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Reef fishes can recognize bleached habitat during settlement: sea anemone bleaching alters anemonefish host selection

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Understanding how bleaching impacts the settlement of symbiotic habitat specialists and whether there is flexibility in settlement choices with regard to habitat quality is essential given our changing climate. We used five anemonefishes (*Amphiprion clarkii*, *Amphiprion latezonatus*, *Amphiprion ocellaris*, *Amphiprion percula* and *Premnas biaculeatus*) and three host sea anemones (*Entacmaea quadricolor*, *Heteractis crispa* and *Heteractis magnifica*) in paired-choice flume experiments to determine whether habitat naive juveniles have the olfactory capabilities to distinguish between unbleached and bleached hosts, and how this may affect settlement decisions. All anemonefishes were able to distinguish between bleached and unbleached hosts, and responded only to chemical cues from species-specific host anemones irrespective of health status, indicating a lack of flexibility in host use. While bleached hosts were selected as habitat, this occurred only when unbleached options were unavailable, with the exception of *A. latezonatus*, which showed strong preferences for *H. crispa* regardless of health. This study highlights the potential deleterious indirect impacts of declining habitat quality during larval settlement in habitat specialists, which could be important in the field, given that bleaching events are becoming increasingly common.

1. Introduction

Coral reefs are becoming increasingly degraded, with local stressors such as declining water quality, overfishing, and outbreaks of invasive species and predators such as crown-of-thorns starfish being exacerbated by the global stressors of climate change and ocean acidification [1–4]. Ocean warming threatens the structure and function of coral reefs through episodes of bleaching, which are increasing in frequency and magnitude due to anthropogenic activity [5]. Bleaching is a general stress response that can be triggered by a variety of causes, and affects a range of organisms, including corals, sea anemones and clams [6,7]. It is characterized by the breakdown of the symbiosis between the animal host and its unicellular dinoflagellate algae (*Symbiodinium* spp.). Algae are expelled from the host and/or the photosynthetic pigments from within the symbionts are reduced, which leads to the animal becoming pale or white in appearance [6–8]. The breakdown in this relationship has implications for host health as these algae provide photosynthate that is used to supply a range of metabolic needs [9]. When stressors are extreme or last for an extended duration, the host may bleach and die; however, when conditions are less severe or persist for a shorter period, the host may bleach, remain alive and reacquire a healthy algal population [6].

While bleaching has obvious direct effects on the impacted organisms, the indirect effects on the ecosystem and other associated species, such as fishes, often vary [10–13]. For species that form obligate symbioses with microhabitats

that are susceptible to bleaching, the impacts are amplified, as habitat quality and health are directly linked to survival. The symbiotic relationship among anemonefishes and their host sea anemones provides an ideal model for further exploring the consequences of habitat degradation, as the former are generally unable to survive in the wild without their hosts [14]. This association occurs on suitable reefs throughout the Indo-Pacific, in both shallow and mesophotic waters [15,16], with the greatest species richness occurring in Papua New Guinea [17]. Host specificity varies among the 28 species of anemonefishes, with some being extreme specialists that use only one host species and others being able to use a range or all 10 hosts [14,15].

All host anemones are susceptible to bleaching, and reductions in the abundance of both the anemones and their resident fish have been recorded following high-temperature anomalies [18–20]. Thus, flexibility in host use by anemonefishes may be crucial for survival during severe habitat degradation and subsequent community shifts [21]. Such plasticity is regularly observed in captivity, where anemonefishes can reside in unnatural sea anemone hosts, and even use other invertebrates such as soft corals as habitat [15,21]. In artificial situations, predation pressure is removed and therefore the protection provided by the anemones to the fish may no longer be vital. However, there has been one long-term field observation of *Amphiprion clarkii* using the soft coral *Lobophytum* sp. after bleaching impacted anemone populations in Japan [21]. As adult anemonefishes are highly site-attached [14,22], this may have occurred due to the need to find habitat within close proximity to the original settlement site.

The replenishment and persistence of coral reef fishes, such as anemonefishes, are reliant upon dispersing individuals finding and becoming established in a suitable habitat. The majority of reef fishes begin life as larvae that disperse among isolated adult populations [23]. The ability to find suitable habitat after this pelagic larval period represents a significant challenge. As predation pressure is estimated to be greatest during settlement [24], and inappropriate habitat selection has critical repercussions to fitness, survival and reproductive success, this process should be subject to strong selection pressure. Coral reef fish larvae have well-developed sensory and swimming capabilities [25], and evidence suggests that both sound [26,27] and olfactory cues such as those derived from resident conspecifics, coral tissues or symbiotic partners [28–35] are used to locate suitable habitat. Field observations and laboratory experiments have shown that larval recruitment of fishes, especially habitat specialists, is impacted by episodes of habitat degradation and bleaching [11–13,36].

Available settlement sites for recruiting anemonefishes will decrease as the incidence and extent of habitat degradation increases. While it is well documented that anemonefishes are able to distinguish among host anemones using chemical cues [34,35,37], it is not known if chemical cues are used by fishes to determine the bleaching status of the symbiotic hosts and if host health impacts settlement preferences. We therefore address the following questions. (i) Can juvenile anemonefishes distinguish between unbleached and bleached host anemones using chemical cues alone? (ii) Will a bleached anemone be selected as habitat? (iii) Are settlement preferences flexible when known host species are bleached and other unbleached non-host species are available?

Table 1. Associations formed by anemonefishes and sea anemone species used in the experiments. Numbers in brackets refer to the number of anemones or anemonefishes that associate with that particular species; tick marks indicate known associations; crosses indicate associations that do not occur in the field [15,38–40].

