

Piecewise Regression with Negative Binomial Type I Error on Real Data using **brms** Custom Family

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Goal

- Fit two piece negative binomial type 1 formulation to data
- From 2023-02-28 version of `nbinom_type1.R`

Negative Binomial distribution parameterized by mean (μ) and overdispersion parameter (θ). This parameterization is referred to as NEGBIN type I (Cameron and Trivedi, 1998) as cited by <https://doi.org/10.1080/03610926.2018.1563164> `## x ~ nbinom_type1(mu, theta)`, where $E(x) = \mu$, $Var(x) = (\theta + 1) \mu$. This should not be confused with the μ and shape parameterization of `nbinom` in R or the ‘alternative’ NB (`neg_binomial_2_...`) in stan. Note using `disp` instead of `theta` because using `theta` gives the error `> Error: Currently ‘dirichlet’ is the only valid prior for simplex parameters. See help(set_prior) for more details.` when trying to fit the model.

Recap

- Earlier work generated poor estimates of x_0 .
- Visualization of data and model fit indicates there’s very little information on x_0 .
- While I can generate predictions of expected values, I can’t generate expected values of the data itself. I expect this is due to fact that we generate parameters which result in $y = \text{NaN}$
- TODO
 - Figure out better model definition that avoids generating NaN values. I expect this can be done by imposing a better prior on `x0|male`.
 - Allow `disp` to vary between males.
- On 3/22/2023 I added the missing $|dg^{-1}(disp)/ddisp| = |-mu/disp^2| = mu/disp^2$ term to likelihood function `## Insights`
- When the `disp` (dispersal or `theta`) gets unrealistically large, we get the emergence of a bimodal distribution at both ends of x_0 values.

Even though we included this value, it is very unlikely to be 25C values. I interpret this to mean that when things are really noisy (high `theta`), one way to interpret the data is that one set of males has a very long (presumably slow) decline. It would be good to look at the correlations via `pairs()`.
- To me this is consistent with the informal knowledge that the
- Two males have fitting issues, “T235” and “T236”. This appears to be due to a bimodal posterior surface where one region has low ‘ x_0 ’ (< 40C), but low ‘ y_0 ’, and the other has a high x_0 and low y_0

Set up

Install libraries

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```
## load libraries
library(MASS) # provides negative binomial fitting: glm.nb
library(stats)
library(tidyverse)
library(brms)
library(loo)
library(ggplot2)
#library(tidybayes)
library(ggpubr)
library(grid)
library(gridExtra)
library(ragg)
library(GGally)
library(cowplot)

ggplot2::theme_set(theme_default(base_size = 10))
#ggplot2::theme_set(theme_default(plot.background = element_rect(color = "black")))

library(broom)
library(viridisLite)
library(cmdstanr)
library(rstan)
options(mc.cores = (parallel::detectCores()-2))
rstan_options(auto_write = TRUE)

## options(ggplot2.continuous.colour="viridis",
##         ggplot2.discrete.colour="viridis",
##         ggplot2.scale_fill_discrete = scale_fill_viridis_d,
##         ggplot2.scale_fill_continuous = scale_fill_viridis_c)

library(reshape2)
library(lme4)
library(latex2exp)
```

Source family

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```
source(".././../custom-brms-families/families/nbinom_type1.R")
```

```
### Load Data
```

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```
supply(file.path("input", dir("input")),
       load, verbose = TRUE, envir = .GlobalEnv)
```

```
## Loading objects:
##   data_ind
## Loading objects:
##   motif_data
##   motif_data_40C
##   motif_stats
##   motif_stats_40C
##   bird_bill_data
## Loading objects:
##   summary_stats
## Loading objects:
##   stats_ind

## $'input/data_ind.Rda'
## [1] "data_ind"
##
## $'input/data.processing_2022-12-15.Rda'
## [1] "motif_data"      "motif_data_40C"  "motif_stats"     "motif_stats_40C"
## [5] "bird_bill_data"
##
## $'input/obs_summary_stats.Rda'
## [1] "summary_stats"
##
## $'input/stats_ind.Rda'
## [1] "stats_ind"
```

