

- Cardiotensor: A Python Library for Orientation
- 2 Analysis and Tractography in 3D Cardiac Imaging
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# Summary

Understanding the architecture of the human heart requires analyzing its microstructural organization across scales. With the advent of high-resolution imaging techniques such as synchrotron-based tomography, it has become possible to visualize entire hearts at micron-scale resolution. However, translating these large, complex volumetric datasets into interpretable, quantitative descriptors of cardiac organization remains a major challenge. Here we present cardiotensor, an open-source Python package designed to quantify 3D cardiomyocyte orientation in whole- or partial-heart imaging datasets. It provides efficient, scalable implementations of structure tensor analysis, enabling extraction of directional metrics such as helical angle (HA), intrusion angle (IA), and fractional anisotropy (FA). The package supports datasets reaching teravoxel-scale and is optimized for high-performance computing environments, including parallel and chunk-based processing pipelines. In addition, cardiotensor includes tractography functionality to reconstruct continuous cardiomyocyte trajectories. This enables fiber-level visualization and structural mapping of cardiac tissue, allowing detailed assessments of anatomical continuity and regional organization.

# Statement of Need

Despite major advances in high-resolution 3D imaging, there is a lack of open-source tools to analyze cardiomyocyte orientation in large volumetric datasets. Most established frameworks were developed for diffusion tensor MRI (DT-MRI), where orientation is inferred from water diffusion. Examples include MRtrix3 (Tournier et al., 2019), DIPY (Garyfallidis et al., 2014), and DSI Studio (Yeh, 2025). While powerful for diffusion-based neuro and cardiac applications (Mekkaoui et al., 2017), these packages are not designed to handle direct image-gradient-based orientation estimation or the teravoxel-scale datasets produced by synchrotron tomography, micro-CT, or 3D optical microscopy.

For non-diffusion imaging modalities, researchers have historically relied on custom structure tensor implementations to estimate fiber orientation directly from image intensity gradients. However, most of these are in-house codes, often unpublished or not generalizable. For example, structure tensor analysis has been applied in the heart using micro-CT (Reichardt et al., 2020), optical microscopy (Dileep et al., 2023; Garcia-Canadilla et al., 2022), and synchrotron tomography (Dejea et al., 2019), but these methods were tailored to specific datasets and lacked scalability or public availability. Existing tools like OrientationJ (Fiji) and OrientationPy (Python) enable 2D and 3D structure tensor analysis for microscopy (Navaee



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et al., 2023). However, they are not optimized for teravoxel-scale datasets, do not compute classical cardiac microstructure descriptors such as HA and IA, and are not integrated with tractography. Cardiotensor uniquely provides all of these capabilities for large-scale cardiac imaging.

For non-diffusion imaging modalities, such as micro-CT (Reichardt et al., 2020), optical microscopy (Dileep et al., 2023; Garcia-Canadilla et al., 2022), and synchrotron tomography (Brunet et al., 2024; Dejea et al., 2019), researchers have historically relied on custom structure tensor implementations to estimate fiber orientation directly from image intensity gradients. However, most of these are in-house codes, often unpublished or not scalable. Existing tools like OrientationJ (Fiji) and OrientationPy (Python) enable 2D and 3D structure tensor analysis (Navaee et al., 2023), but they are not optimized for teravoxel-scale datasets, do not compute classical cardiac microstructure descriptors such as HA and IA, and do not allow tractography.

Cardiotensor addresses this gap by providing an open-source Python package specifically tailored to structure tensor analysis of large cardiac volumes. Rather than relying on diffusion modeling, cardiotensor infers tissue orientation directly from image intensity gradients, making it applicable across a wide range of modalities. Previous studies have demonstrated strong agreement between structure tensor—based orientation and DT-MRI—derived metrics when applied to the same human hearts (Teh et al., 2016). The package supports full pipelines from raw image stacks to fiber orientation maps and tractography. Its architecture is optimized for large datasets, using chunked and parallel processing suitable for high-performance computing environments.

Cardiotensor has already been successfully applied in published work to characterize 3D cardiomyocyte architecture in healthy and diseased human hearts using synchrotron tomography (Brunet et al., 2024) to datasets over a terabyte in size. While cardiotensor was conceived for cardiac imaging, the package is modality- and tissue-agnostic. Any volumetric dataset exhibiting coherent fibrous microstructure can be analyzed, including brain white matter, skeletal muscle, and tendon. This generality makes the library useful for both cardiovascular and broader anatomical or histological studies.

