Methods in Microbiota Research: Coding Lab

Ivan Vujkovic-Cvijin

(Ee-vahn Vooykoveech Tsveeyeen)

Assistant Professor

F. Widjaja Inflammatory Bowel Disease Institute

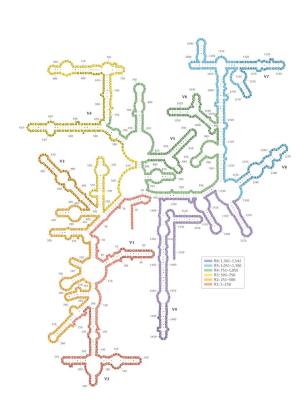
Department of Biomedical Sciences

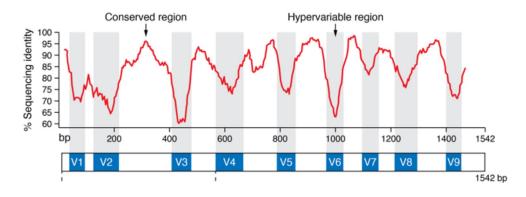
Notes prior to beginning

- Google is a coder's best friend
 - Almost every error message has a solution on some online forum
- Statistical modeling/testing in microbiome science is a constantly evolving area
 - Precedent does not always justify usage methods may be outdated
 - No consensus on best methods more clarity on what not to do
- I am self-taught
 - If I can do it, so can you!

Tools of the Trade

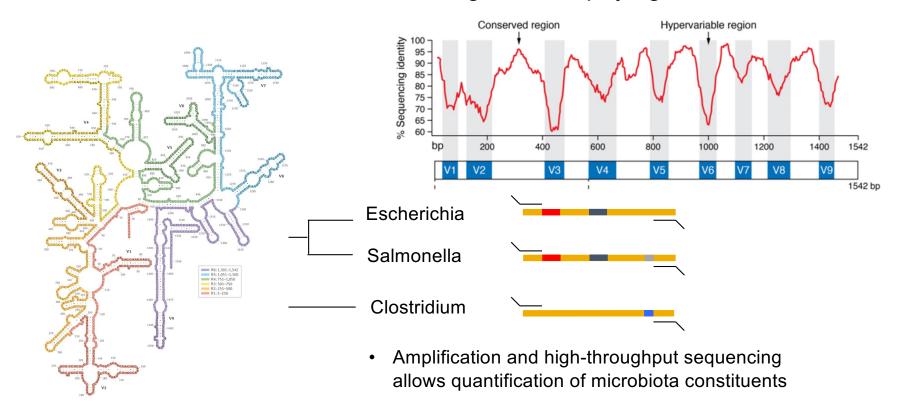
The bacterial 16S ribosomal RNA gene as a phylogenetic barcode





Tools of the Trade

The bacterial 16S ribosomal RNA gene as a phylogenetic barcode



16S rRNA sequencing analysis: dada2

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Read 1 CTAAGACCGGATAGGTA

Read 2 CTAAGACCGGATAGGTA

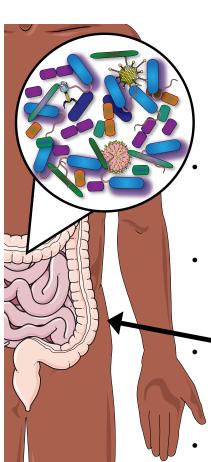
Read 3 CTAAGACCGGGATAGGTA

Read 4 CTAAGACCGGATAGGTA

Read 5 CTAAGACCGGATAGGTA
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Illumina 'quality scores' low high

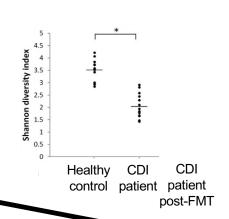
Quality Score Q(X)	Error Probability P(~X)
Q40	0.0001 (1 in 10,000)
Q30	0.001 (1 in 1,000)
Q20	0.01 (1 in 100)
Q10	0.1 (1 in 10)



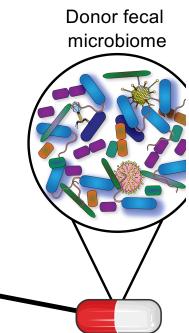
Prototypical microbiome-mediated disease: *Clostridioides difficile* infection

- C. difficile infection is associated with a loss of diversity of the endogenous microbiota
- Is generally treated with antibiotics targeting C. difficile (e.g. vancomycin)
- Antibiotic treatment often does not work, condition becomes chronic(~35%)
- Restoration of the endogenous microbiota via fecal microbial transplantation (FMT) cures up to 94% of recurrent patients (van Nood, NEJM, 2013)

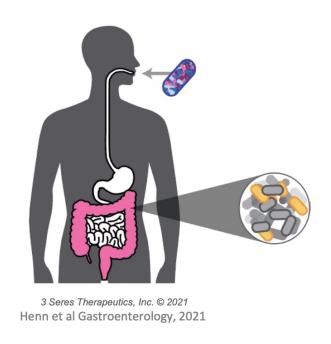
Question: How can we identify gut taxa critical for recurrent CDI cure?

