

AnalysisTools user manual

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1. Introduction

This software package is a set of utilities aimed to analyse trajectories of coarse-grained simulations. It was initially developed to work with [DL_MESO simulation package](#). However, it works with [vtf trajectory files](#), so it can be used to analyse any [vtf](#) trajectories.

The emphasis is placed on assembly of molecules, e.i., self-assembly of polymers. It therefore include utilities to determine if molecules are in aggregates and to calculate various properties of aggregates. The utilities only calculate desired quantities and write them to output text files, there is no plotting or visualisation.

Examples of the resulting data can be seen in the author's thesis (Šindelka, Karel: [The study of the association behavior of the amphiphilic copolymers in solutions containing low molar compounds by means of computer simulations](#), dissertation thesis, Charles University, 2018) as well as in papers in impacted journals (e.g., doi:[10.1039/C8CP05907A](#), [10.1134/S1811238217010052](#), or [10.1007/s00396-017-4090-0](#)).

2. Installation

All programs can be compiled using `cmake` which generates `Makefile` and subsequently running `make`. It requires `C` and `FORTRAN` compilers. The compilation should be done in a separate directory, such as `build`.

To create the `Makefile` and compile all utilities, simply run the following command (assuming `build` is a subdirectory of `AnalysisTools` root directory):

```
cmake -DCMAKE_BUILD_TYPE=Release -G "Unix Makefiles" ../
```

The binaries will be in ‘bin’ subdirectory of ‘build’.

Version for debugging can be compiled when `-DCMAKE_BUILD_TYPE=Debug` is used instead of `-DCMAKE_BUILD_TYPE=Release`.

To compile individual `C` programs using `gcc`, run (assuming you are in a subdirectory of `AnalysisTools` root directory):

```
gcc -O3 -lm ../AnalysisTools.c ../Error.c ../Options.c PATH_TO_PROGRAM -o OUTPUT_NAME.
```

3. Format of input/output files

All utilities read information about the system from **vsf/vcf files** (formatted as described below) and **FIELD** file (input file for **DL_MESO simulation package**). Particle masses and charges are read from the **FIELD** file, but all other structure information (i.e., numbers of beads/molecules, all bonds) is read from the structure file, **vsf** (Section 3.1), and from the coordinate file, **vcf** (Section 3.2). The utilities consider only bead types that are present in the **vcf** coordinate file (i.e., bead types present in the **vsf** file but not in the **vcf** file are not seen by the utilities). A description of how the system data are read is shown in Chapter 5.

Both **vsf** and **vcf** files can be generated using a **traject** utility provided by the DL_MESO simulation package (**traject-v2.5** and **traject-v2.6** provided here are modified versions from earlier DL_MESO simulation package versions). Both structure and coordinates can be in one file (with **vtf** extension) – this file can be used instead of separate **vsf** and **vcf** files.

3.1 vsf structure file

The structure file contains all information about all beads and bonds except for their Cartesian coordinates. The utilities are written for a **vsf** file created by a **traject** utility (for DL_MESO versions from 2.5 to 2.7), but other **vsf** files should work as long as they adhere to the following format.

A **vsf** file is divided into two parts. The first part contains bead definitions. Each line contains the description of a single bead and follows these rules:

- the line starts with **atom** (or just **a**)
- the second string is a bead index number that starts from 0 and increases with every subsequent line (the last bead definition line therefore shows the total number of beads in the simulation)
- the line contains bead name as **name <char(8)>**
- the first bead definition line may contain **default** instead of the index number; every bead that is not explicitly written in the bead definition lines is has the default name
- if the bead is in a molecule, the line contains molecule name (**resname <char(8)>**) and molecule id (**resid <int>**) that starts from 1

- **mass** and **charge** keywords are read if present (otherwise the mass and charge of beads is read from **FIELD**)
- other keywords are ignored

The following is an example of bead definition lines containing all required data:

```
atom default name Bead_A
atom 0 name Bead_B
atom 3 name Bead_C resname Mol_A 1
atom 4 name Bead_C resname Mol_A 1
atom 6 name Bead_C resname Mol_A 2
atom 7 name Bead_C resname Mol_A 2
atom 8 name Bead_D resname Mol_B 3
atom 9 name Bead_D resname Mol_B 3
```

In this example, there are four bead types (named **Bead_A**, **Bead_B**, and **Bead_C**, and **Bead_D**) and 10 beads in all (with indices from 0 to 9). Beads with indices 1, 2, and 5 are of the default type (**Bead_A**). There are two molecule types named **Mol_A** and **Mol_B** with molecule indices 1 to 3. All molecules with the same name must have the same structure, i.e., the same number of beads and the same bond connectivity.

The second part of a **vsf** file contains bonds definitions. Each bond definition line follows these rules:

- the line starts with **bond** (or just **b**)
- bond between two beads is specified by their indices separated by a colon (there cannot be a space between the first number and the colon)

The following is an example of bead definition lines that complement the above-shown bead definition lines:

```
bond 3: 4
# possible comment
bond 6: 7
bond 8: 9
```

In this example, there three bonds between beads with indices 3 and 4, 6 and 7, 8 and 9.

Blank lines and comments (lines beginning with **#**) are allowed in both parts of the **vsf** file.