	<i>E. quadricolor</i> (13)	<i>H. crispa</i> (14)	<i>H. magnifica</i> (12)
<i>A. clarkii</i> (10)	✓	✓	✓
<i>A. latezonatus</i> (3)	✓	✓	✗
<i>A. ocellaris</i> (3)	✗	✗	✓
<i>A. percula</i> (3)	✗	✓	✓
<i>P. biaculeatus</i> (1)	✓	✗	✗

2. Material and methods

Five species of anemonefishes (*Amphiprion clarkii*, *A. latezonatus*, *A. ocellaris*, *A. percula* and *Premnas biaculeatus*) and three species of sea anemone (*Entacmaea quadricolor*, *Heteractis crispa* and *H. magnifica*) were tested to determine the role that chemical cues play in habitat recognition, and the potential settlement preferences of fishes towards unbleached versus bleached host and non-host species (table 1). Using Atema choice flume [28] experiments, we specifically tested responses by fishes to: (i) unbleached anemones versus seawater; (ii) bleached anemones versus seawater; (iii) unbleached versus bleached anemones of the same species; (iv) unbleached anemones versus bleached of different species; and (v) bleached versus bleached anemones of different species.

Experiments were done at Georgia Institute of Technology (GT), USA from the 12–24 October 2014. Each trial used 18–20 habitat naive captive-bred juveniles (all less than two months post-hatching), with a total of between 324 and 360 individuals per species. Fishes were reared at Sustainable Aquatics (Tennessee, USA) from three different breeding pairs of each species to account for genetic variability. Settlement stage juveniles were transported to GT and each species group was housed in individual 10 l aquaria within a 382 l recirculating seawater system with a 13 L:11 D cycle. Before experimentation, fish were fed daily with 0.8 mm dry pellet hatchery diet (Sustainable Aquatics, TN, USA). Water quality measurements were taken daily and 50% water changes were done twice a week to ensure water quality was maintained. Seawater (26°C) used for maintaining the study species and in the experiments was artificial (30 psu, Instant Ocean, Blacksburg, Virginia). Once trials were completed, fishes were placed in different aquaria to ensure they were only used once.

Two unbleached and two bleached individuals (oral disc diameter more than 10 cm) of each species were soaked in 10 l of water for 2 h to generate the chemical cues [35]. The cues created were then used immediately in the trials. Anemones were sourced from a local saltwater aquarium supplier (The Fish Store, <http://www.thefishstore.com>; collection locations remain unknown), and housed in closed 90 l aquaria with carbon filtration, a protein skimmer (Coralife Biocube, Central Aquatics, WI, USA) and actinic lights on a 13 L:11 D cycle. Species were held separately without fish and fed twice weekly with prawn meat. Bleaching status was visually assessed using the colour health monitoring chart developed by Siebeck *et al.* [41]. Unbleached anemones had highly pigmented tentacles, whereas bleached anemones had tentacles that were white and lacked visible pigment. In some cases, anemones arrived bleached from the supplier, whereas in others bleaching was induced by gradually ramping the temperature (1–2°C per day) until full bleaching was achieved.

