Covariation among reproductive traits in flowering plants shapes interactions with floral visitors

- Jose B. Lanuza^{1,2}, Romina Rader¹, Jamie Stavert³, Liam K. Kendall⁴, Manu E. Saunders¹ and Ignasi Bartomeus²
- $_7$ $^{-1}$ School of Environmental and Rural Science, University of New England, Armidale, New
- 8 South Wales 2350, Australia. ² Estación Biológica de Doñana (EBD-CSIC), E-41092 Seville,
- ⁹ Spain. ³ Department of Conservation | Te Papa Atawhai, Auckland, New Zealand. ⁴ Centre for
- Environmental and Climate Science, Lund University, Sölvegatan 37, S-223 62 Lund, Sweden.
- Keywords: life-history strategies | plant reproductive traits | plant-pollinator interac-
- 12 tions

3

Author for correspondence: Jose B. Lanuza | barragansljose@gmail.com

14 Abstract

Globally, plants display enormous variation in life history strategies and trait combinations. However, evidence suggests that evolutionary and physiological constraints limit
the number of plant ecological strategies. Although there have been recent advances in
understanding correlations among plant traits, reproductive traits are rarely considered,
despite their key role in shaping plant life-history strategies and interactions with floral
visitors. Here, using a global dataset of 16 reproductive traits for 1,506 species, we
investigate the reproductive spectrum of flowering plants to identify how it shapes
interactions with floral visitors. We show that over 50% of all trait variation is explained
by the first two reproductive axes, which represent the negative correlation between
flower number and flower size, and the positive correlation between pollinator dependence and floral display size. In empirical plant-pollinator networks, the plant position
within these axes was associated with the number of interacting partners and the visitation rates of the distinct floral visitor guilds, but the unexplained variability found
highlights the need to incorporate other factors along with reproductive traits to fully

explain plant-pollinator interactions. Our study identifies the major reproductive trait correlations in flowering plants and their role in shaping networks of plant-pollinator interactions at a global scale.

1. Introduction

Flowering plants display an astonishing diversity of reproductive structures [1,2] that 33 shape their interactions with animal pollinators [3,4]. The variety of reproductive strategies used by different plant species, from specialized reliance on a single pollinator, to 35 full self-pollination, is incredibly rich. However, not all reproductive trait combinations in plants are possible due to evolutionary and ecological constraints [5,6]. Despite 37 recent advances in theoretical and empirical understanding of correlations between plant reproductive traits at large ecological scales [7–11], studies typically focus on just a few traits, often in isolation. Thus, to develop a comprehensive understanding of the plant reproductive spectrum of trait variation, as done recently for root [12], leaf [13] and wood [14] related traits, we require a multi-trait perspective with broad geographical coverage. Importantly, characterization of the plant reproductive spectrum could reveal key reproductive strategies among different species [6] and improve knowledge of associations between specific reproductive traits and pollinators [10]. With the recent availability of large trait databases [e.g., TRY 15, COMPADRE 16], there has been increased research on plant ecological strategies, which has produced identifi-47 cation of global patterns in plant form and function [7,17–19]. However, studies that 48 look at multiple traits tend to overlook plant reproductive traits [10,20,21], and focus on trait correlations in other plant organs such as morphological and physiological leaf traits [13,22–24]. Despite the lack of a holistic understanding that depicts reproductive trait covariation patterns, there are widely recognized associations between plant reproductive trait pairs (e.g., the negative correlation between flower size and flower number [25,26]; the positive association between flower size and outcrossing rate [27];

or the positive correlation between outcrossing rate and lifespan [28–30]). Although identification of these trait correlations has allowed progress toward a conceptual framework that integrates different floral trait relationships [10], we still lack empirical understanding of how these associations among floral and other reproductive traits (e.g., reproductive biology) shape plant life history strategies.

Trait-based approaches [3,31] and trait-matching analyses [32,33] are powerful for ex-60 ploring the underlying mechanisms driving plant-pollinator interactions. For example, plant traits can define species' network roles [e.g., specialists vs generalists 34] and plant 62 species that occupy reproductive trait space extremes are likely to be more specialized and display tighter trait-matching with pollinators [35,36]. Indeed, the degree of morphological matching between plants and floral visitors often determines whether or not interactions occur, and consequently influences the broader network structure [32,37]. However, it is still unclear how specific reproductive traits (e.g., mating or compatibility system) influence plant-pollinator associations [34,38,39]. Indeed, both morphology 68 and mating system can determine a species' functional role within pollination networks, and the combination of several traits increases power for predicting network structure [40]. Because the species position within the multidimensional trait space can result in different interaction patterns or adaptations to pollinators [see 41 for a purely morphological approach], the use of the reproductive trait space to explore plant-pollinator interactions could help to identify species functional roles and associations of traits with specific pollinators (i.e., pollination syndromes).

Here, we explore the spectrum of reproductive traits in flowering plants, within a global set of plant-pollinator networks, to identify reproductive trait covariation and how this influences the network of interactions between plants and floral visitors. First, we identify the major axes of reproductive trait variation and correlations among traits for plant species. Then, we investigate the associations between plant species' positions in reproductive trait space and the strength of their interactions with different floral visitor guilds within networks. Finally, we determine how the main axes of reproductive trait variation influence plant species' functional roles within pollination networks, using a

range of complementary interaction network metrics.

2. Methods

86 (a) Plant-pollinator network studies

We selected 28 studies from 18 different countries that constituted a total of 64 plantpollinator networks (see table S1 and figure S1). These studies recorded plant-pollinator
interactions in natural systems and were selected so that we had broad geographical
representation across different ecological communities. Although these studies differ
in sampling effort and methodology, all studies provided information about plantpollinator interactions (weighted and non-weighted), which we used to build a database
of plant species that are likely to benefit, to some degree, from animal pollination. Many
of these networks are freely available either as published studies [e.g., 42,43,44] or in
online archives [e.g., Mangal 45,The Web of Life 44]. In total, our network dataset
constituted 60 weighted (number of visits) and four unweighted (interaction presenceabsence) networks, each sampled at a unique location and year, as well as eight metanetworks where interactions were pooled across several locations and multiple years.

99 (b) Taxonomy of plants and pollinators

All species names, genera, families and orders were retrieved and standardized from
the taxonomy data sources NCBI (https://www.ncbi.nlm.nih.gov/taxonomy) for
plants and ITIS (https://www.itis.gov/) for pollinators, using the R package *taxize* [46].
We filled the 'not found' searches manually using http://www.theplantlist.org/ and
http://www.mobot.org/ for plants and http://www.catalogueoflife.org/ for floral
visitors.