3.2 vcf coordinate file

The coordinate file contains Cartesian coordinates of the beads and the size of the cuboid simulation box. Coordinates are read from a **vcf** file containing either ordered timesteps (Section 3.2.1) or indexed timesteps (Section 3.2.2).

An ordered **vcf** file must contain all beads defined in the **vsf** file, while an indexed **vcf** file can contain only a subset of defined beads. Both indexed and ordered **vcf** files contain a line before every timestep specifying the file type – **timestep ordered** or **timestep indexed** (the keyword **timestep** can be omitted). In both ordered and indexed **vcf** files, the size of the simulation box is given by a line **pbx <float> <float> <float>** which is located before the first coordinate block. Only **timestep** and **pbx** lines are read before the first coordinates (everything else is ignored), so **vtf** file can be used instead of a **vcf** file.

The **vcf** file may contain comment lines (beginning with **#**) and blank lines between timesteps, but the coordinate block must be continuous.

3.2.1 Ordered coordinate file

Coordinate lines in ordered **vcf** file contain only the Cartesian coordinates of the beads in the form **<float> <float> <float>**. The beads are written in ascending order of their indices as defined in the **vsf** file. The following is an example of an ordered **vcf** file:

```
# any number of comments or blank lines

timestep ordered
pbx 10 10 10
0.0 0.0 0.0
0.5 0.5 0.5
...

# comments between timesteps
timestep ordered
# another comment
1.0 1.0 1.0
1.5 1.5 1.5
...
```

In this example, the simulation box is cubic with side length of 10. Beginnings of two timesteps are represented by coordinates of the first two beads with indices 0 and 1 (as defined in the **vsf** file).

3.2.2 Indexed coordinate file

Indexed coordinate files contains not only Cartesian coordinates, but also bead indices (preceding the coordinates). Therefore an indexed timestep does not have to contain all beads in the **vsf** structure file. Moreover, the beads do not have to be ordered according to their ascending indices. The following is an example of an ordered **vcf** file:

```
# any number of comments or blank lines

timestep indexed
pbc 10 10 10
  2 0.5 0.5 0.5
21 0.0 0.0 0.0
...

# comments between timesteps
timestep indexed
# another comment
21 1.0 1.0 1.0
  2 1.5 1.5 1.5
...
```

This example is similar to that for the ordered **vcf** file, but two beads have indices 2 and 21 instead of 0 and 1.

3.3 Aggregate file (**agg**)

The aggregate file with **agg** extension is generated using **Aggregates** utility. The file contains information about the number of aggregates in each timestep and which molecules and monomeric beads belong to which aggregate. It serves as an additional input file for utilities that calculate properties of whole aggregates – **agg** file is therefore linked to the **vcf** that was used to generate it.

The **agg** file is a simple text file. The first line contains the command used to generate it – parts of this command can be necessary for subsequent analysis of aggregates. The second line is blank and from the third line the data for individual timesteps are shown. Each timestep follows these rules:

- each timestep starts with **Step:** <int> (only **Step** keyword is read by the utilities)
- the second line contains the number of aggregates in the given timestep and followed by a blank line

- there are two lines for each aggregate:
 - (1) number of molecules in the aggregate followed by their indices taken from the **vsf** file
 - (2) number of monomeric beads in the aggregate followed by their indices taken from the **vsf** file
- no blank or comment lines are allowed inside the aggregate block
- all molecules present in the **vcf** file used to generate this file must be present in every timestep; here, aggregate can also refer to dissolved molecules

Following is an example of an **agg** file:

```
Aggregates in.vcf 1 1 out.agg A
```

```
Step: 1
```

```
2
```

```
2 : 1 3
```

```
3 : 10 100 1000
```

```
1 : 2
```

```
1 : 20
```

```
Step: 2
```

```
1
```

```
3 : 1 2 3
```

```
4 : 10 20 100 2000
```

```
Last Step: 2
```

In this example, command **Aggregates in.vcf 1 1 out.agg A** was used to generate the file (see Section 4.2 for details about this utility). There are two timesteps here – the first contains two aggregates (although one of them is a free, dissolved molecule) and the second a single aggregate. As an example, the aggregate in the second step contains three molecules with indices 1, 2, and 3 (taken from the **vsf** file) and four monomeric beads (i.e., solvent or counterions) with indices 10, 20, 100, and 2000 (again, taken from the **vsf** file).

Besides using this file for further analysis using other utilities, the indices can be used in **vmd** to visualize, e.g., only a specific aggregate.

4. Utilities

All utilities have command line help with short description when `-h` argument is used. Besides `-h`, most of the utilities have several standard command line options that are the same. The standard options can be used with any utility unless stated otherwise.

Standard options

<code>-i <name></code>	use custom <code>vsf</code> file instead of <code>traject.vsf</code>
<code>-v</code>	verbose output that provides information about all bead and molecule types
<code>-V</code>	detailed verbose output that provides information about all individual molecules as well as about bead and molecule types
<code>-s</code>	run silently, i.e., without any output at all (overrides <code>-v</code> and <code>-V</code> options)
<code>--script</code>	do not rewrite terminal line (useful if output is routed to a file)
<code>-h</code>	print help and exit

4.1 AngleMolecules

This utility calculates angles between beads in each molecule of specified molecule type(s). The beads do not have to be connected, so the angle does not have to be between two bonds.

