

# NK cell identification from GBM data

[Code ▾](#)

The GBM data set is downloaded from GEO (GSE84465), including 3587 cells from four primary GBM patients. Darmanis S, Sloan SA, Croote D, Mignardi M et al. Single-Cell RNA-Seq Analysis of Infiltrating Neoplastic Cells at the Migrating Front of Human Glioblastoma. Cell Rep 2017 Oct 31;21(5):1399-1410. PMID: 29091775

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```
GBM=read.csv('GSE84465_GBM_All_data.csv',sep=" ")
```

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```
dim(GBM)
```

```
[1] 23465 3589
```

Run Seurat to identify cluster enriched NK cell

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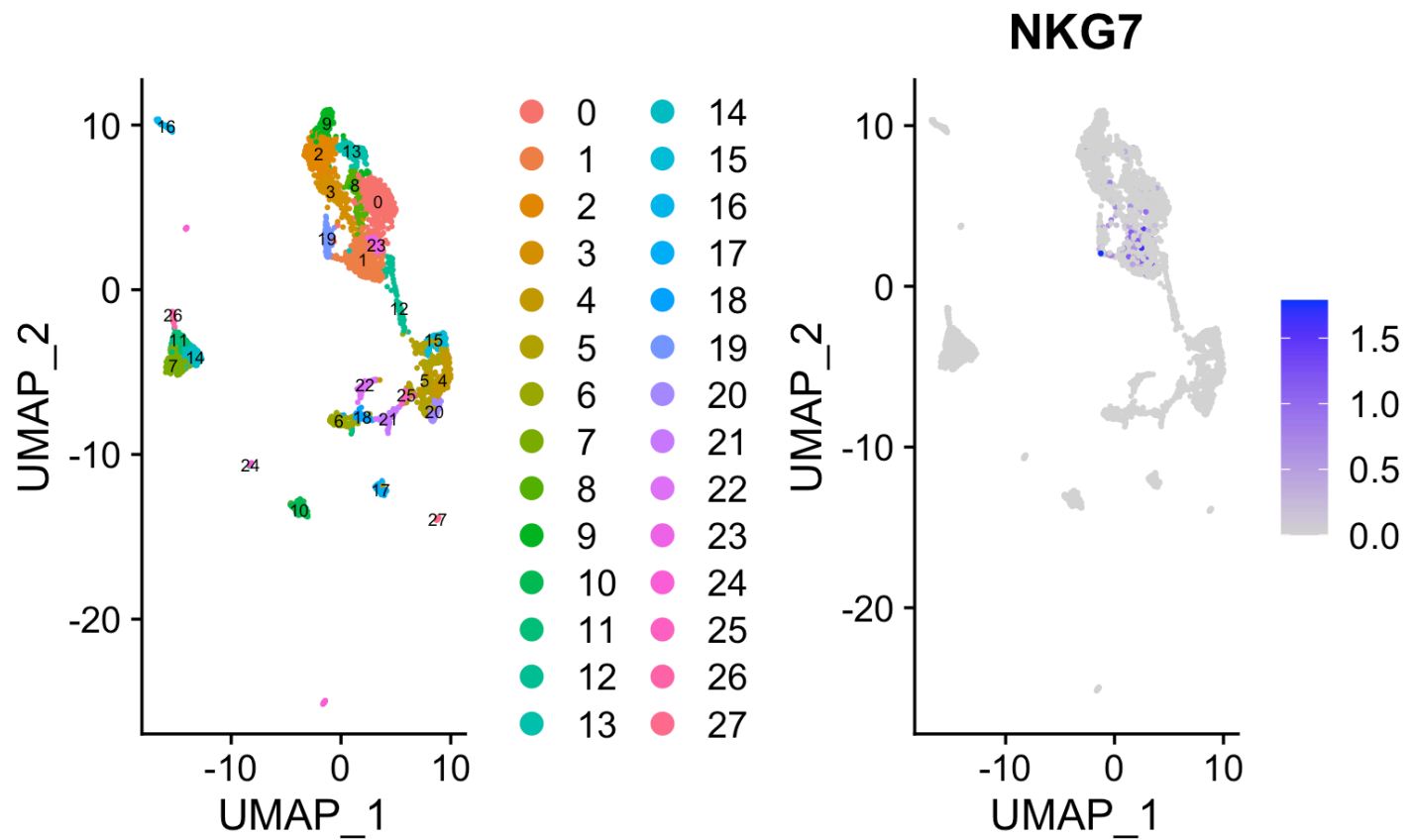
```
library(dplyr)
library(Seurat)
library(cowplot)
library(ggplot2)
```

