# Surf 64 - XP practice Biological Network Inference with Sparse Graphical Models

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Anglet, 27 June 2018

https://github.com/jchiquet/JC2BIM18





# Outline

# Statistical analysis of Networks

Different questions

# Understanding the network topology

- Data = observed network
- Questions: central nodes? cluster structure? small-world property?

# Inferring/Reconstructing the network

- Data = repeated signal observed at each node
- Questions: which nodes are connected?

### Using the network

- Data = a given network + signal on nodes
- Questions: how the epidemic spreads along the network?

### Each to be combined with

covariates, time, heterogeneous data set, missing data, ...

# Automatic reconstruction of biological networks

E. coli regulatory network

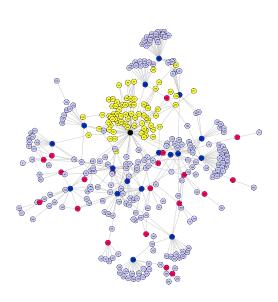
# Target network

Relations between genes and their products

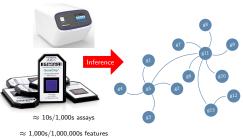
- highly structured
- always incomplete

### Data and method

- transcriptomic data
- Gaussian graphical model with sparse methods



# A challenging problem



# Model point of view

- 1 Nodes (genes, OTUS, ...)
  - fixed variables
- 2 Edges (biological interactions)
  - use (partial) correlations or others fancy statistical concepts
- Oata (intensities, counts)
  - ullet a tidy n imes p dat matrix
- → Quantities and goals well defined

Data point of view: non classical statistics

- (Ultra) High dimensionality  $(n < p, n \ll p)$
- Heterogeneous data

Biological point of view: not well defined goals and questions

- What interaction? Direct? Indirect? Causal?
- Whole network? Subnetwork? Groups of key actors?

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- Network and data modeling Statistical dependence Gaussian Graphical models
- 2 Network inference with sparse GGM
- 3 A tour of the huge package assessing GGM approach
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# Canonical model settings

Biological microarrays in comparable conditions

### **Notations**

- **1** a set  $\mathcal{P} = \{1, \dots, p\}$  of p variables: these are typically the genes (could be proteins);
- ② a sample  $\mathcal{N}=\{1,\dots,n\}$  of individuals associated to the variables: these are typically the microarray (could be sequence counts).

### Basic statistical mode

### This can be view as

- a random vector X in  $\mathbb{R}^p$ , whose jth entry is the jth variable,
- a n-size sample  $(X^1, \ldots, X^n)$ , such as  $X^i$  is the ith microarrays,
  - could be independent identically distributed copies (steady-state
    - could be dependent in a certain way (time-course data)
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# Canonical model settings

Biological microarrays in comparable conditions

### Notations



Stacking  $(X^1,\ldots,X^n)$ , we met the usual individual/variable table  ${\bf X}$ 



$$\mathbf{X} = \begin{pmatrix} x_1^1 & x_1^2 & x_1^3 & \dots & x_1^p \\ \vdots & & & & \\ x_n^1 & x_n^2 & x_1^2 & \dots & x_n^p \end{pmatrix}$$

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sparse Gaussian Graphical Models

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Independence

Definition (Independence of events)

Two events  $\boldsymbol{A}$  and  $\boldsymbol{B}$  are independent if and only if

$$\mathbb{P}(A,B) = \mathbb{P}(A)\mathbb{P}(B),$$

which is usually denoted by  $A \perp \!\!\! \perp B$ . Equivalently,

- $A \perp \!\!\!\perp B \Leftrightarrow \mathbb{P}(A|B) = \mathbb{P}(A)$ ,
- $\bullet \ A \perp \!\!\! \perp B \Leftrightarrow \mathbb{P}(A|B) = \mathbb{P}(A|B^c)$

Example (class vs party)

Table: Joint probability (left) vs. conditional probability (right)

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Example (class vs party)

|             | part   |      | party       |        |      |
|-------------|--------|------|-------------|--------|------|
| class       | Labour | Tory | class       | Labour | Tory |
| working     | 0.42   | 0.28 | working     | 0.60   | 0.40 |
| bourgeoisie | 0.06   | 0.24 | bourgeoisie | 0.20   | 0.80 |

Table: Joint probability (left) vs. conditional probability (right)