```
head(stats_ind)
```

```
## # A tibble: 6 x 9
##   male round n_obs total_round mean_round sd_round cv_round total mean
##   <fct> <dbl> <int>      <int>      <dbl>    <dbl>    <dbl> <int> <dbl>
## 1 T234     1    13        203      40.6     32.0     0.787   601  46.2
## 2 T235     1    13        882     176.    132.     0.748  2333 179.
## 3 T236     1    13        758     152.     46.0     0.303  2095 161.
## 4 T243     1    13        438     87.6     76.4     0.872  1861 143.
## 5 T244     1    13        270      54      14.7     0.272   993  76.4
## 6 T246     1     5        253     50.6     54.6     1.08   253  50.6
```

```
names(stats_ind)
```

```
## [1] "male"      "round"     "n_obs"     "total_round" "mean_round"
## [6] "sd_round"  "cv_round"  "total"     "mean"
```

```
head(data_ind)
```

```
## # A tibble: 6 x 11
##   male index motif_count temp_target temp round trial_round date counter
##   <chr> <int>      <int>      <dbl> <dbl> <dbl>      <dbl> <chr>    <chr>
## 1 T234     1         0        42  43.0     1         1 02/03/22 RAS
## 2 T234     1        30        44  44.5     1         2 02/05/22 RAS
## 3 T234     1        34        27  27.2     1         3 02/07/22 RAS
```

```
## 4 T234      1      87      40 41.1      1      4 02/09/22 RAS
## 5 T234      1      52      35 36.1      1      5 02/11/22 RAS
## 6 T234      1      32      40 39.5      2      1 04/23/22 KIM
## # i 2 more variables: y0_simple_est <dbl>, phi_ind <dbl>
```

```
names(data_ind)
```

```
## [1] "male"      "index"      "motif_count" "temp_target"
## [5] "temp"      "round"      "trial_round" "date"
## [9] "counter"    "y0_simple_est" "phi_ind"
```

Determine reasonable priors for y0

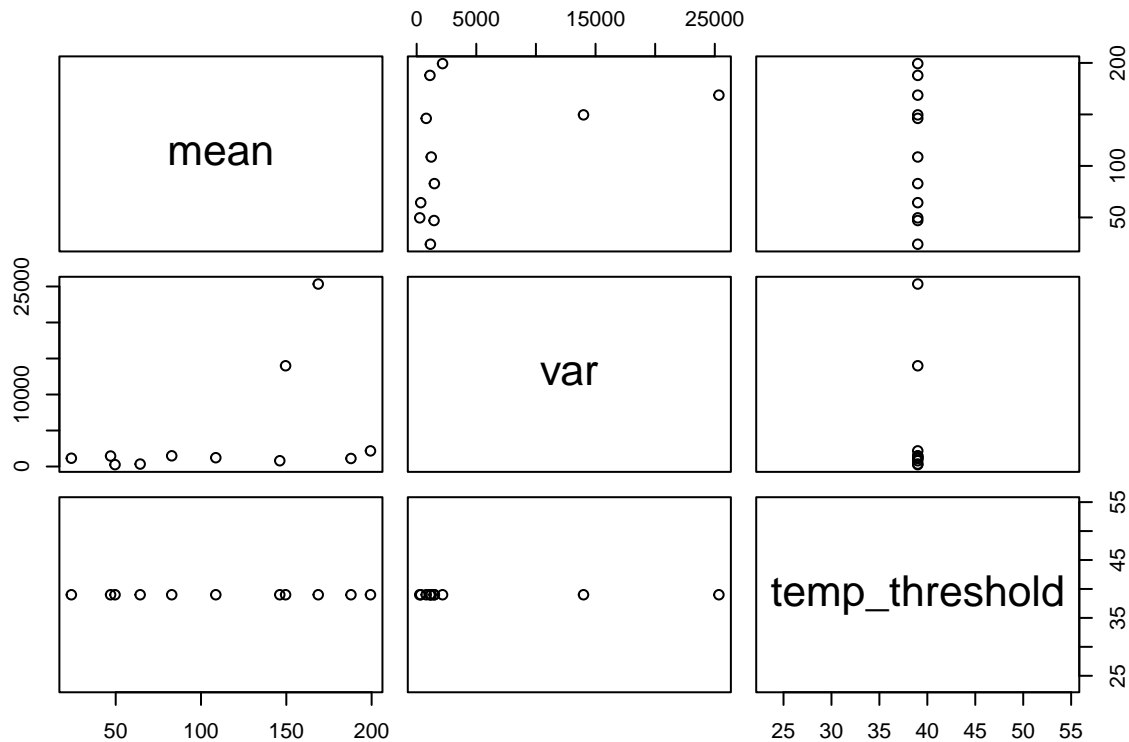
\small

