

Appendix

A1. Data Preparation

```
# R packages
library(ggplot2)
library(tidyverse)
library(Hmisc)
library(PMXForest)
library(patchwork)
library(kableExtra)
library(ggpubr)
library(scales)
library(ggsurvfit)
library(survival)
library(survminer)

# data preparation
data <- read.table("dataset.txt", sep = " ",      # space-separated
                  header = TRUE, na.strings = ".") # treat "." as NA

LLOQ <- 1 # mg/L

data <- data %>% mutate(DOSE = as.numeric(DOSE),
                      EVID = case_when(is.na(CONC) & DOSE!=0 ~ 1,
                                       CONC>=0 ~ 0,
                                       is.na(CONC) & is.na(DOSE) ~ 2),
                      BLQ = case_when(CONC<1 ~ 1,
                                       CONC>=1 ~ 0,
                                       is.na(CONC) ~ 0),
                      CONC_M7 = ifelse(CONC < 1, 0, CONC)) %>%

  group_by(ID) %>%
  fill(c(ALB, BW, CREAT),.direction = "downup") %>%
  arrange(TIME, .by_group = TRUE) %>%
  mutate(ADA = case_when(
    ID %in% 71:75 ~ as.integer(cumany(ADA == 1)),
    TRUE ~ ADA),
    COL = if_else(cummax(COL == 1L) == 1L, 1L, COL)) %>%
  ungroup()

data$DOSE = data$DOSE * data$BW # dose (mg) = dose (mg/kg) * body weight (kg)

write.csv(data, file = "dataset_clean.csv", quote = FALSE,
          na = ".", row.names = FALSE)
```

A2. Final Model

A2.1. Two-Compartment Target-Mediated Drug Disposition Model with Michaelis-Menten Approximation

```
$PROBLEM PMX001 Two Compartment TMDD Model
;; 2 assumptions:
;; 1. Free ligand concentration is much larger than total target conc.
;; 2. Vmax = cst => Rtot = cst => target degradation rate (kout)
;;    = ligand-target complex internalization rate (kint)

$INPUT DUMMY=DROP ID TIME WEEK DOSE=AMT RATE CONC=DROP ADA BW ALB SEX AGE
       CREAT COL EVID BLQ CONC_M7=DV

$DATA dataset_clean.csv IGNORE=@

$ABBR DERIV2=NO

$SUBROUTINES
ADVAN13
TOL=5

$MODEL
NCOMP=3 ; number of compartments
COMP=(DOSE, DEFDOSE, DEFOBS) ; first (central) compartment
COMP=(PERIPH) ; second (peripheral) compartment
COMP=(AUC) ; continuous AUC integration

$PK
IF (EVID.EQ.1) LASTDOSE = TIME
TAD = TIME - LASTDOSE

TVCL = THETA(1)
TVVMAX = THETA(2)
TVKM = THETA(3)
TVV1 = THETA(4)
TVV2 = THETA(5)
TVQ = THETA(6)

CLALB = ((ALB/31.85)**THETA(7))

CL = TVCL * EXP(ETA(1)) * CLALB
VMAX = TVVMAX
KM = TVKM
```

```

V1 = TVV1
V2 = TVV2
Q = TVQ

K10 = CL/V1
K12 = Q/V1
K21 = Q/V2

S1 = V1/1 ; scale prediction based on DOSE (mg) and DV (mg/L)
S2 = V2/1

$THETA ; values are determined in 3 iterations
(0.01,      0.944,      2.3) ; CL (L/d)
(0.5,       1.38,       3) ; VMAX
(0.001,     0.00758,    0.02) ; KM
(0.5,       4.42,       9) ; V1 (L)
(0.00001,   0.000307,  0.0006) ; V2
(0.1,       13,        30) ; Q
(-100,      -3.41223,   50) ; CLALB

$DES DADT(1) = -K10*A(1) -K12*A(1) +K21*A(2) -VMAX*A(1)/(KM*V1 + A(1))
      DADT(2) =          K12*A(1) -K21*A(2)
      cAUC = A(1)/V1
      DADT(3) = cAUC

$ERROR
IPRED = F
LLOQ = 1 ; (mg/L)

; --- Residual error SDs ---
PROP_SD = SIGMA(1)
ADD_SD = SIGMA(2)

; --- BLQ inflation (gentle, stable) ---
IF (BLQ.EQ.1) ADD_SD = ADD_SD + LLOQ

; --- Final residual error ---
W = SQRT(ADD_SD**2+PROP_SD**2)

IF (W.LE.0.000001) W=0.000001 ; Protective code

IRES=DV-IPRED
IWRES=IRES/W
Y = IPRED * (1 + EPS(1)) + EPS(2)