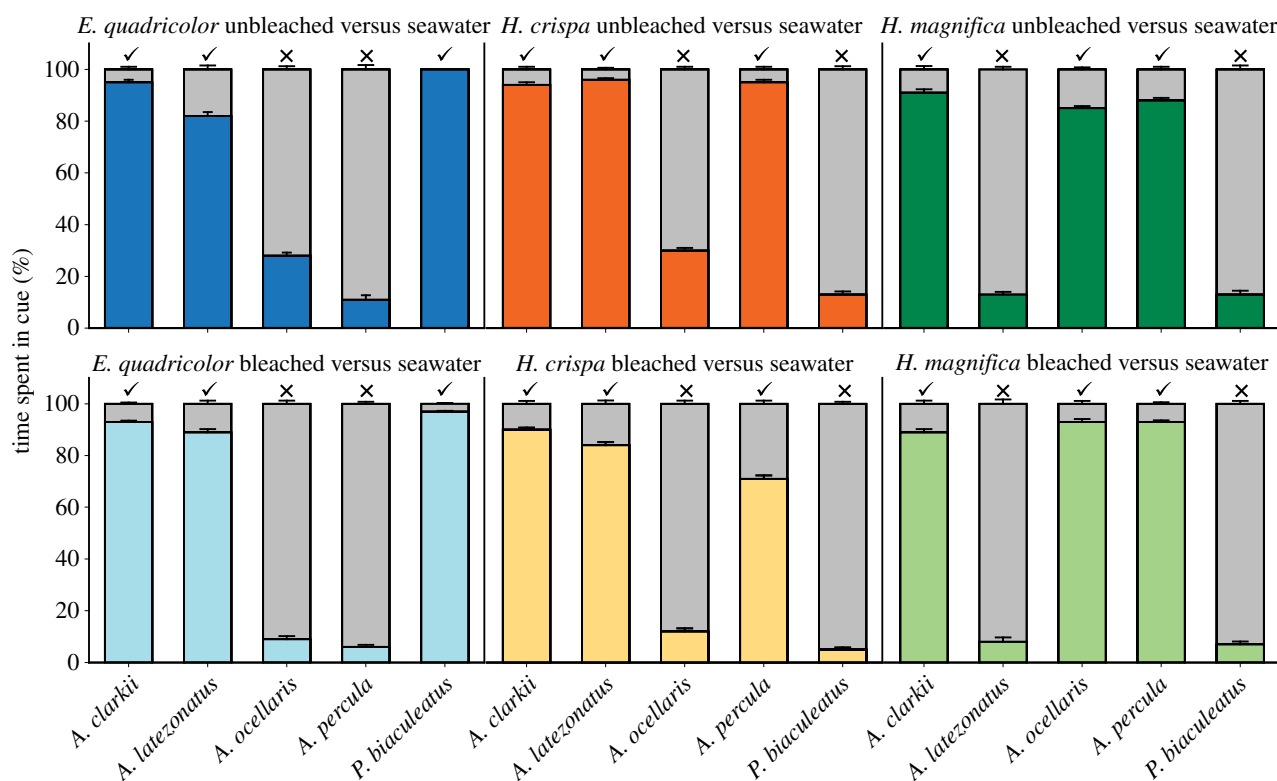


Figure 1. Anemonefish responses to chemical cues from unbleached and bleached sea anemones versus seawater. *Entacmaea quadricolor*: unbleached (dark blue), bleached (light blue); *Heteractis crispa*: unbleached (orange), bleached (yellow); *Heteractis magnifica*: unbleached (dark green), bleached (light green); seawater is represented by grey bars; tick marks above the bars indicate known associations, crosses indicate associations that do not naturally occur in the field. $n = 18–20$, data are means \pm s.e.

Using methods outlined by Gerlach *et al.* [29], a constant gravity-driven flow of 100 ml min^{-1} per channel was supplied from two 9.5 l header tanks (which contained anemone cues or seawater) and maintained using flow meters (flume dimensions: 13 cm length \times 4 cm width, 4 cm depth, working volume 308 ml, see [42] for further details). This flow rate ensured that the juveniles were not struggling to maintain their desired location within the flume, and therefore were able to respond to the chemical preferences being tested. Dye tests were done at each water change to ensure the two channels exhibited parallel water flow, and no areas of turbulence or eddies were present.

An individual fish was released in the downstream end of the flume where it was free to move to either side or swim towards the preferred water source. Each trial consisted of a 2 min habituation period, followed by a 2 min testing period, where the position of the fish on either the right or left side of the chamber was recorded at 5 s intervals (using a stopwatch and direct observation). Then a 1 min rest period was provided; during this time the water sources were switched from one side to the other, a measure to ensure a side preference was not being displayed by the fish. The 2 min habituation period and 2 min testing period were then repeated.

Kolmogorov–Simonov tests were used to compare the proportion of time that individuals spent in the stream of water containing the olfactory cue compared with the proportion of time that individuals spent in one side of the chamber when no cue was present (blank trial, seawater compared with seawater). Additional tests were run to determine significant differences in the strength of preferences between bleached anemones and unbleached anemone cues when either were compared with untreated seawater. To do this Kolmogorov–Simonov tests compared the proportion of time individuals spent in the stream of water containing the olfactory cue of the unbleached anemone against untreated seawater to the proportion of time that individuals spent in the stream of water containing the olfactory cues of the bleached anemone against untreated seawater.

3. Results

(a) Responses of anemonefishes to unbleached and bleached sea anemones versus seawater

All anemonefishes preferentially selected unbleached sea anemones that they associate with in the field over seawater, spending more than 82% of their time in the anemone cue ($p > 0.001$ in all cases; figure 1; electronic supplementary material, table S1). When presented with cues from non-host species, all fishes chose seawater ($p < 0.001$ in all cases), spending less than 30% of their time in the anemone cue. These same patterns were observed regardless of host health with all fish choosing bleached host chemical cues over seawater, and seawater over non-host chemical cues.

While all fishes preferred the chemical cues derived from the host compared with seawater irrespective of the anemone's bleaching status, the strength of the fish's response towards the chemical cues of unbleached host tissue compared with the response towards bleached host tissue often differed (figure 1). *Amphiprion clarkii* was the only species to show no difference in the time spent in cues from known unbleached or bleached hosts versus seawater (*E. quadricolor* $p = 0.23$, *H. crispa* $p = 0.135$, *H. magnifica* $p = 0.771$). By contrast, *A. latezonatus* selected unbleached *H. crispa* $96 \pm 0.6\%$ of the time over seawater, whereas the bleached equivalent was only selected $84 \pm 1.3\%$ of the time ($p < 0.01$); *A. percula* selected unbleached *H. crispa* $95 \pm 1.0\%$ of the time, while spending $71 \pm 1.2\%$ of the time in the bleached *H. crispa* cue ($p < 0.001$); and *P. biaculeatus* spent $100 \pm 0\%$ of their time in cues from unbleached *E. quadricolor* and $97 \pm 0.3\%$ in bleached cues ($p < 0.001$). Interestingly, *A. ocellaris* and *A. percula* spent significantly more time in bleached *H. magnifica* cues versus