106 (c) Plant traits

We selected a total of 19 quantitative and categorical functional traits that included 107 reproductive traits and traits related with plant form and size (see table 1). Reproductive 108 traits consisted of those related to floral and reproductive biology: (i) floral traits 109 included those related to the size of floral organs (e.g., style length), floral display size 110 (e.g., flower number) and floral rewards (e.g., pollen quantity), while (ii) reproductive biology traits represented the plant reproductive system (i.e., the breeding, mating and 112 compatibility system). The remaining traits were related to plant size and form (i.e., plant height, lifespan and life form), which are commonly used to characterize the 114 fast-slow continuum of plant trait variation [7]. For each plant species, we undertook an extensive literature and online search for all traits across a wide range of resources 116 (plant databases, online floras, books, journals and images). From a matrix comprising 117 a total of 28,614 possible cells (considering all traits and plant species: 19 columns × 118 1,506 species), we were able to fill 23,354 cells (81.6% of the dataset, see figure S2 for 119 missing value information for each trait). An extended description of each trait and 120 how it was obtained can be found in Appendix S1. 121

122 (d) Phylogenetic Distance

We calculated the phylogenetic distance between all plant species considered using the function *get_tree* from the package *rtrees* (https://github.com/daijiang/rtrees), which downloads phylogenetic distances from the extended R implementation of the Open Tree of Life [47,48].

127 (e) Data Imputation

To avoid excluding species with trait missing values, we imputed trait missing values with the help of the function *missForest* [49] which allows imputation of datasets with continuous and categorical variables. We accounted for the phylogenetic distance

Table 1. Quantitative and categorical traits used in this study.

	
Traits	Numerical range/Categories
Quantitative	
Plant height	0.01-30m
Flower width	0.50-205mm
Flower length	0.20-195mm
Inflorescence width	0.80-300mm
Style length	0-138mm
Ovules per flower	1-25000
Flowers per plant	$1-1.8 \times 10^5$
Microliters of nectar	$0-160\mu$ l
Milligrams of nectar	0-6.1mg
Nectar concentration	0-100%
Pollen grains per flower	$13-2 \times 10^8$
Autonomous selfing (fruit set)	0-100%
Categorical	
Lifepan	Short-lived and perennial
Life form	Herb, shrub and tree
Flower shape	Brush, campanulate, capitulum,
	open, papilionaceous and tube
Flower symmetry	Actinomorphic and zygomorphic
Nectar	Presence and absence
Compatibility system	Self-incompatible, partially self-compabtile
	and self-compatible
Breeding system	Hermaphrodite, monoecious and dioecious

among species on the imputation process by including the eigenvectors from principal 131 component analysis of the phylogenetic distance (PCoA), which has been shown to 132 improve the performance of missForest [50]. To extract the eigenvectors, we used 133 the function *PVRdecomp* from the package *PVR* [51] based on a previous conceptual 134 framework that considers phylogenetic eigenvectors [52]. We conducted two different 135 imputations, one for the full set of species (1,506 species: 5.79% of missing values), 136 and excluded nectar and pollen traits because of the high percentage of missing values 137 (figure S2), and a second one for the subset of species with data for pollen per flower 138 and microliters of nectar (755 species: 8.01% of missing values). To corroborate that our 139 imputation of missing values did not affect our results, we evaluated the reproductive spectrum (see section below) with and without imputed values and results for the two 141 datasets were consitent (Fig S3 and Fig S4).

43 (f) Plant strategies

We explored the association between the different quantitative plant traits using a 144 phylogenetically informed Principal Component Analysis (pPCA). We did not include the quantitative variables of flower length and inflorescence width because they were 146 highly and moderately correlated to flower width, respectively (Pearson's correlation 147 = 0.72, P < 0.01 and Pearson's correlation = 0.36, P < 0.01), and thus we avoided 148 overemphasizing flower size on the spectrum of trait variation. Prior to the analyses, we excluded outliers and standardized the data. Specifically, due to the high sensitivity 150 of dimensionality reduction to outliers [53,54], we excluded values outside the 2.5th– 151 97.5th percentile range, and thus our final dataset had 1,236 species. Then, we log 152 transformed all continuous variables to reduce the influence of extreme values and 153 then z-transformed (X= 0, SD=1) these variables so that they were within the same 154 numerical range, as recommended for principal component analysis [54]. Although 155 qualitative traits were not included in the dimensionality reduction analysis, we also 156 investigated the statistical association of the different qualitative traits with the main 157 axes of trait variation using an ANOVA and Tukey test. We performed the pPCA using

the function *phyl.pca* from the package *phytools* [55] with the method lambda (λ) that calculates the phylogenetic correlation between 0 (phylogenetic independence) and 1 (shared evolutionary history) and we implemented the mode "covariance" because values for each variables were on the same scale following transformation [56]. Finally, we conducted an additional phylogenetic informed principal component analysis for the subset of species with pollen and nectar quantity data. For this, we included all quantitative traits included in the main pPCA, plus pollen grains and microlitres of nectar per flower.

167 (g) Phylogenetic signal

We calculated the phylogenetic signal of the different quantitative traits on the imputed dataset with the full set of species (N = 1,506) with the package *phytools* [55] and used Pagel's λ as a measurement of the phylogenetic signal. However, for pollen and nectar traits, phylogenetic signal was calculated only on the subset of species that had quantitative information for these traits (N = 755).

173 (h) Network analyses

(i) Visitation patterns

We used Bayesian modeling (see details below) to investigate how the different guilds 175 of floral visitors interacted along the main axes of reproductive trait variation using 176 qualitative and quantitative measures of plant-pollinator interactions. Specifically, 177 interactions between plants and floral visitors in binary networks were defined as the presence or absence of floral visitation, while in weighted networks, interactions 179 were measured as the the number of visits from each insect species to different plant 180 species. Although floral visitors are not always pollinators and the number of visits 181 does not consider the pollination efficiency of each species [57], the number of visits 182 does provide valuable information about the relative contributions different floral 183 visitors make to pollination [58,59]. We divided floral visitors into six main taxonomical

guilds that differ in life form, behaviour and thus likely play similar ecological roles: 185 (i) bees (Hymenoptera-Anthophila), (ii) non-bee-Hymenoptera (Hymenoptera-non-186 Anthophila), (iii) syrphids (Diptera-Syrphidae), (iv) non-syrphid-Diptera (Diptera-non-187 Syrphidae), (v) Lepidoptera, and (vi) Coleoptera. Moreover, because bees had the 188 greatest number of occurrences in networks (2,256 records) and the highest frequency 189 of visits, we further broke this group down into the main bee families (Andrenidae, 190 Apidae, Colletidae, Halictidae, and Megachilidae). In addition, we found that *Apis* 191 mellifera was the floral visitor with the largest proportion of occurrences in networks 192 (7.55% of the total). This is consistent with previous research showing that *A. mellifera* 193 was the most frequent floral visitor in a similar dataset of 80 plant-pollinator networks 194 in natural ecosystems [60]. Hence, to control for the effect of A. mellifera on observed 195 visitation patterns of bees, we conducted an analogous analysis with presence-absence of the interaction and number of visits excluding A. mellifera. We found that A. mellifera, 197 was partly driving some of the observed trends on PC1 (figure S5). However, we did not detect major differences on PC2 and PC3. 199