```
cumulative_count_vs_temp <- list()

for(temp_threshold in 26:45) {
  tmp <- data_ind %>% group_by(male) %>% filter(temp < temp_threshold) %>% summarize(mean = mean(motif_
  mean <- mean(tmp$mean)
  sd <- sd(tmp$mean)
  cumulative_count_vs_temp[[temp_threshold]] <- tmp
  print(paste0("Temp: ", temp_threshold, ", mean: ", mean, ", sd: ", sd))
}
```

```
## [1] "Temp: 26, mean: 63, sd: 89.095454429505"
## [1] "Temp: 27, mean: 126.2, sd: 126.689383927778"
## [1] "Temp: 28, mean: 118.142857142857, sd: 110.701443187564"
## [1] "Temp: 29, mean: 116.071428571429, sd: 111.492312355093"
## [1] "Temp: 30, mean: 116.944444444444, sd: 102.314907407366"
## [1] "Temp: 31, mean: 120.055555555556, sd: 114.670572850134"
## [1] "Temp: 32, mean: 120.257575757576, sd: 104.858697610863"
## [1] "Temp: 33, mean: 120.257575757576, sd: 104.858697610863"
## [1] "Temp: 34, mean: 116.606060606061, sd: 89.5174183151982"
## [1] "Temp: 35, mean: 111.068181818182, sd: 69.0805871808609"
## [1] "Temp: 36, mean: 109.833333333333, sd: 69.568651145629"
## [1] "Temp: 37, mean: 108.231818181818, sd: 69.5451985140861"
## [1] "Temp: 38, mean: 110.348484848485, sd: 68.53139688191"
## [1] "Temp: 39, mean: 111.646176046176, sd: 61.7875535115004"
## [1] "Temp: 40, mean: 112.341666666667, sd: 64.4998385010898"
## [1] "Temp: 41, mean: 113.540928768201, sd: 56.0919858489611"
## [1] "Temp: 42, mean: 116.815308933491, sd: 60.694488706503"
## [1] "Temp: 43, mean: 115.212741046832, sd: 60.7911617164827"
## [1] "Temp: 44, mean: 114.864362152999, sd: 61.2514194145016"
## [1] "Temp: 45, mean: 113.168939393939, sd: 62.0293112972156"
```

```
plot_pairs <- pairs(cumulative_count_vs_temp[[39]] %>% select(-c(male, sd, cv)))
```



```
print(plot_pairs)
```

```
## NULL
```

```
hist <- ggplot(cumulative_count_vs_temp[[39]], aes(mean)) +  
  geom_histogram(bins = 6)
```

```
hist_log <- hist + scale_x_log10()
```

```
plot_grid <- plot_grid(plotlist = list(hist, hist_log))
```

If using a normal prior, go with $\text{mean} = 125$ and $\text{sd} = 125 * 4 = 500$. However, the data doesn't seem to follow any real distribution and its at the motif rather than song scale, as a result each male has its own, unknown scaling factor between motifs and songs (i.e. $1/E(\# \text{ motifs/song})$) so a flat prior is justifiable.

Fit Models

- Code derived from `../2023-02-28_fit.real.data.using.nbinom_type1/brms_two.piece_fit.nbionm_type1.Rmd`

Set up functions, parameters, and results tibble

```
data_stan <- data_ind %>% rename(y = motif_count, x = temp) %>%  
  mutate(male = factor(male))  
males <- unique(data_stan$male)  
nmales <- length(males)
```