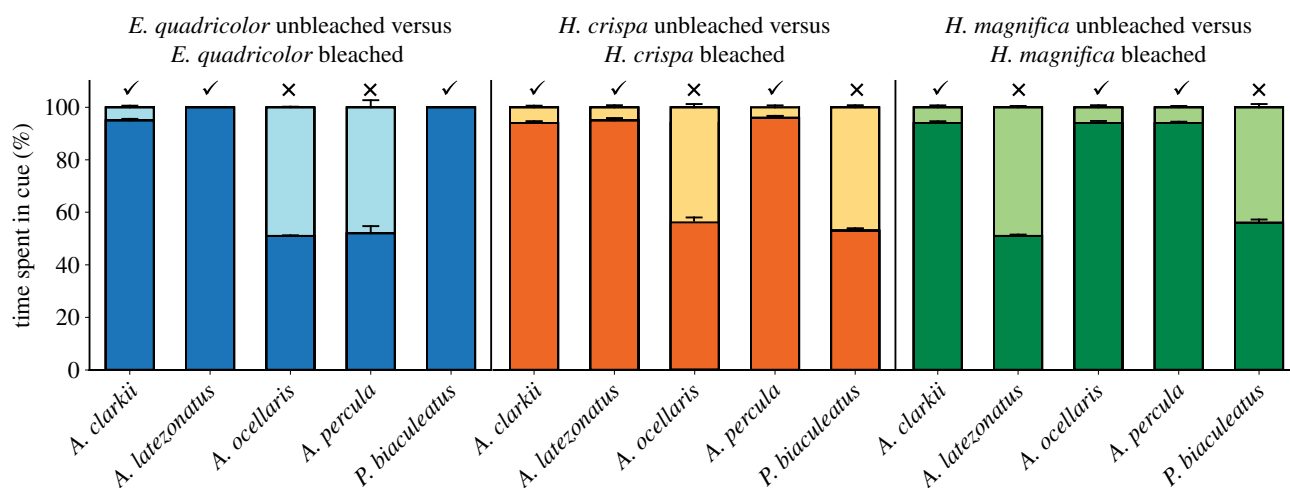


Figure 2. Anemonefish responses to chemical cues from unbleached versus bleached sea anemones of the same species. *Entacmaea quadricolor*: unbleached (dark blue), bleached (light blue); *Heteractis crispa*: unbleached (orange), bleached (yellow); *Heteractis magnifica*: unbleached (dark green), bleached (light green); tick marks above the bars indicate known associations; crosses indicate associations that do not naturally occur in the field. $n = 18–20$, data are means \pm s.e.

seawater than in unbleached cues versus seawater ($p < 0.001$ in both cases), and so did *A. latezonatus* in the *E. quadricolor* versus seawater trials ($p = 0.01$, figure 1).

(b) Responses of anemonefishes to unbleached versus bleached sea anemones of the same species

All anemonefishes displayed very strong preferences for unbleached known host anemones when tested simultaneously against bleached anemones of the same species, spending between 94 and 100% of their time in the cues generated from the former ($p < 0.001$ in all cases; figure 2; electronic supplementary material, table S1). When unbleached and bleached non-hosts of the same species were trialled against each other, fishes displayed no preference for either cue ($p > 0.05$ in all cases; figure 2; electronic supplementary material, table S1).

(c) Responses of anemonefishes to unbleached versus bleached sea anemones of differing species

Amphiprion clarkii and *A. percula* showed strong preferences for unbleached over bleached hosts ($p < 0.001$ in all cases; figure 3; electronic supplementary material, table S1). By contrast, *A. latezonatus* selected *H. crispa* over *E. quadricolor* regardless of health ($p < 0.001$ in both cases). Fishes, with the exception of *A. clarkii*, which uses all sea anemones tested, selected unbleached or bleached hosts rather than non-host species ($p < 0.001$ in all cases). When two non-hosts were tested simultaneously, fishes typically showed no preference towards either cue (figure 3; electronic supplementary material, table S1).

(d) Responses of anemonefishes to bleached versus bleached sea anemones of differing species

Anemonefishes showed differing responses when they were presented with cues from two different known hosts that were bleached (figure 4; electronic supplementary material, table S1). Both *A. clarkii* and *A. percula* showed no preference and spent equal time in the cues produced by bleached hosts ($p > 0.05$ in all cases). By contrast, *A. latezonatus* showed strong preferences for bleached *H. crispa* over bleached

E. quadricolor, spending $71 \pm 1.1\%$ of their time in these cues ($p < 0.001$ in all cases). When a bleached host was tested against a non-host anemone, all fishes selected the bleached host ($p < 0.001$ in all cases); and when two non-hosts were tested against each other no preference was shown for either cue ($p > 0.05$ in all cases; figure 4; electronic supplementary material, table S1).

4. Discussion

Bleaching events are increasing in frequency and severity, and are one of the major contributors to the decline in the diversity of coral reef communities [5,7]. This study used paired-choice flume experiments to examine the consequences of sea anemone bleaching during settlement of an iconic group of reef fishes. We found that juvenile anemonefishes (*Amphiprion clarkii*, *A. latezonatus*, *A. ocellaris*, *A. percula* and *Premnas biaculeatus*) have olfactory capabilities that allow them to distinguish between unbleached and bleached host sea anemones, and that although bleached anemones will be selected as habitat, this generally only occurs when other host options are unavailable. Bleached hosts were always preferentially selected over unbleached non-host sea anemones, indicating that the need for species-specific hosts during settlement is more powerful than the health of the habitat. In some cases, the potential adverse impacts of settling on a bleached host in the field may only be short-term, as depending on the causative stressor, anemones can recover [19].

The ability to recognize bleached sea anemones during settlement is advantageous, as depending on severity, bleaching can lead to host mortality [20,43], and consequent fish mortality due to obligate nature of the symbiosis [14,15,18]. If the bleached host remains alive, post-settlement mortality may not be an issue, as juvenile, sub-adult and adult *A. polymnus* numbers were not affected after mild bleaching in Bootless Bay, Papua New Guinea [19]. However, settlement onto a bleach host may have implications for the future reproductive success of the fish, as habitat degradation can result in a reduced egg production [19].