Conditional independence

# Generalizing to more than two events requires strong assumptions (mutual independence). Better handle with

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Consider the events A = "having low QI", B = "having low weight"

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Estimating  $\mathbb{P}(A,B)$ ,  $\mathbb{P}(A)$  and  $\mathbb{P}(B)$  in a sample would lead to

$$\mathbb{P}(A,B) \neq \mathbb{P}(A)\mathbb{P}(B)$$

<sup>&</sup>lt;sup>1</sup>stupidly

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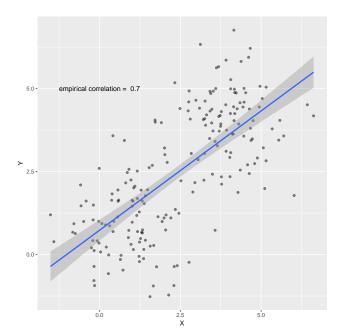
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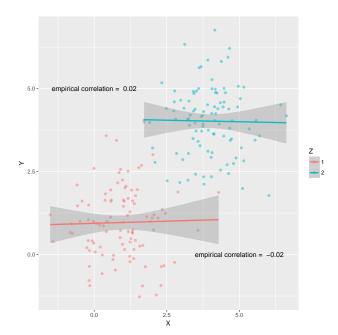
Consider the events A= "having low QI", B= "having low weight". But in fact, introducing C= "having a given age",

$$\mathbb{P}(A, B|C) = \mathbb{P}(A|C)\mathbb{P}(B|C)$$

# Limits of correlation for network reconstruction



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# Correlation networks

# Correlation (association network)

Similar expression profile → high-correlation

- Compute the correlation matrix (Pearson, Spearman, ...)
- Predict an edge between two actors if their absolute correlation is above a given threshold

### Questions

- How to set up the threshold?
- If we target actors with similar profiles, why not clustering?
- Information is drowned (all actors are correlated ...)

# Graphical models

### Definition

A graphical model gives a graphical (intuitive) representation of the dependence structure of a probability distribution, by linking

- **1** a random vector (or a set of random variables.)  $X = \{X_1, \dots, X_p\}$  with distribution  $\mathbb{P}$ ,
- 2 a graph  $\mathcal{G}=(\mathcal{P},\mathcal{E})$  where
  - $\mathcal{P} = \{1, \dots, p\}$  is the set of nodes associated to each variable,
  - $\mathcal E$  is a set of edges describing the dependence relationship of  $X \sim \mathbb P$ .

### Conditional independence graph

It is the undirected graph  $\mathcal{G} = \{\mathcal{P}, \mathcal{E}\}$  where

$$(i,j) \notin \mathcal{E} \Leftrightarrow X_i \perp \!\!\! \perp X_j | \mathcal{P} \setminus \{i,j\}$$

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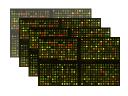
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# The Gaussian case

### The data



$$\mathbf{X} = \begin{pmatrix} x_1^1 & x_1^2 & x_1^3 & \dots & x_1^p \\ \vdots & & & & \\ x_n^1 & x_n^2 & x_1^2 & \dots & x_n^p \end{pmatrix}$$

# Assuming $f_X(\mathbf{X})$ multivariate Gaussian

### Greatly simplifies the inference:

- naturally links independence and conditional independence to the covariance and partial covariance,
- gives a straightforward interpretation to the graphical modeling previously considered.

# Why Gaussianity helps?

Case of 2 variables or size-2 random vector

Let X, Y be two real random variables.

### **Definitions**

$$\operatorname{cov}(X,Y) = \mathbb{E}\Big[\big(X - \mathbb{E}(X)\big)\big(Y - \mathbb{E}(Y)\big)\Big] = \mathbb{E}(XY) - \mathbb{E}(X)\mathbb{E}(Y).$$

$$\rho_{XY} = \operatorname{cor}(X,Y) = \frac{\operatorname{cov}(X,Y)}{\sqrt{\mathbb{V}(X)} \cdot \mathbb{V}(Y)}.$$

# Proposition

- $\operatorname{cov}(X, X) = \mathbb{V}(X) = \mathbb{E}[(X \mathbb{E}X)(Y \mathbb{E}Y)],$
- $\operatorname{cov}(X + Y, Z) = \operatorname{cov}(X, Z) + \operatorname{cov}(X, Z)$ ,
- $\mathbb{V}(X+Y) = \mathbb{V}(X) + \mathbb{V}(Y) + \operatorname{cov}(X,Y)$ .
- $X \perp Y \Rightarrow cov(X, Y) = 0.$
- $X \perp \!\!\! \perp Y \Leftrightarrow \operatorname{cov}(X,Y) = 0$  when X,Y are Gaussian