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```
GBMobj <- CreateSeuratObject(counts = GBM, project = "GBM")
GBMobj <- NormalizeData(object = GBMobj, verbose = FALSE)
GBMobj <- FindVariableFeatures(object = GBMobj, selection.method = "vst", nfeatures = 2000)
GBMobj <- ScaleData(GBMobj)
GBMobj <- RunPCA(GBMobj)
GBMobj <- FindNeighbors(GBMobj, dims = 1:25)
GBMobj <- FindClusters(GBMobj, resolution = 1.7)
GBMobj <- RunUMAP(GBMobj, dims = 1:25)
```

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```
p1 <- DimPlot(GBMobj,label = TRUE,pt.size = 0.2,label.size=2)
p2 <- FeaturePlot(GBMobj, features = c("NKG7"),pt.size=0.2)
p1+p2
```

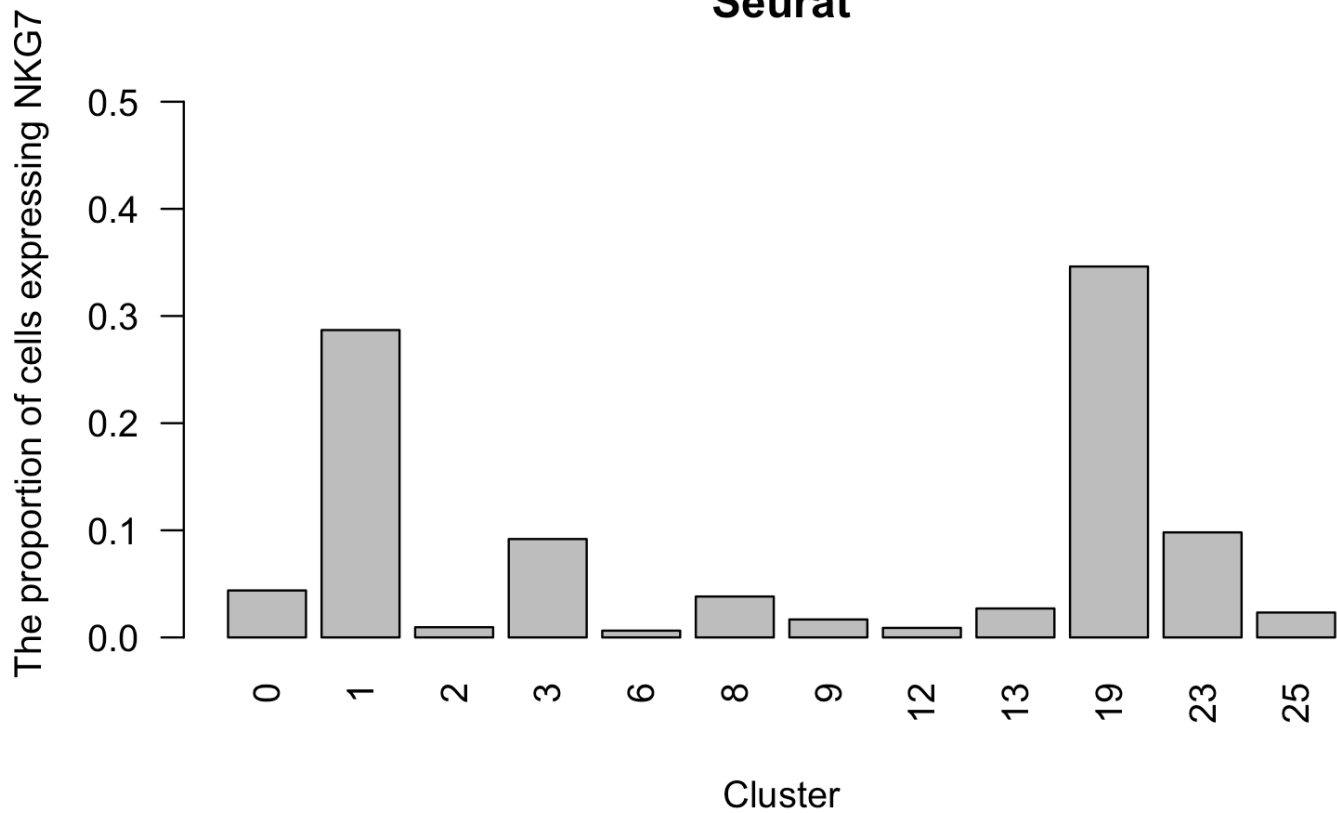


The proportion of cells expressing NK cell marker NKG7 in each cluster

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```
subGBM=subset(GBMobj,NKG7>0)
index=match(names(table(Idsents(subGBM))),names(table(Idsents(GBMobj))))
seurat=table(Idsents(subGBM))/table(Idsents(GBMobj))[index]
barplot(seurat,ylim = c(0,0.55),las=2,main='Seurat',xlab="Cluster",ylab="The proportion of cells expressing NKG7")
```

## Seurat



### Integration of Seurat and SCMarker

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```
library(SCMarker)
GBM1=log(GBM+1)
scMarker.res=ModalFilter(data=GBM1, geneK=10, cellK=10, width=2) # default width = 1 for
UMI data, width =2 for TPM data.
scMarker.res=GeneFilter(obj=scMarker.res)
scMarker.res=getMarker(obj=scMarker.res, k=300, n=30)
```

### Extract markers identified by SCMarker

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```
scmarker=scMarker.res$marker
length(scmarker)
```

```
[1] 840
```

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```
head(scmarker)
```