Another potential deleterious effect of settling onto a bleached host would be the loss of visual crypsis. Fishes are more conspicuous against the white background of a

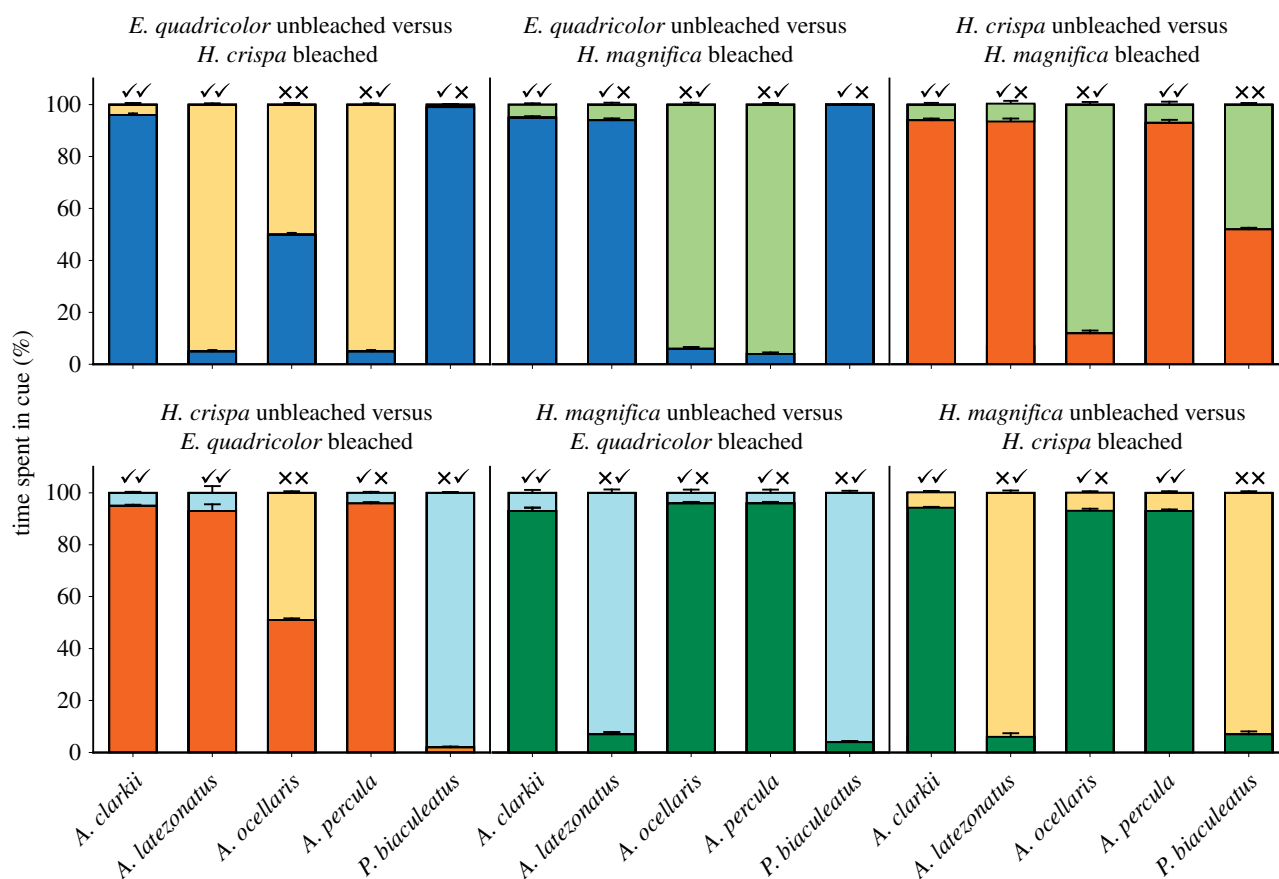


Figure 3. Anemonefish responses to chemical cues from unbleached and bleached sea anemones of differing species. *Entacmaea quadricolor*: unbleached (dark blue), bleached (light blue); *Heteractis crispa*: unbleached (orange), bleached (yellow); *Heteractis magnifica*: unbleached (dark green), bleached (light green); tick marks above the bars indicate known associations; crosses indicate associations that do not naturally occur in the field. $n = 18–20$, data are means \pm s.e.

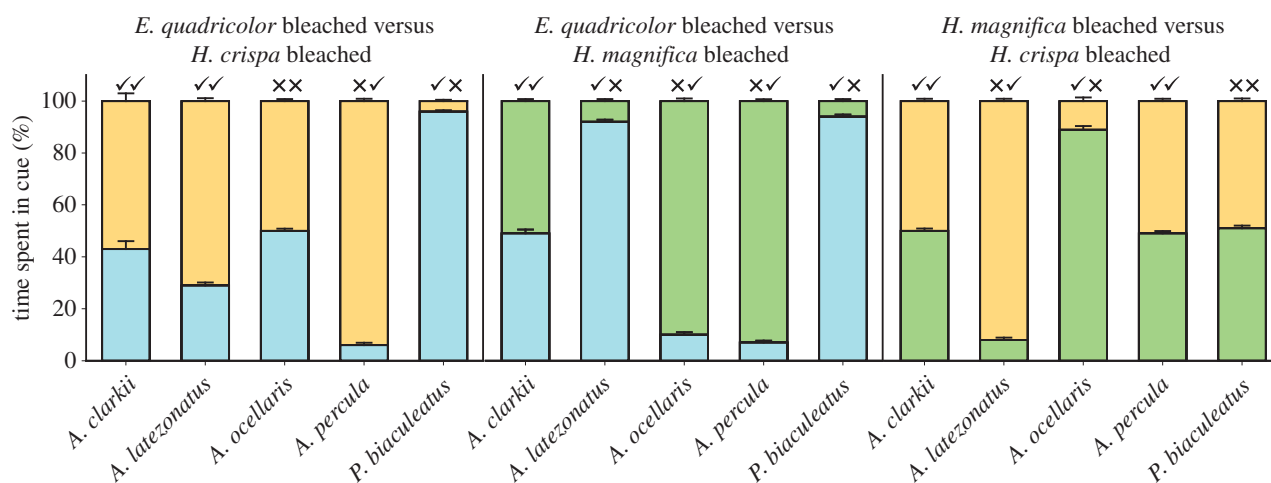


Figure 4. Anemonefish responses to chemical cues from bleached versus bleached sea anemones of differing species. *Entacmaea quadricolor*, light blue; *Heteractis crispa*, light yellow; *Heteractis magnifica*, light green. Tick marks above the bars indicate known associations; crosses indicate associations that do not naturally occur in the field. $n = 18–20$, data are means \pm s.e.

bleached anemone, which could lead to increased predation [19,44]; however, the protection afforded by sheltering in the anemone's nematocyst-laden tentacles [14] may help negate this. While chemical crypsis has not been investigated in anemonefishes, many organisms have evolved mechanisms to blend in with the chemical components of their habitat [45]. Because our study has shown that anemonefishes have the olfactory capabilities to detect differences between bleached and unbleached hosts, bleaching may also alter chemical crypsis. Furthermore, behavioural changes have been documented in *A. akindynos* that occupy bleached

hosts, and it has been suggested that these changes may increase predation susceptibility [44].