We used Bayesian generalized linear mixed models in the R package brms [61] to model 200 the presence-absence of observed interactions and number of visits as a function of 201 the main axes of plant trait variation and their interactions with floral visitor guilds 202 (e.g., number of visits \sim PC1 x FGs + PC2 x FGs + PC3 x FGs). Because we were 203 interested in possible differences in visitation patterns among floral visitor guilds to 204 plants with different reproductive strategies, we included interactions between the 205 main axes of trait variation (PC1, PC2 and PC3) and the different floral visitor guilds 206 in the model. We added a nested random effect of networks within the study system 207 to capture the variation in networks among studies and within networks. Moreover, 208 we included the phylogenetic covariance matrix as a random factor due to the possible 209 shared evolutionary histories of species and therefore lack of independence among 210 them. We specified for the presence or absence of an interaction and number of visits 211 with Bernoulli and a zero inflated negative binomial distributions, respectively. The 212 models were run with 3,000 iterations with previous 1,000 warm up iterations and

with non or very weakly informative priors from the brm function so the priors had negligible influence on the results [61]. We set delta (Δ) to 0.99 to avoid divergent transitions and visualized the posterior predictive checks with the function *pp_check* using the *bayesplot* package [62].

218 (ii) Plant species network roles

We also used Bayesian modeling to investigate how the main axes of trait variation 219 determined different plant species' functional roles within pollination networks. For 220 this, we selected complementary species-level network metrics, commonly applied in 221 bipartite network studies [63], with clear ecological interpretations and are relevant for 222 defining species' functional roles in networks. Specifically, the species-level metrics we 223 included were: (i) sum of visits per plant species; (ii) normalized degree, calculated 224 as the number of links per plant species divided by the total possible number of 225 partners; and (iii) specialization (d') [64], which measures the deviation of an expected 226 random choice of the available interaction partners and ranges between 0 (maximum 227 generalization) and 1 (maximum specialization). Normalized degree and specialization were calculated with the *specieslevel* function in the R package *bipartite* [63]. We modeled 229 each plant species metric (sum of visits, normalized degree and plant specialization) as a function of the three main axes of trait variation (plant species level metric ~ 231 PC1 + PC2 + PC3). For the different response variables in each model (i.e., each plant species level metric), we used different distribution families (zero inflated negative 233 binomial for the sum of visits, weibull for normalized degree, and zero-one inflated beta for specialization). Finally, we used the same random factors, model settings and 235 conducted the same posterior predictive checks for each model as detailed above in the 'visitation patterns' section. 237

Our analyses were conducted on the subset of 60 weighted networks (out of a total of 64 networks) sampled in a unique flowering season and site, which included 556 plant and 1,126 pollinator species. In total, our network dataset (excluding meta- and non-weighted networks) included 2,256 interactions for bees, 1,768 for non-syrphid-

dipterans, 845 for syrphids, 437 for lepidopterans, 432 for coleopterans, and 362 for nonbee-hymenopterans. Sampling methods varied across studies but this was accounted for in analyses by including study identity as a random effect in our models. All analyses were conducted in R version 4.0.3.

3. Results

47 (a) Plant strategies

Our phylogenetically informed principal component analysis captured 51.8% and 248 70.97% of the reproductive trait variation in the first two and three axes, respectively (figure 1 and figure S6). The first principal component (PC1; the flower number - flower 250 size axis) represented 26.72% of the trait variation and indicated a negative correlation between flower number and flower size. The main contributing traits to PC1 were 252 plant height, flower number, ovule number and flower size (loadings > | 0.5|; table 253 S3) but style length also contributed moderately (loading = -0.33). One end of this 254 axis comprised species with high investment in flower number and plant height but 255 small flower size, short style length and low ovule number. For instance, Cornus florida 256 occurred at this end of the spectrum, which, on average, has approximately 10,000 257 flowers, is 7.5 m high, has flowers that are 3 mm wide, styles 3.5 mm long and two 258 ovules per flower. The other end of this spectrum had species that were short in height, 259 with few large flowers that had long styles and many ovules. For example, Petunia 260 axillaris on average, is 0.5 m high and has approximately 10 flowers per plant that are 261 over 50 mm wide with 25 mm long styles and over 200 ovules. The second principal 262 component (PC2; the pollinator dependence axis) encompassed 25.05% of the trait 263 variation and represented variation from low to high autonomous selfing (i.e., high to 264 low pollinator dependence). The main driver of trait variation on PC2 was autonomous 265 selfing (loading = 0.85) but all other traits (except ovule number) made moderate 266 contributions (loadings ranging between 0.27 to 0.4; table S3). Generally speaking, 267 species with high pollinator dependence had larger and many flowers with long styles,

and were tall. Zuccagnia punctata occurred at this extreme of the spectrum, which 269 is a self-incompatible species, completely dependent on animal pollination for seed 270 production, producing approximately 1,500 flowers per plant with an average height 271 of 3 m and 20 mm long styles. In contrast, species with low pollinator dependence 272 tended to have fewer and smaller flowers with shorter styles, and were shorter in 273 height. Veronica peregrina occurred at this end of the spectrum, and is a self-compatible 274 plant that requires little or no pollination by animals, it has approximately 20 flowers 275 per plant, is 0.2 m high and has 0.25 mm long styles. The third principal component 276 (PC3; the style length - pollinator dependence axis) explained a considerable amount of 277 trait variability (19.17%) and represented a negative correlation between style length 278 (loading = -0.66) and pollinator dependence (autonomous selfing loading = |-0.51|), 279 where species with short styles had high pollinator dependence and species with long styles low pollinator dependence. The remaining traits, apart from ovule number, were 281 all positively correlated with style length and negatively correlated with pollinator 282 dependence (loadings from -0.23 to -0.46; table S3). In addition, the pPCA with the 283 subset of species that we had nectar and pollen quantity data for showed that nectar 284 quantity (microlitres of nectar per flower) and pollen grains per flower were positively 285 associated with floral display size. Floral rewards were particularly strongly correlated with flower size but weakly correlated with flower number (figure S4). The first two 287 principal components for this pPCA explained similar variance (45.52%) and similar 288 associations of traits, despite some variability in the loadings (table S4). 289

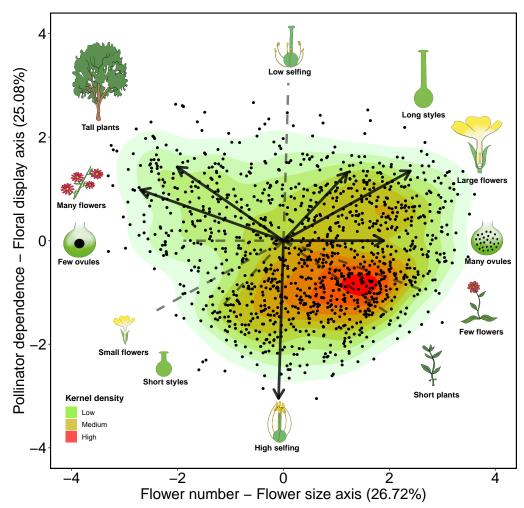


Figure 1. Phylogenetically informed principal component analysis (pPCA) of 1,236 plant species from 28 plant-pollinator network studies. The solid arrows indicate the direction of the different quantitative traits (flower number, plant height, style length, flower size, ovule number and level of autonomous selfing) across the two main axes of trait variation. The length of the arrows indicate the weight of the variables on each principal component and the dashed lines show the opposed direction of trait variation. The icons at both ends of arrows and dashed lines illustrate the extreme form of the trait continuum.