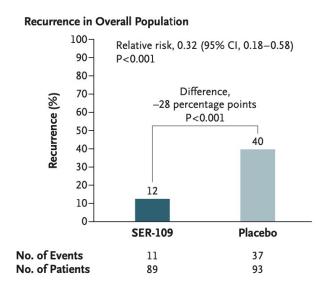


Weingarden et al. Am J Phys 2015



Precision microbiota restoration prevents recurrent CDI

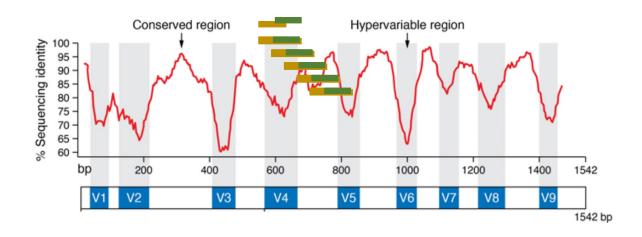




Feuerstadt et al. NEJM. 2022

- Seres Therapeutics identified gut taxa that could help prevent CDI, bottled them up, and made a therapeutic
 - Question: How can we identify these key gut taxa? Let's get coding...

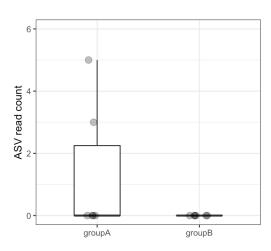
Pre-processing: Chimera filtering

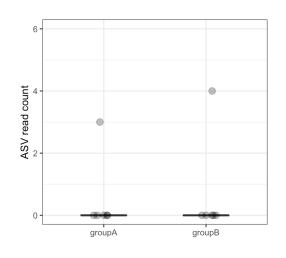


- Chimeras are identifiable if one portion is identical to a more abundant 'parent' sequence and the rest is identical to another 'parent' sequence
 - 'removeBimeraDenovo' function in dada2

Pre-processing: removing taxa with low read counts

Mean fold difference A/B = infinity





Mean fold difference A/B = 0.75

- Random re-sampling (e.g. multiple rarefactions or re-sequencing) of samples causes wide variation in abundance of taxa with low read counts
- Fold changes between groups can be very unstable and unreliable
 - Example solution: filter taxa that are below 0.001% abundance threshold

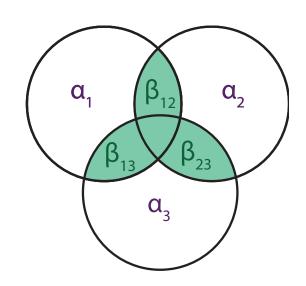
Comparative metrics: alpha vs. beta diversity

Alpha diversity:

- The number of different taxa present in a sample (richness) and/or the relative distribution of these taxa (evenness)
 - Examples: chao1 (richness), Shannon or inverse Simpson (richness and evenness), Pielou's (evenness)
- Each sample gets its own alpha diversity quantification

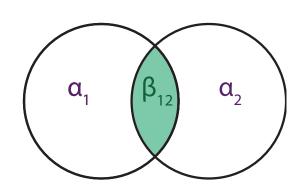
Beta diversity:

- The ecological similarity between two samples
 - Similarity in common taxon membership (ex. Canberra, Bray-Curtis), phylogenetic similarity of taxa within a community (ex. Unweighted UniFrac)
- Each pair of samples gets its own beta diversity quantification



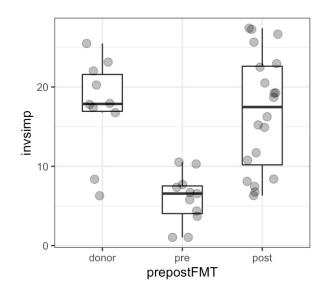
Beta diversity matrix

Subject ID	1	2	3	
1	0	0.22	0.9	
2	0.22	0	0.47	
3	0.9	0.47	0	



Comparative metrics: alpha vs. beta diversity

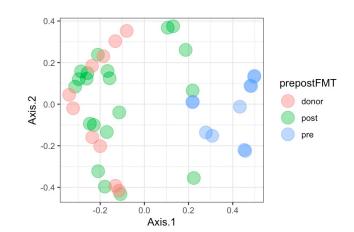
Inverse Simpson (Alpha diversity)



Beta diversity matrix

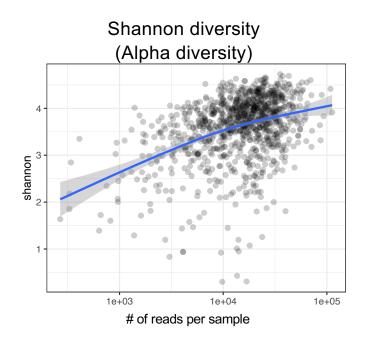
Subject ID	1	2	3	
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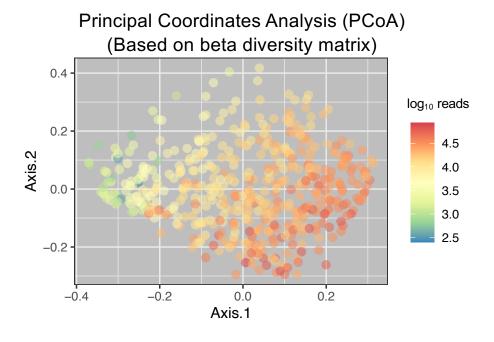
Principal Coordinates Analysis (PCoA) (Based on beta diversity matrix)



Pre vs. post PERMANOVA P = 0.00001

Alpha and beta diversity are skewed by sequencing read depth





Rarefaction: random selection of 'n' reads per sample

Randomly sub-sample 9 candies from each handful (rarefy to 9 candies)

