Sexual selection improves population fitness: a systematic review and meta-analysis

Justin G. Cally[§*], Devi Stuart-Fox[§] and Luke Holman[§]

[*] *justin.g.cally@gmail.com*

Abstract

Sexual selection has manifold ecological and evolutionary consequences, making its net effect on population fitness difficult to predict from first principles. Instead, one can empirically determine whether sexual selection increases or decreases population fitness by manipulating the opportunity for sexual selection in replicate populations, allow evolution to proceed, and then compare the evolved average fitness between the treatment groups. Here, we synthesise 459 effect sizes from 65 pertinent experimental evolution studies using meta-analysis. We find that sexual selection on males tends to elevate the mean and reduce the variance for many fitness traits, especially in females, and for populations evolving under stressful conditions. We conclude that the beneficial population-level consequences of sexual selection typically outweigh the harmful ones and that the effects of sexual selection on total selection, genetic variance, and/or evolvability can differ markedly between sexes and environments. We discuss the implications of these results for conservation and evolutionary biology.

Keywords: Adaptation, Gender load, Monogamy, Mutation load, Sexual conflict.

[§]School of BioSciences, The University of Melbourne, Victoria, Australia.

Introduction

Sexual selection, defined as selection resulting from competition for mates or their gametes[1], is a ubiquitous evolutionary force that has profoundly shaped the natural world. As far back as Darwin[2], researchers have theorised that sexual selection can change the average absolute fitness of individuals in a population, henceforth termed 'population fitness'[3,4]. However, opinion is divided over whether the net effect on population fitness is positive or negative[5–8]. *Prima facie*, one might predict that sexual selection would have no effect on population fitness, since it does not matter which individuals of the faster-reproducing sex (typically males) succeed in mating, so long as some do[9]. However, when genotypes with high mating or fertilisation success also have superior breeding values for traits that affect population fitness (*e.g.*, survival, parental care, female fecundity, or success in interspecific competition), sexual selection is predicted to elevate population fitness by causing a correlated response in these other traits[8]. In essence, the demographically-limiting sex (typically females) benefits from a gene pool that has been purged of harmful alleles through sexual selection on the non-limiting sex (typically males). Theoretically, the benefit to population fitness could be large[10–12].

Conversely, sexual selection can decrease population fitness if male sexually-selected traits are negatively genetically correlated with female fitness, producing intralocus sexual conflict[13–18]. Additionally, sexual selection frequently favours phenotypes that reduce population fitness but benefit the individuals expressing them, such as harassment or infanticide by mate-seeking males[19–21], as well as investment in costly sexual signals and weaponry at the expense of parental care: *inter*locus sexual conflict[22–24]. Given these conflicting theoretical expectations and empirical results, it remains unclear whether sexual selection tends to have a net benefit or cost to population fitness[5–8].

Researchers have investigated the population-level consequences of sexual selection using a range of approaches including macro-evolutionary studies[25–27], analysis of the fossil record[28], quantitative genetics[13,29–32] and especially experimental evolution. In particular, many experimental evolution studies have manipulated the intensity of sexual selection in captive populations, allowed evolution to proceed, and then measured population fitness components such as lifespan, reproductive success, population extinction rate and mutation load. This approach facilitates direct measurement of the net effect of sexual selection on population fitness, at least in the specific populations and ecological conditions under study.

A number of factors might influence the strength and sign of the correlation between sexual selection and population fitness. First, the genetic correlation between female fecundity and male mating/fertilisation success varies in sign and magnitude between species[33] and even between conspecific populations[34], implying that sexual selection on males increases mean female fitness in some species and populations but not others. These inconsistencies could derive from differences in allele frequencies, or environmental differences that alter how genotype relates to phenotype and fitness. Second, it has been hypothesised that populations should display a more positive genetic correlation between male and female fitness – and thus potentially between mating/fertilisation success and population fitness – in novel or fluctuating environments, relative to stable environments[13,32,35,36]. This is because stable environments create consistent selection, preferentially eroding genetic variation at sexually-concordant loci (*i.e.* loci where the fittest genotype is the same in both sexes) and leaving behind variation at sexually antagonistic loci. We know of no systematic reviews of this latter theory, though it is has motivated several recent empirical tests[13,29,31], and is relevant to conservation genetics.

Here, we synthesise the empirical literature on sexual selection and population fitness using formal meta-analysis. We focus exclusively on experimental evolution studies that manipulated the presence or strength of sexual selection on males, and then measured some fitness component, since experiments provide a particularly strong test of the hypothesis that sexual selection affects the average fitness of populations. Our principle aims were to measure the average net effect of sexual selection on the population-level mean and variance for various fitness components, to test whether this effect varies between stressful and benign environments, and identify key moderators of the effect size of sexual selection treatment.

Results

The effect size dataset

We retrieved 459 effect sizes from 65 studies. 92 effect sizes were collected from populations evolving under stressful conditions, while 337 were measured for those evolving in benign conditions. 189 of the effect sizes came from measurements made on males, 219 on females, and the remaining 51 from measurements of a mixed-sex sample of individuals. Most effect sizes in our dataset came from studies that manipulated sexual selection by completely removing it in one treatment via enforced random monogamy ($n = 241$); other effect sizes ($n = 218$) derived from alternative manipulations, such as changing the adult sex ratio. In total, we obtained effect sizes for 22 different fitness traits, with female reproductive success ($n = 102$) and offspring viability ($n = 56$) being the most commonly-measured traits. We classified 171 effect sizes as direct measures of population fitness, 141 as indirect, and the remaining 144 effect sizes as ambiguous (see Methods, Figure 1 and Table S1). Specifically, we scored traits that are likely to correlate with population growth and persistence as direct (*e.g.*, female reproductive success, offspring viability, and extinction rate), those which are not necessarily correlated with population fitness but which do measure individual fitness as indirect (*e.g.*, lifespan, male mating success and ejaculate quality/production); and those for which the relationship to population fitness is unclear as ambiguous (*e.g.*, body size, mating duration, male reproductive success, and early fecundity). Tables S4-S5 give a detailed description of our dataset.

Sexual selection is associated with higher mean values for most fitness components

The grand mean across all types of effect sizes (direct, indirect and ambiguous) was positive (REML $\beta = 0.24$, 95% CIs: 0.055 to 0.43, $p = 0.011$; Bayesian $\beta = 0.25$, 95% CIs: -0.0074 to 0.51, $BF_{>0} = 35$), indicating that sexual selection on males typically had a net positive effect on the majority of populations and fitness components so far studied. Moreover, the effect sizes associated with the manipulation of sexual selection varied between different fitness traits. Sexual selection had a beneficial effect on most fitness traits, but varied across the three relationships to fitness Figure 1; Table S6). Sexual selection elevated fitness for traits that shared an ambiguous relationship to fitness (REML $\beta = 0.21$, 95% CIs: 0.058 to 0.093; Bayesian $\beta = 0.20$, 95% CIs: -0.0016 to 0.39, $BF_{>0} = 38$, $n = 144$) and an indirect relationship to fitness (REML $\beta = 0.24$, 95% CIs: 0.13 to 0.36; Bayesian $\beta = 0.24$, 95% CIs: 0.033 to 0.43, $BF_{>0} = 59$, $n = 141$). Additionally, sexual

selection elevated fitness components directly related to fitness, albeit at a lower magnitude (REML $\beta = 0.13$, 95% CIs: 0.019 to 0.24; Bayesian $\beta = 0.13$, 95% CIs: -0.079 to 0.31, $BF_{>0} = 11$, $n = 174$). A large forest plot with predicted effect sizes for each fitness component is presented in Figure S1 and further detailed with model predictions and individual meta-analyses in Table S8 and S9. Sexual selection significantly reduced two fitness components, namely immunity (REML β = -0.42, 95% CIs: -0.64 to -0.20; Bayesian β = -0.43, 95% CIs: -0.70 to -0.15, $BF_{>0} = 0.0026$; $n = 35$) and body condition (REML β = -1.2, 95% CIs: -1.8 to -0.63; Bayesian β = -1.2, 95% CIs: -1.9 to -0.63, $BF_{>0} < 0.0001$, $n = 1$).

