the trait variation and indicated a negative correlation 277 between flower number and flower size. The main contributing traits to PC1 were 278 plant height, flower number, ovule number and flower size (loadings > | 0.5|; table 279 S3) but style length also contributed moderately (loading = -0.33). One end of this 280 axis comprised species with high investment in flower number and plant height but 281 small flower size, short style length and low ovule number. For instance, Cornus florida 282 occurred at this end of the spectrum, which, on average, has approximately 10,000 283 flowers, is 7.5 m high, has flowers that are 3 mm wide, styles 3.5 mm long and two 284 ovules per flower. The other end of this spectrum had species that were short in height, 285 with few large flowers that had long styles and many ovules. For example, *Petunia* 286 axillaris on average, is 0.5 m high and has approximately 10 flowers per plant that are 287 over 50 mm wide with 25 mm long styles and over 200 ovules. The second principal 288 component (PC2; the autonomous selfing - floral display axis) encompassed 25.05% 289 of the trait variation and represented variation from low to high autonomous selfing 290 (i.e., high to low pollinator dependence). The main driver of trait variation on PC2 was 291 autonomous selfing (loading = 0.85) but all other traits (except ovule number) made 292 moderate contributions (loadings ranging between 0.27 to 0.4; table S3). Generally speaking, species with low autonomous selfing had larger and many flowers with long 294 styles, and were tall. Zuccagnia punctata occurred at this extreme of the spectrum, which 295 is a self-incompatible species, completely dependent on animal pollination for seed 296 production, producing approximately 1,500 flowers per plant with an average height of 297 3 m and 20 mm long styles. In contrast, species with high autonomous selfing tended 298 to have fewer and smaller flowers with shorter styles, and were shorter in height. 299 Veronica peregrina occurred at this end of the spectrum, and is a self-compatible plant 300 that requires little or no pollination by animals, it has approximately 20 flowers per 301 plant, is 0.2 m high and has 0.25 mm long styles. The third principal component (PC3; 302 the style length - autonomous selfing axis) explained a considerable amount of trait 303 variability (19.17%) and represented a positive correlation between style length (loading 304

= -0.66) and autonomous selfing (loading = |-0.51|), where species with short styles 305 had low autonomous selfing and species with long styles high autonomous selfing. 306 The remaining traits, apart from ovule number, were all positively correlated with style 307 length and autonomous selfing (loadings from -0.23 to -0.46; table S3). In addition, the 308 pPCA with the subset of species that we had nectar and pollen quantity data for showed 309 that nectar quantity (microlitres of nectar per flower) and pollen grains per flower were 310 positively associated with floral display size. Floral rewards were particularly strongly 311 correlated with flower size but weakly correlated with flower number (figure S4). The 312 first two principal components for this pPCA explained similar variance (45.52%) and 313 similar associations of traits, despite some variability in the loadings (table S4).

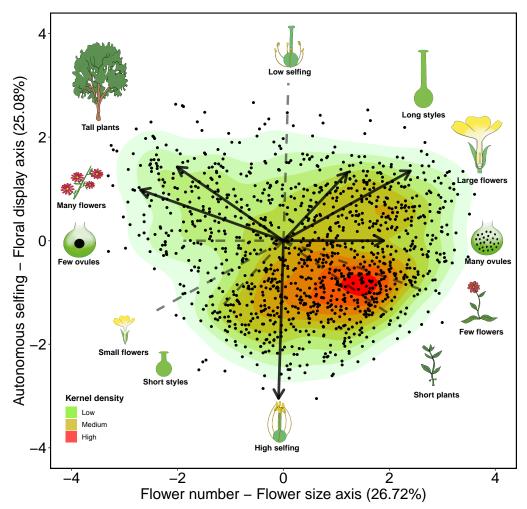


Figure 1. Phylogenetically informed principal component analysis (pPCA) of 1,236 plant species from 28 plant-pollinator network studies. The solid arrows indicate the direction of the different quantitative traits (flower number, plant height, style length, flower size, ovule number and level of autonomous selfing) across the two main axes of trait variation. The length of the arrows indicate the weight of the variables on each principal component and the dashed lines show the opposed direction of trait variation. The icons at both ends of arrows and dashed lines illustrate the extreme form of the trait continuum.

As expected, due to individual trait correlations between qualitative and quantitative traits (e.g., life form and plant height), we found that most categorical traits were clustered on the trait space and statistically associated with the first two axes of trait variation (figure s2 and table s2). However, the position of plants connected to each of the pollinator guilds was scattered without clear aggregation patterns on the trait space (figure s7; but see "Visitation patterns" section below for detailed results from quantitative analyses). Flower symmetry, which was only associated with PC2 (Sum of squares

= 8.51, F-value = 14.72, P < 0.01), and nectar provision, which was independent of PC1 and PC2 (PC1: Sum of squares = 0.37, F-value = 0.29, P = 0.59; PC2: Sum of squares = 323 0.83, F-value = 1.43, P = 0.23) showed a lack of statistical association. In addition, we 324 found statistical differences between the different categorical trait levels within the trait 325 space (figure S8). Regarding self-compatibility, we found larger differences on PC2. For 326 instance, species with unisexual flowers that were self-incompatible, were statistically 327 differentiated from species with partial or full self-compatibility on this axis (figure S8a 328 and figure S8b; Tukey test P < 0.05). Life forms differed statistically across both axes of 329 trait variation and followed a gradient of larger life forms (trees and shrubs) with lower 330 autonomous selfing to smaller ones (herbs) with higher autonomous selfing (figure 331 S8c and figure S8d; Tukey test P < 0.05). Consequently, lifespan also followed this 332 gradient but perennial and short lived species only differed statistically on PC2 (figure 333 S8e and figure S8f; Tukey test P < 0.05). Species with unisexual flowers (monoecious 334 and dioecious) were clustered on both extremes of the first two principal components 335 and had both the lowest autonomous selfing levels and the highest number of flowers 336 (figure S8g and figure S8h; Tukey test P < 0.05). Moreover, we found that campanulate 337 and capitulum flower shapes were differentiated from tube, papilionaceous, open and 338 brush shapes in the trait space; the former had larger flowers and low autonomous selfing levels, while the latter had more flowers and high autonomous selfing (figure S8i 340 and figure S8j; Tukey test P < 0.05). Finally, in terms of flower symmetry, zygomorphic flowers were associated with high levels of autonomous selfing, whereas actinomorphic 342 flowers were more dependent on pollinators (figure S8k and figure S8l; Tukey test *P* < 0.05).