We found that most categorical traits were statistically associated with the first two axes of trait variation (figure 2 and table S2). Flower symmetry, which was only associated with PC2 (Sum of squares = 8.51, F-value = 14.72, P < 0.01), and nectar provision, which was independent of PC1 and PC2 (PC1: Sum of squares = 0.37, F-value = 0.29 , P = 0.59; PC2: Sum of squares = 0.83, F-value = 1.43, P = 0.23) showed a lack of statistical association. In addition, we found statistical differences between the different categorical trait levels within the trait space (figure S7). Regarding self-compatibility,

we found larger differences on PC2. For instance, species with unisexual flowers that 297 were self-incompatible, were statistically differentiated from species with partial or full 298 self-compatibility on this axis (figure S7a and figure S7b; Tukey test P < 0.05). Life forms 299 differed statistically across both axes of trait variation and followed a gradient of larger 300 life forms (trees and shrubs) with higher pollinator dependence to smaller ones (herbs) 301 with lower pollinator dependence (figure S7c and figure S7d; Tukey test P < 0.05). 302 Consequently, lifespan also followed this gradient but perennial and short lived species 303 only differed statistically on PC2 (figure S7e and figure S7f; Tukey test P < 0.05). Species 304 with unisexual flowers (monoecious and dioecious) were clustered on both extremes of 305 the first two principal components and had both the highest pollinator dependence and 306 highest number of flowers (figure S7g and figure S7h; Tukey test P < 0.05). Moreover, 307 we found that campanulate and capitulum flower shapes were differentiated from tube, papilionaceous, open and brush shapes in the trait space; the former had larger flowers 309 and high pollinator dependence, while the latter had more flowers and lower pollinator dependence (figure S7i and figure S7j; Tukey test P < 0.05). Finally, in terms of flower 311 symmetry, zygomorphic flowers were associated with lower pollinator dependence, 312 whereas actinomorphic flowers were more dependent on pollinators (figure S7k and 313 figure S7l; Tukey test P < 0.05).

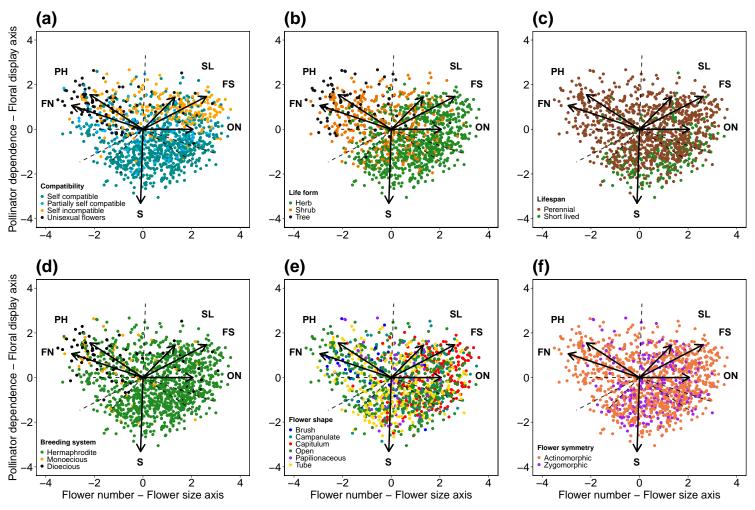


Figure 2. Location of the different qualitative traits in the trait space. The figure panel is composed of the traits that showed statistical association with the first two axes of trait variation: compatibility system (a), life form (b), lifespan (c), breeding system (d), flower shape (e) and flower symmetry (f). The solid arrows indicate the direction of variation of the different quantitative traits shown in figure 1: flower number (FN), plant height (PH), style length (SL), flower size (FS), ovule number (ON) and the level of autonomous selfing (S).

315 (b) Phylogenetic signal

We found a strong phylogenetic signal (P < 0.01) for most quantitative traits (table S5). The traits that showed the highest phylogenetic signal were ovule number ($\lambda = 1$), pollen grains per flower ($\lambda = 1$) and plant height ($\lambda = 0.96$), followed by flower length ($\lambda = 0.75$), flower width ($\lambda = 0.73$), number of flowers per plant ($\lambda = 0.69$) and nectar concentration ($\lambda = 0.65$). The traits that had a moderate phylogenetic signal included inflorescence width ($\lambda = 0.57$), style length ($\lambda = 0.49$) and autonomous selfing ($\lambda = 0.34$). Finally, microliters of nectar per flower showed the lowest phylogenetic signal of all the traits we measured ($\lambda = 0.14$).

324 (c) Visitation patterns

The main axes of trait variation (PC1, PC2 and PC3) partly explained the presenceabsence of interaction partners (conditional $R^2 = 0.26$; marginal $R^2 = 0.20$) but little of 326 the total number of visits (conditional $R^2 = 0.31$; marginal $R^2 = 0.06$). However, we 327 found relevant differences across the different floral visitor guilds for both the presence-328 absence of interactions and number of visits (figure 3). On the flower number - flower 329 size axis, we found that plants with many small flowers tended to have interaction 330 partners that were Coleoptera, non-bee-Hymenoptera and Diptera (figure 3a), whereas 331 plants with few large flowers mostly had interaction partners that were bees and 332 Lepidoptera. We found similar trends for visitation rates on the flower number - flower 333 size axis, but bees and syrphids visited plant species with many small flowers more 334 frequently (figure 3d). On the pollinator dependence axis, we found that all plant 335 species with higher pollinator dependence had many interaction partners and recieved 336 many visits from all floral visitor guilds (figure 3b and figure 3e). Finally, on the style 337 length - pollinator dependence axis, plant species with short styles and high pollinator 338 dependence had more interaction partners for all guilds, except bees which interacted 339 more frequently with plant species with long styles and low pollinator dependence (figure 3c). Plants with long styles and low pollinator dependence interacted more

³⁴² frequently with Lepidoptera and non-bee-Hymenoptera (figure 3f).