The roles of environmental stress and sex

We found that the sex of the individuals measured (male, female, or a mixture), and the conditions under which the population evolved (stressful or benign) interacted to affect the relationship between sexual selection and fitness (Table 1, 2). Sexual selection on males significantly improved female fitness, and the beneficial effect of sexual selection was significantly stronger for females from populations evolving under stressful conditions (*e.g.* a food source to which they were not well-adapted) than under benign conditions (Figure 2a, Table 2). Sexual selection had a positive but non-significant effect on male fitness, and in contrast to females, fitness benefits were significantly weaker in stressful than benign environments (Figure 2a, Table 2). Consistent with the different consequences of sexual selection for female and male fitness, the mean effect size in mixed-sex samples was non-significantly positive, and there was no significant difference between benign versus stressful conditions (Figure 2a, Table 2). Overall, our results indicate that the positive effect of sexual selection on fitness is greater for females than males, and the difference between the sexes is magnified in stressful environments. When we used an alternative measure of effect size (log response ratio or lnRR), the results aligned with those of Hedges' g in that sexual selection elevates population fitness, with its effect magnified for females evolving in stressful environments (Figure S3 and Table S16).

Other moderator variables that we examined had minimal impacts on effect size (Figure S2, Table S10). Specifically, effect size did not depend on whether or not the study was conducted blind (Figure S6), nor on the number of generations for which the experimental evolution study was run (Figure S7, S8).

The effect size estimates we recovered were highly heterogeneous ($I^2 = 95.2\%$, 95% CIs: 94.4% to 95.9%), reflecting the large differences in experimental procedures, study species, and fitness components included in our meta-analysis[37,38]. Heterogeneity stemmed mostly from between-study differences ($I^2_{study} = 36\%$, 95% CIs: 26.5% to 45.4%) rather than differences between fitness components and taxon ($I^2_{fitness\ components} = 0.4\%$, 95% CIs: 0.2% to 0.9%; $I^2_{taxon} = 1.4\%$, 95% CIs: 0.2% to 3.5%). Variation among taxa is explored further in the ESM (Figure S2, Table S11).

Sexual selection reduces phenotypic variance, for female traits in stressful environments

By applying meta-analysis to log coefficient of variation ratios[39], we found evidence that sexual selection reduces phenotypic variation under certain conditions (Figure 2b). Specifically, phenotypic variance was significantly reduced by sexual selection for fitness components measured in females under stressful conditions ($lnCVR$ = -0.78, 95% CIs: -1.23 to -0.34, $n = 27$). By contrast, we found no significant effect of sexual

selection on phenotypic variance in males, or for either sex under benign conditions (Figure 2b; Tables S13-S16). However, similar to the results in females, there was a non-significant trend for a reduction in phenotypic variance in mixed-sex samples measured under stressful conditions ($lnCVR$ = -0.76, 95% CIs: -1.22 to -0.31; Figure 2b). A meta-analysis using the log variability ratio (lnVR), which does not account for the mean-variance relationship present in the dataset (Figure S4), suggested sexual selection reduces variance for mixed-sex samples in stressful conditions, but not females, results of this meta-analysis can be found in the Supplementary Material (Figure S5 and Table S21).

As in the meta-analysis of trait means, there was high heterogeneity in the estimates of $lnCVR$ (I^2 = 98.9%, 95% CIs: 98.7% to 99.1%). Heterogeneity in the dataset was due to variability between studies fitness components ($I^2_{fitness\ components}$ = 12.7%, 95% CIs: 4.5% to 23.1%) and taxon (I^2_{taxon} = 7.7%, 95% CIs: 1.3% to 18.4%), as well as large amounts of residual heterogeneity (78.6 %, 95 % CIs: 65.9% to 89%). Using the REML approach heterogeneity associated with the Study ID random effect was estimated at zero (I^2_{study} = 0 %), however the Bayesian approach suggests heterogeneity between studies may be zero or small (sd_{study} = 0.07, 95 % CIs: 0 to 0.19).

Publication bias

The funnel plot of effect sizes was asymmetrical, suggesting that some publication bias might be present (Figure 3a; Egger's test: $z = 5.9$, $p < 0.0001$). Specifically, there was a moderate excess of low-powered studies in which sexual selection had a more positive effect on the fitness component than average, implying that low-powered studies are more likely to be published if they report statistically significant fitness benefits of sexual selection (though funnel plots are not a decisive evidence of publication bias[40]). There was no significant relationship between effect size and journal impact factor (Figure 3b; t_{437} = 1.2, p = 0.23) or year of publication (Figure 3c; t_{437} = -1.2, p = 0.24); thus, we found no evidence that effect size dictates the likelihood of publication in high-profile journals, or that effect sizes have diminished as the field has matured[41,42].

Discussion

Our meta-analysis revealed that populations evolving under sexual selection often have higher values for multiple fitness traits, relative to populations where sexual selection on males was experimentally removed or weakened. Sexual selection had beneficial effects for traits that were ambiguously or indirectly related to fitness, but had smaller, variable effects on direct measures of fitness such as extinction rate, reproductive success (defined as the number of offspring produced) and the proportion of viable offspring. Fitness traits related to immunocompetence were an exception: sexual selection typically resulted in weaker immunity. This result is interesting in light of the hypothesised trade-off between sexually-selected phenotypes and immunity, for example, due to immunosuppressive effects of sex hormones[43,44]. Furthermore, the overall benefit of sexual selection was greater for females than males, and this sex difference was magnified in stressful environments. Consistent with stronger selection on female fitness under stress, female and mixed-sex samples showed reduced phenotypic variance when sexual selection was applied under stressful as opposed to benign conditions. These results suggest that sexual selection may contribute to population persistence under stressful conditions, such as fluctuating environmental change[35] or spatial variability[45], particularly since female reproductive output is often a limiting factor in population growth[46].

The results of the meta-analysis support predictions that sexual selection on males can improve population fitness and accelerate adaptation[10–12,35,47–49]. Specifically, our results support arguments that positive genetic correlations between male mating success and female non-sexual traits are common, allowing females to benefit from a genome that has been 'purged' of deleterious alleles through sexual selection on males[8,10,11]. What is less clear is why sexual selection had a larger effect on female trait means and variances as opposed to males (the sex in which sexual selection is typically strongest). Below we discuss possible explanations for this result, in light of the core principle that the extent of adaptation depends on additive genetic variance, the structure of genetic covariance, and the strength of selection[50,51]. However, these explanations are speculative and *post hoc*, as the result is arguably opposite to what one would predict, (*i.e.* that manipulating sexual selection on males should affect the phenotype of males more than females).

Firstly, the response of male fitness to sexual selection may be constrained in species where the heritability of male fitness is influenced by the sex determination system. Males are heterogametic in many of the species in our sample (*e.g.* due to XY or XO sex determination); this can reduce father-to-son heritability relative to mother-to-daughter, since sons never inherit the larger sex chromosome from their fathers[16,52–54], which might slow adaptation in male relative to female traits[54,55]. However, X-linkage is also likely to inflate the contribution of X-linked genes to fitness variance in the heterogametic sex. Under this view, X-linkage facilitates adaptation by increasing the efficacy of selection against recessive deleterious alleles and selection for recessive beneficial alleles[56,57]. Perhaps as a result of these conflicting predictions, there seems to be no clear consensus regarding sex-based differences in heritability. For instance, a systematic review found no difference in mean heritability between male and female traits, although there was a male-biased skew in heritability of reproductive traits[58]. Conversely, a recent association study in humans found a number of traits showing higher heritability in females than males[59].

Secondly, selection on males might be weaker than selection on females, resulting in slower adaptation following the experimental manipulation of sexual selection. This explanation may initially seem implausible, because net selection on males is generally assumed to be stronger than on females, due to (actual or

hypothesised) sex differences in sexual selection and inter-individual variance in fitness[60–63]. However, an oft-overlooked aspect is that selection might frequently be 'softer' on males and 'harder' on females[64], because the local competitive environment is usually more important for males than it is for females. For instance, a mediocre male genotype can have high fitness provided it outcompetes its local rivals, while low-fitness female genotypes are likely to produce few offspring even when competing with other low-fitness females. Therefore, improvements in genetic quality should have stronger diminishing returns in males, possibly contributing to our finding that the genetic consequences of sexual selection lead to greater fitness benefits for females. Though this argument is speculative, we note that many experimental evolution designs exaggerate the sex difference in the nature of selection, relative to expectations for large, natural populations[65–67]. For example, many studies[68–72] have evolved insects in small sub-populations, each containing one female and multiple males, whose progeny are then mixed and randomly sampled to create the next generation; this design ensures that successful males simply needed to outcompete their rival(s) in the same sub-population (soft selection), while each female's reproductive output is measured against the entire female population (hard selection).