The angle is specified by three bead indices (according to the order of beads in the molecule in `vsf` – similarly to the `-c` option in `DensityMolecules`, i.e., Section 4.7). More than one angle can be specified (i.e., a multiple of three numbers have to be supplied to the `-a` option.). For example, assuming indices 1 2 3 are specified (default if `-a` option is not used), the angle will be between lines defined by beads with indices 1 2 and 2 3. The angle is calculated in degrees and is between 0 and 180°.

For now, `AngleMolecules` writes angles in all individual molecules in each timestep into the output file – it does not calculate averages or distribution of angles.

Usage:

AngleMolecules <input> <mol name(s)> <options>

Mandatory arguments

<input>	input coordinate file (either vcf or vtf format)
<mol name(s)>	molecule name(s) to calculate angles for

Non-standard options

--joined	specify that <input> contains joined coordinates (i.e., periodic boundary conditions for molecules do not have to be removed)
-a <name> <ints>	output file (default: angle.txt) and a multiple of three indices for angle calculation (default: 1 2 3)
-st <int>	starting timestep for calculation (default: 1)

4.2 Aggregates and Aggregates-NotSameBeads

These utilities determine which molecules belong to which aggregates according to a simple criterion: two molecules belong to the same aggregate if they share at least a specified number of contact pairs. A contact pair is a pair of two beads belonging to different molecules which are closer than a specified distance.

The number of contact pairs, the distance, and bead type(s) to use for aggregate determination are all arguments of the utilities. Any molecule type(s) can be excluded from aggregate determination (and therefore from the output **agg** file).

While the **Aggregates** utility uses all possible pairs of given bead types, **Aggregates-NotSameBeads** does not use same-type pairs. For example, if bead types A and B are given, **Aggregates** will use all three possible bead type pairs (i.e., A-A, A-B, and B-B), but **Aggregates-NotSameBeads** will use only A-B bead type pairs.

Also, periodic boundary conditions can be removed from whole aggregates and the new coordinates saved to an indexed **vcf** file. Therefore aggregates will not be split by simulation box boundaries when viewing in **vmd**.

Usage:

Aggregates (or **Aggregates-NotSameBeads**) <input> <distance> <contacts>
 <output.agg> <bead type name(s)> <options>

Mandatory arguments

<input>	input coordinate file (either vcf or vtf format)
---------	--

<code><distance></code>	minimum distance for two beads to be in contact (thus constituting a contact pair)
<code><contacts></code>	minimum number of contact pairs between two molecules to be in one aggregate
<code><output.agg></code>	output agg file (must end in .agg) with aggregate information
<code><bead type(s)></code>	bead type name(s) to use for determining contact pairs (at least two for Aggregates-NotSameBeads)
<code><options></code>	
Non-standard options	
<code>-x <mol name(s)></code>	exclude specified molecule type(s) from aggregate determination (and from the output agg file)
<code>-j <output.vcf></code>	output vcf file with coordinates of joined aggregates (i.e., without periodic boundary conditions)

4.3 Average

This utility uses the binning method to analyse data in a text file. It does not use any of the standard options.

Average calculates average, statistical error, and estimate of the autocorrelation time τ . Empty lines and comments (lines beginning with **#**) are skipped. **Average** prints to standard output (i.e., the screen) four numbers: `<n_blocks>` `<average>` `<std error>` `<tau estimate>`:

<code><n_blocks></code>	number of blocks used for the binning analysis
<code><average></code>	simple arithmetic average
<code><std error></code>	one- σ statistical error
<code><tau estimate></code>	estimate of autocorrelation time τ

A way to quickly estimate a ‘real’ value of τ is to use a wide range of `<n_blocks>` value and plot the `<tau estimate>` values as a function of `<n_blocks>`. Because the number of data points in one block of the binning analysis should be significantly larger than τ (e.g., ten times larger), plotting $f(x) = (\text{number of data lines in the file}) / (10x)$ will produce an exponential function that intersects the `<tau estimate>` line. A value of `<tau estimate>` near the intersection (but to the left, where the exponential is above `<tau average>`) can be used as a good estimate of τ .

Usage:

Average <input> <column> <discard> <n_blocks>

Mandatory arguments

<input>	input text file
<column>	column number in <input> for data analysis
<discard>	number of lines to discard from the beginning of <input>
<n_blocks>	number of blocks for binning analysis

4.4 BondLength

This utility calculates a distribution of bond lengths in specified molecule(s). For each of the specified molecule type(s), **BondLength** calculates bond lengths between all different types of connected bead pairs.

For example, assume two linear molecule types **Mol_1** and **Mol_2** both composed of bead types **A** and **B**. **Mol_1** is connected like this: **A-A-B**; **Mol_2** like this: **A-B-B**. If both molecule types are used, **BondLength** calculates for each molecule type distribution of lengths for bonds **A-A**, **A-B**, and **B-B** (separate for each molecule even though the molecules share the same bead types).

Furthermore, **BondLength** can also calculate distribution of distances between specified beads in each of the specified molecules. The **-d** option takes as arguments pairs of bead indices (according to the order of beads in the molecule in **vsf** – similarly to the **-c** option in **DensityMolecules**, i.e., Section 4.7). More than one pair can be specified. These indices are the same for all <mol name(s)>. If an index higher than the number of beads in the molecule is provided, the utility takes the last bead of the molecule (i.e., the highest index). For example, using **-d file.txt 1 2 1 999** would write two distributions for each <mol name(s)> into <file.txt>: distribution of distances between the first and second beads in each <mol name(s)> and between the first and last (or 999th bead).