```

```

$OMEGA 0.366 ; IIV CL

$SIGMA      ; residual variability
0.327      ; EPS(1), proportional
1E-13 FIX  ; EPS(2); additive, required by M7+ censoring method

$EST
METHOD=1 INTERACTION; FOCE-I
MAXEVAL=9999
SIG=3
SIGL=3
PRINT=5

$COVARIANCE PRINT=E UNCONDITIONAL MATRIX=S

$TABLE ; output table for standard outcomes
ID TIME TAD DV EVID PRED IPRED WRES IWRES RES IRES CWRES CRES NOPRINT
ONEHEADER FILE=run16_sdtab

$TABLE ; output table for parameters and covariates
ID cAUC CL VMAX KM V1 V2 Q K10 K12 K21 ADA SEX COL BW ALB AGE CREAT NOPRINT
NOAPPEND ONEHEADER FILE=run16_pa_cov

```

A2.2. Two-Compartment Model with Linear Elimination

```
$PROBLEM PMX001 Two Compartment Linear Elimination

$INPUT DUMMY=DROP ID TIME WEEK DOSE=AMT RATE CONC=DROP ADA BW ALB SEX AGE
       CREAT COL EVID BLQ CONC_M7=DV

$DATA dataset_clean.csv IGNORE=@

$ABBR DERIV2=NO

$SUBROUTINES
ADVAN13
TOL=5

$MODEL
NCOMP=3
COMP=(DOSE, DEFDOSE, DEFOBS)
COMP=(PERIPH)
COMP=(AUC)

$PK
IF (EVID.EQ.1) LASTDOSE = TIME
TAD = TIME - LASTDOSE

TVCL = THETA(1)
TVV1 = THETA(2)
TVV2 = THETA(3)
TVQ = THETA(4)

IF(ADA.EQ.0) CLADA = 1 ; Most common
IF(ADA.EQ.1) CLADA = ( 1 + THETA(5))
V1ALB = ( 1 + THETA(6)*(ALB - 31.84))

CL = TVCL * EXP(ETA(1)) * CLADA
V1 = TVV1 * EXP(ETA(2)) * V1ALB
V2 = TVV2
Q = TVQ

K10 = CL/V1
K12 = Q/V1
K21 = Q/V2

S1 = V1/1 ; scale prediction based on DOSE (mg) and DV (mg/L)
S2 = V2/1
```

```

$THETA ; values are determined in 3 iterations
(0.001, 1.41, 3) ; CL (L/d)
(0.001, 4.73, 7) ; V1 (L)
(0.01, 0.231, 0.8) ; V2
(0.001, 2.15, 5) ; Q
(0.001, 5.78, 8) ; CLADA
(-0.140, -0.127, -0.001) ; V1ALB1

$DES DADT(1) = -K10*A(1) -K12*A(1) +K21*A(2) ; ODE central compartment
      DADT(2) = K12*A(1) -K21*A(2) ; ODE peripheral compartment
      cAUC = A(1)/V1 ; calculates cumulative AUC
      DADT(3) = cAUC ; continuous AUC integration

$ERROR
IPRED = F
LLOQ = 1 ; (mg/L)

; --- Residual error SDs ---
PROP_SD = SIGMA(1)
ADD_SD = SIGMA(2)

; --- BLQ inflation (gentle, stable) ---
IF (BLQ.EQ.1) ADD_SD = ADD_SD + LLOQ

; --- Final residual error ---
W = SQRT(ADD_SD**2+PROP_SD**2)

IF (W.LE.0.000001) W=0.000001 ; Protective code

IRES=DV-IPRED
IWRES=IRES/W
Y = IPRED * (1 + EPS(1)) + EPS(2)

$OMEGA
0.608 ; IIV CL
0.375 ; IIV V1

$SIGMA ; residual variability
0.289 ; EPS(1), proportional
1E-13 FIX ; EPS(2); additive, required by M7+ censoring method

$EST
METHOD=1 INTERACTION; FOCE-I
MAXEVAL=9999
SIG=3