Our results suggest that the greater benefit of sexual selection to females than males is magnified in stressful environments. Recent work has emphasised that environmental stress should reduce the strength of sexually antagonistic selection relative to selection that is concordant between sexes. Theoretical models reaching this conclusion[35,73] have been supported by some empirical work[13,36,74]; for example, one study found that high fitness males produced low fitness daughters under benign conditions but high fitness daughters under stress[36]. However, other quantitative genetic studies have shown that stressful conditions do not always reduce sexual antagonism[29,32,75]. Variation in effects of sexual selection in stressful environments may be due to potentially variable responses amongst taxa[33] and environments. Notably, Connallon & Hall[35] predict that the dynamics of environmental change alter the strength of sexual antagonism; for instance, gradual directional selection may facilitate indefinite sexual antagonism, while rapid cyclical change can swiftly remove it. Our meta-analysis suggests that under directional selection imposed by environmental stress, sexual antagonism is likely dampened; allowing sexual selection to facilitate adaptation and persistence.

Although our meta-analysis revealed an overall positive effect of sexual selection, the variation in effect size across the dataset is high, as is often the case for studies in ecology and evolution[37]. Most of the heterogeneity was between studies (potentially due to differences between study designs and populations), while the taxon, number of generations of evolution, and use of blinding had less impact on effect size. Experimental evolution studies cover relatively few taxa, and most focus on easy-to-culture invertebrates with similar mating systems and sex determination. However, a meta-analysis of macroevolutionary studies on sexual selection and speciation rate found no significant taxon-based differences across a diverse sample of vertebrates and invertebrates (fish, insects, birds, spiders, reptiles, mammals)[76], perhaps suggesting that our results would generalise to other taxa. On the other hand, biological differences between taxa could change the relationship between sexual selection and population fitness. For example, species where males and females have radically different morphology might have a reduced inter-sexual genetic correlation for fitness, such that sexual selection has fewer pleiotropic benefits for females, while for species with sexually-selected male parental care, sexual selection might help the population by conferring high fitness to caring fathers.

Our findings have implications for fundamental and applied research. For example, the beneficial population-level consequences of sexual selection have been proposed as one possible resolution to the long-standing evolutionary puzzle regarding sexual reproduction[77]. If sufficiently strong, these benefits can more than compensate for the costs of sexual reproduction, and prevent sexual populations from being outcompeted by

asexual mutants[10,11,78]. Sexual selection is also important for conservation[6] and captive breeding programs[79]. Within captive breeding programs, genetic diversity is often managed through the enforced monogamy of a strategically selected (genetically diverse) breeding pair[79]. Captive breeding programs may benefit from allowing sexual selection of 'good genes' or more compatible genes[80], or by increasing maternal investment by females paired with 'attractive' males[81–83]. Additionally, our findings imply that anthropogenic environmental changes that reduce the opportunity for sexual selection, such as eutrophication, pesticides, artificial light and noise pollution, could reduce the genetic quality of the population, and potentially compromise its long-term persistence[84–88]. Equally, our results support recent evidence that human activities that directly counteract sexual selection, such as selective harvesting of the largest or most ornamented males, can lower population fitness[89]. Based on the weight of evidence from experimental evolution, we suggest that sexually-selected populations may be more resilient to environmental change, including anthropogenic environmental pressures, over relevant time scales.

Methods

Literature search

We searched *ISI Web of Science* and *Scopus* on 9th June 2017 for peer-reviewed, English language studies that manipulated the presence or strength of sexual selection using experimental evolution, and then measured some proxy of population fitness. A detailed list of search terms is given in the Electronic Supplementary Material (ESM).

After removing duplicates, we read the titles and abstracts of the remaining 1,015 papers and removed those that did not fit our inclusion criteria (typically because they did not present primary experimental evolution data). This left 130 papers, for which we read the full text and applied the inclusion criteria outlined in the PRISMA diagram (Figure 4). Briefly, we included studies that 1) were conducted in a dioecious animal, 2) experimentally manipulated the strength of sexual selection (*e.g.*, via experimentally-enforced random monogamy or an altered sex ratio) for at least one generation, and 3) measured a trait that we judged to be a potential correlate of population fitness. This third criterion is the most subjective, because there is rarely enough data to determine whether a particular trait is (or is not) correlated with population fitness. We therefore relied on our best judgement when deciding what outcomes were correlated with population fitness. We categorised the fitness outcomes into three categories: ambiguous, indirect and direct (detailed in Table S1). Briefly, ambiguous measures of fitness were those that are reported to have an unclear or variable association with fitness (*e.g.*, body size, mating duration, early fecundity and male reproductive success). Indirect fitness components were those that are often used as a proxy of fitness but do not directly measure aspects of success in reproduction or population viability (*e.g.*, lifespan, mating success and ejaculate quality/production). Finally, direct measures of fitness (female/mixed sex reproductive success, offspring viability and extinction rate) are those that measure fitness through components of reproduction or long-term viability. The ESM describes why each of the 130 papers was included or excluded (Table S2).

Of these 130 papers, 62 were excluded based on the PRISMA criteria (Figure 4). Additionally, three papers presented insufficient information to calculate effect size. In these cases, we contacted the authors and

attempted to obtain the missing data, with partial success. The final meta-analysis included data from 65 papers.

Data extraction

From each paper, we first attempted to extract the arithmetic means, standard deviations and sample sizes of each of the different treatment groups, which facilitate calculation of effect size (see below). Typically, there were two or three treatments, which varied in the strength of sexual selection on males through manipulations to the adult sex ratio, in these cases we considered treatments with the greater male to female ratio to be the high sexual selection treatment group. For some papers, summary statistics were not written down, but were presented in a figure such as a bar chart: in these cases, we extracted the data using *WebPlotDigitizer* v.3.12[90]. If the treatment means were not reported (and the raw data were unavailable), we instead calculated effect size from test statistics comparing treatment means (*e.g.*, F, t, z or χ^2 values), which we used to estimate effect size using several formulae (see below).

Where possible, we extracted data for each independent replicate or experimental evolution line within a study; otherwise, we used pooled treatment means. For studies that repeatedly measured the same population across multiple generations, we only extracted data for the last reported generation.

In addition to the data used to calculate effect size, we collected a set of moderator variables for each paper (see Table S3 and associated ESM). The moderators were selected due to their ready availability, and because we hypothesised that they might explain some of the observed heterogeneity in effect size. A key moderator was whether the environmental conditions that a population evolved under were stressful (*e.g.*, elevated mutation load, novel/sub-optimal food source, increased sub-lethal temperatures). Additionally, we collected details for each effect size on: sex (male, female or a mixed sample of both), taxon (flies, beetles, mice, nematodes, mites, crickets and guppies), the presence/absence of blind methodology, and number of generations a treatment group underwent experimental evolution. In the interests of creating a useful data resource, we also recorded details about each experiment that were not formally analysed due to a shortage of data, such as the type of sexual selection that was manipulated (pre-copulatory, post-copulatory, or both) and the male to female ratio, which is included in the ESM (Table S3).

Effect size calculation

For each measurement of each pair of treatments, we estimated the standardised effect size Hedges' g[91]. Similar to Cohen's d, Hedges' g expresses the difference in means in terms of standard deviations (making it dimensionless), but it is more robust to unequal sampling and small sample sizes[92]. For comparisons of extracted treatment means, we calculated Hedges' g using the `mes` function in the *compute.es* R package[93]. To calculate Hedges' g from test statistics, we used the `fes`, `chies`, and `tes` functions in the *compute.es* package (for F, χ^2 and t statistics, respectively). The `propes` function was used to calculate effect size from a difference in proportions; in two cases[94,95], a proportion was equal to one (producing infinite effect sizes), and so we subtracted one from the numerator when estimating Hedges' g. In all cases, we selected a direction for the effect size calculation such that in our meta-analysis, negative effect sizes indicate that the removal of sexual selection was associated with higher fitness trait values, and positive effect sizes indicate higher

fitness when sexual selection was elevated or left intact. We also inverted the sign of effect sizes pertaining to measurements that are expected to be negatively related to population fitness (*e.g.*, parasite load, mutation load, extinction risk/rate, mating latency (males), and rate of senescence). Because many of our 65 papers measured multiple fitness outcomes, studied multiple replicate populations, or had three or more sexual selection treatments, we calculated a total of 459 effect sizes.

Additionally, using studies that presented means, standard deviations and sample sizes ($n = 352$) we were able to calculate an alternative measure of effect size: the log response ratio ($lnRR$)[96,97]. The $lnRR$ was used as a supplement to Hedges' g because it relaxes the assumption in equal variances between control and treatment groups (homoscedasticity).