Usage:

BondLength <input> <width> <output> <mol name(s)> <options>

Mandatory arguments

<input>	input coordinate file (either vcf or vtf format)
<width>	width of each bin of the distribution
<output>	output file with distribution of bond lengths
<mol name(s)>	molecule name(s) to calculate bond lengths for

Non-standard options

<code>-st <int></code>	starting timestep for calculation (default: 1)
<code>-d <output> <int(s)></code>	write distribution of distances between specified beads in each specified molecule to <code><output></code>

4.5 Config

This utility creates DL_MESO CONFIG file. It requires input coordinate file with all beads; otherwise the utility will still run, but will produce incomplete CONFIG.

Usage:

`Config <input.vcf> <options>`

Mandatory argument

<code><input></code>	input coordinate file (either <code>vcf</code> or <code>vtf</code> format)
----------------------------	--

Non-standard option

<code>-st <int></code>	timestep for creating CONFIG file from (default: last step)
------------------------------	---

There is also utility `Config_from_xyz` which takes a `xyz` coordinate file and creates the DL_MESO CONFIG file. Because `xyz` file does not contain information about the simulation box, the resulting CONFIG file must be modified manually – `Config_from_xyz` prints `x`, `y`, and `z` into the output file where box dimensions should be.

Usage:

`Config_from_xyz <input.xyz> <options>`

Mandatory argument

<code><input></code>	input <code>xyz</code> coordinate file
----------------------------	--

Non-standard option

<code>-st <int></code>	timestep for creating CONFIG file from (default: last step)
------------------------------	---

4.6 DensityAggregates

This utility calculates radial density profiles (RDP, or radial number densities) for bead types in an aggregate with specified size (the number of molecules or aggregation number, A_S) from its centre of mass.

$RDP_i(r)$ of bead type i , where r is distance from an aggregate's centre of mass, is the number of these beads in a spherical shell between the distances r and $r + dr$ (in `DensityAggregates`, dr is the `<width>` argument) divided by the volume of this shell. The distance written in the output file is always $r + 0.5dr$. Besides RDP, the output file also contains radial number profiles (i.e., the number of beads in the spherical shell not divided by the shell's volume) and one-sigma errors for both RDP and radial number profiles. The output file also contains a header describing what is in each column.

Instead of 'true' aggregate size, a number of molecules of specific type(s) can be used (`-m` option). For example, an aggregate containing 1 `Mol_A` molecule and 2 `Mol_B` molecules (i.e., three molecules in all) can be specified in several ways:

- (1) with `<agg size(s)>` of 3;
- (2) with `<agg size(s)>` of 3 and `-m Mol_A Mol_B`;
- (3) with `<agg size(s)>` of 1 and `-m Mol_A`; or
- (4) with `<agg size(s)>` of 2 and `-m Mol_B`.

Care must be taken when different molecule types share the same bead type. If one bead type is in more molecule types, the resulting density for that bead type will be the sum of its densities from all molecule types it appears in. The `-x` option can overcome this – specific molecule types can be excluded from density calculations, i.e., density of beads in the excluded molecule types will not be calculated. For example, assume two molecule types – `Mol_1` and `Mol_2`. `Mol_1` contains bead types A and B; `Mol_2` contains bead types A and C. Depending on whether and how the `-x` option is used, the utility will calculate:

- (1) densities of A, B, and C beads (density of A beads is a sum from both molecules), if no `-x` is used;
- (2) densities of only A and B beads (with A beads only from `Mol_1`), if `-x Mol_2` is used;
- (3) densities of only A and C beads (with A beads only from `Mol_2`), if `-x Mol_1` is used; or
- (4) no densities at all if `-x Mol_1 Mol_2` is used.

Therefore, to be able to plot density of A beads from `Mol_1` and `Mol_2` separately, (2) and (3) should be used (i.e., `DensityAggregates` should be run twice).

Usage:

```
DensityAggregates <input> <output.agg> <width> <output.rho> <agg
size(s)> <options>
```

Mandatory arguments

<code><input></code>	input coordinate file (either <code>vcf</code> or <code>vtf</code> format)
<code><input.agg></code>	input <code>agg</code> file

<code><width></code>	width of each bin of the distribution
<code><output.rho></code>	output file(s) (one per aggregate size) with automatic <code>#.rho</code> ending (<code>#</code> is aggregate size)
<code><agg size(s)></code>	aggregate size(s) (the number of molecules in an aggregate or the aggregation number, A_S) to calculate density for
Non-standard options	
<code>--joined</code>	specify that <code><input></code> contains joined coordinates (i.e., periodic boundary conditions for aggregates do not have to be removed)
<code>-n <int></code>	number of bins to average to get smoother density (default: 1)
<code>-st <int></code>	starting timestep for calculation (default: 1)
<code>-m <mol name(s)></code>	instead of 'true' aggregate size, use the number of specified molecule type(s) in an aggregate
<code>-x <mol name(s)></code>	exclude specified molecule type(s) (i.e., do not calculate density for beads in molecules <code><mol name(s)></code>)

4.7 DensityMolecules

This utility works similarly to **DensityAggregates**, only instead for whole aggregates, RDPs are calculated for individual molecules. Similarly to **DensityAggregates**, the output file(s) also contain statistical errors and radial number profiles.