```

xmax <- 46 # maximum value for x0
xignore <- 39 # x value above which data is ignored in one_piece model

stan_two_piece_func <- paste0(" real two_piece(real x, real x0, real y0) {
  real xmax = ", xmax, "; ## paste in value for xmax\n
  real y;

  if(x0 > xmax) {
    y = log(0);
  } else {
    y = y0 * (xmax - fmax(x0, x))/(xmax - x0);
  }
  return(y);
} ")

stan_one_piece_func <- paste0(" real one_piece(real y0) {
  return(y0);
} ")

stan_asymptotic_func <- paste0(" real asymp(real x, real phi, real y0) {
  real xmax = ", xmax, "; ## paste in value for xmax\n
  return(y0 * (1 - exp( - phi * (xmax - x))) );
} ")

## Function to drop chains, such as those that get stuck on a suboptimal posterior peak
## Taken from:
remove_chains <- function(brm_fit, chains_to_drop) {
  # brm_fit is the output of brms::brm

  sim <- brm_fit$fit@sim # Handy shortcut
  sim$samples <- sim$samples[-chains_to_drop]

  # Update the meta-info
  sim$chains <- sim$chains - length(chains_to_drop)
  sim$warmup2 <- sim$warmup2[-chains_to_drop]

  # Add the modified sim back to x
  brm_fit$fit@sim <- sim
  brm_fit
}

## Function to find row(s) in tbl that match criteria

which_tbl_row <- function(filter_male = FALSE, x0_flag = "individual", y0_flag = "individual", disp_flag,
  which( tbl$filter_male %in% filter_male &
    tbl$x0_flag %in% x0_flag &
    tbl$y0_flag %in% y0_flag &
    tbl$disp_flag %in% disp_flag &
    tbl$disp_value %in% disp_value &
    tbl$model %in% model
    # tbl$sampling_dist %in% sampling &
  )
}

```

Set up Dataframe for fit results

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```
models <- c("one_piece", "two_piece", "asymptotic")[2:3] #, "one_piece") #, "asymptotic")
sampling_dists <- c("nbinom_type1") ##, "com_poisson") ## lognormal doesn't work since the counts can be 0
flags_x0 <- c("uniform_1",
              "groups_1", ## this doesn't work with x0_Intercept prior, suggests error in priors
              # "groups_2a",
              # "groups_2b",
              "individual")

flags_y0 <- c("uniform_1", "groups_1", "individual")[3]

values_disp <- switch(1,
                      c(0.01), # 0.125 is a good value
                      c(0.01, 0.1), #, 0.25), # used in exploring model behavior
                      list(0.1, "flat"), #, 0.1, 1) # doesn't work yet
                      c("flat"))

flags_disp <- c("uniform_1", "groups_1", "individual")

## whether to filter males with large disp values estimated using one piece model
filter_male <- c(TRUE, FALSE)

N <- length(data)

fit_tbl <- crossing(model = models,
                    #sampling_dist = sampling_dists,
                    x0_flag = flags_x0,
                    y0_flag = flags_y0,
                    disp_value = values_disp,
                    disp_flag = flags_disp,
                    desc = "NA_character",
                    filter_male = filter_male,
                    #y0_group_list = list(NA), #tbl_tmp, #list(NA),
                    x0_group_list = list(NA),
                    fit = list(NA),
                    llik = list(NA),
                    r_eff = list(NA),
                    loo = list(NA)
                    )
```

Run fits

\small

```
run_fits <- FALSE
force_load <- FALSE #reload fit_tbl even if it already exists
save_fits <- FALSE
```

```

if(run_fits) {
  infile_tbl <- NULL
  cur_time <- gsub(" ", "_", Sys.time()) %>% gsub(":", ".", .)
  outfile_tbl <- file.path(output_dir, "tibbles", paste0("fit_tbl_", cur_time, ".Rda"))
} else {
  infile <- last(dir(file.path(output_dir, "tibbles"), "fit_tbl.*"))
  infile_tbl <- file.path(output_dir, "tibbles", infile)
  outfile_tbl <- NULL
}

sampling = "nbinom_type1"
prior_shape_y0 = "flat"

flags_x0_used <- c("individual", "groups_1", "uniform_1") %>% rev()#
flags_y0_used <- c("individual")
values_disp_used <- values_disp
flags_disp_used <- c("individual", "groups_1", "uniform_1")[3] # /> rev() /> first()
models_used <- c("one_piece", "two_piece", "asymptotic")[2] #c("one_piece", "two_piece") #, "two_piece"
shape_y0_prior <- "flat" # flat or normal

## These males produce bimodal posteriors and interfere with model fitting
## Ideally, we'd do a preliminary analysis without them and then include them later.

male_exclude = c("T235", "T236")

fit_index <- 0

for(male_filter in c(FALSE, TRUE)) {
  for(model in models_used) {

    print(model)
    switch(model,
      two_piece = {
        ## Note issues in non-convergence are related to bimodality of posterior surface.
        stan_func <- stan_two_piece_func
        warmup <- 3000 # floor(3/4 * iter)
        iter <- warmup + 2000
        adapt_delta <- 0.99
        thin <- 4
      },
      one_piece = {
        stan_func <- stan_one_piece_func
        warmup <- 2000
        iter <- warmup + 2000
        thin <- 4
        adapt_delta <- 0.7
      },
      asymptotic = {
        stan_func <- stan_asymptotic_func
        warmup <- 2000
        iter <- warmup + 2000
      }
    )
  }
}