The additional models for the most represented bee families, for both presence-absence of interactions (marginal $R^2 = 0.29$; conditional $R^2 = 0.19$) and the number of visits (marginal $R^2 = 0.30$; conditional $R^2 = 0.03$), showed that the Apidae family was the main driver of the observed patterns. The contrast between interaction presence/absence and number of visits for bees on PC1 (figure 3a and figure 3d) was driven by the Andrenidae family, which had more interaction partners but made fewer visits to plant species with few large flowers (figure S8).

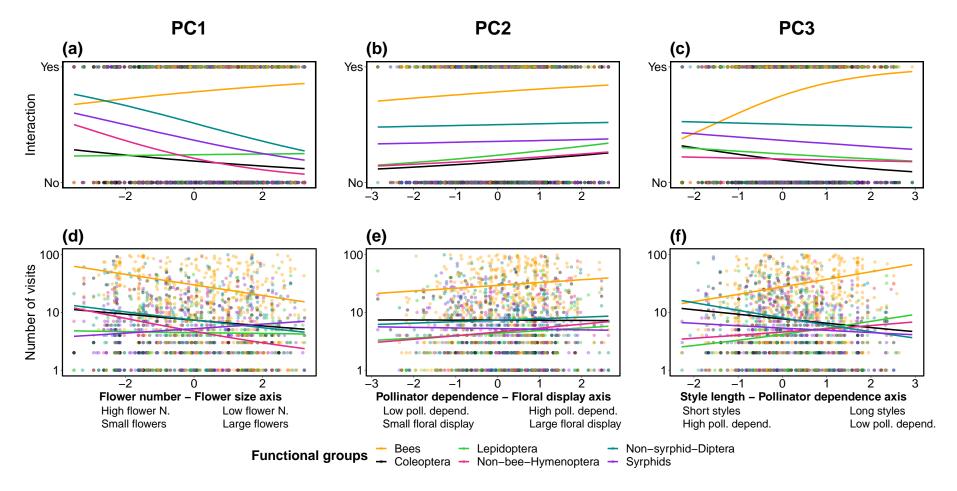


Figure 3. Fitted posterior estimates of the presence-absence of interaction (a, b and c) and number of visits (d, e and f) of the different floral visitor guilds in relation to the main axes of plant reproductive trait variation (PC1, PC2 and PC3). PC1 represents the flower number - flower size axis, PC2 represents the pollinator dependence axis and PC3 represents the style length - pollinator dependence axis. For visualization purposes due to large differences between number of visits from bees and the rest of the pollinator guilds, the number of visits was log-transformed (Y-axis of lower panel).

(d) Plant species network roles

Variance within plant species-level network metrics was poorly explained by the three 351 main axes of trait variation (figure S9; number of visits \sim PCs, conditional $R^2 = 0.11$, 352 marginal $R^2 = 0.02$; normalized degree ~ PCs, conditional $R^2 = 0.24$, marginal $R^2 = 0.02$; 353 and, specialization ~ PCs, conditional $R^2 = 0.37$, marginal $R^2 = 0.03$). Overall, the most 354 notable trends were found on the flower number - flower size axis (PC1) and the style length - pollinator dependence axis (PC3) for number of visits and specialization. On 356 PC1, the number of visits per flower was higher for plant species with more flowers but 357 was lower for plant species with larger flowers (figure S9a). However, specialization 358 (d') was higher for plant species with large flowers but lower for plant species with small flowers (figure S9g). On PC3, the number of visits was lower for plants with 360 shorter styles and higher pollinator dependence but higher for plant species with longer 361 styles and lower pollinator dependence (figure S9c). Again, specialization showed the 362 opposite trend to the number of visits (figure S9i).

4. Discussion

We found strong covariation among traits within the flowering plant reproductive spectrum, which shapes interactions with floral visitors at a global scale. Despite the 366 enormous variability in plant reproductive structures, the first two and three axes 367 captured over 50% and 70% of trait variation, respectively, and these axes encapsulated 368 correlations among flower number and flower size, pollinator dependence and floral 369 display, and style length and pollinator dependence. Although explained trait variation 370 on the first two axes was less than previous studies of morphological and physiological 371 traits [17,19], our results align with the largest and most recent study of plant life 372 strategies that included reproductive, form and size related traits [7]. The different 373 plant reproductive axes that we identified were associated with different numbers of 374 interacting partners and visitation rates of the distinct floral visitor guilds. However, 375 the unexplained variability found highlights the need to account for other local factors, such as species relative abundances [33], along with reproductive traits, to better explain plant-pollinator associations.

Our study provides strong evidence for several widely discussed trait associations 379 with the help of a great diversity of phylogenetic lineages (170 families and over 380 1,200 species). Specifically, by using a multitrait approach, we are able to link the 381 reproductive spectrum of plant trait variation with the previously described "fast-382 slow continuum" in plant [7] and animal [65] life-history strategies. This fast-slow 383 continuum was illustrated by the associations between reproductive traits and plant 384 height, life form and lifespan traits. For instance, although the negative correlation 385 between flower number and flower size has received substantial attention [25,26,66], it 386 has only been investigated across a few hundred Angiosperm species in the absence of a multitrait perspective. We found support for this trait association and show that other 388 reproductive traits (e.g., style length or ovule number) covary along this flower number 389 flower size axis. Our study also complements previous work linking plant breeding 390 systems and life forms [8,67] by showing that most species with unisexual flowers also have woody life forms and many flowers. Moreover, we identify the previously 392 documented positive association between outcrossing rate and floral display size [27], 393 where species with high pollinator dependence also allocate large resource quantities to 394 attract floral visitors. Although we did not include floral rewards in our main analysis 395 due to insufficient data, we did find that plant species with large floral displays and, 396 in particular, large flowers, tended to have higher pollen and nectar quantity. Thus, 397 we demonstrate that globally, reproductive trait covariation in flowering plants likely 398 imposes constraints on the broader spectrum of plant trait variation. 399

Overall, plant reproductive traits explained interaction patterns among and within floral visitor guilds suggesting that plant reproductive strategies partly mediate these associations across spatial large scales. For example, we found that all floral visitor guilds interacted more frequently with large flowered and pollinator dependent plant species, which aligns with previous findings [68–70]. As predicted by evolutionary life-history theory [71], larger investment in floral display and rewards to attract

pollinators occurs when pollinator dependence is high. However, the possible range of 406 reproductive strategies in flowering plants is constrained by the cost of reproductive 407 organs, trait associations and selective pressures exerted by different floral visitor guilds 408 [72]. For instance, smaller flowers are visited less overall, but they are inexpensive 409 to produce (e.g., have a low ovule number) and low visitation rate at the individual 410 flower level can be compensated by overall visitation at the plant level by producing 411 many flowers resulting in a larger quantity of small fruits [26]. In addition, because 412 floral traits can be associated with their animal pollinators to optimise pollen transfer, 413 the correlation between distinct suites of reproductive traits and floral visitor guilds 414 could indicate the existence of pollination syndromes [3,4,31,73]. For instance, bees 415 had a higher number of interacting partners on plant species with large flowers but 416 interacted more frequently with species with small flowers while syrphid flies showed the opposite trend. This could be reflective of differences in the ecology and behaviour 418 between these pollinator guilds [74,75] that ultimately exert different selective pressures, leading divergent plant reproductive strategies. In any case, it is worth noting that 420 local factors (e.g., species relative abundances or phenologies) likely contribute to the 421 variability in the association between the plant reproductive spectrum and floral visitors 422 [33,76,77].

