rGriffin

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2018-09-16

Introduction

Boolean networks allow us to give a mechanistic explanation to how cell types emerge from regulatory networks. However, inferring the regulatory network and its functions is complex problem, as the available information is often incomplete. refriffin uses available biological information (regulatory interactions, cell types, mutants) codified as a set of restrictions and returns the Boolean Networks that satisfy that restrictions. This Boolean networks can then be used to study the biological system.

The rGriffin package is an R connector to <u>Griffin</u> (Gene Regulatory Interaction Formulator For Inquiring Networks), a java library for inference and analysis of Boolean Network models. Griffin takes as inputs biologically meaningful constraints and turns them into a symbolic representation. Using a SAT engine, Griffin explores the Boolean Network search space, finding all satisfying assignments that are compatible with the specified constraints. TherGriffin package includes a number of functions to interact with the BoolNet package.

Queries

The first step is to attach rGriffin. This will initialize the Java Virtual Machine and start Griffin with the default JVM options.

```
library("rGriffin")
```

If you want to initialize the JVM with different options like more memory see the functions <code>init.griffin()</code>. It is also possible to modify the default parameters changing the file "rGriffin/java/jvm-param.R".

Create a query

All queries start with a topology that describes the nodes and its interactions. The function create.gquery.graph() takes a dataframe with columns for: source node, target node, and type of interaction. It also takes a vector with the node names.

Depending on the sign the interactions can **positive** or **negative**. If in every condition the regulation will have the same sign the interaction is **ambiguous**. However, if you are not sure if the regulation is positive or

negative in all contexts you can say that the interaction is **ambiguous**. Depending on the degree of confidence in the existence of the interaction, the interactions can be: **mandatory** if you are sure the interaction will happen or **optional** if you suspect the interaction exists but you are not sure.

The valid types of interctions are:

- false: Contradiction
- MA: Mandatory, ambiguous
- MPU (or +): Mandatory, positive, unambiguous
- MPPA: Mandatory, positive, possibly ambiguous
- MNU (or -): Mandatory, negative, unambiguous
- MNPA: Mandatory, negative, possibly ambiguous
- · MUSU: Mandatory, unknown sign, unambiguous
- MUSPA: Mandatory, unknown sign, possibly ambiguous
- · NR: No regulation
- OA: Optional, ambiguous
- o OPU: Optional, positive, unambiguous
- · OPPA: Optional, positive, possibly ambiguous
- · ONU: Optional, negative, unambiguous
- ONPA: Optional, negative, possibly ambiguous
- OUSU: Optional, unknown sign, unambiguous
- true: Tautology

For example, suppose a network where:

- a activates b
- **b** and **c** inhibit each other
- b and c may have a positive self-regulatory loops.

We can codify this information as:

```
genes = c('a', 'b', 'c')
inter = data.frame(source=c('a', 'b', 'b', 'c', 'c'),
                 target=c('b','b','c','b','c'),
                 type=c('+','OPU','-','-','OPU'),
                   stringsAsFactors = F )
inter
#>
   source target type
       а
#> 1
#> 2
#> 3
       b
b
               b OPU
                c -
#> 4
       С
                c OPU
#> 5
```

We then create the query q. This creates an instance of the class query in the JVM.

```
q = createGqueryGraph(inter, genes)
q
#> [1] "Java-Object{mx.unam.iimas.griffin.r.RGriffinQuery@1bba400}"
```