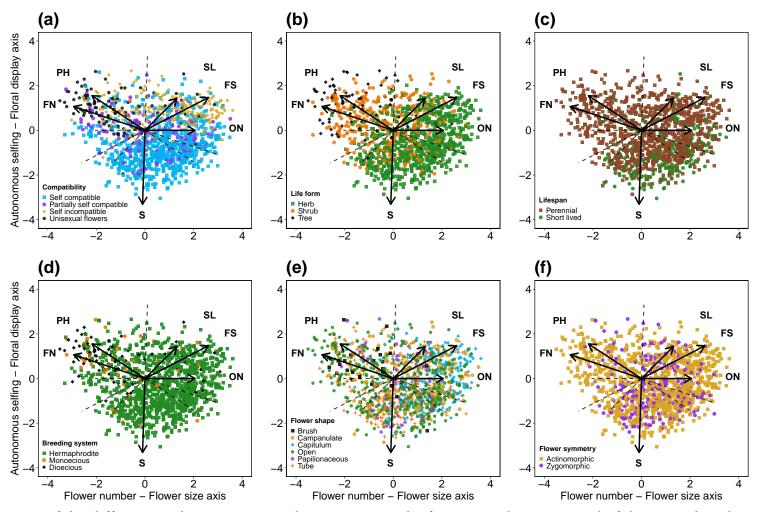


Figure 2. Location of the different qualitative traits in the trait space. The figure panel is composed of the traits that showed statistical association with the first two axes of trait variation: compatibility system (a), life form (b), lifespan (c), breeding system (d), flower shape (e) and flower symmetry (f). The solid arrows indicate the direction of variation of the different quantitative traits shown in figure 1: flower number (FN), plant height (PH), style length (SL), flower size (FS), ovule number (ON) and the level of autonomous selfing (S).

345 (b) Phylogenetic signal

We found a strong phylogenetic signal (P < 0.01) for most quantitative traits (table S5). The traits that showed the highest phylogenetic signal were ovule number ($\lambda = 1$), pollen grains per flower ($\lambda = 1$) and plant height ($\lambda = 0.96$), followed by flower length ($\lambda = 0.75$), flower width ($\lambda = 0.73$) and number of flowers per plant ($\lambda = 0.69$). The traits that had a moderate phylogenetic signal included inflorescence width ($\lambda = 0.57$), style length ($\lambda = 0.49$) and autonomous selfing ($\lambda = 0.34$). Finally, microliters of nectar per flower showed the lowest phylogenetic signal of all the traits we measured ($\lambda = 0.14$).

353 (c) Visitation patterns

The main axes of trait variation (PC1, PC2 and PC3) partly explained the presence-354 absence of interaction partners (conditional $R^2 = 0.26$; marginal $R^2 = 0.20$) but little of the total number of visits (conditional $R^2 = 0.31$; marginal $R^2 = 0.06$). However, we found 356 relevant differences across the different pollinator guilds for both the presence-absence 357 of interactions and number of visits (figure 3). On the flower number - flower size 358 axis, we found that plants with many small flowers tended to have interaction partners 359 that were Coleoptera, non-bee-Hymenoptera and Diptera (figure 3a), whereas plants 360 with few large flowers mostly had interaction partners that were bees and Lepidoptera. 361 We found similar trends for visitation rates on the flower number - flower size axis, 362 but bees and syrphids visited plant species with many small flowers more frequently 363 (figure 3d). On the autonomous selfing - floral display axis, we found that all plant 364 species with lower autonomous selfing had many interaction partners and recieved 365 many visits from all pollinator guilds (figure 3b and figure 3e). Finally, on the style length - autonomous selfing axis, plant species with short styles and low autonomous 367 selfing had more interaction partners for all guilds, except bees which interacted more frequently with plant species with long styles and high autonomous selfing (figure 3c). 369 Plants with long styles and high autonomous selfing interacted more frequently with Lepidoptera and non-bee-Hymenoptera (figure 3f).

The additional models for the most represented bee families, for both presence-absence of interactions (marginal $R^2 = 0.29$; conditional $R^2 = 0.19$) and the number of visits (marginal $R^2 = 0.30$; conditional $R^2 = 0.03$), showed that the Apidae family was the main driver of the observed patterns. The contrast between interaction presence/absence and number of visits for bees on PC1 (figure 3a and figure 3d) was driven by the Andrenidae family, which had more interaction partners but made fewer visits to plant species with few large flowers (figure S9).

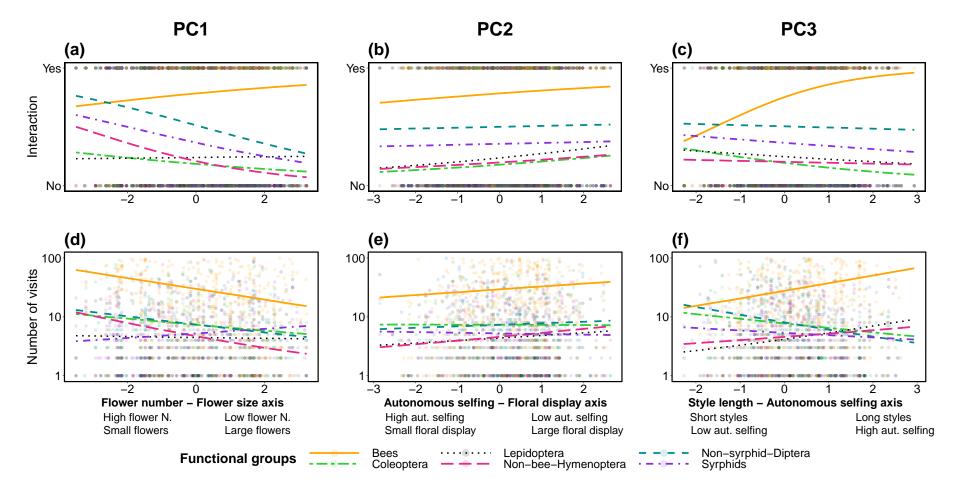


Figure 3. Fitted posterior estimates of the presence-absence of interaction (a, b and c) and number of visits (d, e and f) of the different pollinator guilds in relation to the main axes of plant reproductive trait variation (PC1, PC2 and PC3). PC1 represents the flower number - flower size axis, PC2 represents the autonomous selfing - floral display axis and PC3 represents the style length - autonomous selfing axis. For visualization purposes due to large differences between number of visits from bees and the rest of the pollinator guilds, the number of visits was log-transformed (Y-axis of lower panel).