Our study provides a comprehensive assessment of reproductive trait covariation patterns in flowering plants, using a global dataset of traits for plant species within 425 plant-pollinator networks. We show the importance of floral visitors in mediating 426 reproductive trait correlations and identify the need to consider broader plant re-427 productive strategies, rather than just flower morphology, to understand patterns of 428 plant-pollinator interactions. Further, we provide empirical evidence of reproductive 429 trait associations that complements trait covariation studies for other plant organs, 430 highlighting the need to include reproductive traits in the global spectrum of plant 431 form and function. Future work incorporating underrepresented areas of the world 432 and additional relevant reproductive traits (e.g., flower life span and colour) will be valuable for enhancing our understanding of the plant reproductive spectrum and

associated plant-pollinator interactions.

References

- Barrett SCH. 2002 The evolution of plant sexual diversity. *Nat. Rev. Genet.* 3, 274–284. (doi:10.1038/nrg776)
- 2. Schiestl FP, Johnson SD. 2013 Pollinator-mediated evolution of floral signals.

 Trends Ecol. Evol. 28, 307–315. (doi:10.1016/j.tree.2013.01.019)
- Fenster CB, Armbruster WS, Wilson P, Dudash MR, Thomson JD. 2004 Pollination Syndromes and Floral Specialization. *Annu. Rev. Ecol. Evol. Syst.* **35**, 375–403. (doi:10.1146/annurev.ecolsys.34.011802.132347)
- 4. Dellinger AS. 2020 Pollination syndromes in the 21st century: Where do we stand and where may we go? *New Phytol.* **228**, 1193–1213. (doi:10.1111/nph.16793)
- 5. Stearns SC. 1989 Trade-offs in life-history evolution. *Funct. Ecol.* **3**, 259–268. (doi:10.2307/2389364)
- Agrawal AA. 2020 A scale-dependent framework for trade-offs, syndromes, and specialization in organismal biology. *Ecology* **101**, e02924. (doi:10.1002/ecy.2924)
- 7. Salguero-Gómez R, Jones OR, Jongejans E, Blomberg SP, Hodgson DJ, Mbeau-Ache C, Zuidema PA, de Kroon H, Buckley YM. 2016 Fast-slow continuum and reproductive strategies structure plant life-history variation worldwide. *Proc. Natl. Acad. Sci. U.S.A.* 113, 230–235. (doi:10.1073/pnas.1506215112)
- Friedman J. 2020 The evolution of annual and perennial plant life histories: Ecological correlates and genetic mechanisms. *Annu. Rev. Ecol. Evol. Syst.* **51**, 461–481. (doi:10.1146/annurev-ecolsys-110218-024638)
- Paterno GB, Silveira CL, Kollmann J, Westoby M, Fonseca CR. 2020 The maleness of larger angiosperm flowers. *Proc. Natl. Acad. Sci.* 117, 10921–10926.
 (doi:10.1073/pnas.1910631117)

- RS, Silveira FAO. 2021 Towards the flower economics spectrum. *New Phytol.* **229**, 665–672. (doi:10.1111/nph.16823)
- Song B *et al.* 2022 Global analysis of floral longevity reveals latitudinal gradients and biotic and abiotic correlates. *New Phytologist* (doi:10.1111/nph.18271)
- Laughlin DC *et al.* 2021 Root traits explain plant species distributions along climatic gradients yet challenge the nature of ecological trade-offs. *Nat. Ecol. Evol.*, 1–12. (doi:10.1038/s41559-021-01471-7)
- Wright IJ *et al.* 2004 The worldwide leaf economics spectrum. *Nature* **428**, 821–827. (doi:10.1038/nature02403)
- Chave J, Coomes D, Jansen S, Lewis SL, Swenson NG, Zanne AE. 2009 Towards a worldwide wood economics spectrum. *Ecol. Lett.* **12**, 351–366. (doi:10.1111/j.1461-0248.2009.01285.x)
- Kattge J *et al.* 2011 TRY a global database of plant traits. *Glob. Chang. Biol.* 17,
 2905–2935. (doi:10.1111/j.1365-2486.2011.02451.x)
- Salguero-Gómez R *et al.* 2015 The compadre Plant Matrix Database: An open online repository for plant demography. *J. Ecol.* **103**, 202–218. (doi:10.1111/1365-2745.12334)
- Díaz S *et al.* 2016 The global spectrum of plant form and function. *Nature* 529,
 167–171. (doi:10.1038/nature16489)
- Bruelheide H *et al.* 2018 Global trait–environment relationships of plant communities. *Nat. Ecol. Evol.* **2**, 1906–1917. (doi:10.1038/s41559-018-0699-8)
- Carmona CP *et al.* 2021 Erosion of global functional diversity across the tree of life. *Sci. Adv.* 7, eabf2675. (doi:10.1126/sciadv.abf2675)
- E-Vojtkó A, Bello F de, Durka W, Kühn I, Götzenberger L. 2020 The neglected importance of floral traits in trait-based plant community assembly. *J. Veg. Sci.*31, 529–539. (doi:10.1111/jvs.12877)