```

```
PRINT=5
```

```
$COVARIANCE PRINT=E UNCONDITIONAL MATRIX=S
```

```
$TABLE ; output table for standard outcomes
```

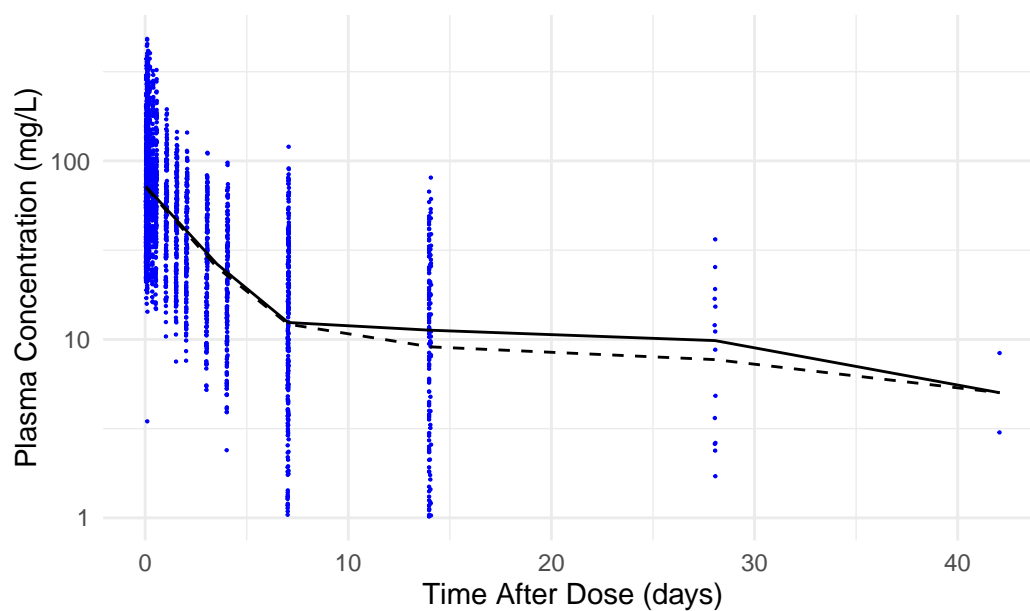
```
ID TIME TAD DV cAUC EVID PRED IPRED WRES IWRES RES IRES CWRES CRES NOPRINT  
ONEHEADER FILE=run12_sdtab
```

```
$TABLE ; output table for parameters and covariates
```