(d) Plant species network roles

Variance within plant species-level network metrics was poorly explained by the three 380 main axes of trait variation (figure S10; number of visits \sim PCs, conditional $R^2 = 0.11$, 381 marginal $R^2 = 0.02$; normalised degree ~ PCs, conditional $R^2 = 0.24$, marginal $R^2 = 0.02$; 382 and, selectivity ~ PCs, conditional $R^2 = 0.37$, marginal $R^2 = 0.03$). Overall, the most 383 notable trends were found on the flower number - flower size axis (PC1) and the style length - autonomous selfing axis (PC3) for number of visits and selectivity. On PC1, 385 the number of visits per flower was higher for plant species with more flowers but 386 was lower for plant species with larger flowers (figure S10a). However, selectivity (d') 387 was higher for plant species with large flowers but lower for plant species with small flowers (figure S10g). On PC3, the number of visits was lower for plants with shorter 389 styles and lower autonomous selfing but higher for plant species with longer styles and higher autonomous selfing (figure S10c). Again, selectivity showed the opposite trend 391 to the number of visits (figure S10i).

4. Discussion

We found strong covariation among traits within the flowering plant reproductive spectrum, which shapes interactions with pollinators at a global scale. Despite the 395 enormous variability in plant reproductive structures, the first two and three axes 396 captured over 50% and 70% of trait variation, respectively, and these axes encapsulated 397 correlations among flower number and flower size, autonomous selfing and floral 398 display, and style length and autonomous selfing. Although explained trait variation 399 on the first two axes was less than previous studies of morphological and physiological 400 traits (Carmona et al., 2021; Díaz et al., 2016), our results align with the two largest 401 and most recent studies of plant life strategies that included reproductive, form and 402 size related traits (E-Vojtkó et al., 2022; Salguero-Gómez et al., 2016). The different 403 plant reproductive axes that we identified were associated with different numbers of 404 interacting partners and visitation rates of the distinct pollinator guilds. However, the unexplained variability found highlights the need to account for other local factors, such as species relative abundances (Bartomeus et al., 2016), along with reproductive traits, to better explain plant-pollinator associations.

Our study provides strong evidence for several widely discussed trait associations 409 with the help of a great diversity of phylogenetic lineages (170 families and over 410 1,200 species). Specifically, by using a multitrait approach, we are able to link the 411 reproductive spectrum of plant trait variation with the previously described "fast-slow 412 continuum" in plant (Salguero-Gómez et al., 2016) and animal (Healy et al., 2019) 413 life-history strategies. This fast-slow continuum was illustrated by the associations 414 between reproductive traits and plant height, life form and lifespan traits. For instance, 415 although the negative correlation between flower number and flower size has received substantial attention (Kettle et al., 2011; Sargent et al., 2007; Worley et al., 2000), it has 417 only been investigated across a few hundred Angiosperm species and generally lacking 418 a multitrait perspective. We found support for this trait association and show that other 419 reproductive traits (e.g., style length or ovule number) covary along this flower number - flower size axis. Our study also complements previous work linking plant breeding 421 systems and life forms (Friedman, 2020; Petit & Hampe, 2006) by showing that most 422 species with unisexual flowers also have woody life forms and many flowers. Moreover, 423 we identify the previously documented positive association between outcrossing rate 424 and floral display size (Goodwillie et al., 2010), where species with low autonomous 425 selfing levels also allocate large resource quantities to attract pollinators. 426

Although we did not include floral rewards in our main analysis due to insufficient data, we did find that plant species with large floral displays and, in particular, large flowers, tended to have higher pollen and nectar quantity. Thus, we demonstrate that globally, reproductive trait covariation in flowering plants likely imposes constraints on the broader spectrum of plant trait variation.

Overall, plant reproductive traits explained interaction patterns among and within pollinator guilds suggesting that plant reproductive strategies partly mediate these

associations across spatial large scales. For example, we found that all pollinator guilds 434 interacted more frequently with species with large floral displays and low ability to 435 self pollinate, which aligns with previous findings (Hegland & Totland, 2005; Kaiser-436 Bunbury et al., 2014; Lázaro et al., 2013). As predicted by evolutionary life-history 437 theory (Stebbins, 1970), larger investment in floral display and rewards to attract pol-438 linators occurs when pollinator dependence is high. Indeed, the possible range of 439 reproductive strategies in flowering plants is constrained by the cost of reproductive 440 organs, trait associations and selective pressures exerted by different pollinator guilds 441 (Galen, 1999). For instance, smaller flowers are visited less overall, but they are in-442 expensive to produce (e.g., have a low ovule number) and low visitation rate at the individual flower level can be compensated by overall visitation at the plant level by 444 producing many flowers resulting in a larger quantity of small fruits (Kettle et al., 2011). In addition, we found that a substantial number of species with high autonomous self-446 ing had large styles and were associated with a high number of interactions with bees. These species seem to rely on animal pollinators as indicated by their high investment 448 in sizable styles that can help prevent pollen competition (Ashman & Arceo-Gómez, 2013; Lanuza et al., 2021; Ramesha et al., 2011) while having reproductive assurance in 450 their absence. For instance, the annual herb Scabiosa atropurpurea has flowers grouped in clusters with relatively long styles and a single ovule per flower that is likely to 452 be fertilised with a single legitimate floral visit but avoids reproductive failure with 453 autonomous self pollination. Similarly, zygomorphic flowers are specialised floral 454 structures that promote precise pollen transfer (Gong & Huang, 2009; Sargent, 2004) 455 but they were associated with high levels of autogamy. Our findings are consistent 456 with the idea that reproductive assurance is not rare within specialised pollination 457 systems (Fenster & Marten-Rodriguez, 2007) and highlight the need to explore the 458 timing (e.g., early versus delayed) and selfing mechanisms (see Goodwillie & Weber, 459 2018) in relation to other reproductive traits to better understand reproductive trait 460 correlations and plant-pollinator interactions. 461