For the meta-analysis testing whether sexual selection affects phenotypic variance (as opposed to the mean), we estimated the difference in variance between each pair of treatments using the natural logarithm of the ratio between the coefficient of variation for each group (termed $lnCVR$)[39]: $ln(CVfitness_{SS\ high}/CVfitness_{SS\ low})$. The use of $lnCVR$ allows us to determine the effects of sexual selection on phenotypic variance, with the coefficient of variation implicitly controlling for the mean-variance relationship seen in the dataset (Figure S4). As a supplement, we also calculated the natural logarithm of the absolute ratio between the absolute variation for each group ($lnVR$) in order to assess the impact of sexual selection on trait variance, irrespective of their magnitudes[39]. The calculation of $lnCVR$ and $lnVR$ relies on the availability of arithmetic means, standard deviations and sample sizes for the two treatment groups[39,98], and so we were only able to calculate $lnCVR$ and $lnVR$ for 354 of 459 comparisons.

Mixed-effects meta-analysis

Firstly, we obtained a weighted mean effect size (Hedges' g) for the entire dataset, using both Bayesian and restricted maximum likelihood (REML) approaches for completeness. The weighted mean was obtained by fitting a model with no moderator variables (*i.e.* fixed effects), but fitness component (*e.g.*, body size, female reproductive success), study ID, and taxon as random/group-level effects. That is, we separately model correlations between different effect sizes sourced from the same study, taxon, or pertaining to the same fitness component, and account for these interdependencies when estimating the overall effect. Given the small number of phylogenetically diverse species, we did not utilise phylogenetic corrections within the models. In our meta-analyses we report Bayes Factors (BF), giving the likelihood ratio that the focal effect size differs from zero $BF_{>0}$.

Secondly, we fixed the relationship to fitness class (Ambiguous, Indirect or Direct) as a moderator variable in Bayesian and REML models (whilst maintaining study and taxon as group-level effects) to derive predictions for effect size within each of the three fitness-relationship classes, using the relevant `predict` functions for each of the `R` packages used (see below). This meta-analysis was then supplemented by another model where we fixed fitness component as a moderator variable (*e.g.*, immunity, lifespan, offspring viability and female reproductive success); predictions for this model on the 22 fitness components were derived as above. Alternatively, to assess the impact of sexual selection on each fitness component independently of one another we conducted seperate meta-analyses ($n = 18$); subset for each fitness trait with more than three effect sizes. These models were were intercept only REML models with study and taxon as group-level effects. Further details on model parameters can be found within the ESM.

Thirdly, we measured the impact of environment, sex and their interaction on the effect size (Hedges' g, $lnRR$, $lnCVR$ and $lnVR$ in the ESM) associated with the manipulation of sexual selection, by fitting these predictors as moderators in a pair of separate mixed-effects meta-analyses. These meta-analyses were restricted to effect sizes calculated from 'unambiguous' outcomes (*i.e.* those scored as being directly or indirectly related to population fitness), as well as those where we were able to define the environmental conditions as either stressful or benign (Hedges' g: $n = 330$; $lnRR$, $lnCVR$ & $lnVR$: $n = 269$). We again fit study ID, fitness component, and taxon as random/group level effects. Models investigating other moderators such as number of generations and blinding are presented in the ESM.

For our meta-analyses investigating the effects of environment and sex on the magnitude and variance of fitness-related traits we provide estimates of heterogenity present in the dataset. We use the statistic I^2 as an estimate of the proportion of variance in effect size that is due to differences between levels of a random effect (*e.g.* studies)[99]. I^2 is prefered over other statistics as it is independent of sample size, is easily interperatable, and can be partitioned between random effects[37]. Within ecology and evolution heterogeneity in datasets is often high, with the mean I^2 from 86 studies above 90%[38].

Meta-analyses fit by REML were implemented in the *metafor* `R` package[100], while their Bayesian equivalents used the `R` package *brms* to run models in Stan[101].

Publication bias

We tested for publication bias via funnel plots, using Egger's test to quantify plot asymmetry[102,103]. Additionally, we tested for time-lag bias[41], in which effect size magnitudes decline over time as more data are collected. Additionally, we assessed a potential source of publication bias through the correlation between effect size and journal impact factor[42], which can arise if null or countervailing results are more difficult to publish (impact factors were from *InCites Journal Citation Reports*).

Code availability

The code used to perform this meta-analysis is freely available on Github (https://justincally.github.io/SexualSelection/). Here the code is available through a neatly compiled html, with the R Markdown file found within the corresponding github repository: (https://github.com/JustinCally/SexualSelection).

Data availability

All data are freely available on Github (https://github.com/JustinCally/SexualSelection). The repository provides a source data file that was used to create all results, tables and figures.

References

1. Andersson, M. B. *Sexual selection.* (Princeton University Press, 1994).

2. Darwin, C. *The descent of man and selection in relation to sex.* **1**, (John Murray, 1871).

3. Kimura, M. On the change of population fitness by natural selection. *Heredity* **12**, 145–167 (1958).

4. Lande, R. Natural selection and random genetic drift in phenotypic evolution. *Evolution* **30**, 314–334 (1976).

5. Candolin, U. & Heuschele, J. Is sexual selection beneficial during adaptation to environmental change? *Trends in Ecology & Evolution* **23**, 446–452 (2008).

6. Holman, L. & Kokko, H. The consequences of polyandry for population viability, extinction risk and conservation. *Philosophical Transactions of the Royal Society B-Biological Sciences* **368**, (2013).

7. Kokko, H. & Brooks, R. Sexy to die for? Sexual selection and the risk of extinction. *Annales Zoologici Fennici* **40**, 207–219 (2003).

8. Whitlock, M. C. & Agrawal, A. F. Purging the genome with sexual selection: Reducing mutation load through selection on males. *Evolution* **63**, 569–582 (2009).

9. Rankin, D. J. & Kokko, H. Do males matter? The role of males in population dynamics. *Oikos* **116**, 335–348 (2007).

10. Agrawal, A. F. Sexual selection and the maintenance of sexual reproduction. *Nature* **411**, 692–695 (2001).

11. Siller, S. Sexual selection and the maintenance of sex. *Nature* **411**, 689–692 (2001).

12. Whitlock, M. C. Fixation of new alleles and the extinction of small populations: Drift load, beneficial alleles, and sexual selection. *Evolution* **54**, 1855–1861 (2000).

13. Berger, D. *et al.* Intralocus sexual conflict and environmental stress. *Evolution* **68**, 2184–2196 (2014).

14. Bonduriansky, R. & Chenoweth, S. F. Intralocus sexual conflict. *Trends in Ecology & Evolution* **24**, 280–8 (2009).

15. Pennell, T. M. & Morrow, E. H. Two sexes, one genome: The evolutionary dynamics of intralocus sexual conflict. *Ecology and Evolution* **3**, 1819–1834 (2013).

16. Pischedda, A. & Chippindale, A. K. Intralocus sexual conflict diminishes the benefits of sexual selection. *PLOS Biology* **4**, e356 (2006).

17. Harano, T., Okada, K., Nakayama, S., Miyatake, T. & Hosken, D. J. Intralocus sexual conflict unresolved by sex-limited trait expression. *Current Biology* **20**, 2036–2039 (2010).

18. Plesnar Bielak, A., Skrzynecka, A. M., Miler, K. & Radwan, J. Selection for alternative male reproductive tactics alters intralocus sexual conflict. *Evolution* **68**, 2137–2144 (2014).

19. Berger, D. *et al.* Intralocus sexual conflict and the tragedy of the commons in seed beetles. *The American Naturalist* **188**, E98–E112 (2016).

20. Chenoweth, S. F., Appleton, N. C., Allen, S. L. & Rundle, H. D. Genomic evidence that sexual selection

411 impedes adaptation to a novel environment. *Current Biology* **25**, 1860–1866 (2015).

412 21. Rankin, D. J., Dieckmann, U. & Kokko, H. Sexual conflict and the tragedy of the commons. *The*
413 *American Naturalist* **177**, 780–791 (2011).

414 22. Fromhage, L., Elgar, M. A. & Schneider, J. M. Faithful without care: The evolution of monogyny.
415 *Evolution* **59**, 1400–1405 (2007).

416 23. Kokko, H. & Jennions Michael, D. Parental investment, sexual selection and sex ratios. *Journal of*
417 *Evolutionary Biology* **21**, 919–948 (2008).

418 24. Trivers, R. *Parental investment and sexual selection.* **136**, (Biological Laboratories, Harvard University,
419 1972).

420 25. Doherty, P. F. *et al.* Sexual selection affects local extinction and turnover in bird communities. *Proceedings*
421 *of the National Academy of Sciences of the United States of America* **100**, 5858–5862 (2003).