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# The bivariate Gaussian distribution

### The Covariance Matrix

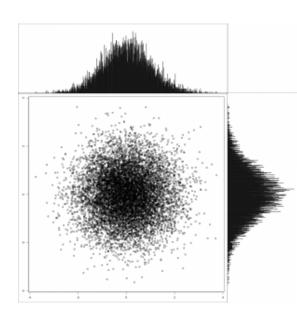
Let

$$X \sim \mathcal{N}(\mathbf{0}, \mathbf{\Sigma}),$$

with unit variance and  $\rho_{XY}=0$ 

$$\Sigma = \begin{pmatrix} 1 & 0 \\ 0 & 1 \end{pmatrix}.$$

The shape of the 2-D distribution evolves accordingly.



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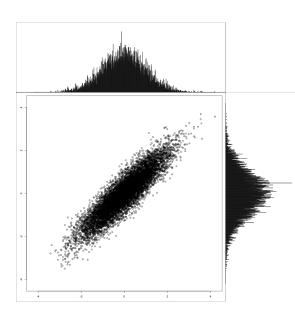
Let

$$X \sim \mathcal{N}(\mathbf{0}, \mathbf{\Sigma}),$$

with unit variance and  $\rho_{XY}=0.9$ 

$$\Sigma = \begin{pmatrix} 1 & 0.9 \\ 0.9 & 1 \end{pmatrix}.$$

The shape of the 2-D distribution evolves accordingly.



# Generalization: multivariate Gaussian vector

Now need partial covariance and partial correlation

Let X, Y, Z be real random variables.

**Definitions** 

$$cov(X, Y|Z) = cov(X, Y) - cov(X, Z)cov(Y, Z)/\mathbb{V}(Z).$$

$$\rho_{XY|Z} = \frac{\rho_{XY} - \rho_{XZ}\rho_{YZ}}{\sqrt{1 - \rho_{XZ}^2}\sqrt{1 - \rho_{YZ}^2}}.$$

 $\rightsquigarrow$  Give the interaction between X and Y once removed the effect of Z.

Proposition

When X, Y, Z are jointly Gaussian, then

$$\mathrm{cov}(X,Y|Z) = 0 \Leftrightarrow \mathrm{cor}(X,Y|Z) = 0 \Leftrightarrow X \perp\!\!\!\perp Y|Z$$

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# Important properties of Gaussian vectors

Proposition (Gaussian vector and conditioning)

Consider a Gaussian vector with the following decomposition

$$Z = \begin{pmatrix} Z_1 \\ Z_2 \end{pmatrix} \sim \mathcal{N}(\mathbf{0}, \mathbf{\Sigma}), \quad \mathbf{\Sigma} = \begin{pmatrix} \mathbf{\Sigma}_{11} & \mathbf{\Sigma}_{12} \\ \mathbf{\Sigma}_{21} & \mathbf{\Sigma}_{22} \end{pmatrix}, \quad \mathbf{\Omega} = \mathbf{\Sigma}^{-1} = \begin{pmatrix} \mathbf{\Omega}_{11} & \mathbf{\Omega}_{12} \\ \mathbf{\Omega}_{21} & \mathbf{\Omega}_{22} \end{pmatrix}.$$

Then,

$$Z_2|Z_1=\mathbf{z}\sim\mathcal{N}\left(-\mathbf{\Omega}_{22}^{-1}\mathbf{\Omega}_{21}\mathbf{z},\mathbf{\Omega}_{22}^{-1}
ight)$$

and

$$\Omega_{22}^{-1} = \Sigma_{22} - \Sigma_{21} \Sigma_{11}^{-1} \Sigma_{12}.$$

# Corollary

Partial correlations are related to the inverse of the covariance matrix:

$$cor(Z_i, Z_j | Z_k, k \neq i, j) = -\frac{\Omega_{ij}}{\sqrt{\Omega_{ii}\Omega_{jj}}}$$

# Gaussian Graphical Model: canonical settings

Biological experiments in comparable Gaussian conditions

Profiles of a set  $\mathcal{P} = \{1, \dots, p\}$  of genes is described by  $X \in \mathbb{R}^p$  such as

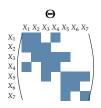
- **1**  $X \sim \mathcal{N}(\boldsymbol{\mu}, \boldsymbol{\Sigma})$ , with  $\boldsymbol{\Theta} = \boldsymbol{\Sigma}^{-1}$  the precision matrix.
- 2 a sample  $(X^1,\ldots,X^n)$  of exp. stacked in an  $n\times p$  data matrix  ${\bf X}$ .