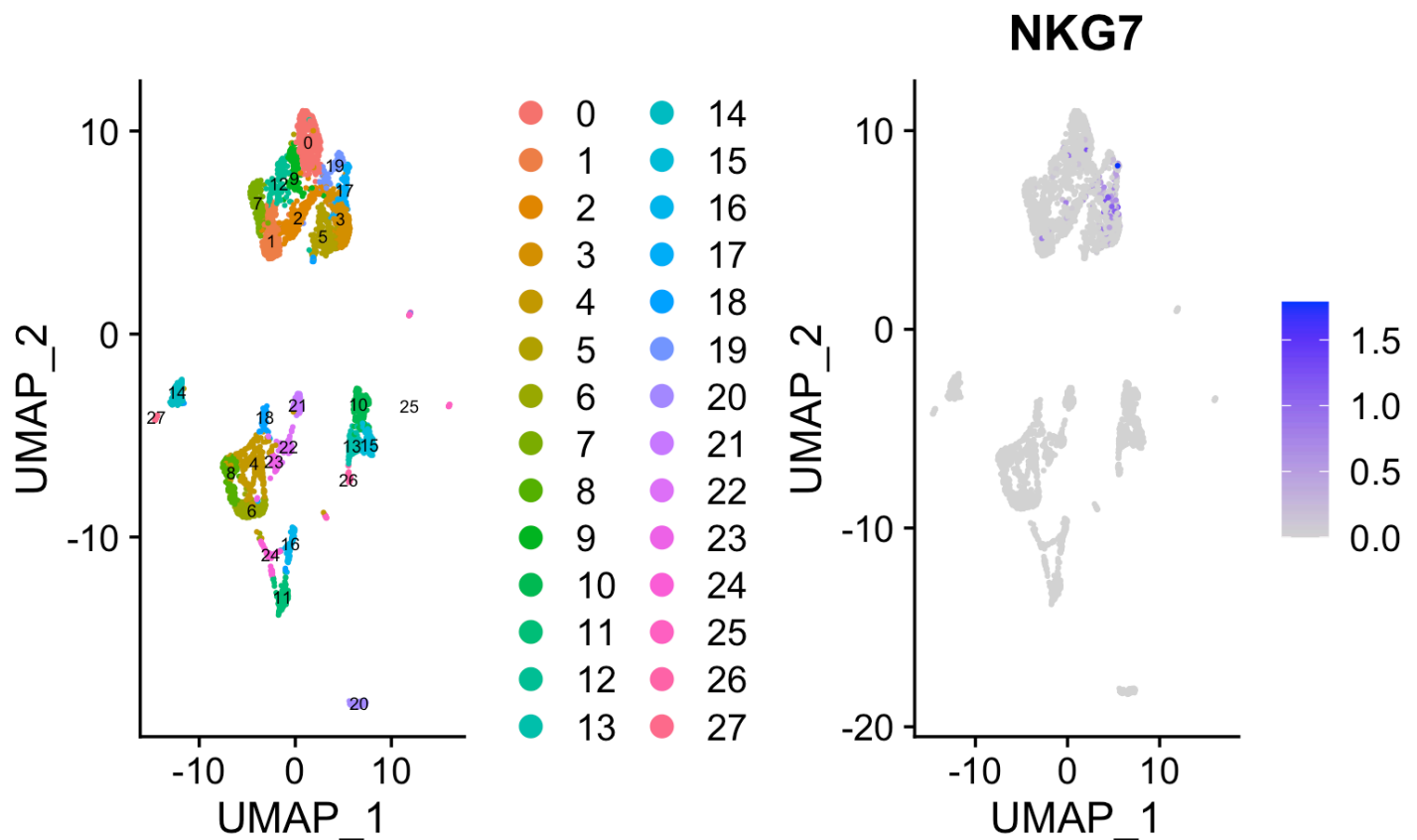
```
[1] "ABL2" "ACSL1" "ADORA3" "AIF1" "AKR1B1" "ALOX5"
```

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```
all.genes <- rownames(GBMobj)
GBMscmarker <- ScaleData(GBMobj, features = all.genes)
###Run PCA based on union of markers from SCMarker and Seurat
GBMscmarker <- RunPCA(GBMscmarker, features = union(scmarker, VariableFeatures(GBMobj))
)
GBMscmarker <- FindNeighbors(GBMscmarker, dims = 1:25)
GBMscmarker <- FindClusters(GBMscmarker, resolution = 1.7)
GBMscmarker <- RunUMAP(GBMscmarker, dims = 1:25)
```

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```
p1 <- DimPlot(GBMscmarker, label = TRUE, pt.size = 0.2, label.size=2)
p2 <- FeaturePlot(GBMscmarker, features = c("NKG7"), pt.size=0.2)
p1+p2
```