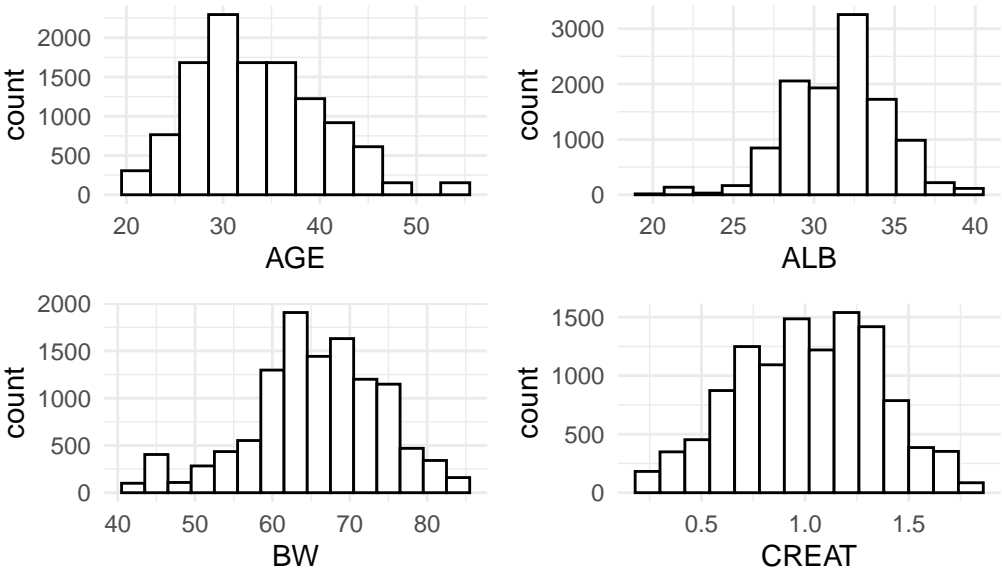
```
ID cAUC CL V1 V2 Q K10 K12 K21 ADA SEX COL BW ALB AGE CREAT NOPRINT NOAPPEND  
ONEHEADER FILE=run12_pa_cov
```