# Conditional independence structure

$$(i,j) \notin \mathcal{E} \Leftrightarrow X_i \perp X_j | X_{\setminus \{i,j\}} \Leftrightarrow \Theta_{ij} = 0.$$

# Graphical interpretation

$$\mathcal{G} = (\mathcal{P}, \mathcal{E})$$
 $X_1$ 
 $X_3$ 
 $X_5$ 
 $X_6$ 
 $X_7$ 



# Gaussian Graphical Model and Linear Regression

### Linear regression viewpoint

Gene expression  $X_i$  is linearly explained by the other genes':

$$X_i | X_{\setminus i} = -\sum_{j \neq i} \frac{\Theta_{ij}}{\Theta_{ii}} X_j + \varepsilon_i, \quad \varepsilon_i \sim \mathcal{N}(0, \Omega_{ii}^{-1}), \quad \varepsilon_i \perp X$$

Conditional on its neighborhood, other profiles do not give additional insights

$$X_i|X_{\backslash i} = \sum_{j \in \mathsf{neighbors}(i)} \beta_j X_j + \varepsilon_i \quad \mathsf{with} \ \beta_j = -\frac{\Theta_{ij}}{\Theta_{ii}}.$$

→ "Neighborhood" selection

# Gaussian Graphical Model and AR process (1)

### Time course data

Time course- data experiment can be represented as a multivariate vector  $X=(X_1,\ldots,X_p)\in\mathbb{R}^p$ , generated through a first order vector autoregressive process VAR(1):

$$X^t = \mathbf{\Theta}X^{t-1} + \mathbf{b} + \boldsymbol{\varepsilon}^t, \quad t \in [1, n]$$

where  $\varepsilon^t$  is a white noise to ensure the Markov property and  $X^0 \sim \mathcal{N}(0, \Sigma^0)$ .

Consequence: a Gaussian Graphical Model

- Each  $X^t | X^{t-1} \sim \mathcal{N}(\theta X^{t-1}, \Sigma)$
- or, equivalently,  $X_i^t|X^{t-1} \sim \mathcal{N}(\Theta_i X^{t-1}, \Sigma)$

where  $\Sigma$  is known and  $\Theta_j$  is the jth row of  $\Theta$ .

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# Gaussian Graphical Model and AR process (2)

Interpretation as a GGM

The VAR(1) as a covariance selection model

$$\theta_{ij} = \frac{\operatorname{cov}\left(X_i^t, X_j^{t-1} | X_{\mathcal{P} \setminus j}^{t-1}\right)}{\operatorname{var}\left(X_j^{t-1} | X_{\mathcal{P} \setminus j}^{t-1}\right)},$$

#### Graphical Interpretation

 $\leadsto$  The matrix  $\Theta = (\theta_{ij})_{i,j \in \mathcal{P}}$  encodes the network  $\mathcal{G}$  we are looking for.



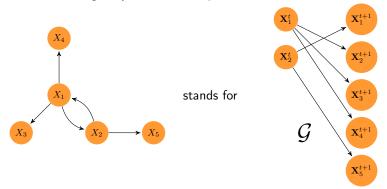


conditional dependency between  $X_j^{t-1}$  and  $X_i^t$  or non-null partial correlation between  $X_j^{t-1}$  and  $X_i^t$   $\theta_{ij} \neq 0$ 

# Gaussian Graphical Model and AR process (3)

#### Graphical interpretation

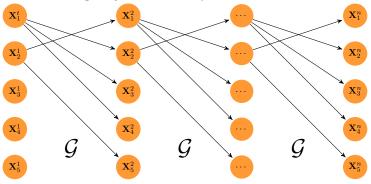
- Follow-up of one single experiment/individual;
- ② Close enough time-points to ensure
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# Gaussian Graphical Model and AR process (3)