By default, the utility calculates RDPs from the molecule's centre of mass, but specified bead in the molecule can be used instead (`-c` option). The bead is specified by its index inside the molecule as defined in the **vsf** file. For example, assume that molecule's beads in **vsf** are ordered **A**, **B**, **C**. Then, bead **A** is 1, bead **B** is 2, and **C** is 3. The index for the bead RDP should be calculated from is an argument of the `-c` option. The number of arguments of `-c` must be equal to the number of specified molecule type(s). The order for the arguments is taken from the **vsf** file, not from the **DensityMolecules** command. For example, assume that RDPs for molecules **Mol_A** (with centre at the third bead) and **Mol_B** (with centre at the centre of mass) should be calculated. The form of the `-c` option depends on the order of when the first molecules **Mol_A** and **Mol_B** appear in the **vsf** file:

- (1) if **Mol_A** is above **Mol_B** in **vsf**, then it must be `-c 3 x` (where **x** means the centre of mass);
- (2) if **Mol_B** is above **Mol_A** in **vsf**, then it must be `-c x 3`.

Usage:

```
DensityMolecules <input> <width> <output> <mol name(s)> <options>
```

Mandatory arguments	
<code><input></code>	input coordinate file (either <code>vcf</code> or <code>vtf</code> format)
<code><width></code>	width of each bin of the distribution
<code><output></code>	output file(s) (one per molecule type) with automatic <code><mol_name>.rho</code> ending
<code><mol_name(s)></code>	molecule name(s) to calculate density for
Non-standard options	
<code>--joined</code>	specify that <code><input></code> contains joined coordinates (i.e., periodic boundary conditions for molecules do not have to be removed)
<code>-n <int></code>	number of bins to average to get smoother density (default: 1)
<code>-st <int></code>	starting timestep for calculation (default: 1)
<code>-c <int>/x</code>	use specified bead (<code><int></code>) or centre of molecule's mass (<code>x</code>) as centre (i.e., as $r = 0$)

4.8 DihedralMolecules

This utility calculates angles between specified planes in each molecule of specified molecule type(s). The planes in a molecule are arbitrary, so they can represent true dihedral angles or improper dihedrals.

The angle is specified by six bead indices (according to the order of beads in the molecule in `vsf` – similarly to the `-c` option in `DensityMolecules`, i.e., Section 4.7). The first three indices specify one plane and the next three the other. For example, assuming indices 1 2 3 4 5 6, the first plane is specified by the first three beads in the molecule; second plane by the next three beads (beads 4 through 6). The default indices (i.e., if `-a` option is not used) are 1 2 3 2 3 4.

More than one angle can be specified (i.e., a multiple of six numbers have to be supplied to the `-a` option.).

For now, `DihedralMolecules` writes angles in all individual molecules in each timestep into the output file – it does not calculate averages or distribution of angles.

Usage:

`DihedralMolecules <input> <mol_name(s)> <options>`

Mandatory arguments	
<code><input></code>	input coordinate file (either <code>vcf</code> or <code>vtf</code> format)
<code><mol_name(s)></code>	molecule name(s) to calculate angles for

Non-standard options

<code>--joined</code>	specify that <code><input></code> contains joined coordinates (i.e., periodic boundary conditions for molecules do not have to be removed)
<code>-a <name> <ints></code>	output file (default: <code>dihedral.txt</code>) and a multiple of six indices for angle calculation (default: 1 2 3 2 3 4)
<code>-st <int></code>	starting timestep for calculation (default: 1)

4.9 DistrAgg

This utility calculates average aggregate mass and aggregation number for each timestep (i.e., time evolution) and the averages over all timesteps. It calculates number, weight, and z averages. It also calculates distribution functions of aggregation sizes and volumes.

For a quantity \mathcal{O} , the number, weight, and z averages, $\langle \mathcal{O} \rangle_n$, $\langle \mathcal{O} \rangle_w$, and $\langle \mathcal{O} \rangle_z$, respectively, are defined as

$$\langle \mathcal{O} \rangle_n = \frac{\sum_i \mathcal{O}_i}{N}, \quad \langle \mathcal{O} \rangle_w = \frac{\sum_i m_i \mathcal{O}_i}{\sum_i m_i}, \text{ and } \langle \mathcal{O} \rangle_z = \frac{\sum_i m_i^2 \mathcal{O}_i}{\sum_i m_i^2}, \quad (4.1)$$

where N is the total number of measurements, e.g., the total number of aggregates if a utility calculates per-aggregate averages (or molecules if a utility calculates per-molecule averages); m_i is mass of an aggregate i (or molecule i).

Per-timestep averages are written to the `<output avg>` and overall averages are appended as comments (with commented legend) to both `<output avg>` and `<output distr>` files.

Number, weight, and z distribution functions of aggregate sizes, $F_n(A_S)$, $F_w(A_S)$, and $F_z(A_S)$ respectively, are defined as

$$\begin{aligned} F_n &= \frac{N_{A_S}}{\sum_{A_S} N_i} = \frac{N_{A_S}}{N}, \\ F_w &= \frac{N_{A_S} m_{A_S}}{\sum_{A_S} N_i m_i} = \frac{N_{A_S} m_{A_S}}{\sum_{i=1}^N m_i} = \frac{N_{A_S} m_{A_S}}{M}, \text{ and} \\ F_z &= \frac{N_{A_S} m_{A_S}^2}{\sum_{A_S} N_i m_i^2} = \frac{N_{A_S} m_{A_S}^2}{\sum_{i=1}^N m_i^2}, \end{aligned} \quad (4.2)$$

where N_{A_S} and m_{A_S} stand for the number and mass, respectively, of aggregates with aggregate size A_S ; M is the total mass of all aggregates. The equations are normalised so that $\sum F_x(A_S) = 1$.