- Rüger N, Comita LS, Condit R, Purves D, Rosenbaum B, Visser MD, Wright SJ, Wirth C. 2018 Beyond the fast–slow continuum: Demographic dimensions structuring a tropical tree community. *Ecol. Lett.* **21**, 1075–1084. (doi:10.1111/ele.12974)
- Shipley B, Lechowicz MJ, Wright I, Reich PB. 2006 Fundamental trade-offs generating the worldwide leaf economics spectrum. *Ecology* **87**, 535–541. (doi:10.1890/05-1051)
- Donovan LA, Maherali H, Caruso CM, Huber H, Kroon H de. 2011 The evolution of the worldwide leaf economics spectrum. *Trends in Ecology & Evolution* **26**, 88–95. (doi:10.1016/j.tree.2010.11.011)
- Osnas JL, Lichstein JW, Reich PB, Pacala SW. 2013 Global leaf trait relationships: Mass, area, and the leaf economics spectrum. *Science* **340**, 741–744. (doi:10.1126/science.1231574)
- Sargent RD, Goodwillie C, Kalisz S, Ree RH. 2007 Phylogenetic evidence for a flower size and number trade-off. *Am. J. Bot.* **94**, 2059–2062. (doi:10.3732/ajb.94.12.2059)
- ⁴⁸⁷ 26. Kettle CJ, Maycock CR, Ghazoul J, Hollingsworth PM, Khoo E, Sukri RSH, Burslem DFRP. 2011 Ecological Implications of a Flower Size/Number Trade-Off in Tropical Forest Trees. *PLoS One* **6**, e16111. (doi:10.1371/journal.pone.0016111)
- Goodwillie C *et al.* 2010 Correlated evolution of mating system and floral display traits in flowering plants and its implications for the distribution of mating system variation. *New Phytol.* **185**, 311–321. (doi:10.1111/j.1469-8137.2009.03043.x)
- Barrett SCH. 2003 Mating strategies in flowering plants: The outcrossing-selfing paradigm and beyond. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* **358**, 991–1004. (doi:10.1098/rstb.2003.1301)
- Munoz F, Violle C, Cheptou P-O. 2016 CSR ecological strategies and plant mating systems: Outcrossing increases with competitiveness but stress-tolerance is related to mixed mating. *Oikos* 125, 1296–1303. (doi:10.1111/oik.02328)

- Moeller DA *et al.* 2017 Global biogeography of mating system variation in seed plants. *Ecol. Lett.* **20**, 375–384. (doi:10.1111/ele.12738)
- Ago 31. Rosas-Guerrero V, Aguilar R, Martén-Rodríguez S, Ashworth L, Lopezaraiza-Mikel M, Bastida JM, Quesada M. 2014 A quantitative review of pollination syndromes: Do floral traits predict effective pollinators? *Ecol. Lett.* **17**, 388–400. (doi:10.1111/ele.12224)
- Stang M, Klinkhamer PGL, Waser NM, Stang I, van der Meijden E. 2009 Size-specific interaction patterns and size matching in a plant–pollinator interaction
 web. *Ann. Bot.* 103, 1459–1469. (doi:10.1093/aob/mcp027)
- Bartomeus I, Gravel D, Tylianakis JM, Aizen MA, Dickie IA, Bernard-Verdier M. 2016 A common framework for identifying linkage rules across different types of interactions. *Funct. Ecol.* **30**, 1894–1903. (doi:10.1111/1365-2435.12666)
- Tur C, Castro-Urgal R, Traveset A. 2013 Linking Plant Specialization to Dependence in Interactions for Seed Set in Pollination Networks. *PLoS One* 8, e78294.
 (doi:10.1371/journal.pone.0078294)
- Junker RR, Blüthgen N, Brehm T, Binkenstein J, Paulus J, Schaefer HM, Stang M. 2013 Specialization on traits as basis for the niche-breadth of flower visitors and as structuring mechanism of ecological networks. *Funct. Ecol.* 27, 329–341. (doi:10.1111/1365-2435.12005)
- ⁵⁰⁷ 36. Coux C, Rader R, Bartomeus I, Tylianakis JM. 2016 Linking species functional roles to their network roles. *Ecol. Lett.* **19**, 762–770. (doi:10.1111/ele.12612)
- Towards a mechanistic understanding of ecological networks. *Oecologia* **170**, 233–242. (doi:10.1007/s00442-012-2290-3)
- Sargent RD, Ackerly DD. 2008 Plant–pollinator interactions and the assembly of plant communities. *Trends Ecol. Evol.* **23**, 123–130. (doi:10.1016/j.tree.2007.11.003)

- Devaux C, Lepers C, Porcher E. 2014 Constraints imposed by pollinator behaviour on the ecology and evolution of plant mating systems. *J. Evol. Biol.* 27, 1413–1430. (doi:10.1111/jeb.12380)
- 515 40. Eklöf A *et al.* 2013 The dimensionality of ecological networks. *Ecol. Lett.* **16**, 577–583. (doi:10.1111/ele.12081)
- Dehling DM, Jordano P, Schaefer HM, Böhning-Gaese K, Schleuning M. 2016

 Morphology predicts species' functional roles and their degree of specialization in plant–frugivore interactions. *Proc. Royal Soc. B* **283**, 20152444.

 (doi:10.1098/rspb.2015.2444)
- Carvalheiro LG *et al.* 2014 The potential for indirect effects between co-flowering plants via shared pollinators depends on resource abundance, accessibility and relatedness. *Ecol. Lett.* **17**, 1389–1399. (doi:10.1111/ele.12342)
- Olesen JM, Bascompte J, Dupont YL, Jordano P. 2007 The modularity of pollination networks. *PNAS* **104**, 19891–19896. (doi:10.1073/pnas.0706375104)
- Fortuna MA, Stouffer DB, Olesen JM, Jordano P, Mouillot D, Krasnov BR, Poulin R, Bascompte J. 2010 Nestedness versus modularity in ecological networks:

 Two sides of the same coin? *J. Anim. Ecol.* **79**, 811–817. (doi:10.1111/j.1365-2656.2010.01688.x)
- Poisot T *et al.* 2016 Mangal making ecological network analysis simple. *Ecogra- phy* **39**, 384–390. (doi:10.1111/ecog.00976)
- 527 46. Chamberlain S *et al.* 2020 Taxize: Taxonomic information from around the web.

 R package version 0.9.99. At https://CRAN.r-project.org/package=taxize.
- 529 47. Smith SA, Brown JW. 2018 Constructing a broadly inclusive seed plant phylogeny.

 Am. J. Bot. 105, 302–314. (doi:10.1002/ajb2.1019)
- Jin Y, Qian H. 2019 V.PhyloMaker: An R package that can generate very large phylogenies for vascular plants. *Ecography* **42**, 1353–1359. (doi:10.1111/ecog.04434)