Because floral traits can be associated with their animal pollinators to optimise pollen

transfer, the correlation between distinct suites of reproductive traits and pollinator 463 guilds could indicate the existence of pollination syndromes (Dellinger, 2020; Faegri & 464 Van Der Pijl, 2013; Fenster et al., 2004; Rosas-Guerrero et al., 2014). For instance, bees 465 had a higher number of interacting partners on plant species with large flowers but 466 interacted more frequently with species with small flowers while syrphid flies showed 467 the opposite trend. This could be reflective of differences in the ecology and behaviour 468 between these pollinator guilds (Földesi et al., 2021; Rader et al., 2020) that ultimately 469 exert different selective pressures, leading divergent plant reproductive strategies. 470 In any case, it is worth noting that local factors (e.g., species relative abundances or 471 phenologies) likely contribute to the variability in the association between the plant 472 reproductive spectrum and pollinators (Bartomeus et al., 2016; Encinas-Viso et al., 473 2012; Vázquez et al., 2007). Taking both aspects together, we suggest that while plant reproductive traits do shape their interactions with pollinators, pollinator syndromes 475 at a macroecological scale are not distinct compartmentalised items, but form part of a multifaceted context dependent continuum. 477

Our study provides a comprehensive assessment of reproductive trait covariation pat-478 terns in flowering plants, using a global dataset of traits for plant species within plant-479 pollinator networks. We show the importance of pollinators in mediating reproductive 480 trait correlations and identify the need to consider broader plant reproductive strate-481 gies, rather than just flower morphology, to understand patterns of plant-pollinator 482 interactions. Further, we provide empirical evidence of reproductive trait associations 483 that complement trait covariation studies for other plant organs, highlighting the need 484 to include reproductive traits in the global spectrum of plant form and function. Future 485 work incorporating underrepresented areas of the world and additional relevant repro-486 ductive traits (e.g., flower life span and colour) from a broader range of life forms will 487 be valuable for enhancing our understanding of the plant reproductive spectrum and 488 associated plant-pollinator interactions. 489

References

```
Abdi, H., & Williams, L. J. (2010). Principal component analysis. WIREs Comp. Stats.,
491
       2(4), 433–459. https://doi.org/10.1002/wics.101
492
   Agrawal, A. A. (2020). A scale-dependent framework for trade-offs, syndromes, and
493
       specialization in organismal biology. Ecology, 101(2), e02924. https://doi.org/10.1
494
       002/ecy.2924
495
    Ashman, T.-L., & Arceo-Gómez, G. (2013). Toward a predictive understanding of the
496
       fitness costs of heterospecific pollen receipt and its importance in co-flowering
497
       communities. American Journal of Botany, 100(6), 1061–1070. https://doi.org/10.373
498
       2/ajb.1200496
499
   Ballantyne, G., Baldock, K. C. R., & Willmer, P. G. (2015). Constructing more informative
500
       plant-pollinator networks: Visitation and pollen deposition networks in a heathland
501
       plant community. Proc. Royal Soc. B, 282(1814), 20151130. https://doi.org/10.1098/
502
       rspb.2015.1130
503
   Barrett, S. C. H. (2002). The evolution of plant sexual diversity. Nat. Rev. Genet., 3(4),
504
       274–284. https://doi.org/10.1038/nrg776
505
   Barrett, S. C. H. (2003). Mating strategies in flowering plants: The outcrossing-selfing
506
       paradigm and beyond. Philos. Trans. R. Soc. Lond. B Biol. Sci., 358(1434), 991–1004.
507
       https://doi.org/10.1098/rstb.2003.1301
508
   Bartomeus, I., Gravel, D., Tylianakis, J. M., Aizen, M. A., Dickie, I. A., & Bernard-Verdier,
       M. (2016). A common framework for identifying linkage rules across different types
510
       of interactions. Funct. Ecol., 30(12), 1894–1903. https://doi.org/10.1111/1365-
511
       2435.12666
512
   Blüthgen, N., Menzel, F., & Blüthgen, N. (2006). Measuring specialization in species
       interaction networks. BMC Ecol., 6(1), 9. https://doi.org/10.1186/1472-6785-6-9
514
   Bruelheide, H., Dengler, J., Purschke, O., Lenoir, J., Jiménez-Alfaro, B., Hennekens,
       S. M., Botta-Dukát, Z., Chytry, M., Field, R., Jansen, F., et al. (2018). Global trait-
516
       environment relationships of plant communities. Nat. Ecol. Evol., 2(12), 1906–1917.
```

```
https://doi.org/10.1038/s41559-018-0699-8
518
    Bürkner, P.-C. (2017). Brms: An R package for Bayesian multilevel models using Stan. J.
519
       Stat. Softw., 80(1), 1–28. https://doi.org/10.18637/jss.v080.i01
520
    Carmona, C. P., Tamme, R., Pärtel, M., Bello, F. de, Brosse, S., Capdevila, P., González-M,
521
       R., González-Suárez, M., Salguero-Gómez, R., Vásquez-Valderrama, M., & Toussaint,
522
       A. (2021). Erosion of global functional diversity across the tree of life. Sci. Adv.,
523
       7(13), eabf2675. https://doi.org/10.1126/sciadv.abf2675
524
    Carvalheiro, L. G., Biesmeijer, J. C., Benadi, G., Fründ, J., Stang, M., Bartomeus, I., Kaiser-
525
       Bunbury, C. N., Baude, M., Gomes, S. I. F., Merckx, V., Baldock, K. C. R., Bennett,
526
       A. T. D., Boada, R., Bommarco, R., Cartar, R., Chacoff, N., Dänhardt, J., Dicks, L. V.,
527
       Dormann, C. F., ... Kunin, W. E. (2014). The potential for indirect effects between co-
528
       flowering plants via shared pollinators depends on resource abundance, accessibility
529
       and relatedness. Ecol. Lett., 17(11), 1389–1399. https://doi.org/10.1111/ele.12342
530
    Chamberlain, S., Szoecs, E., Foster, Z., Arendsee, Z., Boettiger, C., Ram, K., Bartomeus,
       I., Baumgartner, J., O'Donnell, J., Oksanen, J., Tzovaras, B. G., Marchand, P., Tran,
532
       V., Salmon, M., Li, G., & Grenié, M. (2018). Package "PVR." R package version 0.3. At
533
       https://CRAN.r-project.org/package=PVR.
534
    Chamberlain, S., Szoecs, E., Foster, Z., Arendsee, Z., Boettiger, C., Ram, K., Bartomeus,
       I., Baumgartner, J., O'Donnell, J., Oksanen, J., Tzovaras, B. G., Marchand, P., Tran, V.,
536
       Salmon, M., Li, G., & Grenié, M. (2020). Taxize: Taxonomic information from around the
537
       web. R package version 0.9.99. At https://CRAN.r-project.org/package=taxize.
538
    Chave, J., Coomes, D., Jansen, S., Lewis, S. L., Swenson, N. G., & Zanne, A. E. (2009).
539
       Towards a worldwide wood economics spectrum. Ecol. Lett., 12(4), 351–366. https:
540
       //doi.org/10.1111/j.1461-0248.2009.01285.x
541
    Coux, C., Rader, R., Bartomeus, I., & Tylianakis, J. M. (2016). Linking species functional
542
       roles to their network roles. Ecol. Lett., 19(7), 762–770. https://doi.org/10.1111/ele.
543
       12612
544
    Dehling, D. M., Jordano, P., Schaefer, H. M., Böhning-Gaese, K., & Schleuning, M. (2016).
545
       Morphology predicts species' functional roles and their degree of specialization
546
```