#### Graphical interpretation

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## Some families of methods for network reconstruction

#### Test-based methods

- Tests the nullity of each entries
- Combinatorial problem when  $p > 30 \dots$

#### Sparsity-inducing regularization methods

- induce sparsity with the  $\ell_1$ -norm penalization
- Use results from convex optimization
- Versatile and computationally efficient

#### Bayesian methods

- Compute the posterior probability of each edge
- Usually more computationally demanding
- For special graphs, computation gets easier

## Inference: maximum likelihood estimator

The natural approach for parametric statistics

Let X be a random vector with distribution defined by  $f_X(x; \Theta)$ , where  $\Theta$  are the model parameters.

Maximum likelihood estimator

$$\hat{\boldsymbol{\Theta}} = \arg\max_{\boldsymbol{\Theta}} \ell(\boldsymbol{\Theta}; \mathbf{X})$$

where  $\ell$  is the log likelihood, a function of the parameters:

$$\ell(\boldsymbol{\Theta}; \mathbf{X}) = \log \prod_{i=1}^{n} f_X(\mathbf{x}_i; \boldsymbol{\Theta}),$$

where  $\mathbf{x}_i$  is the *i*th row of  $\mathbf{X}$ .

#### Remarks

- This a convex optimization problem,
- ullet We just need to detect non zero coefficients in  $oldsymbol{\Theta}$

# The multivariate Gaussian log-likelihood

Let  $\mathbf{S} = n^{-1}\mathbf{X}^{\mathsf{T}}\mathbf{X}$  be the empirical variance-covariance matrix:  $\mathbf{S}$  is a sufficient statistic of  $\mathbf{\Theta}$ .

#### The log-likelihood

$$\ell(\mathbf{\Theta}; \mathbf{S}) = \frac{n}{2} \log \det(\mathbf{\Theta}) - \frac{n}{2} \operatorname{Trace}(\mathbf{S}\mathbf{\Theta}) + \frac{n}{2} \log(2\pi).$$

- $\leadsto$  The MLE =  $\mathbf{S}^{-1}$  of  $\mathbf{\Theta}$  is not defined for n < p and never sparse.
- The need for regularization is huge.

# Application to GGM: the "Graphical-Lasso"

#### A penalized likelihood approach

$$\hat{\boldsymbol{\Theta}}_{\lambda} = \operatorname*{arg\ max}_{\boldsymbol{\Theta} \in \mathbb{S}_{+}} \ell(\boldsymbol{\Theta}; \mathbf{X}) - \lambda \|\boldsymbol{\Theta}\|_{\ell_{1}}$$

#### where

- ℓ is the model log-likelihood,
- $\|\cdot\|_{\ell_1}$  is a penalty function tuned by  $\lambda > 0$ .
  - 1 regularization (needed when  $n \ll p$ ),
  - 2 selection (sparsity induced by the  $\ell_1$ -norm),
- solved in R-packages glasso, quic, huge  $(\mathcal{O}(p^3))$

# Application to GGM: "Neighborhood selection"

A close cousin, thank to the relationship between Gaussian vector and linear regression

Remember that

$$X_i|X_{\backslash i} = \sum_{j \in \mathsf{neighbors}(i)} \beta_j X_j + \varepsilon_i \quad \mathsf{with} \ \beta_j = -\frac{\Theta_{ij}}{\Theta_{ii}}.$$

#### A penalized least-square approach

Let  $\mathbf{X}_i$  be the ith column of the data matrix (i.e data associated to variable (gene) i), and  $\mathbf{X}_{\backslash i}$  deprived of colmun i. We select the neighbors of variable i by solving

$$\widehat{\boldsymbol{\beta}}^{(i)} = \operatorname*{arg\ min}_{\boldsymbol{\beta} \in \mathbb{R}^{p-1}} \frac{1}{n} \left\| \mathbf{X}_i - \mathbf{X}_{\setminus i} \, \boldsymbol{\beta} \right\|_2^2 + \lambda \left\| \boldsymbol{\beta} \right\|_1$$

- not symmetric, not positive-definite
- + p Lasso solved with Lars-like algorithms ( $\mathcal{O}(npd)$  for d neighbors).