- Stekhoven DJ, Bühlmann P. 2012 MissForest—non-parametric missing value imputation for mixed-type data. *Bioinformatics* **28**, 112–118. (doi:10.1093/bioinformatics/btr597)
- Penone C, Davidson AD, Shoemaker KT, Marco MD, Rondinini C, Brooks TM, Young BE, Graham CH, Costa GC. 2014 Imputation of missing data in life-history trait datasets: Which approach performs the best? *Methods Ecol. Evol.* **5**, 961–970. (doi:10.1111/2041-210X.12232)
- 537 51. Chamberlain S *et al.* 2018 Package "PVR". R package version 0.3. At https://CRAN.r-project.org/package=PVR.
- Diniz-Filho JAF, Bini LM, Rangel TF, Morales-Castilla I, Olalla-Tárraga MÁ,
 Rodríguez MÁ, Hawkins BA. 2012 On the selection of phylogenetic eigenvectors for ecological analyses. *Ecography* 35, 239–249. (doi:10.1111/j.1600-0587.2011.06949.x)
- 53. Serneels S, Verdonck T. 2008 Principal component analysis for data containing outliers and missing elements. *Comput. Stat. Data Anal.* **52**, 1712–1727. (doi:10.1016/j.csda.2007.05.024)
- 54. Legendre P, Legendre L. 2012 *Numerical ecology*. 2nd edn. Elsevier, Amsterdam.
- 545 Sevell LJ. 2012 Phytools: An R package for phylogenetic comparative biology (and other things). *Methods Ecol. Evol.* 3, 217–223. (doi:10.1111/j.2041-210X.2011.00169.x)
- 547 56. Abdi H, Williams LJ. 2010 Principal component analysis. *WIREs Comp. Stats.* **2**, 433–459. (doi:10.1002/wics.101)
- 57. Ballantyne G, Baldock KCR, Willmer PG. 2015 Constructing more informative plant–pollinator networks: Visitation and pollen deposition networks in a heathland plant community. *Proc. Royal Soc. B* **282**, 20151130. (doi:10.1098/rspb.2015.1130)

- 58. Vázquez DP, Morris WF, Jordano P. 2005 Interaction frequency as a surrogate for the total effect of animal mutualists on plants. *Ecol. Lett.* **8**, 1088–1094. (doi:10.1111/j.1461-0248.2005.00810.x)
- Vázquez DP, Lomáscolo SB, Maldonado MB, Chacoff NP, Dorado J, Stevani EL,
 Vitale NL. 2012 The strength of plant–pollinator interactions. *Ecology* 93, 719–725.
 (doi:10.1890/11-1356.1)
- Hung K-LJ, Kingston JM, Albrecht M, Holway DA, Kohn JR. 2018 The worldwide importance of honey bees as pollinators in natural habitats. *Proc. Royal Soc. B* 285, 20172140. (doi:10.1098/rspb.2017.2140)
- 557 61. Bürkner P-C. 2017 Brms: An R package for Bayesian multilevel models using Stan. *J. Stat. Softw.* **80**, 1–28. (doi:10.18637/jss.v080.i01)
- Gabry J, Simpson D, Vehtari A, Betancourt M, Gelman A. 2019 Visualization in Bayesian workflow. *J. R. Stat. Soc. Ser. A Stat. Soc.* **182**, 389–402. (doi:10.1111/rssa.12378)
- Dormann CF, Gruber B, Fründ J. 2008 Introducing the bipartite package:
 Analysing ecological networks. *R News* **8/2**.
- Blüthgen N, Menzel F, Blüthgen N. 2006 Measuring specialization in species interaction networks. *BMC Ecol.* **6**, 9. (doi:10.1186/1472-6785-6-9)
- Healy K, Ezard THG, Jones OR, Salguero-Gómez R, Buckley YM. 2019 Animal life history is shaped by the pace of life and the distribution of age-specific mortality and reproduction. *Nat. Ecol. Evol.* 3, 1217–1224. (doi:10.1038/s41559-019-0938-7)
- Worley AC, Baker AM, Thompson JD, Barrett SC. 2000 Floral display in narcissus:

 Variation in flower size and number at the species, population, and individual levels. *Int. J. Plant Sci.* **161**, 69–79. (doi:10.1086/314225)
- 67. Hampe 2006 Some evolutionary RJ, A. consequences of 569 187–214. being a tree. Annu. Rev. Ecol. Evol. Syst. 37, (doi:10.1146/annurev.ecolsys.37.091305.110215) 570

- Hegland SJ, Totland Ø. 2005 Relationships between species' floral traits and pollinator visitation in a temperate grassland. *Oecologia* **145**, 586–594. (doi:10.1007/s00442-005-0165-6)
- Lázaro A, Jakobsson A, Totland Ø. 2013 How do pollinator visitation rate and seed set relate to species' floral traits and community context? *Oecologia* 173, 881–893. (doi:10.1007/s00442-013-2652-5)
- Kaiser-Bunbury CN, Vázquez DP, Stang M, Ghazoul J. 2014 Determinants of the microstructure of plant–pollinator networks. *Ecology* 95, 3314–3324.
 (doi:10.1890/14-0024.1)
- Stebbins GL. 1970 Adaptive radiation of reproductive characteristics in angiosperms, i: Pollination mechanisms. *Annual Review of Ecology and Systematics*,
 307–326. (doi:10.1146/annurev.es.01.110170.001515)
- Galen C. 1999 Why do flowers vary? The functional ecology of variation in flower size and form within natural plant populations. *Bioscience* **49**, 631–640. (doi:10.2307/1313439)
- ⁵⁸¹ 73. Faegri K, Van Der Pijl L. 2013 *Principles of pollination ecology*. 3rd edn. Elsevier.

582

- Rader R, Cunningham S, Howlett B, Inouye D. 2020 Non-bee insects as visitors and pollinators of crops: Biology, ecology, and management. *Annu. Rev. Entomol.* 65, 391–407. (doi:10.1146/annurev-ento-011019-025055)
- Földesi R, Howlett BG, Grass I, Batáry P. 2021 Larger pollinators deposit more pollen on stigmas across multiple plant species—a meta-analysis. *J. Appl. Ecol.*58, 699–707. (doi:10.1111/1365-2664.13798)
- Vázquez DP, Melián CJ, Williams NM, Blüthgen N, Krasnov BR, Poulin R. 2007
 Species abundance and asymmetric interaction strength in ecological networks.
 Oikos 116, 1120–1127. (doi:10.1111/j.0030-1299.2007.15828.x)

- 589 77. Encinas-Viso F, Revilla TA, Etienne RS. 2012 Phenology drives mutualistic network structure and diversity. *Ecol. Lett.* **15**, 198–208. (doi:10.1111/j.1461-0248.2011.01726.x)
- Data accessibility. All data and code used to conduct this study are available in the online repositories of Zenodo (https://doi.org/10.5281/zenodo.6705621) and Github (https://github.com/JoseBSL/Reproductive-traits).
- Authors' contributions. JBL, RR and IB designed the study. JBL collated the data and conducted analysis with guidance of JS, LKK and IB. JBL wrote the manuscript with contributions of all authors.
- **Conflict of interest declaration.** We declare we have no competing interests.
- Funding. This study was supported by the European project SAFEGUARD (101003476 H2020-SFS-2019-2) and JBL was supported by a University of New England IPRA postgraduate scholarship.
- Acknowledgements. We thank all researchers that made their data openly available and sent it upon request. We also thank Bryony Wilcox, Greg Bible, Mercedes Sanchez-Lanuza and David Ragel for their help with data collection. We thank Jason Tylianakis for his insightful comments on the manuscript before submission. Finally, we thank Marcos Méndez, Susan Kalisz, Amanda Benoit, May Berenbaum and an anonymous reviewer for thoughtful feedback and useful comments on earlier versions of this manuscript.