While all five anemonefish species were attracted to bleached hosts rather than seawater, the strength of this attraction often varied when compared with the responses found during trials that used unbleached hosts. Some fishes spent more time in unbleached host cues versus seawater than in bleached cues versus seawater, and vice versa. *Amphiprion clarkii* was the only species that spent equal time in cues from unbleached or bleached hosts versus seawater. Because host anemones are typically rare on reefs [20,46–49], anemonefishes

have relatively short pelagic larval durations of 7–22 days [50–52], and predation pressure is likely to be high during this time; settling on the first host encountered, even if bleached, may be beneficial as rejecting bleached habitat may mean that settlement does not occur. Our study provides laboratory-based comparative observations and insights into how anemonefishes may respond to bleaching, which are further supported by field observations, as Saenz-Agudelo *et al.* [19] found no evidence of *A. polymnus* larvae avoiding bleached anemones in the field. By contrast, field experiments have shown that some other pomacentrids prefer live coral over bleached habitat at the settlement stage [36], and that recruitment can be reduced during large-scale bleaching events [12].

Anemonefishes showed differing responses when presented with cues from two bleached hosts. *Amphiprion clarkii*, a generalist species that uses all 10 host anemones, and *A. percula*, which is more specialized in terms of host use [15], both showed no preference between bleached hosts that were tested against each other. By contrast, *A. latezonatus*, which uses *E. quadricolor* and *H. crista* as habitat [38], showed a strong preference for the latter species, and was the only fish to preferentially select a bleached over an unbleached host. *Heteractis crista* has been suggested to serve as important nursery habitat for *A. latezonatus* in the Solitary Islands Marine Park, Australia (A. Scott and H. Malcolm 2016, personal communication), and similarly for *A. bicinctus* in the Red Sea [53].

5. Conclusion

This laboratory-based study furthers our understanding of the information used during coral reef fish larvae settlement,

specifically investigating the role of habitat quality in the selection of settlement sites by habitat specialists. All five anemonefishes tested were able to recognize bleached habitat using chemical cues alone, and all but *A. latezonatus* preferentially selected unbleached versus bleached host anemones when tested concurrently. Anemonefishes did not show any flexibility in terms of host use regardless of bleaching status, which could be deleterious to populations during severe habitat degradation events that lead to host mortality. The ability of anemonefishes to determine the health status of their species-specific host could be particularly important given that bleaching events are becoming more common, and recruitment occurs during the warmer months [54–56] when thermal bleaching is most likely to occur [5]. These findings highlight how specialists that associate with bleaching-susceptible habitat may potentially respond to the indirect impacts of climate change.

Ethics. Animal care and experimental protocols complied with animal ethics regulations and approvals from Georgia Institute of Technology. Ethics approval number A14035.

Data accessibility. The full dataset supporting this article is available from the institutional repository at Southern Cross University (http://epubs.scu.edu.au/data_collections/).

Authors' contributions. A.S. and D.L.D. conceived the research idea, designed the study, performed the research, collected the data and wrote the manuscript; D.L.D. performed the statistical analyses. Both authors gave final approval for publication.

Competing interests. We declare no competing interests.