Distribution of volume fractions of aggregates, $\phi(A_S)$, is defined (assuming all beads have the same volume) as

$$\phi(A_S) = \frac{N_{A_S} m_{A_S}}{\sum_{i=1}^N n_i} = \frac{N_{A_S} m_{A_S}}{n}, \quad (4.3)$$

where n_i is the number of beads in aggregate i and n is the total number of beads in all aggregates. If all beads have unit mass (as is often the case in dissipative particle dynamics), the volume distribution, $\phi(A_S)$, is the equal to the number distribution, $F_n(A_S)$. These distribution are written into the `<output distr>` file.

Lastly, **DistrAgg** can calculate number distribution of composition for aggregates with specified size(s) (`-c` option). This is a number distribution of the numbers of different molecule types in the aggregate. For example, if the simulation box contains molecule types **Mol_A** and **Mol_B**, aggregates with the same size can contain different numbers of these molecules, or different ratios of the numbers of **Mol_A** to **Mol_B** molecules, $\xi = N_{\text{Mol_A}}/N_{\text{Mol_B}}$. For now, **DistrAgg** can calculate this distribution only for aggregates containing two molecule types. The composition distribution is defined as

$$F_n(\xi) = \frac{N_{\xi, A_S}}{N_{A_S}}, \quad (4.4)$$

where N_{ξ, A_S} is the number of aggregate with aggregate size A_S and ratio ξ ; N_{A_S} is the total number of aggregates with aggregate size A_S .

The definition of aggregate size is flexible. If none of `-m`, `-x`, `--only` options is used, aggregate size is the ‘true’ aggregation number, A_S , i.e., the number of all molecules in the aggregate; if `-m` is used, aggregate size is the sum of only specified molecule type(s); if `-x` is used, aggregates containing only specified molecule type(s) are disregarded; if `--only` is used, only aggregates composed of the specified molecule type(s) are taken into account. For example, consider a system in the following table containing three aggregates composed of various numbers of three different molecule types:

Molecule types	Aggregate composition
Mol_A	Agg_1: 1 Mol_A + 2 Mol_B + 3 Mol_C = 6 molecules
Mol_B	Agg_2: 1 Mol_A + 2 Mol_B = 3 molecules
Mol_C	Agg_3: 1 Mol_A = 1 molecule

Here is a list of several possibilities depending on the option(s) used:

- (1) if none of `-m`, `-x`, `--only` is used, all three aggregates are counted and their sizes are their aggregation numbers, i.e., $A_S = 6, 3$, and 1
- (2) if `-m Mol_A Mol_B` is used, all three aggregates are counted, but their size is the sum of only **Mol_A** and **Mol_B** molecules: **Agg_1** – 3; **Agg_2** – 3; **Agg_3** – 1

- (3) if `-m Mol_B Mol_C` is used, `Agg_3` is not counted, because its size would be zero; `DistrAgg` would detect only two aggregates with sizes: `Agg_1 - 5`; `Agg_2 - 2`
- (4) if `-x Mol_A Mol_B` is used, `Agg_2` and `Agg_3` are not counted, because neither contains anything else than `Mol_A` and/or `Mol_B`; `DistrAgg` would detect only one aggregate with size: `Agg_1 - 6`
- (5) if `-x Mol_A Mol_B` is combined with `-m Mol_A Mol_B`, `DistrAgg` would again detect only `Agg_1`, but its size would be taken as 3
- (6) if `--only Mol_A Mol_B` is used, `Agg_1` is not counted, because it contains a molecule not specified by `--only`; `DistrAgg` would detect only two aggregates with sizes: `Agg_2 - 3`; `Agg_3 - 1`
- (7) if `--only Mol_A Mol_B` is combined with `-m Mol_A`, the two detected aggregates have sizes: `Agg_2 - 1`; `Agg_3 - 1`

Usage:

`DistrAgg <input.agg> <output distr> <output avg> <options>`

Mandatory arguments

<code><input.agg></code>	input agg file
<code><output distr></code>	output file with distribution of aggregate sizes
<code><output avg></code>	output file with per-timestep averages

Non-standard options

<code>-st <int></code>	starting timestep for calculation (default: 1)
<code>-n <int> <int></code>	use aggregate sizes in given range
<code>-m <mol name(s)></code>	use number of specified molecule type(s) as aggregate size
<code>-x <mol name(s)></code>	exclude aggregates containing only specified molecule type(s)
<code>--only <mol name></code>	use only aggregates composed of specified molecule type
<code>-c <output> <int(s)></code>	save composition distribution for specified aggregate size(s) to <code><output></code> file

4.10 GyrationAggregates

This utility calculates the gyration tensor and its eigenvalues (or the roots of the tensor's characteristic polynomial) for all aggregates. Using the eigenvalues, the utility calculates shape descriptors: radius of gyration, asphericity, acylindricity,

and relative shape anisotropy.