422 26. Morrow, E. H. & Fricke, C. Sexual selection and the risk of extinction in mammals. *Proceedings of the*
423 *Royal Society B-Biological Sciences* **271**, 2395–2401 (2004).

424 27. Morrow, E. H. & Pitcher, T. E. Sexual selection and the risk of extinction in birds. *Proceedings of the*
425 *Royal Society B-Biological Sciences* **270**, 1793–1799 (2003).

426 28. Martins, M. J. F., Puckett, T. M., Lockwood, R., Swaddle, J. P. & Hunt, G. High male sexual investment
427 as a driver of extinction in fossil ostracods. *Nature* **556**, 366–369 (2018).

428 29. Holman, L. & Jacomb, F. The effects of stress and sex on selection, genetic covariance, and the evolutionary
429 response. *Journal of Evolutionary Biology* **30**, 1898–1909 (2017).

430 30. Lewis, Z., Wedell, N. & Hunt, J. Evidence for strong intralocus sexual conflict in the Indian meal moth,
431 *Plodia interpunctella*. *Evolution* **65**, 2085–2097 (2011).

432 31. Martinossi-Allibert, I., Arnqvist, G. & Berger, D. Sex-specific selection under environmental stress in
433 seed beetles. *Journal of Evolutionary Biology* **30**, 161–173 (2017).

434 32. Martinossi-Allibert, I. *et al.* The consequences of sexual selection in well-adapted and maladapted
435 populations of bean beetles. *Evolution* **72**, 518–530 (2017).

436 33. Poissant, J., Wilson Alastair, J. & Coltman David, W. Sex-specific genetic variance and the evolution of
437 sexual dimorphism: A systematic review of cross-sex genetic correlations. *Evolution* **64**, 97–107 (2010).

438 34. Collet, J. M. *et al.* Rapid evolution of the intersexual genetic correlation for fitness in *Drosophila*
439 *melanogaster*. *Evolution* **70**, 781–795 (2016).

440 35. Connallon, T. & Hall, M. D. Genetic correlations and sex-specific adaptation in changing environments.
441 *Evolution* **70**, 2198 (2016).

442 36. Long, T. A. F., Agrawal, A. F. & Rowe, L. The effect of sexual selection on offspring fitness depends on
443 the nature of genetic variation. *Current Biology* **22**, 204–208 (2012).

444 37. Nakagawa, S., Noble, D. W. A., Senior, A. M. & Lagisz, M. Meta-evaluation of meta-analysis: Ten
445 appraisal questions for biologists. *BMC Biology* **15**, 18 (2017).

446 38. Senior, A. M. *et al.* Heterogeneity in ecological and evolutionary meta-analyses: Its magnitude and

implications. *Ecology* **97**, 3293–3299 (2016).

39. Nakagawa, S. *et al.* Meta-analysis of variation: Ecological and evolutionary applications and beyond. *Methods in Ecology and Evolution* **6**, 143–152 (2015).

40. Lau, J., Ioannidis, J. P. A., Terrin, N., Schmid, C. H. & Olkin, I. Evidence based medicine: The case of the misleading funnel plot. *BMJ: British Medical Journal* **333**, 597 (2006).

41. Jennions, M. D. & Møller, A. P. Relationships fade with time: A meta-analysis of temporal trends in publication in ecology and evolution. *Proceedings: Biological Sciences* **269**, 43–48 (2002).

42. Murtaugh, P. A. Journal quality, effect size, and publication bias in meta-analysis. *Ecology* **83**, 1162–1166 (2002).

43. Folstad, I. & Karter, A. J. Parasites, bright males, and the immunocompetence handicap. *The American Naturalist* **139**, 603–622 (1992).

44. Foo, Y. Z., Nakagawa, S., Rhodes, G. & Simmons, L. W. The effects of sex hormones on immune function: A meta-analysis. *Biological Reviews* **92**, 551–571 (2016).

45. Harts, A. M. F., Schwanz, L. E. & Kokko, H. Demography can favour female-advantageous alleles. *Proceedings of the Royal Society B: Biological Sciences* **281**, (2014).

46. Crowley, P. H. Sexual dimorphism with female demographic dominance: Age, size, and sex ratio at maturation. *Ecology* **81**, 2592–2605 (2000).

47. Lorch, P. D., Proulx, S., Rowe, L. & Day, T. Condition-dependent sexual selection can accelerate adaptation. *Evolutionary Ecology Research* **5**, 867–881 (2003).

48. Proulx, S. R. Matings systems and the evolution of niche breadth. *The American Naturalist* **154**, 89–98 (1999).

49. Proulx, S. R. Niche shifts and expansion due to sexual selection. *Evolutionary Ecology Research* **4**, 351–369 (2002).

50. Blows, M. W. & Hoffmann, A. A. A reassessment of genetic limits to evolutionary change. *Ecology* **86**, 1371–1384 (2005).

51. Lande, R. Quantitative genetic-analysis of multivariate evolution, applied to brain - body size allometry. *Evolution* **33**, 402–416 (1979).

52. Connallon, T. Genic capture, sex linkage, and the heritability of fitness. *The American Naturalist* **175**, 564–576 (2010).

53. Hastings, I. M. Manifestations of sexual selection may depend on the genetic basis of sex determination. *Proceedings of the Royal Society of London. Series B: Biological Sciences* **258**, 83 (1994).

54. Reeve, H. K. & Pfennig, D. W. Genetic biases for showy males: Are some genetic systems especially conducive to sexual selection? *Proceedings of the National Academy of Sciences* **100**, 1089 (2003).

55. Sturgill, D., Zhang, Y., Parisi, M. & Oliver, B. Demasculinization of X chromosomes in the Drosophila genus. *Nature* **450**, 238–241 (2007).

56. Charlesworth, B., Coyne, J. A. & Barton, N. H. The relative rates of evolution of sex chromosomes and

483 autosomes. *The American Naturalist* **130**, 113–146 (1987).

484 57. Charlesworth, B., Campos, J. L. & Jackson, B. C. Faster-X evolution: Theory and evidence from
485 Drosophila. *Molecular Ecology* **27**, 3753–3771 (2018).

486 58. Wyman, M. J. & Rowe, L. Male bias in distributions of additive genetic, residual, and phenotypic
487 variances of shared traits. *The American Naturalist* **184**, 326–337 (2014).

488 59. Ge, T., Chen, C.-Y., Neale, B. M., Sabuncu, M. R. & Smoller, J. W. Phenome-wide heritability analysis
489 of the UK Biobank. *PLOS Genetics* **13**, e1006711 (2017).

490 60. Agrawal, A. F. Are males the more 'sensitive' sex? *Heredity* **107**, 20–1 (2011).

491 61. Mallet, M. A. & Chippindale, A. K. Inbreeding reveals stronger net selection on *Drosophila melanogaster*
492 males: Implications for mutation load and the fitness of sexual females. *Heredity* **106**, 994–1002 (2011).

493 62. Mallet, M. A., Bouchard, J. M., Kimber, C. M. & Chippindale, A. K. Experimental mutation-accumulation
494 on the X chromosome of *Drosophila melanogaster* reveals stronger selection on males than females. *BMC*
495 *Evolutionary Biology* **11**, 156 (2011).

496 63. Sharp, N. P. & Agrawal, A. F. Male-biased fitness effects of spontaneous mutations in *Drosophila*
497 *melanogaster*. *Evolution* **67**, 1189–1195 (2012).

498 64. Li, X.-Y. & Holman, L. Evolution of female choice under intralocus sexual conflict and genotype-
499 by-environment interactions. *Philosophical Transactions of the Royal Society B: Biological Sciences* **373**,
500 20170425 (2018).

501 65. Gavrilets, S. Rapid evolution of reproductive barriers driven by sexual conflict. *Nature* **403**, 886 (2000).

502 66. Kokko, H. & Rankin, D. J. Lonely hearts or sex in the city? Density-dependent effects in mating systems.
503 *Philosophical Transactions of the Royal Society B: Biological Sciences* **361**, 319–334 (2006).

504 67. Martin, O. Y. & Hosken, D. J. Costs and benefits of evolving under experimentally enforced polyandry or
505 monogamy. *Evolution* **57**, 2765–2772 (2003).

506 68. Crudgington, H. S., Beckerman, A. P., Brustle, L., Green, K. & Snook, R. R. Experimental removal and
507 elevation of sexual selection: Does sexual selection generate manipulative males and resistant females? *The*
508 *American Naturalist* **165**, S72–S87 (2005).