```



```

        thin <- 4
        adapt_delta <- 0.9
    }

    )
  for(displ_flag in flags_displ_used) {

    print(stan_func)

    for(x0_flag in flags_x0_used) {

      for(y0_flag in flags_y0_used) {

        ## define variable for labeling figures
        x0_label <- ifelse(model == "one_piece", "NA", x0_flag)

        print_get_prior <- TRUE ## reset value
        print_prior_summary <- TRUE

        for(displ_value in values_displ_used) {

          ## used when loading fits
          fit_index <- fit_index + 1
          ## Set up variables for saving model and fit

          desc_short <- paste0("x0: " , x0_label, "; y0: ", y0_flag, "; displ_flag: ", displ_flag)
          desc <- paste0(sampling, "; ", model, "; ", desc_short)

          filename_desc <- gsub("_", "-", desc) %>%
            gsub("; ", "_", .) %>%
            gsub(":" , "-", .)

          #stan_model_name <- sub(filename_desc, "_displ-prior-[0-9.]+"_fil
          stan_model_name <- filename_desc #sub("_displ-prior-[0-9.]+"_fil

          curr_row <- which_tblr_row(male_filter,
                                x0_flag,
                                y0_flag,
                                displ_flag,
                                displ_value,
                                model,
                                fit_tblr)

          fit_tblr[ curr_row, ]$desc <- desc

          print(desc)

          if(run_fits) {

            print("Fitting Models")
            switch(sampling,
                  "nbinom_type1"= {

```

```

family <- nbinom_type1(link = "identity", link_disp = "identity")
adapt_delta <- adapt_delta #0.95 ## will decreasing value increa
iter <- iter
warmup <- warmup
thin <- thin
n_cores <- 6 ## set to 1 if getting errors from stan in order to
n_chains <- n_cores
stanvar_func <-
  stanvar(scode = paste(
    stan_func,
    stan_nbinom_type1, sep = "\n"),
    block = "functions")
}
)

## Refresh data in case x0_group or y0_group are all set to 1
data <- data_stan
if(male_filter) data <- data %>% filter(!(male %in% male_exclude))

males_used <- unique(data$male)

if(model == "one_piece") data <- data %>% filter(x < xignore)

print("Set flags based on fitted model structure")
if(x0_flag %in% c("uniform_1", "groups_1")) data <- mutate(data, x0_group =
if(y0_flag %in% c("uniform_1", "groups_1")) data <- mutate(data, y0_group =
if(x0_flag %in% c("individual")) data <- mutate(data, x0_group = male)
if(y0_flag %in% c("individual")) data <- mutate(data, y0_group = male)

## Note we need to put a tibble into a list because row updates, even if do
## just one cell, require a list format.
## Haven't defined "y0_group" or "x0_group" variables
if(FALSE) {
  fit_tbl[[curr_row, "x0_group_list"]] <- list(unique(data[, c("male", "x0_group")]))
  fit_tbl[[curr_row, "y0_group_list"]] <- list(unique(data[, c("male", "y0_group")]))
}

print("Define parameter formulas")
x0_form <- switch(x0_flag,
  uniform_1 = formula(x0 ~ 0 + Intercept),
  uniform_2 = formula(x0 ~ 0 + x0_group),
  ## `0 + Intercept` avoids prior being defined on centered
  groups_1 = formula(x0 ~ 0 + Intercept + (1|male)),
  groups_2 = formula(x0 ~ 0 + Intercept + (1|male) + x0_group),
  individual = formula(x0 ~ 0 + male) ## Do not use 1 + mal
)

phi_form <- formula(deparse(x0_form) %>% gsub("x0", "phi", .))

y0_form <- switch(y0_flag,
  uniform_1 = formula(y0 ~ 0 + Intercept),
  ## `0 + Intercept` avoids prior being defined on centered
  uniform_2 = formula(y0 ~ 0 + y0_group),