If you want to see the query use the print method.

```
print(q)
#> Gene Regulation Specification
```

```
#> Set of genes: [a, b, c]
#> geneOrder: null
#> Known regulations: None
#> Hypothetical regulations: None
#> Gene Interactions: [GeneInteraction [source=a, target=b, type=MPU], GeneInteraction
[source=b, target=b, type=OPU], GeneInteraction [source=b, target=c, type=MNU],
GeneInteraction [source=c, target=b, type=MNU], GeneInteraction [source=c, target=c,
type=OPU]]
#> Stable states: None
#> Prohibited states: null
   Known Regulators: None
#> Hypothetical Regulators: None
#> Extended regulators: None
#> Known Regulations: None
#> Hypothetical Regulations: None
#> Cyclic attractors: None
#> Input-output pairs: []
#> Subspaces: None
#> Mutant experiments: None
#> State transition rules: None
#> exactRegulations: false
#> additionalSteadyStates: false
#> divideQueryByTopology: false
#> topologyIteratorType: SEQUENTIAL
#> limitTopologyRange: false
#> limitToNumberOfNetworks: -1
#> topologyRange: None
#> topologicalDistanceRadius: 1
#> blockSteadyAPosteriory: false
#> additonalCycles: false
#> ambiguityNotAllowed: true
#> numberOfCycles:0
```

Add restrictions to the query

It is possible to add more restrictions to the query. It is important to remember that if there are more restrictions we expect less networks.

For example, suppose that we have some information about the expected cell types. We can add this restrictions as attractors.

For example, suppose that we know that the attractors are:

```
• Only express \mathbf{b} => c(0,1,0)
```

Only express c => c(0,0,1)

We can also add partial attractors where we lack information

• Do not express **a** or **c** but we have NO information on **b** => c(0, '*', 0)

We codify this information as:

We can add this information to the query with the add.gquery.attractors() function:

```
q = addGqueryAttractors(q, attr)
```

You can add aditional restrictions like:

- add.gquery.cycle() Add target cycle
- add.gquery.mutant() Add mutant with attractors
- $\verb| o add.gquery.prohibited.attractors()| \textbf{Add prohibited attractors} \\$
- add.gquery.transition() Add a transition between two succesive states
- add.gquery.trapspace() Add a trapspace

Run the query

Once you have created the query with run.gquery(). This function will return all the networks that satisfy the restrictions

```
nets = runGquery(q)
print(nets)
#> [1] "targets, factors\na, false\nb, ((((!a&b)&!c))|((a&!b)&!c))|((a&b)&!c))\nc, (!b&c)\n"
#> [2] "targets, factors\na, false\nb, (((((!a&b)&!c))|((a&!b)&!c))|((a&b)&!c))|((a&b)&c))\nc, (!b&c)\n"
#> [3] "targets, factors\na, false\nb, (((((!a&b)&!c))|((!a&b)&c))|((a&!b)&!c))|((a&b)&!c))|((a&b)&c))\nc, (!b&c)\n"
#> [4] "targets, factors\na, false\nb, ((((!a&b)&!c))|((a&b)&!c))|((a&b)&c))\nc, (!b&c)\n"
#> [5] "targets, factors\na, false\nb, (((((!a&b)&!c))|((a&!b)&!c))|((a&!b)&c))|((a&b)&!c))|((a&b)&c))\nc, (!b&c)\n"
```

The function run.gquery() includes multiple options that can be seen in the documentation. Some of the most important are:

- allow.hypothesis activate or deactivate hypothetical regulations
- allow.additional.states allows networks with additional fixed-point attractors to those specified in the query
- allow.additional.cycles allows networks with additional cyclic attractors to those specified in the query
- return.network.limit limit the maximum number of networks that the query will return

Connect to other packages

BoolNet

The R package includes various functions to import and export data to BoolNet.