509 69. Crudgington, H. S., Fellows, S. & Snook, R. R. Increased opportunity for sexual conflict promotes harmful
510 males with elevated courtship frequencies. *Journal of Evolutionary Biology* **23**, 440–446 (2010).

511 70. Crudgington, H. S., Fellows, S., Badcock, N. S. & Snook, R. R. Experimental manipulation of sexual
512 selection promotes greater male mating capacity but does not alter sperm investment. *Evolution* **63**, 926–938
513 (2009).

514 71. Holland, B. & Rice, W. R. Experimental removal of sexual selection reverses intersexual antagonistic
515 coevolution and removes a reproductive load. *Proceedings of the National Academy of Sciences of the United*
516 *States of America* **96**, 5083–5088 (1999).

517 72. Pitnick, S., Miller, G. T., Reagan, J. & Holland, B. Males' evolutionary responses to experimental removal

518 of sexual selection. *Proceedings of the Royal Society B-Biological Sciences* **268**, 1071–1080 (2001).

519 73. Connallon, T. The geography of sex-specific selection, local adaptation, and sexual dimorphism. *Evolution*
520 **69**, 2333–2344 (2015).

521 74. Punzalan, D., Delcourt, M. & Rundle, H. D. Comparing the intersex genetic correlation for fitness across
522 novel environments in the fruit fly, *Drosophila serrata*. *Heredity* **112**, 143 (2013).

523 75. Delcourt, M., Blows, M. W. & Rundle, H. D. Sexually antagonistic genetic variance for fitness in an
524 ancestral and a novel environment. *Proceedings of the Royal Society B: Biological Sciences* **276**, 2009 (2009).

525 76. Kraaijeveld, K., Kraaijeveld-Smit, F. J. L. & Maan, M. E. Sexual selection and speciation: The
526 comparative evidence revisited. *Biological Reviews* **86**, 367–377 (2011).

527 77. Lehtonen, J., Jennions, M. D. & Kokko, H. The many costs of sex. *Trends in Ecology & Evolution* **27**,
528 172–8 (2012).

529 78. Kleiman, M. & Hadany, L. The evolution of obligate sex: The roles of sexual selection and recombination.
530 *Ecology and Evolution* **5**, 2572–83 (2015).

531 79. Charge, R., Teplitsky, C., Sorci, G. & Low, M. Can sexual selection theory inform genetic management of
532 captive populations? A review. *Evolutionary Applications* **7**, 1120–1133 (2014).

533 80. Russell, T. *et al.* MHC diversity and female age underpin reproductive success in an Australian icon; the
534 Tasmanian Devil. *Scientific Reports* **8**, 4175 (2018).

535 81. Cunningham, E. J. A. & Russell, A. F. Egg investment is influenced by male attractiveness in the mallard.
536 *Nature* **404**, 74 (2000).

537 82. Loyau, A. & Lacroix, F. Watching sexy displays improves hatching success and offspring growth through
538 maternal allocation. *Proceedings of the Royal Society B: Biological Sciences* **277**, 3453 (2010).

539 83. Loyau, A., Saint Jalme, M., Mauget, R. & Sorci, G. Male sexual attractiveness affects the investment
540 of maternal resources into the eggs in peafowl (*Pavo cristatus*). *Behavioral Ecology and Sociobiology* **61**,
541 1043–1052 (2007).

542 84. Arellano-Aguilar, O. & Macias Garcia, C. Exposure to pesticides impairs the expression of fish ornaments
543 reducing the availability of attractive males. *Proceedings of the Royal Society B: Biological Sciences* **275**,
544 1343–1351 (2008).

545 85. Botha, L. M., Jones, T. M. & Hopkins, G. R. Effects of lifetime exposure to artificial light at night on
546 cricket (*Teleogryllus commodus*) courtship and mating behaviour. *Animal Behaviour* **129**, 181–188 (2017).

547 86. Candolin, U., Salesto, T. & Evers, M. Changed environmental conditions weaken sexual selection in
548 sticklebacks. *Journal of Evolutionary Biology* **20**, 233–239 (2007).

549 87. Swaddle, J. P. & Page, L. C. High levels of environmental noise erode pair preferences in zebra finches:
550 Implications for noise pollution. *Animal Behaviour* **74**, 363–368 (2007).

551 88. Wong, B. B. M., Candolin, U. & Lindström, K. Environmental deterioration compromises socially enforced
552 signals of male quality in three-spined sticklebacks. *The American Naturalist* **170**, 184–189 (2007).

553 89. Knell, R. J. & Martínez-Ruiz, C. Selective harvest focused on sexual signal traits can lead to extinction

554 under directional environmental change. *Proceedings of the Royal Society B: Biological Sciences* **284**, (2017).

555 90. Rohatgi, A. WebPlotDigitalizer: HTML5 based online tool to extract numerical data from plot images.
556 *https://automeris.io/WebPlotDigitizer/* **Version 3.17**, (2011).

557 91. Hedges, L. V. & Olkin, I. *Statistical methods for meta-analysis.* (Academic Press, Inc, 1985).

558 92. Rosenberg, M. S., Rothstein, H. R. & Gurevitch, J. Effect sizes: Conventional choices and calculations.
559 in *Handbook of meta-analysis in ecology and evolution* (eds. Koricheva, J., Gurevitch, J. & Mengersen, K.) p.
560 61–71 (Princeton University Press, 2013).

561 93. Del Re, M. Compute.es: Compute effect sizes. *R Package* **0.2-4**, (2013).

562 94. Firman, R. C., Gomendio, M., Roldan, E. R. S. & Simmons, L. W. The coevolution of ova defensiveness
563 with sperm competitiveness in house mice. *The American Naturalist* **183**, 565–572 (2014).

564 95. Plesnar-Bielak, A., Skrzynecka, A. M., Prokop, Z. M. & Radwan, J. Mating system affects population
565 performance and extinction risk under environmental challenge. *Proceedings of the Royal Society B-Biological*
566 *Sciences* **279**, 4661–4667 (2012).

567 96. Hedges, L. V., Gurevitch, J. & Curtis, P. S. The meta-analysis of response ratios in experimental ecology.
568 *Ecology* **80**, 1150–1156 (1999).

569 97. Osenberg, C. W., Sarnelle, O. & Cooper, S. D. Effect size in ecological experiments: The application of
570 biological models in meta-Analysis. *The American Naturalist* **150**, 798–812 (1997).

571 98. Senior, A. M., Gosby, A. K., Lu, J., Simpson, S. J. & Raubenheimer, D. Meta-analysis of variance: An
572 illustration comparing the effects of two dietary interventions on variability in weight. *Evolution, Medicine,*
573 *and Public Health* **2016**, 244–255 (2016).

574 99. Higgins, J. P. T. & Thompson, S. G. Quantifying heterogeneity in a meta-analysis. *Statistics in Medicine*
575 **21**, 1539–1558 (2002).

576 100. Viechtbauer, W. Conducting meta-analyses in R with the metafor package. *Journal of Statistical*
577 *Software* **36**, 1–48 (2010).

578 101. Bürkner, P.-C. Brms: An R package for Bayesian multilevel models using Stan. *Journal of Statistical*
579 *Software* **80**, 1–28 (2016).

580 102. Egger, M., G., D. S., Schneider, M. & Minder, C. Bias in meta-analysis detected by a simple, graphical
581 test. *BMJ* **315**, 629–634 (1997).

582 103. Sterne, J. A. C. & Egger, M. Regression methods to detect publication and other bias in meta-analysis.
583 in *Publication bias in meta-analysis* (eds. Rothstein, H. R., Sutton, A. J. & Borenstein, M.) **1**, 99–110 (John
584 Wiley & Sons, Ltd, 2005).

Acknowledgements

We are very grateful to Paco Garcia-Gonzalez for sharing a collection of literature that helped to guide our study, and to Tim Connallon for comments on the manuscript.

Author contributions

All authors designed the study; JGC collected the data; JGC and LH conducted the meta-analyses; JGC wrote the first draft of the manuscript; and all authors contributed substantially to all further revisions.

Competing interests

The authors declare no competing interests.