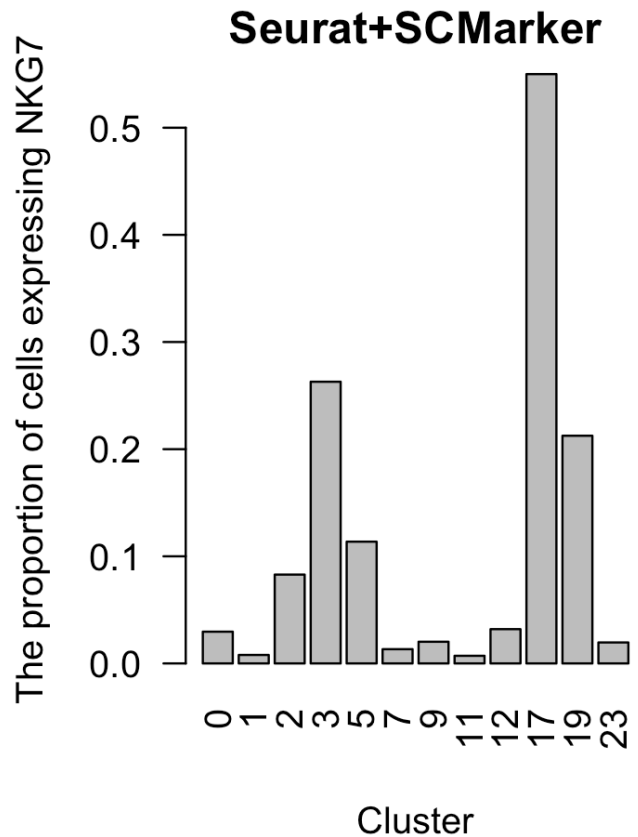
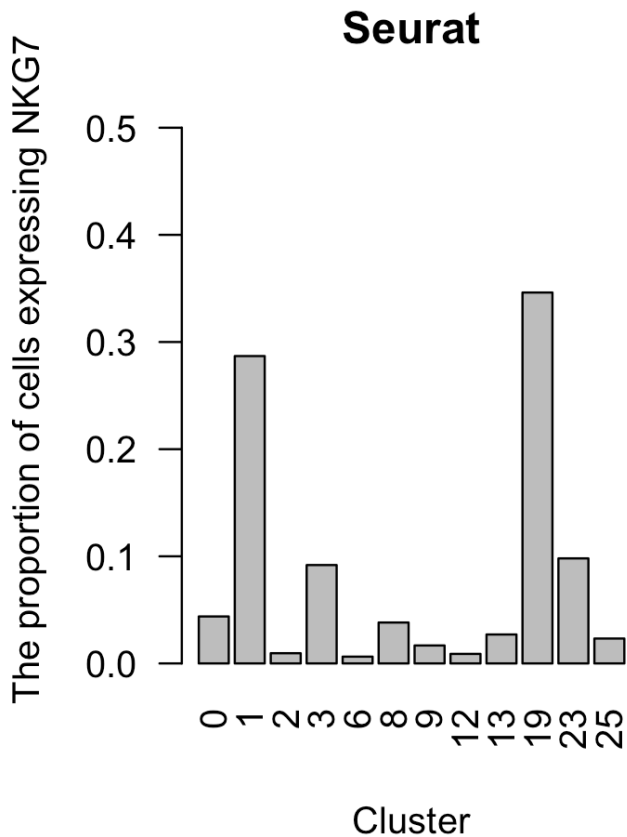
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```

subGBMmarker=subset(GBMscmarker,NKG7>0)
index=match(names(table(Idsents(subGBMmarker))),names(table(Idsents(GBMscmarker))))
scmarkercluster=table(Idsents(subGBMmarker))/table(Idsents(GBMscmarker))[index]
par(mfrow=c(1,2))
barplot(seurat,ylim = c(0,0.55),las=2,main='Seurat',xlab="Cluster",ylab="The proportion of cells expressing NKG7")
barplot(scmarkercluster,ylim = c(0,0.55),las=2,main='Seurat+SCMarker',xlab="Cluster",ylab="The proportion of cells expressing NKG7")

```



FindMarker for each cluster

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```

cluster=unique(Ids(GBMscmarker))
cluster=as.numeric(as.character(cluster[order(cluster)]))
gene=c()
for (i in cluster){
  cluster1.markers <- FindMarkers(GBMscmarker, only.pos = TRUE,ident.1 = i, min.pct =
0.25)
  if (length(grep("ERCC-",row.names(cluster1.markers)))!=0){
    cluster1.markers=cluster1.markers[-grep("ERCC-",row.names(cluster1.markers)),]
  }
  cluster1.markers=cluster1.markers[order(cluster1.markers$p_val_adj),]
  gene=union(gene[!is.na(gene)],row.names(cluster1.markers)[1:6])
}

```

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```
"NKG7" %in% gene
```

```
[1] TRUE
```

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```

DoHeatmap(GBMscmarker, features = gene,size=3) + NoLegend()+ theme(axis.text.y = element_text(size = 3))

```