A3. Exploratory Data Analysis

Concentration–Time Profile of PMX001



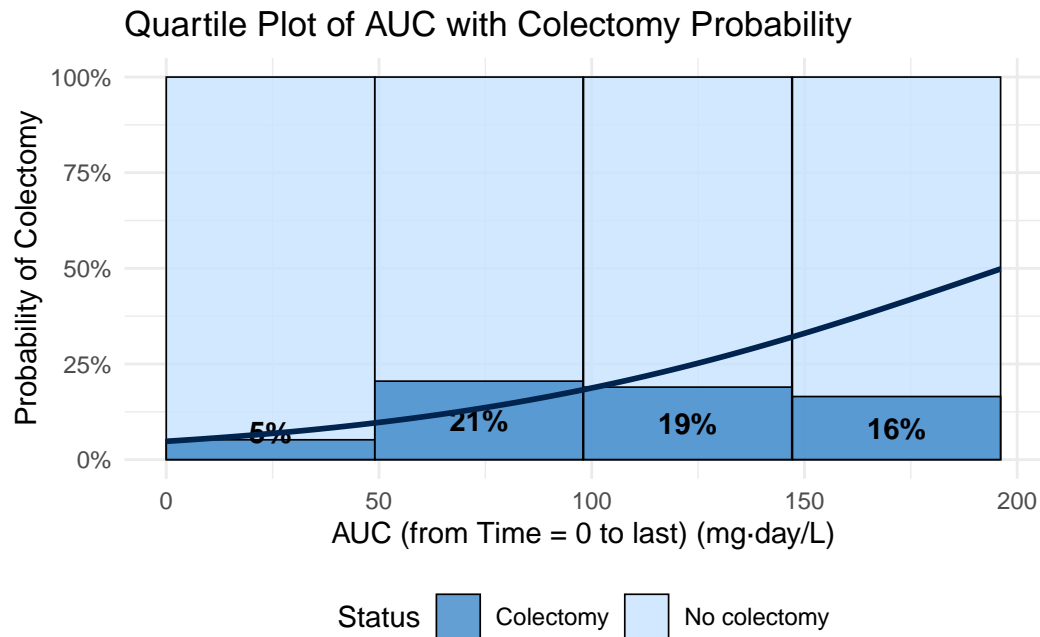
Histograms of Continuous Covariates



A4. Exposure-Response Quartile Quartile Plot

```
# A tibble: 2 x 6
```

COL	n	median_cAUC	IQR_cAUC	mean_cAUC	sd_cAUC
<fct>	<int>	<dbl>	<dbl>	<dbl>	<dbl>
1 No Colectomy	10539	1.37	32.2	22.9	38.2
2 Colectomy	936	48.4	61.3	56.5	41.7



```
library(dplyr)
library(ggplot2)
library(scales)
library(tidyr)

# --- define shared breaks (4 bins -> 5 break points) ---
brks <- seq(min(dat3$AUCO_tlast, na.rm = TRUE),
            max(dat3$AUCO_tlast, na.rm = TRUE),
            length.out = 5)

dat3 <- dat3 %>%
  mutate(
    # factor by the shared breaks (include lowest to avoid NA for min)
    quantile = cut(AUCO_tlast, breaks = brks, right = TRUE, include.lowest = TRUE)
  )

# --- summary per bin, with exact min/max taken from the shared breaks ---
summary_df <- dat3 %>%
  filter(!is.na(quantile)) %>%
  group_by(quantile) %>%
  summarise(
    AUCO_tlast_min = min(AUCO_tlast, na.rm = TRUE),
    AUCO_tlast_max = max(AUCO_tlast, na.rm = TRUE),
    prob_col = mean(COL == "Colectomy", na.rm = TRUE),
    .groups = "drop"
  ) %>%
```

```

# Replace the computed min/max with the exact interval bounds from `brks`
# so adjacent bins share boundaries perfectly.
# We can do this by mapping factor levels back to brks:
mutate(bin_id = as.integer(quantile)) %>%
mutate(
  AUCO_tlast_min = brks[bin_id],
  AUCO_tlast_max = brks[bin_id + 1],
  perc_label = percent(prob_col, accuracy = 1),
  x_mid = (AUCO_tlast_min + AUCO_tlast_max) / 2,
  y_label = pmax(prob_col * 0.5, 0.07)
)

rect_df <- bind_rows(
  summary_df %>%
    transmute(quantile, AUCO_tlast_min, AUCO_tlast_max, ymin = 0, ymax = 1,
              type = "No colectomy"),
  summary_df %>%
    transmute(quantile, AUCO_tlast_min, AUCO_tlast_max, ymin = 0, ymax = prob_col,
              type = "Colectomy")
)

# prediction curve (unchanged)
curve_df <- data.frame(
  AUCO_tlast = seq(min(dat3$AUCO_tlast, na.rm = TRUE),
                    max(dat3$AUCO_tlast, na.rm = TRUE),
                    length.out = 100)
)

logit_model <- glm(COL ~ AUCO_tlast, data = dat3, family = binomial)
curve_df$pred_prob <- predict(logit_model, newdata = curve_df, type = "response")

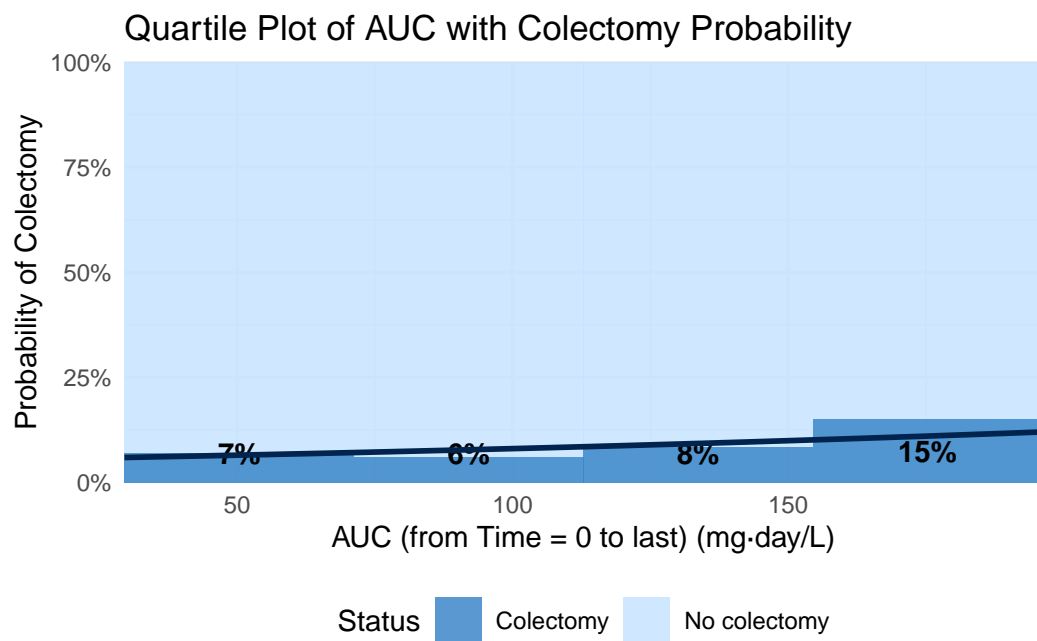
ggplot(rect_df) +
  geom_rect(aes(xmin = AUCO_tlast_min, xmax = AUCO_tlast_max,
                ymin = ymin, ymax = ymax, fill = type),
            # remove borders so adjacent bins visually connect
            color = NA, size = 0, alpha = 0.85) +
  geom_text(data = summary_df,
            aes(x = x_mid, y = y_label, label = perc_label),
            color = "black", fontface = "bold", size = 4) +
  geom_line(data = curve_df,
            aes(x = AUCO_tlast, y = pred_prob),
            color = "#00244D", size = 1) +
  scale_y_continuous(labels = scales::percent_format(), limits = c(0, 1),
                     expand = expansion(mult = 0)) + # no vertical padding (optional)
  scale_x_continuous(expand = expansion(mult = 0)) + # no horizontal padding -> bins fl
  scale_fill_manual(