Tables and Figures

Table 1: Multilevel meta-analysis model results

Parameters	Estimate	SE	LCI	UCI	z	p
REML Model						
Intercept	0.19	0.12	-0.05	0.43	1.53	0.13
Both Sexes	0	0.07	-0.14	0.15	0.04	0.97
Female Sex	**0.11**	0.03	0.05	0.17	3.73	0.00
Stressed Environment	**-0.16**	0.04	-0.24	-0.07	-3.63	0.00
Both Sexes x Stressed Environment	**0.18**	0.09	0.01	0.36	2.07	0.04
Female Sex x Stressed Environment	**0.26**	0.05	0.16	0.37	5.11	0.00
Bayesian Model						
Intercept	0.188	0.18	-0.17	0.52		
Both Sexes	0.003	0.07	-0.14	0.14		
Female Sex	**0.113**	0.03	0.05	0.17		
Stressed Environment	**-0.156**	0.04	-0.24	-0.07		
Both Sexes x Stressed Environment	**0.182**	0.09	0.01	0.35		
Female Sex x Stressed Environment	**0.264**	0.05	0.16	0.37		

* Moderator variables whose 95% confidence intervals do not cross zero are shown in bold.
† Hedges' g is the response variable for this model

Table 2: Hypothesis tests showing how sex and environmental stress interact to modulate effect size

Condition	Test	Statistical Approach	Estimate	SE	LCI	UCI
In benign environments	Female > Male	**REML**	**0.113**	0.03	0.05	0.17
		Bayesian	**0.113**	0.03	0.05	0.17
In stressful environments		**REML**	**0.377**	0.05	0.29	0.47
		Bayesian	**0.377**	0.05	0.29	0.47
For Females	Stressful > Benign	**REML**	**0.108**	0.04	0.04	0.18
		Bayesian	**0.109**	0.04	0.04	0.18
For Males	Benign > Stressful	**REML**	**0.156**	0.04	0.07	0.24
		Bayesian	**0.156**	0.04	0.07	0.24
For Both	Stressful > Benign	REML	0.028	0.08	-0.13	0.18
		Bayesian	0.026	0.08	-0.13	0.18

* Moderator variables whose 95% confidence intervals do not cross zero are shown in bold.
† Hedges' g is the response variable for this model

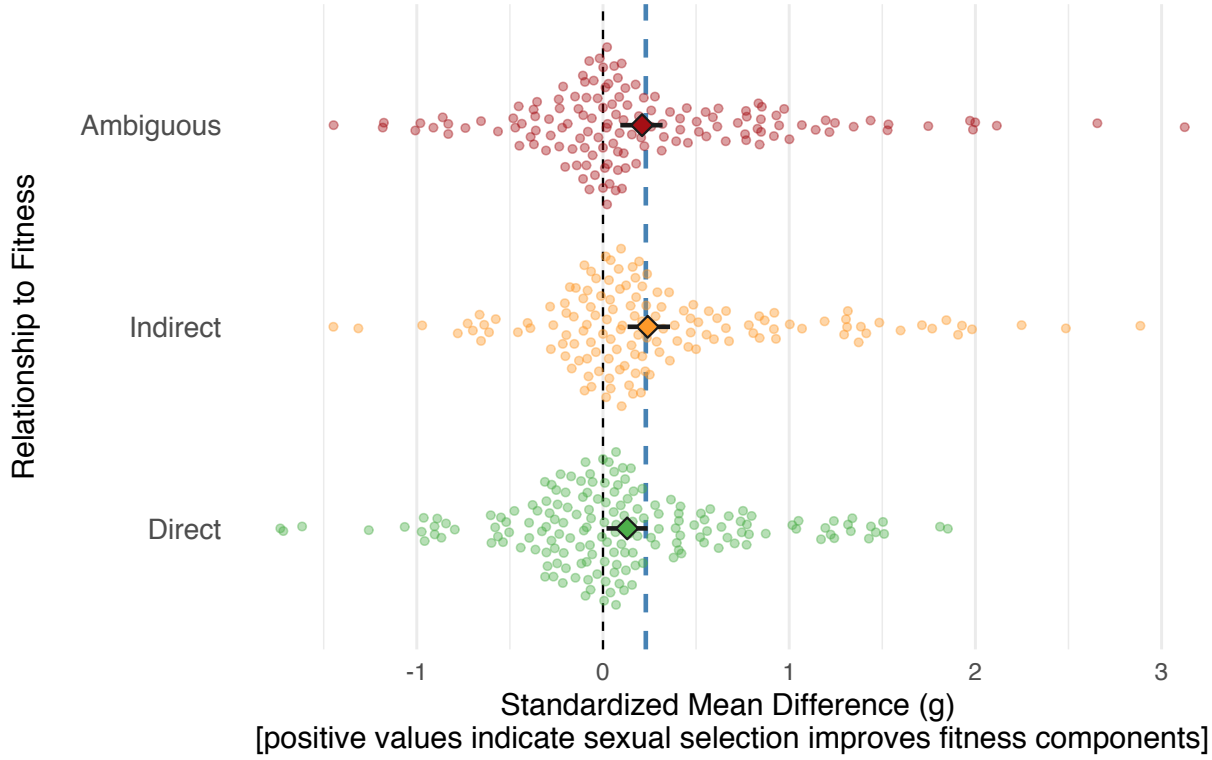

Figure 1: The effect of sexual selection on direct, indirect and ambiguous fitness components. The effect sizes used in this meta-analysis ($n = 459$) were grouped into either direct, indirect or ambiguous measures of fitness. Overall, effect sizes were more often positive than negative. Predicted average values are presented as a diamond with 95 % CIs for each fitness-relationship category. The estimates presented here are from REML models with the grand mean across all effect sizes ($\beta = 0.25$) shown as the blue dotted line. Predictions from both Bayesian and REML models can be found in Table S7).

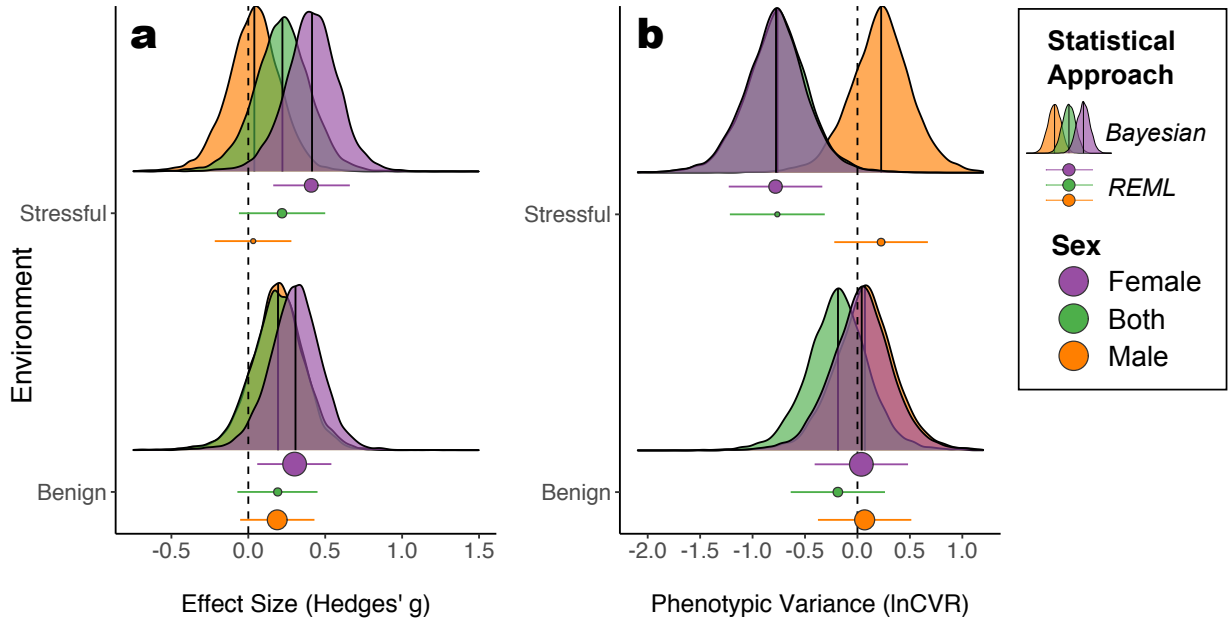

Figure 2: The roles of environmental stress and sex. (a) Sexual selection tends to increase the population mean values of fitness traits, especially for female traits and for populations living under stressful conditions. (b) Under stressful conditions, sexual selection tends to reduce the phenotypic variance in fitness traits, especially for traits measured in females or mixed-sex individuals. The points with error bars show the mean effect sizes and their 95% CIs, determined from a meta-regression fit using restricted maximum likelihood (REML); the point sizes are proportional to the number of effect sizes (see Table S12 and S14). Results from Bayesian meta-regression are shown as posterior prediction density curves, with vertical lines indicating the median.