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References

- Hughes TP. 1994 Catastrophes, phase shifts, and large-scale degradation of a Caribbean coral reef. *Science* **265**, 1547–1551. (doi:10.1126/science.265.5178.1547)
- Pandolfi JM *et al.* 2003 Global trajectories of the long-term decline of coral reef ecosystems. *Science* **301**, 955–958. (doi:10.1126/science.1085706)
- Hoegh-Guldberg O *et al.* 2007 Coral reefs under rapid climate change and ocean acidification. *Science* **318**, 1737–1742. (doi:10.1126/science.1152509)
- Done TJ. 1992 Phase shifts in coral reef communities and their ecological significance. *Hydrobiologia* **247**, 121–132. (doi:10.1007/BF00008211)
- Hoegh-Guldberg O. 1999 Climate change, coral bleaching and the future of the world's coral reefs. *Mar. Freshwater Res.* **50**, 839–866. (doi:10.1071/MF99078)
- Glynn PW. 1996 Coral reef bleaching: facts, hypotheses and implications. *Glob. Change Biol.* **2**, 495–509. (doi:10.1111/j.1365-2486.1996.tb00063.x)
- Brown B. 1997 Coral bleaching: causes and consequences. *Coral Reefs* **16**, S129–S138. (doi:10.1007/s003380050249)
- Hoegh-Guldberg O, Smith GJ. 1989 The effect of sudden changes in temperature, light and salinity on the population density and export of zooxanthellae from the reef corals *Stylophora pistillata* Esper and *Seriatopora hystrix* Dana. *J. Exp. Mar. Biol. Ecol.* **129**, 279–303. (doi:10.1016/0022-0981(89)90109-3)
- Muscattine L, Porter JW. 1977 Reef corals: mutualistic symbioses adapted to nutrient-poor environments. *Bioscience* **27**, 454–460. (doi:10.2307/1297526)
- Bonin M. 2012 Specializing on vulnerable habitat: *Acropora* selectivity among damselfish recruits and the risk of bleaching-induced habitat loss. *Coral Reefs* **31**, 287–297. (doi:10.1007/s00338-011-0843-2)
- Jones GP, McCormick MI, Srinivasan M, Eagle JV. 2004 Coral decline threatens fish biodiversity in marine reserves. *Proc. Natl Acad. Sci. USA* **101**, 8251–8253. (doi:10.1073/pnas.0401277101)
- Booth DJ, Beretta GA. 2002 Changes in a fish assemblage after a coral bleaching event. *Mar. Ecol. Prog. Ser.* **245**, 205–212. (doi:10.3354/meps245205)
- Feary DA, Almany GR, McCormick MI, Jones GP. 2007 Habitat choice, recruitment and the response of coral reef fishes to coral degradation. *Oecologia* **153**, 727–737. (doi:10.1007/s00442-007-0773-4)
- Fautin DG. 1991 The anemonefish symbiosis: what is known and what is not. *Symbiosis* **10**, 23–46.
- Fautin DG, Allen GR. 1997 *Anemonefishes and their host sea anemones: a guide for aquarists and divers*. Perth, Australia: Western Australian Museum.
- Bridge T, Scott A, Steinberg D. 2012 Abundance and diversity of anemonefishes and their host sea anemones at two mesophotic sites on the Great Barrier Reef, Australia. *Coral Reefs* **31**, 1057–1062. (doi:10.1007/s00338-012-0916-x)
- Fautin DG. 1988 Sea anemones (Actiniaria and Corallimorpharia) of Madang Province. *Sci. New Guinea* **14**, 22–29.
- Hattori A. 2002 Small and large anemonefishes can coexist using the same patchy resources on a coral reef, before habitat destruction. *J. Anim. Ecol.* **71**, 824–831. (doi:10.1046/j.1365-2656.2002.00649.x)
- Saenz-Agudelo P, Jones G, Thorrold S, Planes S. 2011 Detrimental effects of host anemone bleaching on anemonefish populations. *Coral Reefs* **30**, 497–506. (doi:10.1007/s00338-010-0716-0)