- in plant–frugivore interactions. *Proc. Royal Soc. B*, 283(1823), 20152444. https:
- //doi.org/10.1098/rspb.2015.2444
- Dellinger, A. S. (2020). Pollination syndromes in the 21st century: Where do we stand
- and where may we go? *New Phytol.*, 228(4), 1193–1213. https://doi.org/10.1111/np
- 551 h.16793
- Devaux, C., Lepers, C., & Porcher, E. (2014). Constraints imposed by pollinator be-
- haviour on the ecology and evolution of plant mating systems. J. Evol. Biol., 27(7),
- 1413–1430. https://doi.org/10.1111/jeb.12380
- Díaz, S., Kattge, J., Cornelissen, J. H. C., Wright, I. J., Lavorel, S., Dray, S., Reu, B., Kleyer,
- M., Wirth, C., Colin Prentice, I., Garnier, E., Bönisch, G., Westoby, M., Poorter, H.,
- Reich, P. B., Moles, A. T., Dickie, J., Gillison, A. N., Zanne, A. E., ... Gorné, L. D.
- (2016). The global spectrum of plant form and function. *Nature*, 529(7585), 167–171.
- https://doi.org/10.1038/nature16489
- Diniz-Filho, J. A. F., Bini, L. M., Rangel, T. F., Morales-Castilla, I., Olalla-Tárraga, M.
- Å., Rodríguez, M. Å., & Hawkins, B. A. (2012). On the selection of phylogenetic
- eigenvectors for ecological analyses. *Ecography*, 35(3), 239–249. https://doi.org/10
- .1111/j.1600-0587.2011.06949.x
- Donovan, L. A., Maherali, H., Caruso, C. M., Huber, H., & Kroon, H. de. (2011). The
- evolution of the worldwide leaf economics spectrum. Trends in Ecology & Evolution,
- ⁵⁶⁶ 26(2), 88–95. https://doi.org/10.1016/j.tree.2010.11.011
- Dormann, C. F., Gruber, B., & Fründ, J. (2008). Introducing the bipartite package:
- Analysing ecological networks. *R News*, 8/2.
- Eklöf, A., Jacob, U., Kopp, J., Bosch, J., Castro-Urgal, R., Chacoff, N. P., Dalsgaard,
- B., Sassi, C. de, Galetti, M., Guimaraes, P. R., et al. (2013). The dimensionality of
- ecological networks. *Ecol. Lett.*, 16(5), 577–583. https://doi.org/10.1111/ele.12081
- Encinas-Viso, F., Revilla, T. A., & Etienne, R. S. (2012). Phenology drives mutualistic
- network structure and diversity. Ecol. Lett., 15(3), 198–208. https://doi.org/10.111
- 1/j.1461-0248.2011.01726.x
- E-Vojtkó, A., Bello, F. de, Durka, W., Kühn, I., & Götzenberger, L. (2020). The neglected

```
importance of floral traits in trait-based plant community assembly. J. Veg. Sci.,
576
       31(4), 529–539. https://doi.org/10.1111/jvs.12877
577
   E-Vojtkó, A., Junker, R. R., Bello, F. de, & Götzenberger, L. (2022). Floral and reproduc-
578
       tive traits are an independent dimension within the plant economic spectrum of
579
       temperate central europe. New Phytologist. https://doi.org/10.1111/nph.18386
580
   Faegri, K., & Van Der Pijl, L. (2013). Principles of pollination ecology (3rd edn). Elsevier.
581
   Fenster, C. B., Armbruster, W. S., Wilson, P., Dudash, M. R., & Thomson, J. D. (2004).
582
       Pollination Syndromes and Floral Specialization. Annu. Rev. Ecol. Evol. Syst., 35(1),
583
       375–403. https://doi.org/10.1146/annurev.ecolsys.34.011802.132347
584
   Fenster, C. B., & Marten-Rodriguez, S. (2007). Reproductive assurance and the evolution
585
       of pollination specialization. International Journal of Plant Sciences, 168(2), 215–228.
586
       https://doi.org/10.1086/509647
   Földesi, R., Howlett, B. G., Grass, I., & Batáry, P. (2021). Larger pollinators deposit more
588
       pollen on stigmas across multiple plant species—a meta-analysis. J. Appl. Ecol.,
589
       58(4), 699–707. https://doi.org/10.1111/1365-2664.13798
590
   Fortuna, M. A., Stouffer, D. B., Olesen, J. M., Jordano, P., Mouillot, D., Krasnov, B. R.,
       Poulin, R., & Bascompte, J. (2010). Nestedness versus modularity in ecological
592
       //doi.org/10.1111/j.1365-2656.2010.01688.x
594
   Friedman, J. (2020). The evolution of annual and perennial plant life histories: Ecological
595
       correlates and genetic mechanisms. Annu. Rev. Ecol. Evol. Syst., 51, 461–481.
596
       https://doi.org/10.1146/annurev-ecolsys-110218-024638
   Gabry, J., Simpson, D., Vehtari, A., Betancourt, M., & Gelman, A. (2019). Visualization
598
       in Bayesian workflow. J. R. Stat. Soc. Ser. A Stat. Soc., 182(2), 389–402. https:
599
       //doi.org/10.1111/rssa.12378
600
   Galen, C. (1999). Why do flowers vary? The functional ecology of variation in flower
601
       size and form within natural plant populations. Bioscience, 49(8), 631–640. https:
602
       //doi.org/10.2307/1313439
603
```