#### Model selection

#### Cross-validation

Optimal in terms of prediction, not in terms of selection

#### Information based criteria

- GGMSelect (Girault et al, '12) selects among a family of candidates.
- Adapt IC to sparse high dimensional problems, e.g.

$$\mathsf{EBIC}_{\gamma}(\widehat{\boldsymbol{\Theta}}_{\lambda}) = -2\mathsf{loglik}(\widehat{\boldsymbol{\Theta}}_{\lambda}; \mathbf{X}) + |\mathcal{E}_{\lambda}|(\mathsf{log}(n) + 4\gamma \, \mathsf{log}(p)),$$

#### Resampling/subsampling

Keep edges frequently selected on an range of  $\lambda$  after sub-samplings

- Stability Selection (Meinshausen and Bühlman, 2010, Bach 2008)
- Stability approach to Regularization Selection (StaRS) (Liu, 2010).

# Concluding remark about GGM

#### Sparse GGM

- + very solid statistical and computational framework
- + competitive to other inference methods (DREAM 5 benchmark, 2012)
- performances remain questionable on real data, as for other methods
- → Network inference is a very difficult problem
- Some biological questions can be answered without network inference

## Outline

- Network and data modeling Statistical dependence Gaussian Graphical models
- 2 Network inference with sparse GGM
- 3 A tour of the huge package assessing GGM approach
- 4 Basic network analysis of transcriptomics exposome data set

## Assess the standard GGMs approaches

Full analysis can be found at <a href="http://julien.cremeriefamily.info/exposome.html">http://julien.cremeriefamily.info/exposome.html</a>

```
suppressMessages(library(huge, quietly = TRUE))
```

- Simulated data
  - Test that an approach is working under some simple conditions
  - · Especially usefull when the approach has no underlying model
  - Essential sanity check
- ② Breast cancer data (pinpoint interesting genes/pathways)
  - Several hundred breast cancers (estrogen receptor + and -)
  - Several thousand genes
  - Goal: How can GGMs approaches help?

# Simple simulations (network with hubs)

```
set.seed(11)
n <- 80; d <- 10;
rd.net <- huge.generator(
   n, ## number of samples
   d, ## number of genes
graph="hub", ## type of net
g = 2, ## number of group)
verbose=FALSE)</pre>
```

# Simple simulations (network with hubs)

plot(rd.net)









## Inference using GGMs and correlation

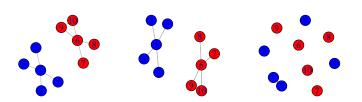
#### Inference

#### Selection

```
## glasso, mb and ct
glasso.sel <- huge.select(glasso, "stars", verbose=F)
mb.sel <- huge.select(mb, "stars", verbose=F)
corthr.sel <- huge.select(corthr, "stars", verbose=F)</pre>
```

# Inference using GGMs and correlation (results)

```
gr.glasso <- graph.adjacency(glasso.sel$refit)
V(gr.glasso)$label.cex <- 2
V(gr.glasso)$color <- rep(c("blue", "red"), each=5)
par(mfrow=c(1, 3))
plot(gr.glasso, vertex.size=30, edge.arrow.mode = "-")
plot(gr.mb, vertex.size=30, edge.arrow.mode = "-")
plot(gr.cor, vertex.size=30, edge.arrow.mode = "-")</pre>
```



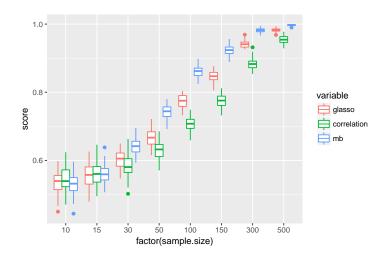
## A bit of code to run a simulation

```
suppressMessages(require(reshape2))
one.simu <- function(i) {
 lbd.c \leftarrow seq(1, 0, -10^{-2});
  d <- 25; seg.n <- c(10, 15, 30, 50, 100, 150, 300, 500)
  out <- data.frame(t(sapply(seq.n, function(n) {</pre>
   exp <- huge.generator(n, d, graph="cluster",
                         g=3, prob=1, verbose=F)
  gl <- huge(exp$data, method="glasso", nlambda=50, verbose=F)
  mb <- huge(exp$data, method="mb", nlambda=50, verbose=F)
   cthr <- huge(exp$data, method="ct", lambda=lbd.c, verbose=F)
  res.cthr <- perf.auc(perf.roc(cthr$path, exp$theta))
  res.gl <- perf.auc(perf.roc(gl$path, exp$theta))
  res.mb <- perf.auc(perf.roc(mb$path, exp$theta))
  return(setNames(c(res.gl,res.cthr,res.mb,n,i),
   c("glasso", "correlation", "mb", "sample size", "simu")))
  })))
return(melt(out, measure.vars = 1:3, value.name = "score"))}
```