It is possible to obtain the topology of a BooleanNetwork object using get.net.topology(). This function determines the sign of each regulation as positive '+', negative '-' or ambiguous 'MA'. All regulations between nodes are considered mandatory. The function can also detect Non-functional regulations 'NR'.

```
library(BoolNet)
data("cellcycle")
topology <- getNetTopology(cellcycle)</pre>
topology
#>
     Source Target Interaction
#> 1
       CycD
              CycD
#> 2
       CycD
                Rb
       CycE
                Rb
#> 4
       CycA
                Rb
        p27
                Rb
#> 5
       CycB
                Rb
#> 6
#> 7
               E2F
        Rb
#> 8
       CycA
               E2F
#> 9
        p27
               E2F
#> 10
       CycB
               E2F
#> 11
        Rb
               CycE
        E2F
#> 12
              CycE
#> 13
        Rb
              CycA
#> 14
        E2F
              CycA
#> 15
       CycA
              CycA
#> 16 Cdc20
              CycA
       Cdh1
#> 17
              CycA
#> 18 UbcH10
              CycA
#> 19
       CycD
               p27
#> 20
       CycE
               p27
#> 21
       CycA
               p27
#> 22
        p27
               p27
#> 23
       CycB
               p27
#> 24
       CycB Cdc20
       CycA
#> 25
              Cdh1
#> 26
       p27
              Cdh1
#> 27 Cdc20
              Cdh1
       CycB
#> 28
              Cdh1
#> 29
       CycA UbcH10
#> 30 Cdc20 UbcH10
#> 31
       Cdh1 UbcH10
#> 32 UbcH10 UbcH10
#> 33
       CycB UbcH10
#> 34 Cdc20
              CycB
#> 35 Cdh1
              CycB
```

It is also possible to convert an AttractorInfo object into a data.frame using attractor2dataframe.

```
cc.attr <- getAttractors(cellcycle)
cycle <- attractorToDataframe(cc.attr)</pre>
```

```
cycle <- cycle[cellcycle$genes] #remove info columns
cycle
   CycD Rb E2F CycE CycA p27 Cdc20 Cdh1 UbcH10 CycB
#>
     0 1
              0
                0
                    1
#> 1
#> 2
     1 0 0
                 1
                   0
                         0
                            0
                                 0
                                     0
              1
#> 3
     1 0 0
              0 1
                    0
                                 1
                                     1
                      1
                          0
#> 4
     1 0 0 0 1 0
                                 1
                                     1
#> 5
     1 0 0
            0 0 0
                        1
                          1
                                 1
                                   0
     1 0 1 0 0 0 0 1
#> 7
     1 0
          1 1 0 0 0 1
                                 0
                                     0
#> 8
     1 0
          1
              1
                 1
                    0
                         0
                            1
                                 0
                                     0
```

Using this functions, it is possible to create a query that corresponds to a Boolean network. As we expect a cyclic attractor we will use add.gquery.cycle().

```
q <- createGqueryGraph(topology, cellcycle$genes)
q <- addGqueryCycle(q, cycle)</pre>
```

It is also possible to export the networks generated by run.gquery() directly to BoolNet with the option return = "BoolNet". This option generates an iterator object, that returns the BooleanNetwork objects one by one using the function iterators::nextElem().

For example, if we use the three node example network:

```
nets = runGquery(q, return = "BoolNet")
#> Warning in boolnetModelIterator(controller): iterators is not attached
iterators::nextElem(nets)
#> Boolean network with 3 genes
#>
#> Involved genes:
#> a b c
#>
#> Transition functions:
#> a = 0
#> b = ((((!a&b)&!c))|((a&!b)&!c))
#> c = (!b&c)
#>
#> Knocked-out and over-expressed genes:
#> a = 0
```

If there are no more available networks the <code>nextElem()</code> method will rise an error: <code>Error</code> in <code>obj\$nextElem()</code> : <code>StopIteration</code>.

Other

It is possible to plot the network topology dataframe with the R package igraph. This dataframe can also be used to import and export the network topology to other resources like the python library networkx or to the software Cytoscape.

It is possible to export the network functions as an SBML file using the BoolNet function toSBML().

References

Muñoz, S., Carrillo, M., Azpeitia, E., & Rosenblueth, D. A. (2018). Griffin: A Tool for Symbolic Inference of Synchronous Boolean Molecular Networks. Frontiers in genetics, 9, 39.