```

```

  name = "Status",
  values = c("No colectomy" = "#C7E3FF", "Colectomy" = "#3C87C9")
) +
labs(
  x = "AUC (from Time = 0 to last) (mg·day/L)",
  y = "Probability of Colectomy",
  title = "Quartile Plot of AUC with Colectomy Probability"
) +
theme_minimal() +
theme(legend.position = "bottom")

```



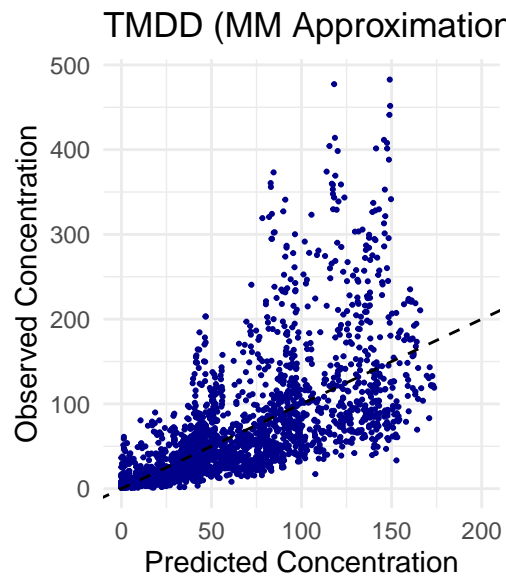
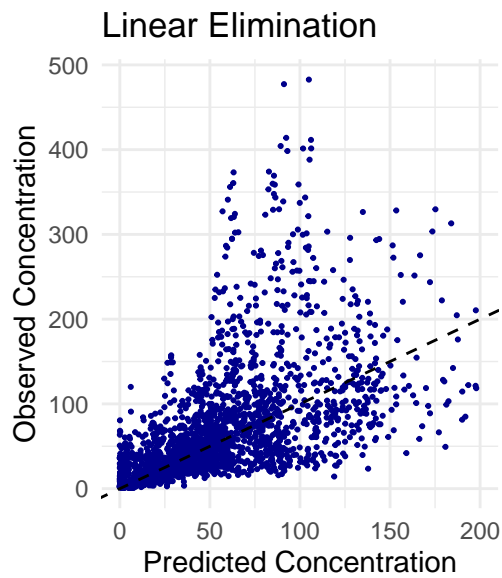
```

ggsave("QQcorr.png", width = 5, height = 4, units = "in")

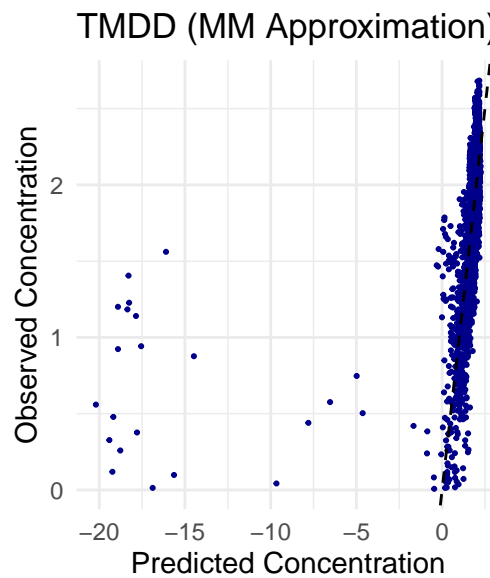
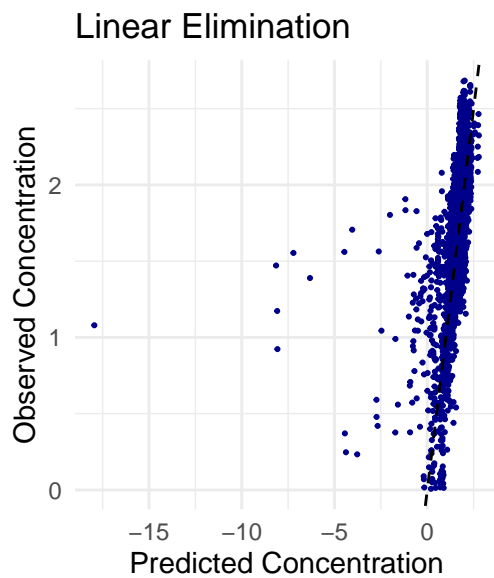
```