Gong, Y.-B., & Huang, S.-Q. (2009). Floral symmetry: Pollinator-mediated stabilizing

604

```
selection on flower size in bilateral species. Proceedings of the Royal Society B: Biological
605
       Sciences, 276(1675), 4013–4020. https://doi.org/10.1098/rspb.2009.1254
606
   Goodwillie, C., Sargent, R. D., Eckert, C. G., Elle, E., Geber, M. A., Johnston, M. O.,
607
       Kalisz, S., Moeller, D. A., Ree, R. H., Vallejo-Marin, M., & Winn, A. A. (2010).
608
       Correlated evolution of mating system and floral display traits in flowering plants
609
       and its implications for the distribution of mating system variation. New Phytol.,
610
       185(1), 311–321. https://doi.org/10.1111/j.1469-8137.2009.03043.x
611
   Goodwillie, C., & Weber, J. J. (2018). The best of both worlds? A review of delayed
612
       selfing in flowering plants. American Journal of Botany, 105(4), 641–655. https://
613
       //doi.org/10.1002/ajb2.1045
614
   Healy, K., Ezard, T. H. G., Jones, O. R., Salguero-Gómez, R., & Buckley, Y. M. (2019).
615
       Animal life history is shaped by the pace of life and the distribution of age-specific
616
       mortality and reproduction. Nat. Ecol. Evol., 3(8), 1217–1224. https://doi.org/10.1
617
       038/s41559-019-0938-7
618
   Hegland, S. J., & Totland, Ø. (2005). Relationships between species' floral traits and
619
       pollinator visitation in a temperate grassland. Oecologia, 145(4), 586–594. https://
620
       //doi.org/10.1007/s00442-005-0165-6
621
   Hung, K.-L. J., Kingston, J. M., Albrecht, M., Holway, D. A., & Kohn, J. R. (2018). The
       worldwide importance of honey bees as pollinators in natural habitats. Proc. Royal
623
       Soc. B, 285(1870), 20172140. https://doi.org/10.1098/rspb.2017.2140
   Ibanez, S. (2012). Optimizing size thresholds in a plant-pollinator interaction web:
625
       Towards a mechanistic understanding of ecological networks. Oecologia, 170(1),
626
       233–242. https://doi.org/10.1007/s00442-012-2290-3
627
   Jin, Y., & Qian, H. (2019). V.PhyloMaker: An R package that can generate very large
628
       phylogenies for vascular plants. Ecography, 42(8), 1353–1359. https://doi.org/10.1
629
       111/ecog.04434
630
   Junker, R. R., Blüthgen, N., Brehm, T., Binkenstein, J., Paulus, J., Schaefer, H. M., &
631
       Stang, M. (2013). Specialization on traits as basis for the niche-breadth of flower
632
       visitors and as structuring mechanism of ecological networks. Funct. Ecol., 27(2),
633
```

```
329–341. https://doi.org/10.1111/1365-2435.12005
   Kaiser-Bunbury, C. N., Vázquez, D. P., Stang, M., & Ghazoul, J. (2014). Determinants
635
       of the microstructure of plant–pollinator networks. Ecology, 95(12), 3314–3324.
636
       https://doi.org/10.1890/14-0024.1
637
   Kattge, J., Díaz, S., Lavorel, S., Prentice, I. C., Leadley, P., Bönisch, G., Garnier, E.,
638
       Westoby, M., Reich, P. B., Wright, I. J., Cornelissen, J. H. C., Violle, C., Harrison, S. P.,
639
       Bodegom, P. M. V., Reichstein, M., Enquist, B. J., Soudzilovskaia, N. A., Ackerly, D.
640
       D., Anand, M., ... Wirth, C. (2011). TRY – a global database of plant traits. Glob.
641
       Chang. Biol., 17(9), 2905–2935. https://doi.org/10.1111/j.1365-2486.2011.02451.x
642
   Kettle, C. J., Maycock, C. R., Ghazoul, J., Hollingsworth, P. M., Khoo, E., Sukri, R. S.
       H., & Burslem, D. F. R. P. (2011). Ecological Implications of a Flower Size/Number
644
       Trade-Off in Tropical Forest Trees. PLoS One, 6(2), e16111. https://doi.org/10.1371/
645
       journal.pone.0016111
646
   Lanuza, J. B., Bartomeus, I., Ashman, T.-L., Bible, G., & Rader, R. (2021). Recipient
       and donor characteristics govern the hierarchical structure of heterospecific pollen
648
       competition networks. Journal of Ecology, 109(6), 2329–2341. https://doi.org/10.111
       1/1365-2745.13640
650
   Laughlin, D. C., Mommer, L., Sabatini, F. M., Bruelheide, H., Kuyper, T. W., McCormack,
       M. L., Bergmann, J., Freschet, G. T., Guerrero-Ramírez, N. R., Iversen, C. M., et al.
652
       (2021). Root traits explain plant species distributions along climatic gradients
653
       yet challenge the nature of ecological trade-offs. Nat. Ecol. Evol., 1–12. https://
654
       //doi.org/10.1038/s41559-021-01471-7
655
   Lázaro, A., Jakobsson, A., & Totland, Ø. (2013). How do pollinator visitation rate and
656
       seed set relate to species' floral traits and community context? Oecologia, 173(3),
657
       881–893. https://doi.org/10.1007/s00442-013-2652-5
658
   Legendre, P., & Legendre, L. (2012). Numerical ecology (2nd edn). Elsevier, Amsterdam.
659
   Moeller, D. A., Runquist, R. D. B., Moe, A. M., Geber, M. A., Goodwillie, C., Cheptou, P.-
660
       O., Eckert, C. G., Elle, E., Johnston, M. O., Kalisz, S., Ree, R. H., Sargent, R. D., Vallejo-
661
       Marin, M., & Winn, A. A. (2017). Global biogeography of mating system variation
662
```