```

```

groups_1 = formula(y0 ~ 0 + Intercept + (1| male)),
groups_2 = formula(y0 ~ 0 + (1| male) + y0_group),
individual = formula(y0 ~ 0 + male)
## ~-1 + ` gives me the error:
## Warning in parallel::mclapply(1:chains, FUN = callFun
## 4 function calls resulted in an error
## Error in FUN(X[[i]], ...) :
## trying to get slot "mode" from an object (class "try-
## should I use `0 +` or ~-1 +` ?
)

threshold_form <- switch(model,
  two_piece = x0_form,
  one_piece = NULL,
  asymptotic = phi_form
)

disp_form <- switch(disp_flag,
  uniform_1 = NULL, #formula(disp ~ 1),
  groups_1 = formula(disp ~ 0 + Intercept + (1|male)),
  individual = formula(disp ~ 0 + male)
)

nlform <- switch(model,
  two_piece = bf(y ~ two_piece(x, x0, y0), nl = TRUE),
  one_piece = bf(y ~ one_piece(y0), nl = TRUE),
  asymptotic = bf(y ~ asymp(x, phi, y0), nl = TRUE)
) +
  threshold_form +
  disp_form +
  y0_form

print("Define priors")

# pass disp_value via stanvar argument
stanvar_prior <- stanvar(disp_value, name = "disp_value")

prior_string <- if(disp_value == "flat") {
  "uniform(0, 20)"
} else {
  # encode non-flat prior here, which force recompiling when disp_
  #paste0("exponential(", disp_value, ")")
  # pass disp_value via stanvar argument
  # Allows disp_value to be changed w/o recompiling
  "exponential(disp_value)"
}

## x0 only used in two_piece model
x0_prior <- switch(x0_flag,
  uniform_1 = NULL,
  uniform_2 = NULL,
  groups_1 = prior(student_t(3, 0, 66.7), lb = 0, ub = 10,

```

```

        groups_2 = NULL,
        individual = NULL
    )

x0_priors <- prior(uniform(32, 44.5), lb = 32, ub = 44.5, nlpar = "x0") + x0_priors

phi_priors <- prior(uniform(0.1, 100), lb = 0.01, ub = 17, nlpar = "phi")

y0_priors <- switch(prior_shape_y0,
    ## Values based on calculations at top of file using `t`
    normal = prior(normal(125, 500), nlpar = "y0", lb = 10,
    # flat prior
    # - consistent with fact we're working with motifs, not songs
    # - avoids bimodal posterior sampling issues with T235 and 236
    flat = prior(uniform(10, 1000), nlpar = "y0", lb = 10,
    )

threshold_priors <- switch(model,
    two_piece = x0_priors,
    one_piece = NULL,
    asymptotic = phi_priors
    )

disp_priors <- switch(disp_flag,
    uniform_1 = set_prior(prior_string, class = "disp", lb = 10,
    uniform_2 = NULL,
    groups_1 = set_prior(prior_string,
        class = "b", dpar = "disp", lb = 10,
        set_prior("uniform(0.1, 5)", class = "sd", dpar = "disp", lb = 10,
    groups_2a = NULL,
    groups_2b = NULL,
    individual = set_prior(prior_string,
        dpar = "disp", lb = 0, ub = 20,
    )
    )

prior <- switch(model,
    one_piece = {
        y0_priors + disp_priors
    },
    threshold_priors + y0_priors + disp_priors
    )

if(print_get_prior) {
    tmp <- get_prior(nlform,
        data = data,
        family = family
    )

    print(tmp,
        max.print = 500)
    print_get_prior <- FALSE # will get reset
}

