

# simExTargId (simultaneous experiment - MS/MS target identification)

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The following illustrates the `simExTargId` works with example (.RAW) data files of human plasma obtained on a Thermo FT-ICR-MS:

It shows how to set up the early warning email notification process first to monitor a dummy raw data directory on your computer's hard drive. Then half the example .RAW data files can be moved to the dummy raw data directory to initiate the **simExTargId** autonomous metabolomic data-analysis process. Then another set of example .RAW data files can then be moved in to the raw data directory to simulate a real-time collection of LC-MS data on a mass spectrometer workstation.

## 1. Set the real-time email notifier

The email notifier must be run as a separate monitoring function in another R session. This is so that the raw data directory on the mass spectrometer workstation can be closely monitored and a stoppage in the run or any files which are much smaller in size (greater than 3 \* median absolute deviation) than the majority of the files can be immediately identified and a warning email sent. If this function was run from within the main **simExTargId** function there might be a delay in informing the user of an unexpected instrument stoppage or spray quality issues for example when file-conversion, peak-picking or another lengthy process is taking place.

```
# select directory where example directories for raw files can be created and also results
# output saved
# For example the C: directory
# create a dummy raw MS1 profiling data directory
# for example
studyName <- paste0(gsub(" ", "", format(Sys.time(), "%Y %m %d")), "_simExTargId_Example")
dummyRawDir <- paste0("C:/", studyName)
# create directory
dir.create(dummyRawDir)
# replace email address with a vector of one or more email addresses of
# people in your laboratory group
emailNotifier(rawDir=dummyRawDir, emailAddress='johndoe@emailprovider.com',
              emailTime=5)
```

## 2. Set the main simExTargId function running

Set the main **simExTargId** function running in a different R session to the **emailNotifier** function. Make sure that you have the **MSConvert** software in your path if using Windows and that you are able to successfully run the command `> MSConvert` in your system shell. This is necessary for the automatic mzXML/mzML file conversion process and **simExTargId** will not work successfully without it.

```
# simExTargId extdata directory
extdataDir <- system.file("extdata", package="simExTargId")
# list blank files
```

```

blanksRaw <- list.files(extdataDir, pattern="blank", full.names=TRUE)
# list plasma IPA extract files
samplesRaw <- list.files(extdataDir, pattern="sample", full.names=TRUE)
# covariates file
coVariates <- paste(extdataDir, "coVariates.csv", sep="/")
# illustrative metabolite database table
metabDb <- paste(extdataDir, 'exampleMetabDatabase.csv', sep='/')
# identify number of virtual cores for parallel processing using parallel package
nCores <- parallel::detectCores()

# as no qc files then use sample files twice to illustrate the
# peakMonitor function

# move the blank files into the temporary directory to start the process
blankRawCopies <- paste(dummyRawDir, basename(blanksRaw), sep="/")
file.copy(from=blanksRaw, to=blankRawCopies)
# set the file time to simulate the files having been acquired at least 5 mins
# since last modification
setTheTime <- function(fileCopy, time){
  Sys.setFileTime(fileCopy, Sys.time() - time)
}
# apply to newly copied files
sapply(blankRawCopies, setTheTime, 240)

# move the plasma samples twice first time rename as QC
# and set the file time less than 5 mins
samplesRawCopies <- paste(dummyRawDir, basename(samplesRaw), sep="/")
file.copy(from=samplesRaw, to=samplesRawCopies)
qcFiles <- gsub('sample', 'qc', samplesRawCopies)
file.rename(from=samplesRawCopies, to=qcFiles)
file.copy(from=samplesRaw, to=samplesRawCopies)

# apply to newly copied files
sapply(c(samplesRawCopies, qcFiles), setTheTime, 300)

# Start simExTargId function
simExTargId(rawDir=dummyRawDir, studyName = studyName, analysisDir='C:/',
            coVar=coVariates, metab=metabDb, nCores=nCores, ionMode='nega',
            minFiles=3)

```

simExTargId will wait until at least five minutes after the raw data file was last modified, to ensure that the file acquisition has completed.

After at least 3 samples of each class found in the second column of the co-variates table, retention time alignment, grouping, zero-filling, then pre-processing, PCA analysis, stats analysis and data-deconvolution will occur.

### 3. peakMonitor (shiny application)

A database table of previously identified metabolites can be monitored in real-time and analytical CV% and signal attenuation affects monitored using the shiny application **peakMonitor**

```
peakMonitor(analysisDir=paste0(dummyRawDir, "_analysis/NEG/output/peakMonitor"))
```

#### 4. targetId (shiny application)

During a run the output of the statistical analyses can be viewed using the shiny application **targetId** a zip file containing a .csv file for each statistical test after setting the thresholds can be downloaded and used to guide and plan further MS/MS experiments.

*# this command will open the application in your web-browser*

```
targetId(analysisDir=paste0(dummyRawDir, "_analysis/NEG/output/04.stats"))
```