634

- in seed plants. Ecol. Lett., 20(3), 375–384. https://doi.org/10.1111/ele.12738
- 664 Munoz, F., Violle, C., & Cheptou, P.-O. (2016). CSR ecological strategies and plant
- mating systems: Outcrossing increases with competitiveness but stress-tolerance is
- related to mixed mating. Oikos, 125(9), 1296–1303. https://doi.org/10.1111/oik.02
- 667 328
- Olesen, J. M., Bascompte, J., Dupont, Y. L., & Jordano, P. (2007). The modularity of
- pollination networks. PNAS, 104(50), 19891–19896. https://doi.org/10.1073/pnas.0
- 670 706375104
- Osnas, J. L., Lichstein, J. W., Reich, P. B., & Pacala, S. W. (2013). Global leaf trait
- relationships: Mass, area, and the leaf economics spectrum. Science, 340(6133),
- 741–744. https://doi.org/10.1126/science.1231574
- Paterno, G. B., Silveira, C. L., Kollmann, J., Westoby, M., & Fonseca, C. R. (2020). The
- maleness of larger angiosperm flowers. *Proc. Natl. Acad. Sci.*, 117(20), 10921–10926.
- https://doi.org/10.1073/pnas.1910631117
- Penone, C., Davidson, A. D., Shoemaker, K. T., Marco, M. D., Rondinini, C., Brooks, T.
- M., Young, B. E., Graham, C. H., & Costa, G. C. (2014). Imputation of missing data
- in life-history trait datasets: Which approach performs the best? Methods Ecol. Evol.,
- 5(9), 961–970. https://doi.org/10.1111/2041-210X.12232
- Petit, R. J., & Hampe, A. (2006). Some evolutionary consequences of being a tree. Annu.
- Rev. Ecol. Evol. Syst., 37, 187–214. https://doi.org/10.1146/annurev.ecolsys.37.091
- 683 305.110215
- Poisot, T., Baiser, B., Dunne, J. A., Kéfi, S., Massol, F., Mouquet, N., Romanuk, T. N.,
- Stouffer, D. B., Wood, S. A., & Gravel, D. (2016). Mangal making ecological network
- analysis simple. *Ecography*, 39(4), 384–390. https://doi.org/10.1111/ecog.00976
- Rader, R., Cunningham, S., Howlett, B., & Inouye, D. (2020). Non-bee insects as visitors
- and pollinators of crops: Biology, ecology, and management. *Annu. Rev. Entomol.*,
- 65, 391–407. https://doi.org/10.1146/annurev-ento-011019-025055
- Ramesha, B., Yetish, M., Ravikanth, G., Ganeshaiah, K., Ghazoul, J., & Shaanker, R. U.
- (2011). Stylish lengths: Mate choice in flowers. *Journal of Biosciences*, 36(2), 229–234.

```
https://doi.org/10.1007/s12038-011-9057-6
692
   Revell, L. J. (2012). Phytools: An R package for phylogenetic comparative biology (and
693
       other things). Methods Ecol. Evol., 3(2), 217–223. https://doi.org/10.1111/j.2041-
694
       210X.2011.00169.x
695
   Roddy, A. B., Martínez-Perez, C., Teixido, A. L., Cornelissen, T. G., Olson, M. E., Oliveira,
696
       R. S., & Silveira, F. A. O. (2021). Towards the flower economics spectrum. New
697
       Phytol., 229(2), 665–672. https://doi.org/10.1111/nph.16823
698
   Rosas-Guerrero, V., Aguilar, R., Martén-Rodríguez, S., Ashworth, L., Lopezaraiza-
699
       Mikel, M., Bastida, J. M., & Quesada, M. (2014). A quantitative review of pollination
700
       syndromes: Do floral traits predict effective pollinators? Ecol. Lett., 17(3), 388–400.
701
       https://doi.org/10.1111/ele.12224
702
   Rüger, N., Comita, L. S., Condit, R., Purves, D., Rosenbaum, B., Visser, M. D., Wright, S.
       J., & Wirth, C. (2018). Beyond the fast–slow continuum: Demographic dimensions
704
       structuring a tropical tree community. Ecol. Lett., 21(7), 1075–1084. https://doi.org/
705
       10.1111/ele.12974
706
   Salguero-Gómez, R., Jones, O. R., Archer, C. R., Buckley, Y. M., Che-Castaldo, J., Caswell,
707
       H., Hodgson, D., Scheuerlein, A., Conde, D. A., Brinks, E., Buhr, H. de, Farack,
708
       C., Gottschalk, F., Hartmann, A., Henning, A., Hoppe, G., Römer, G., Runge, J.,
       Ruoff, T., ... Vaupel, J. W. (2015). The compadre Plant Matrix Database: An
710
       open online repository for plant demography. J. Ecol., 103(1), 202–218. https://
711
       //doi.org/10.1111/1365-2745.12334
712
   Salguero-Gómez, R., Jones, O. R., Jongejans, E., Blomberg, S. P., Hodgson, D. J., Mbeau-
       Ache, C., Zuidema, P. A., de Kroon, H., & Buckley, Y. M. (2016). Fast-slow continuum
714
       and reproductive strategies structure plant life-history variation worldwide. Proc.
715
       Natl. Acad. Sci. U.S.A., 113(1), 230–235. https://doi.org/10.1073/pnas.1506215112
716
   Sargent, R. D. (2004). Floral symmetry affects speciation rates in angiosperms. Proceed-
717