20. Hobbs J-PA, Frisch AJ, Ford BM, Thums M, Saenz-Agudelo P, Furby KA, Berumen ML. 2013 Taxonomic, spatial and temporal patterns of bleaching in anemones inhabited by anemonefishes. *PLoS ONE* **8**, e70966. (doi:10.1371/journal.pone.0070966)
21. Arvedlund M, Takemura A. 2005 Long-term observation *in situ* of the anemonefish *Amphiprion clarkii* (Bennett) in association with a soft coral. *Coral Reefs* **24**, 698. (doi:10.1007/s00338-005-0007-3)
22. Buston P. 2003 Forcible eviction and prevention of recruitment in the clown anemonefish. *Behav. Ecol.* **14**, 576–582. (doi:10.1093/beheco/arg036)
23. Caley M, Carr M, Hixon M, Hughes T, Jones G, Menge B. 1996 Recruitment and the local dynamics of open marine populations. *Annu. Rev. Ecol. Syst.* **27**, 477–500. (doi:10.1146/annurev.ecolsys.27.1.477)
24. Almany GR, Webster MS. 2006 The predation gauntlet: early post-settlement mortality in reef fishes. *Coral Reefs* **25**, 19–22. (doi:10.1007/s00338-005-0044-y)
25. Leis JM, Siebeck U, Dixon DL. 2011 How Nemo finds home: the neuroecology of dispersal and of population connectivity in larvae of marine fishes. *Integr. Comp. Biol.* **51**, 826–843. (doi:10.1093/icb/ict004)
26. Leis J, Carson-Ewart B, Hay A, Cato D. 2003 Coral-reef sounds enable nocturnal navigation by some reef-fish larvae in some places and at some times. *J. Fish Biol.* **63**, 724–737. (doi:10.1046/j.1095-8649.2003.00182.x)
27. Simpson SD, Meekan M, Montgomery J, McCauley R, Jeffs A. 2005 Homeward sound. *Science* **308**, 221. (doi:10.1126/science.1107406)
28. Atema J, Kingsford MJ, Gerlach G. 2002 Larval reef fish could use odour for detection, retention and orientation to reefs. *Mar. Ecol. Prog. Ser.* **241**, 151–160. (doi:10.3354/meps241151)
29. Gerlach G, Atema J, Kingsford MJ, Black KP, Miller-Sims V. 2007 Smelling home can prevent dispersal of reef fish larvae. *Proc. Natl Acad. Sci. USA* **104**, 858–863. (doi:10.1073/pnas.0606777104)
30. Dixon DL, Jones GP, Munday PL, Planes S, Pratchett MS, Srinivasan M, Syms C, Thorrold SR. 2008 Coral reef fish smell leaves to find island homes. *Proc. R. Soc. B* **275**, 2831–2839. (doi:10.1098/rspb.2008.0876)
31. Sweatman H. 1988 Field evidence that settling coral reef fish larvae detect resident fishes using dissolved chemical cues. *J. Exp. Mar. Biol. Ecol.* **124**, 163–174. (doi:10.1016/0022-0981(88)90170-0)
32. Lecchini D, Planes S, Galzin R. 2007 The influence of habitat characteristics and conspecifics on attraction and survival of coral reef fish juveniles. *J. Exp. Mar. Biol. Ecol.* **341**, 85–90. (doi:10.1016/j.jembe.2006.10.006)
33. Lecchini D, Shima J, Banaigs B, Galzin R. 2005 Larval sensory abilities and mechanisms of habitat selection of a coral reef fish during settlement. *Oecologia* **143**, 326–334. (doi:10.1007/s00442-004-1805-y)
34. Elliott JK, Elliott JM, Mariscal RN. 1995 Host selection, location, and association behaviors of anemonefishes in field and settlement experiments. *Mar. Biol.* **122**, 377–389. (doi:10.1007/BF00350870)
35. Dixon D, Munday P, Pratchett M, Jones G. 2011 Ontogenetic changes in responses to settlement cues by anemonefish. *Coral Reefs* **30**, 903–910. (doi:10.1007/s00338-011-0776-9)
36. McCormick M, Moore J, Munday P. 2010 Influence of habitat degradation on fish replenishment. *Coral Reefs* **29**, 537–546. (doi:10.1007/s00338-010-0620-7)
37. Miyagawa K. 1989 Experimental analysis of the symbiosis between anemonefishes and sea anemones. *Ethology* **80**, 19–46. (doi:10.1111/j.1439-0310.1989.tb00727.x)
38. Scott A, Rushworth K, Dalton S, Smith S. In press. Subtropical anemonefish *Amphiprion latezonatus* recorded in two additional species of host sea anemone. *Marine Biodiversity*. (doi:10.1007/s12526-12015-10390-12520)
39. Allen GR, Drew J, Kaufman L. 2008 *Amphiprion barberi*, a new species of anemonefish (Pomacentridae) from Fiji, Tonga, and Samoa. *Aqua Int. J. Ichthyol.* **14**, 105–114.
40. Allen GR, Drew J, Fenner D. 2010 *Amphiprion pacificus*, a new species of anemonefish (Pomacentridae) from Fiji, Tonga, Samoa, and Wallis Island. *Aquaculture* **16**, 129–138.
41. Siebeck U, Marshall N, Klüter A, Hoegh-Guldberg O. 2006 Monitoring coral bleaching using a colour reference card. *Coral Reefs* **25**, 453–460. (doi:10.1007/s00338-006-0123-8)
42. Munday PL, Dixon DL, Donelson JM, Jones GP, Pratchett MS, Devitsina GV, Døving KB. 2009 Ocean acidification impairs olfactory discrimination and homing ability of a marine fish. *Proc. Natl Acad. Sci. USA* **106**, 1848–1852. (doi:10.1073/pnas.0809996106)
43. Hill R, Scott A. 2012 The influence of irradiance on the severity of thermal bleaching in sea anemones that host anemonefish. *Coral Reefs* **31**, 273–284. (doi:10.1007/s00338-011-0848-x)
44. Lönnstedt OM, Frisch AJ. 2014 Habitat bleaching disrupts threat responses and persistence in anemonefish. *Mar. Ecol. Prog. Ser.* **517**, 265–270. (doi:10.3354/meps11031)
45. Brooker RM, Munday PL, Chivers DP, Jones GP. 2015 You are what you eat: diet-induced chemical crypsis in a coral-feeding reef fish. *Proc. R. Soc. B* **282**, 20141887. (doi:10.1098/rspb.2014.1887)
46. Dunn DF. 1981 The clownfish sea anemones: Stichodactylidae (Coelenterata: Actiniaria) and other sea anemones symbiotic with pomacentrid fishes. *Trans. Am. Phil. Soc.* **71**, 1–115. (doi:10.2307/1006382)
47. Hirose Y. 1985 Habitat, distribution and abundance of coral reef sea-anemones (Actiniidea and Stichodactylidae) in Sesoko Island, Okinawa, with notes on expansion and contraction behavior. *Galaxea* **4**, 113–127.
48. Scott A, Baird A. 2015 Trying to find Nemo: low abundance of sea anemones and anemonefishes on central and southern mid-shelf reefs in the Great Barrier Reef. *Mar. Biodivers.* **45**, 327–331. (doi:10.1007/s12526-014-0245-0)
49. Chadwick NE, Arvedlund M. 2005 Abundance of giant sea anemones and patterns of association with anemonefish in the northern Red Sea. *J. Mar. Biol. Assoc. UK* **85**, 1287–1292. (doi:10.1017/S0025315405012440)
50. Jones GP, Planes S, Thorrold SR. 2005 Coral reef fish larvae settle close to home. *Curr. Biol.* **15**, 1314–1318. (doi:10.1016/j.cub.2005.06.061)
51. Wellington G, Victor B. 1989 Planktonic larval duration of one hundred species of Pacific and Atlantic damselfishes (Pomacentridae). *Mar. Biol.* **101**, 557–567. (doi:10.1007/BF00541659)
52. Thresher RE, Colin PL, Bell LJ. 1989 Planktonic duration, distribution and population structure of western and central Pacific damselfishes (Pomacentridae). *Copeia* **1989**, 420–434. (doi:10.2307/1445439)
53. Huebner L, Dailey B, Titus B, Khalaf M, Chadwick N. 2012 Host preference and habitat segregation among Red Sea anemonefish: effects of sea anemone traits and fish life stages. *Mar. Ecol. Prog. Ser.* **464**, 1–15. (doi:10.3354/meps09964)
54. Doherty PP. 1991 Spatial and temporal patterns in recruitment. In *The ecology of fishes on coral reefs* (ed. P. Sale), pp. 261–293. San Diego, CA: Academic Press.
55. Doherty P, Williams D. 1988 The replenishment of coral reef fish populations. *Oceanogr. Mar. Biol. Annu. Rev.* **26**, 487–551.
56. Booth DJ, Brosnan DM. 1995 The role of recruitment dynamics in rocky shore and coral reef fish communities. *Adv. Ecol. Res.* **26**, 309–385. (doi:10.1016/S0065-2504(08)60068-9)