The eigenvalues, λ_x^2 , λ_y^2 , and λ_z^2 , (sorted so that $\lambda_x^2 \leq \lambda_y^2 \leq \lambda_z^2$) are also written to output file(s), because their square roots represent half-axes of an equivalent ellipsoid.

The radius of gyration, R_G , is defined as

$$R_G^2 = \lambda_x^2 + \lambda_y^2 + \lambda_z^2. \quad (4.5)$$

The asphericity, b , and the acylindricity, c , are defined, respectively, as

$$b = \lambda_z^2 - \frac{1}{2}(\lambda_x^2 + \lambda_y^2) = \frac{3}{2}\lambda_z^2 - \frac{R_G^2}{2} \quad \text{and} \quad c = \lambda_y^2 - \lambda_x^2. \quad (4.6)$$

Number average of all properties and, additionally, weight and z averages for the radius of gyration are calculated (see Equation (4.1) in Section 4.9 for general definitions of averages). Per-timestep averages (i.e., time evolution) are written to the <output> file. To save averages for aggregate sizes, **-ps** option can be used.

The shape descriptors are by default calculated for all beads in the aggregates, but **-bt** option can be used to specify which bead types to use.

Similarly to **DistrAgg**, the definition of aggregate size is flexible – see Section 4.9 for explanation of the **-m** and **-x** options.

Usage:

GyrationAggregates <input> <input.agg> <output> <options>

Mandatory arguments

<input>	input coordinate file (either vcf or vtf format)
<input.agg>	input agg file
<output>	output file with per-timestep averages

Non-standard options

--joined	specify that <input> contains joined coordinates (i.e., periodic boundary conditions for aggregates do not have to be removed)
-bt <bead name(s)>	bead type(s) used for calculation
-ps <name>	output file with per-size averages
-m <mol name(s)>	instead of ‘true’ aggregate size, use the number of specified molecule type(s) in an aggregate
-x <mol name(s)>	exclude aggregates containing only specified molecule type(s)
-n <int> <int>	use only aggregate sizes in given range
-st <int>	starting timestep for calculation (default: 1)

4.11 GyrationMolecules

This utility calculates shape descriptors similarly to `GyrationAggregates` (Section 4.10), but for individual molecules instead for whole aggregates.

Usage:

```
GyrationMolecules <input> <output> <mol name(s)> <options>
```

Mandatory arguments

<input>	input coordinate file (either <code>vcf</code> or <code>vtf</code> format)
<output>	output file(s) (one per molecule type) with automatic <code>-<mol name>.txt</code> ending
<mol name(s)>	molecule name(s) to calculate shape descriptors for

Non-standard options

<code>--joined</code>	specify that <input> contains joined coordinates (i.e., periodic boundary conditions for molecules do not have to be removed)
<code>-bt <bead name(s)></code>	bead type(s) to be used for calculation
<code>-st <int></code>	starting timestep for calculation (default: 1)

4.12 JoinAggregates

This utility is meant for cases when non-standard option `-j` is omitted from `Aggregates` (or `Aggregates-NotSameBeads`) command. `JoinAggregates` uses the provided `vcf` and `agg` files to join aggregates, i.e., to remove periodic boundary conditions and save the new coordinates into a `vcf` file. The utility reads `Aggregates` command from the `agg` file to determine distance and number of contact pairs for aggregate check (see Section 4.2 for details on `Aggregates` utility).

Usage:

```
JoinAggregates <input> <input.agg> <output.vcf> <options>
```

Mandatory arguments

<input>	input coordinate file (either <code>vcf</code> or <code>vtf</code> format)
<input.agg>	input <code>agg</code> file
<output.vcf>	output <code>vcf</code> coordinate file with indexed coordinates

Non-standard options

<code>-st <int></code>	starting timestep for calculation (default: 1)
------------------------------	--

4.13 JoinRuns

This utility joins two independent simulation runs of the same system. That is, the system contains identical beads and molecules, but these beads and molecules are numbered differently in the `vsf` and `vcf` files from different simulations. The two input `vcf` files must contain the same timestep type (i.e., both indexed or both ordered) and the same number of beads (i.e., if one bead type is absent from one `vcf` file, it must be absent from the second one as well).

Usage:

```
JoinRuns <1st input> <2nd input> <2nd vsf> <output> <bead type(s)>
<options>
```

Mandatory arguments

<code><1st input></code>	input coordinate file from the first simulation (either <code>vcf</code> or <code>vtf</code> format)
<code><2nd input></code>	input coordinate file from the second simulation (either <code>vcf</code> or <code>vtf</code> format)
<code><2nd vsf></code>	input structure file from the second simulation (structure file from the first simulation is <code>traject.vsf</code> ; changeable via <code>-i</code> option)
<code><output></code>	output <code>vcf</code> coordinate file with indexed coordinates
<code><bead type(s)></code>	bead type names to save

Non-standard options

<code>--join</code>	join molecules by removing periodic boundary conditions
<code>-st1 <int></code>	starting timestep first run (default: 1)
<code>-st2 <int></code>	starting timestep second run (default: 1)
<code>-sk1 <int></code>	number of steps skip per one used for first run (default: 0)
<code>-sk2 <int></code>	number of steps skip per one used for second run (default: 0)

4.14 PairCorrel

This utility calculates pair correlation function (pcf) between specified bead types. All bead type pairs are used – if A and B bead types, A-A, A-B, and B-B bead type

pairs are used. Right now, the pcfs are not correctly normalised.