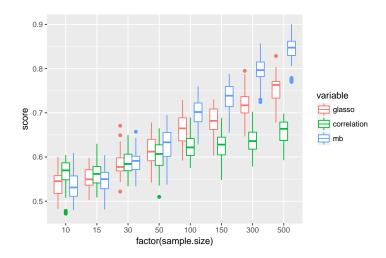
## Run

```
suppressMessages(library(parallel))
res <- do.call(rbind, mclapply(1:40, one.simu, mc.cores=4))</pre>
```

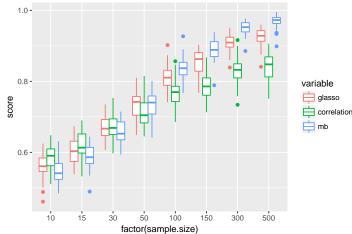
# Simulation results (cluster - clique)



# Simulation results (cluster, connection probability of 0.5)



# Simulation results (random, connection probability of 0.3)



correlation

## Breast cancer: transcriptomics for ER+ and ER- tumors

We look at a large public datasets from Guedj et al. 2011 with two main subgroups

- Estrogen receptor positive
- Estrogen receptor negative

```
load ("huge/breast_cancer_guedj11.RData")
load ("huge/gen_name.RData")
gene.name <- unlist(gene.name)
data.raw <- expr
table(class.ER)

## class.ER
## ERm ERp
## 162 375</pre>
```

## Filtering Unknown genes

```
toDiscard <- which(gene.name == "Not.Known")
gene.name <- gene.name[-toDiscard]
data.raw <- data.raw[-toDiscard, ]</pre>
```

#### We get

```
dim(data.raw)
## [1] 41248 537
```

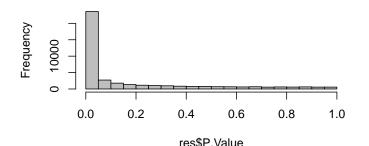
## Differential analysis

#### Do we detect some gene expression differences ?

## Many genes are differentially expressed

- The histogram of p-values looks good
- This is a well known fact (ER+ and ER- are very different)

#### P-values ER- vs ER+



## What to do with this list of genes?

#### ESR1 has the most significant p-values

```
gene.name[order(res$adj.P.Val)[1]]
## 205225_at
## "ESR1"
```

#### Network analysis

- Could we find partners of ESR1 that are specific to ER+?
- We cannot infer a network on 41000 genes (Verzelen 2011)
  - → Most differentially expressed genes
  - → Most varying genes
  - → Look at a specific pathway . . .

## Selecting some probes

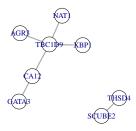
#### Take the 20 most differentially expressed plus some random

```
## Error in graph.from.adjacency_matrix(net_Mspec_): could not find function
"graph.from_adjacency_matrix"
## Error in graph.from_adjacency_matrix(net_Pspec_): could not find function
"graph.from_adjacency_matrix"
```

# MAGDEN 4 NLPPI THISD DIVALII (GR) (ATA)3 (MB). FEXALII TECHNOLOGY TECHNOLOGY

ER+ specific

#### ER-specific



## FOXA1, ESR1, GATA3 a well known interaction

- FOXA1 is a key determinant of estrogen receptor function and endocrine response. Antoni Hurtado et al. 2011 (Nat. Genet.):
  - $\leadsto$  "FOXA1 is a key determinant that can influence differential interactions between ER and chromatin"
- Q GATA3 acts upstream of FOXA1 in mediating ESR1 binding by shaping enhancer accessibility. Theodorou et al. 2013 (Genome Res.)
- 3 Estrogen receptor regulation of carbonic anhydrase XII through a distal enhancer in breast cancer. Barnett DH et al 2008 (Cancer Res.)
  - → "we show that CA12 is robustly regulated by estrogen via ER alpha in breast cancer cells"

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#### **Tutorial**

Let us have a look together at

http://julien.cremeriefamily.info/doc/teachings/exposome/ transcriptomics\_networks\_inference.html