       ings of the Royal Society of London. Series B: Biological Sciences, 271(1539), 603–608.
718
       https://doi.org/10.1098/rspb.2003.2644
719
   Sargent, R. D., & Ackerly, D. D. (2008). Plant–pollinator interactions and the assembly
```

```
of plant communities. Trends Ecol. Evol., 23(3), 123–130. https://doi.org/10.1016/j.
721
       tree.2007.11.003
722
   Sargent, R. D., Goodwillie, C., Kalisz, S., & Ree, R. H. (2007). Phylogenetic evidence
723
       for a flower size and number trade-off. Am. J. Bot., 94(12), 2059–2062. https:
724
       //doi.org/10.3732/ajb.94.12.2059
725
   Schiestl, F. P., & Johnson, S. D. (2013). Pollinator-mediated evolution of floral signals.
726
       Trends Ecol. Evol., 28(5), 307–315. https://doi.org/10.1016/j.tree.2013.01.019
727
   Serneels, S., & Verdonck, T. (2008). Principal component analysis for data containing
728
       outliers and missing elements. Comput. Stat. Data Anal., 52(3), 1712–1727. https:
729
       //doi.org/10.1016/j.csda.2007.05.024
730
   Shipley, B., Lechowicz, M. J., Wright, I., & Reich, P. B. (2006). Fundamental trade-
731
       offs generating the worldwide leaf economics spectrum. Ecology, 87(3), 535–541.
732
       https://doi.org/10.1890/05-1051
733
   Smith, S. A., & Brown, J. W. (2018). Constructing a broadly inclusive seed plant
       phylogeny. Am. J. Bot., 105(3), 302–314. https://doi.org/10.1002/ajb2.1019
735
   Song, B., Sun, L., Barrett, S. C., Moles, A. T., Luo, Y.-H., Armbruster, W. S., Gao, Y.-
       Q., Zhang, S., Zhang, Z.-Q., & Sun, H. (2022). Global analysis of floral longevity
737
       reveals latitudinal gradients and biotic and abiotic correlates. New Phytologist.
       https://doi.org/10.1111/nph.18271
739
   Stang, M., Klinkhamer, P. G. L., Waser, N. M., Stang, I., & van der Meijden, E. (2009).
740
       Size-specific interaction patterns and size matching in a plant-pollinator interaction
741
       web. Ann. Bot., 103(9), 1459–1469. https://doi.org/10.1093/aob/mcp027
   Stearns, S. C. (1989). Trade-offs in life-history evolution. Funct. Ecol., 3(3), 259–268.
743
       https://doi.org/10.2307/2389364
744
   Stebbins, G. L. (1970). Adaptive radiation of reproductive characteristics in an-
745
       giosperms, i: Pollination mechanisms. Annual Review of Ecology and Systematics,
746
       307–326. https://doi.org/10.1146/annurev.es.01.110170.001515
747
   Stekhoven, D. J., & Bühlmann, P. (2012). MissForest—non-parametric missing value
748
       imputation for mixed-type data. Bioinformatics, 28(1), 112–118. https://doi.org/10.1
749
```

```
Tur, C., Castro-Urgal, R., & Traveset, A. (2013). Linking Plant Specialization to Depen-
751
       dence in Interactions for Seed Set in Pollination Networks. PLoS One, 8(10), e78294.
752
       https://doi.org/10.1371/journal.pone.0078294
753
    Vázquez, D. P., Lomáscolo, S. B., Maldonado, M. B., Chacoff, N. P., Dorado, J., Stevani,
754
       E. L., & Vitale, N. L. (2012). The strength of plant–pollinator interactions. Ecology,
755
       93(4), 719–725. https://doi.org/10.1890/11-1356.1
756
    Vázquez, D. P., Melián, C. J., Williams, N. M., Blüthgen, N., Krasnov, B. R., & Poulin,
757
       R. (2007). Species abundance and asymmetric interaction strength in ecological
758
       networks. Oikos, 116(7), 1120–1127. https://doi.org/10.1111/j.0030-1299.2007.1582
759
       8.x
760
    Vázquez, D. P., Morris, W. F., & Jordano, P. (2005). Interaction frequency as a surrogate
       for the total effect of animal mutualists on plants. Ecol. Lett., 8(10), 1088–1094.
762
       https://doi.org/10.1111/j.1461-0248.2005.00810.x
    Worley, A. C., Baker, A. M., Thompson, J. D., & Barrett, S. C. (2000). Floral display
764
       in narcissus: Variation in flower size and number at the species, population, and
765
       individual levels. Int. J. Plant Sci., 161(1), 69–79. https://doi.org/10.1086/314225
766
    Wright, I. J., Reich, P. B., Westoby, M., Ackerly, D. D., Baruch, Z., Bongers, F., Cavender-
       Bares, J., Chapin, T., Cornelissen, J. H. C., Diemer, M., Flexas, J., Garnier, E., Groom,
768
       P. K., Gulias, J., Hikosaka, K., Lamont, B. B., Lee, T., Lee, W., Lusk, C., ... Villar,
769
```

093/bioinformatics/btr597

750

770

R. (2004). The worldwide leaf economics spectrum. Nature, 428(6985), 821–827.

https://doi.org/10.1038/nature02403