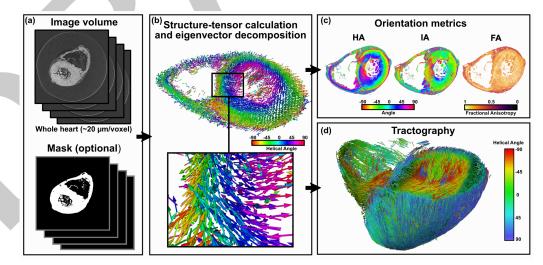


Figure 1: Cardiotensor pipeline for 3D cardiac orientation analysis and tractography. (a) Input whole- or partial-heart volume with optional myocardial mask. (b) Local cardiomyocyte orientation estimated via 3D structure tensor and eigenvector decomposition. The third eigenvector field (smallest eigenvalue) is visualized as arrows color-coded by helical angle (HA); inset shows structure tensor orientation in the ventricular septum. (c) Transformation to a cylindrical coordinate system enables computation of voxel-wise helical angle (HA), intrusion angle (IA), and fractional anisotropy (FA) maps. (d) Streamline tractography reconstructs continuous cardiomyocyte trajectories, color-coded by HA.



#### Implementation

Cardiotensor is implemented in Python and designed to efficiently process very large 3D cardiac imaging datasets. It relies primarily on NumPy (Van Der Walt et al., 2011) for numerical computation, with I/O accelerated by tifffile (Gohlke, 2025), Glymur (Evans, 2025), and OpenCV (Bradski, 2000). Dask (Rocklin, 2015) is used exclusively to parallelize file reading, while the core computations rely on Python's multiprocessing module for local parallelism. The package builds on the structure-tensor library (Jeppesen et al., 2021) to calculate the 3D structure tensor and eigenvector decomposition.

78 The package supports multiple use cases:

- Command-line workflows, which automate batch processing from a configuration file of terabyte-scale heart volumes and produce results as live plots or files saved to disk.
- Embedded use in larger Python analysis workflows, enabling flexible scripting and scalable execution on cluster environments.

Efficient computation is achieved through a chunk-based processing strategy with padding, which avoids edge artifacts. This architecture allows parallelization across computing clusters by splitting volumes into independent jobs, enabling cardiotensor to process whole-heart volumes in hours rather than days while maintaining practical memory requirements.

### 87 Architecture

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Cardiotensor is organized into five main modules, designed for clarity and scalability:

- orientation: Computes local cardiomyocyte (or other texture feature) orientation using a chunked 3D structure tensor pipeline, including eigenvalue decomposition, cylindrical coordinate rotation, and calculation of helical angle (HA), intrusion angle (IA), and fractional anisotropy (FA).
- tractography: Generates and filters streamlines tracing cardiomyocyte trajectories from the orientation field for fiber-level reconstruction and analysis.
- analysis: Provides a GUI for regional quantification and plotting transmural profile.
- **visualization**: Supports interactive 3D visualization of vector fields and streamlines, HA color-coding, and export to VTK/ParaView for large-scale rendering.
- utils: Contains general utilities for I/O, image preprocessing, configuration parsing, and vector math, supporting the entire pipeline.

This modular architecture ensures reproducibility, maintainability, and easy integration into larger cardiac imaging workflows.

#### Documentation and Usage

The documentation for cardiotensor is available online at:

https://josephbrunet.github.io/cardiotensor

105 The main components of the documentation are:

- Step-by-step walkthroughs for installation, first steps, and a guided example covering all available commands. A small example dataset and its corresponding mask are provided with the package.
- In-depth explanations of the core algorithms used in cardiotensor, including structure tensor theory, helical angle calculation, fractional anisotropy (FA), and tractography integration.
- Reference guides for the command-line interface, configuration file format, and public API.



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### References

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- Bradski, G. (2000). The opency library. *Dr. Dobb's Journal: Software Tools for the Professional Programmer*, 25(11), 120–123.
- Brunet, J., Cook, A. C., Walsh, C. L., Cranley, J., Tafforeau, P., Engel, K., Arthurs, O., Berruyer, C., Burke O'Leary, E., Bellier, A., Torii, R., Werlein, C., Jonigk, D. D., Ackermann, M., Dollman, K., & Lee, P. D. (2024). Multidimensional Analysis of the Adult Human Heart in Health and Disease Using Hierarchical Phase-Contrast Tomography. *Radiology*, 312(1), e232731. https://doi.org/10.1148/radiol.232731
- Dejea, H., Garcia-Canadilla, P., Cook, A. C., Guasch, E., Zamora, M., Crispi, F., Stampanoni,
   M., Bijnens, B., & Bonnin, A. (2019). Comprehensive Analysis of Animal Models of
   Cardiovascular Disease using Multiscale X-Ray Phase Contrast Tomography. Scientific
   Reports, 9(1), 6996. https://doi.org/10.1038/s41598-019-43407-z
- Dileep, D., Syed, T. A., Sloan, T. F., Dhandapany, P. S., Siddiqi, K., & Sirajuddin, M. (2023).