```

```

stan_code <- file.path(output_dir,
                        "stan", "code", paste0(stan_model_name, ".stan"))
                        #make_stancode( .... save_model = stan_code)

fit <- brm(nlform,
          data = data,
          ## `link` refers to the mapping of the expectation of the distribution
          ## link_shape corresponds to `phi` of `stan`'s
          ## Negbinomial_2
          ## Defining `phi = mu/theta` creates a quasipoisson
          ## distribution with overdispersion parameter (1 +theta)
          family = family, #negbinomial(link = "identity", link_shape = "identity")
          prior = prior,
          stanvar = stanvar_func + stanvar_prior, ## pass prior values here
          iter = iter,
          warmup = warmup,
          thin = thin,
          silent = ifelse(interactive(), 1, 2), # 0, 1, or 2. 1 is default
          control = list(adapt_delta = adapt_delta,
                         max_treedepth = 12
                         ##model_name = desc ## Incorrect way to set this.
                         ),
          ## Ideally save model to avoid need to recompile
          stan_model_args = list(
            model_name = file.path(output_dir, "stan", "binary", stan_model_name)
          ),
          #sample_prior = "no", ## note improper priors not sampled
          ## Only print out sampling progress if in interactive mode
          refresh = ifelse(interactive(), max(iter/5, 1), 0),
          chains = n_chains,
          cores = n_cores,
          save_model = stan_code
        )

print("Prior Summary")
print(prior_summary(fit))
print("Fit Information")
print(desc)
print(fit)

          #fit_exp <- expose_functions(fit) , vectorize = TRUE)
          #fit_cr <- add_criterion(fit_exp, c("loo", "waic"))
fit_tbl[[curr_row, "fit"]] <- list(fit)
## Print current warnings
warnings(summary)
## Clear warnings()

## End if(run_fit)
} else {
  print("Working with Pre-existing Fits")

  ## Try to assign from local memory.

```

```

fit <- fit_tbl[[curr_row, "fit"]][[1]]

## Load fit if undefined or desired
if(is.na(list(fit)) | force_load) {
  print("Loading Models")
  #if(fit_index ==1)
  load(file = infile_tbl)
  fit <- fit_tbl[[curr_row, "fit"]][[1]]
}

data <- fit$data
males_used <- unique(data$male)

}

if(is.na(list(fit))) {
  warning(paste0("model fit ", desc, "does not exist.\n Skipping. "))
}else {
  ## Print and plot results, regardless of which fits one uses
  print(desc)
  if(print_prior_summary) {
    print("Fit Prior Information")
    print(prior_summary(fit)) # %>% filter(nlpar!="y0")
    print_prior_summary <- TRUE
  }
  print("Fit Information")
  print(summary(fit)) #, pars = "x0*") %>% filter(nlpar!="y0")
  fit_stan <- fit$fit
  #clean up variable names
  fit_stan_rename <-
    fit_stan %>%
    setNames(gsub("b_", "", names(.)) %>%
      gsub("(x0|phi|y0)_male(T[0-9]{3})", "\\2\\1", .) %>%
      gsub("__", "_", .) %>%
      gsub("r_male_(x0|phi|y0)\\[(T[0-9]{3}),Intercept\\]", "\\2\\1", .) %>%
      gsub("\\.", " ", .))

  ##
  vars_fit <- names(fit_stan_rename) %>% na.omit(.) %>% sort(., decreasing = TRUE)
  ncol <- 4
  hist <- stan_hist(fit_stan_rename,
    pars = vars_fit,
    bins = 30,
    ncol = ncol) +
    ggtitle(desc_short)
  print(hist)
  filename <- paste0("histogram_", filename_desc, ".pdf")
  ggsave(filename = filename, path = file.path(output_dir, "figures"),
    width = 8, height = 11, units = "in",
    scale = 0.4,
    dpi=300)

  list_plot <- list()

```



```
print(outfile_tbl)
```

```
## NULL
```

```
if(save_fits) save(fit_tbl, file = outfile_tbl)
```

Exit rendering

```
\small
```

```
knitr::knit_exit()
```