A5. Goodness of Fit Plots

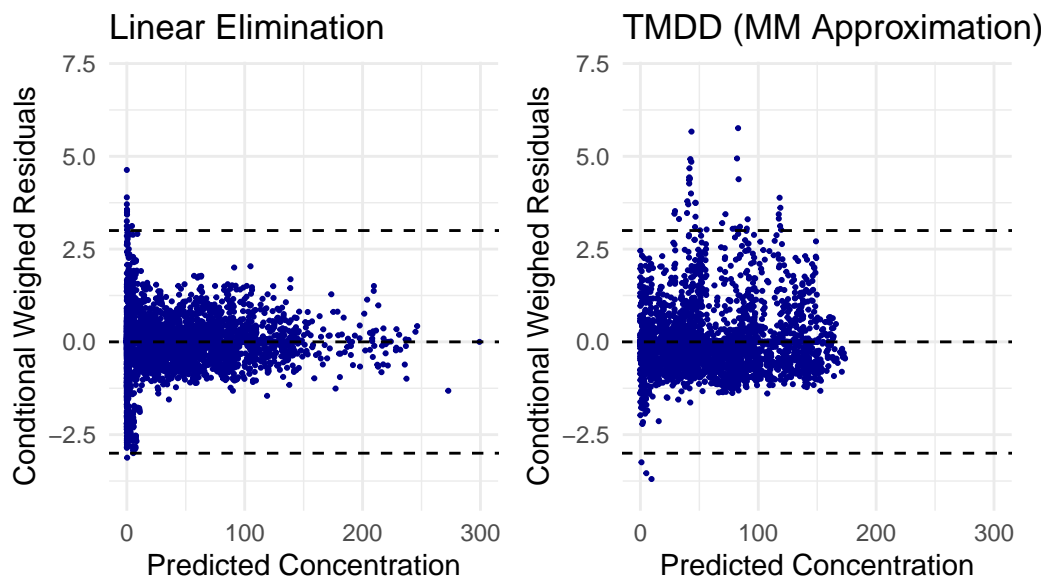
Population Predicted versus Observed Concentrations in Two-Compartment