  Cardiomyocyte orientation recovery at micrometer scale reveals long-axis fiber continuum in heart walls. *The EMBO Journal*, 42(19), e113288. https://doi.org/10.15252/embj. 2022113288
- Evans, J. G. (2025). Quintusdias/glymur. https://github.com/quintusdias/glymur
- Garcia-Canadilla, P., Mohun, T. J., Bijnens, B., & Cook, A. C. (2022). Detailed quantification of cardiac ventricular myocardial architecture in the embryonic and fetal mouse heart by application of structure tensor analysis to high resolution episcopic microscopic data.

  Frontiers in Cell and Developmental Biology, 10, 1000684. https://doi.org/10.3389/fcell. 2022.1000684
- Garyfallidis, E., Brett, M., Amirbekian, B., Rokem, A., Van Der Walt, S., Descoteaux, M., Nimmo-Smith, I., & Dipy Contributors. (2014). Dipy, a library for the analysis of diffusion MRI data. Frontiers in Neuroinformatics, 8. https://doi.org/10.3389/fninf.2014.00008
- Gohlke, C. (2025). Tifffile. https://github.com/cgohlke/tifffile
- Jeppesen, N., Mikkelsen, L. P., Dahl, A. B., Christensen, A. N., & Dahl, V. A. (2021). Quantifying effects of manufacturing methods on fiber orientation in unidirectional composites



- using structure tensor analysis. Composites Part A: Applied Science and Manufacturing, 149, 106541. https://doi.org/10.1016/j.compositesa.2021.106541
- Mekkaoui, C., Reese, T. G., Jackowski, M. P., Bhat, H., & Sosnovik, D. E. (2017). Diffusion MRI in the heart. *NMR in Biomedicine*, 30(3), e3426. https://doi.org/10.1002/nbm.3426
- Navaee, F., Khornian, N., Longet, D., Heub, S., Boder-Pasche, S., Weder, G., Kleger, A., Renaud, P., & Braschler, T. (2023). A three-dimensional engineered cardiac in vitro model:
  Controlled alignment of cardiomyocytes in 3D microphysiological systems. *Cells*, *12*(4), 576.
- Reichardt, M., Töpperwien, M., Khan, A., Alves, F., & Salditt, T. (2020). Fiber orientation in a whole mouse heart reconstructed by laboratory phase-contrast micro-CT. *Journal of Medical Imaging*, 7(02), 1. https://doi.org/10.1117/1.JMI.7.2.023501
- Rocklin, M. (2015). Dask: Parallel Computation with Blocked algorithms and Task Scheduling. 126–132. https://doi.org/10.25080/Majora-7b98e3ed-013
- Teh, I., McClymont, D., Zdora, M.-C., Whittington, H. J., Davidoiu, V., Lee, J., Lygate, C. A., Rau, C., Zanette, I., & Schneider, J. E. (2016). Validation of diffusion tensor MRI measurements of cardiac microstructure with structure tensor synchrotron radiation imaging. *Journal of Cardiovascular Magnetic Resonance*, 19(1), 31. https://doi.org/10.1186/s12968-017-0342-x
- Tournier, J.-D., Smith, R., Raffelt, D., Tabbara, R., Dhollander, T., Pietsch, M., Christiaens, D., Jeurissen, B., Yeh, C.-H., & Connelly, A. (2019). MRtrix3: A fast, flexible and open software framework for medical image processing and visualisation. *NeuroImage*, 202, 116137. https://doi.org/10.1016/j.neuroimage.2019.116137
- Van Der Walt, S., Colbert, S. C., & Varoquaux, G. (2011). The NumPy Array: A Structure for Efficient Numerical Computation. *Computing in Science & Engineering*, 13(2), 22–30. https://doi.org/10.1109/MCSE.2011.37
- Yeh, F.-C. (2025). DSI Studio: An integrated tractography platform and fiber data hub for accelerating brain research. *Nature Methods*. https://doi.org/10.1038/s41592-025-02762-8