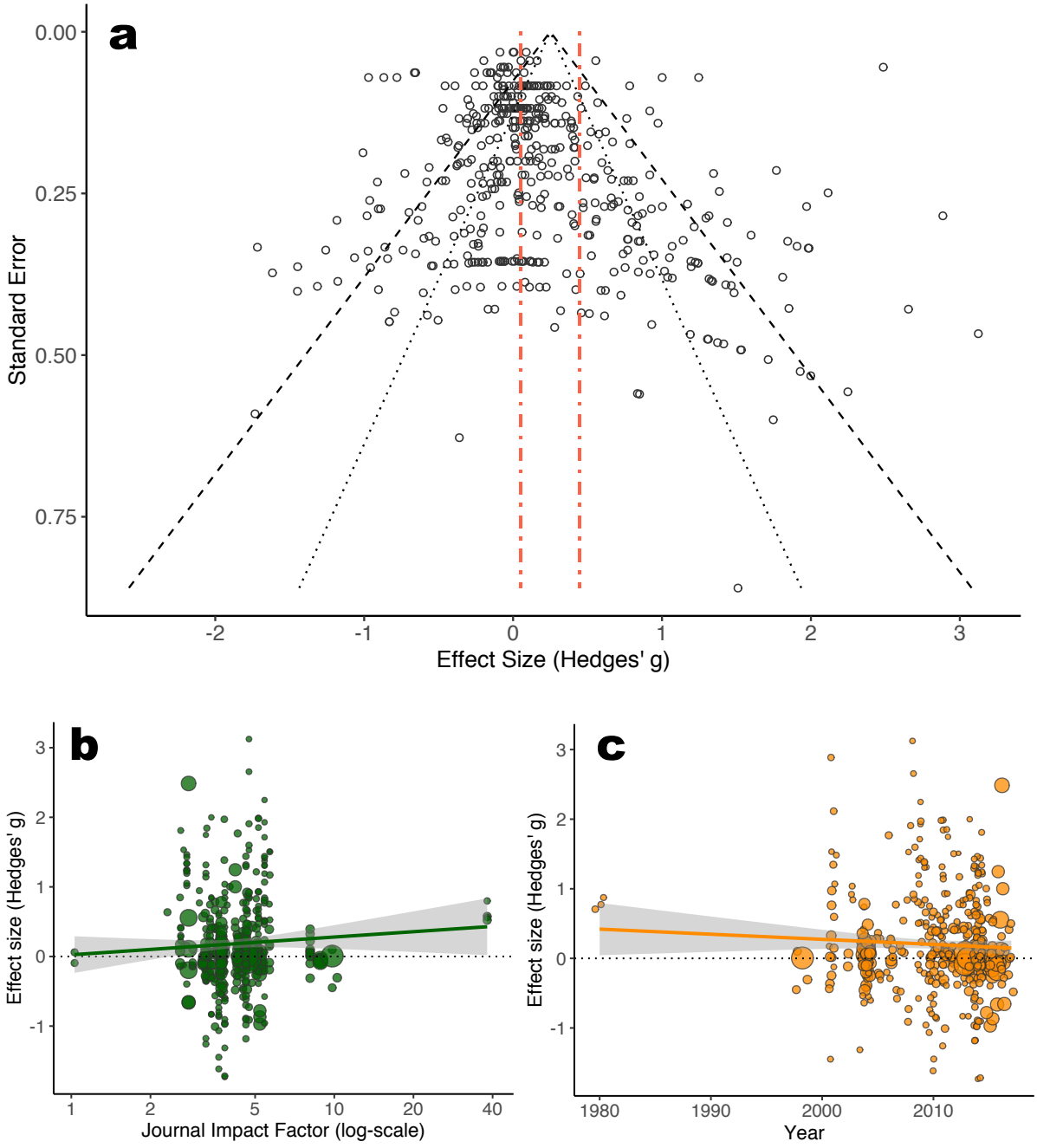

Figure 3: Tests for publication bias in the dataset. Tests of publication bias are mixed and suggest publication bias may be present. (a) Inspection and statistical tests of the funnel plot reveal large amounts of heterogeneity in the dataset with asymmetry from increased low-powered, large effect studies. (b) No significant correlation exists between journal impact factor and effect size, (c) additionally there is no significant correlation between effect size and year of publication when testing for the time-lag bias. For (b) and (c) point size is proportional to the precision of the effect size (*i.e.* the inverse of its variance). For (a), the dashed red lines represent the 95 % CIs of the grand mean estimate for all effect sizes and the black dotted and dashed lines depict the 95 % and 99 % CIs for the dataset. The Grey envelopes in (b) and (c) represent the 95 % CI of the linear regression.

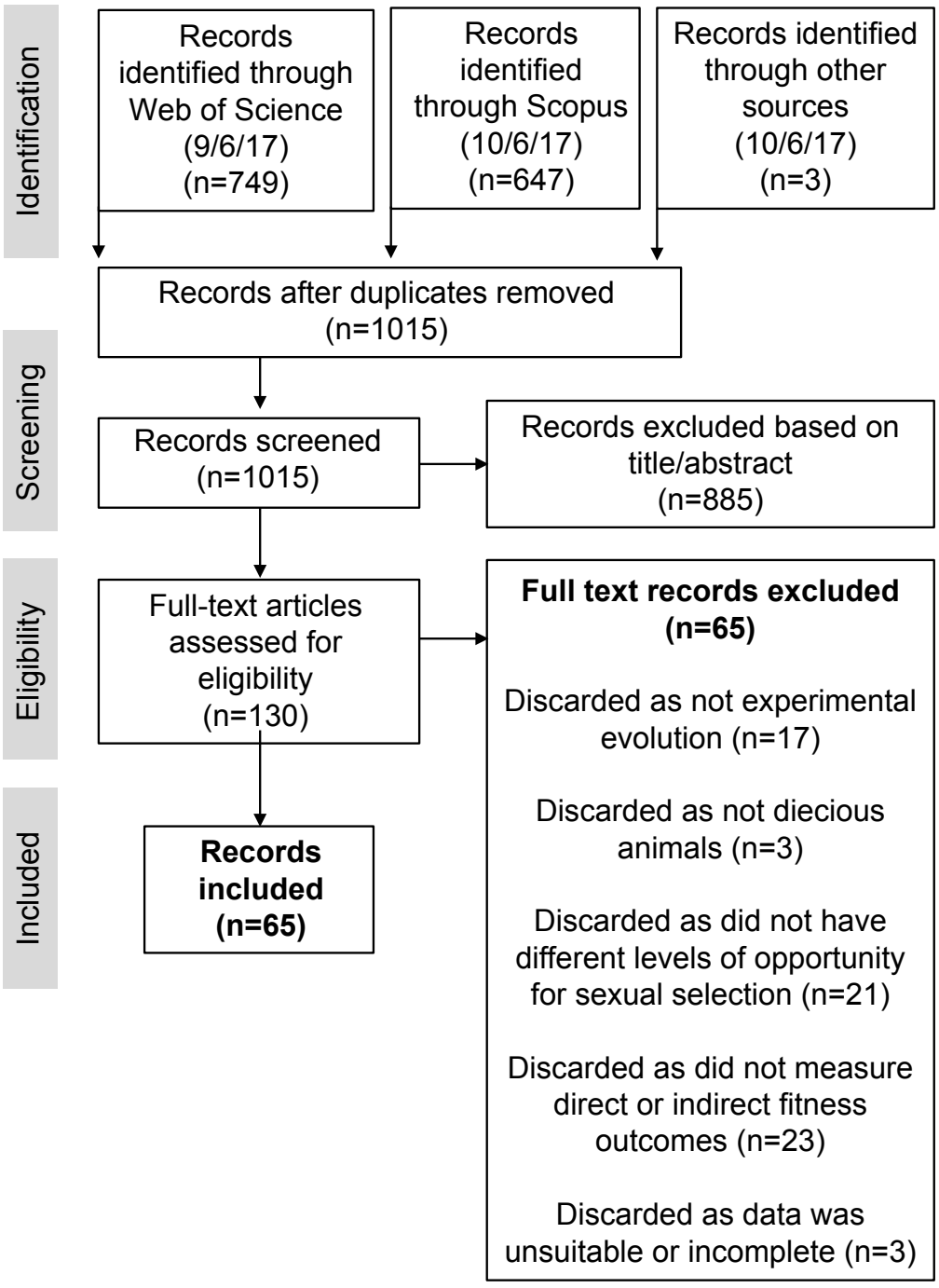

Figure 4: PRISMA diagram. Flow of inclusion and exclusion of studies identified during the literature search, presented as a PRISMA diagram with number of published papers in brackets.