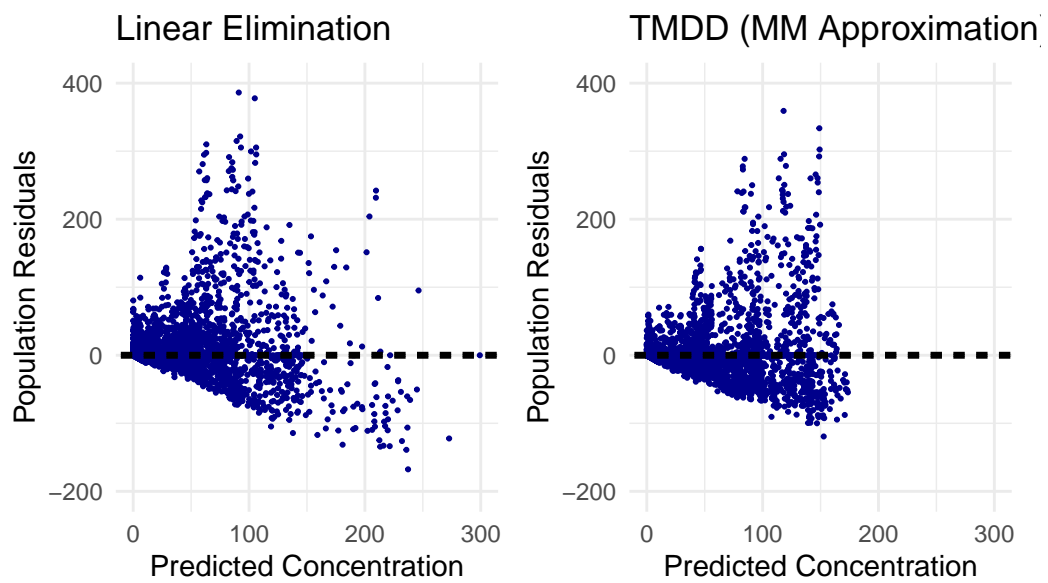
Log-Scaled Population Predicted versus Observed Concentrations



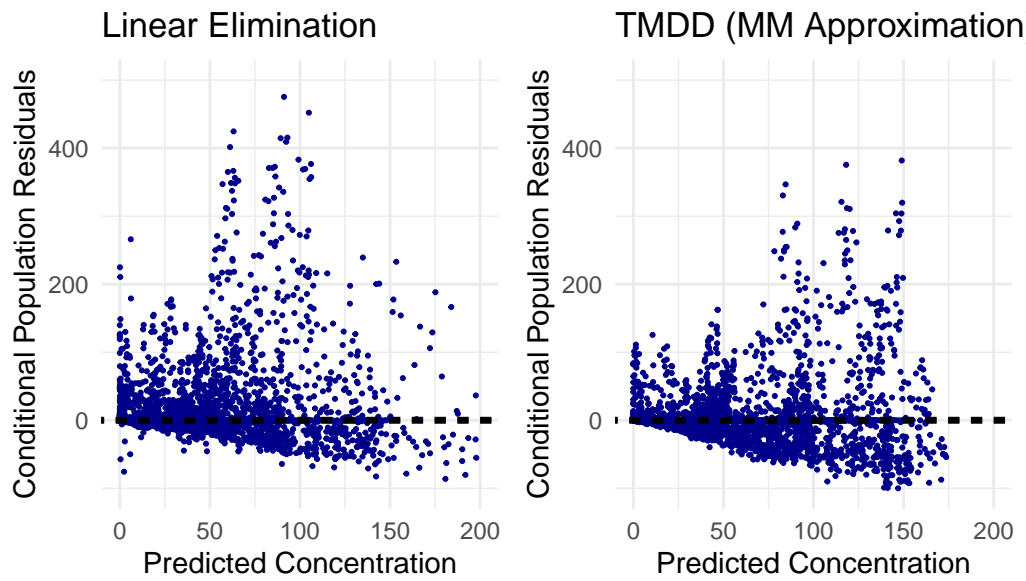
Conditional Weighed Residuals versus Population Predicted Conce



Population Residuals versus Population Predicted Concentrations ii



Conditional Population Residuals versus Population Predicted Conc



Conditional Weighed Residuals versus Time After Dose in Two-Col