The utility do not recognise between beads of the same type that are in different molecules, so a pcf will be a sum of the beads from different molecule types.

Usage:

```
PairCorrel <input> <width> <output> <bead name(s)> <options>
```

Mandatory arguments

<input>	input coordinate file (either vcf or vtf format)
<width>	width of each bin of the pair correlation functions
<output>	output file with pair correlation functions
<bead name(s)>	bead type(s) used for calculation

Non-standard options

-n <int>	number of bins to average to get smoother pair correlation function (default: 1)
-st <int>	starting timestep for calculation (default: 1)

4.15 PotentialAggregates

This utility should be working, but it needs more testing.

This utility calculates electrostatic potential as a function of distance from the centre of mass of specified aggregate size(s). It places a virtual particle with charge $q = 1$ at several places on the surface of ever increasing sphere and calculates electrostatic potential acting on that virtual particle.

At long range, the potential is calculated using Coulomb potential,

$$U_{ij}^{\text{long}} = \frac{l_B q_i q_j}{r_{ij}}, \quad (4.7)$$

where l_B is the Bjerrum length, q_i and q_j are charges of particles i and j , and r_{ij} is interparticle distance. At short range, the potential is for now calculated using potential between two charges with exponentially decreasing charge density,

$$U_{ij}^{\text{short}} = U_{ij}^{\text{long}} [1 - (1 + \beta r_{ij}) e^{-2\beta r_{ij}}], \quad (4.8)$$

where $\beta = \frac{5r_c}{8\lambda}$ (r_c is cut off distance and λ is smearing constant). The utility takes into account periodic images of the simulation box.

For now, parameters for the potential are hard coded in the source code: the Bjerrum length is **bjerrum=1.1** (aqueous solution), cut-off is **r_c=3**, charge smearing constant **lambda=0.2**, and number of periodic images of the simulation box is **images=5**. The parameters can be changed but the utility must then be recompiled.

The aggregate size can be modified using `-m` options similarly to `DensityAggregates` (Section 4.6).

Usage:

```
PotentialAggregates <input> <input.agg> <width> <output> <agg size(s)>
<options>
```

Mandatory arguments

<code><input></code>	input coordinate file (either <code>vcf</code> or <code>vtf</code> format)
<code><input.agg></code>	input <code>agg</code> file
<code><width></code>	width of each bin of the distribution
<code><output></code>	output file(s) (one per aggregate size) with automatic <code>agg#.txt</code> ending (<code>#</code> is aggregate size)
<code><agg size(s)></code>	aggregate size(s) for calculation of electrostatic potential

Non-standard options

<code>--joined</code>	specify that <code><input></code> contains joined coordinates (i.e., periodic boundary conditions for aggregates do not have to be removed)
<code>-st <int></code>	starting timestep for calculation (default: 1)
<code>-m <mol name(s)></code>	instead of 'true' aggregate size, use the number of specified molecule type(s) in an aggregate

4.16 SelectedVcf

This utility creates a new `vcf` coordinate file containing only beads of specified types with an option of removing periodic boundary conditions (i.e., joining molecules).

The selected bead types are printed as comments at the beginning of the output `vcf` file.

Also, specified molecules can be excluded which is useful when the same bead type is shared between more molecule types. However, as of now, no utilities can read a `vcf` file that does not contain all beads of a given type, so this can be used only for `vmd` visualization.

Lastly, `xyz` coordinate file can also be created from the selected bead type(s).

Usage:

```
SelectedVcf <input> <output> <bead type(s)> <options>
```

Mandatory arguments

<input>	input coordinate file (either vcf or vtf format)
<output.vcf>	output vcf coordinate file with indexed coordinates
<bead type(s)>	bead type names to save (can be omitted if -r is used)

Non-standard options

-r	reverse function, i.e., exclude <bead type(s)> instead of including them; if no <bead type(s)> are specified, all bead types are used (requires <input> with all bead types)
--join	join molecules by removing periodic boundary conditions
-st <int>	starting timestep for calculation (default: 1)
-sk <int>	number of steps skip per one used (default: 0)
-x <name(s)>	exclude molecules of specified name(s) – do not use if output.vcf is further analysed
-xyz <name>	save coordinates to xyz file – does not take into account -x option

4.17 **traject**

This utility is from the DL_MESO simulation package and comes in three version for 2.5, 2.6, and 2.7 versions of DL_MESO. For the latest DL_MESO version, the utility is unmodified and therefore is not included here. The utilities **traject-v2_5** and **traject-v2_6** shipped with DL_MESO 2.5 and 2.6 were modified to generate separate **vsf** and **vcf** file (the **traject** for DL_MESO 2.7 does this natively with **-sc** command line option).

Usage of 2.5 and 2.6 versions (see DL_MESO manual for the description of latest version):

traject-v2_5 <int> or **traject-v2_6 <int>**

Mandatory argument

<int>	number of computer cores used for the simulation run (equals the number of HISTORY files)
--------------------	--

4.18 TransformVsf

This not-very-useful utility just rewrites **vsf** file for better visualization with **vmd**. The output **vsf** file contains not only bead name and index, but its charge and mass as well.

Usage:

```
TransformVsf <output.vsf> <options>
```

Mandatory argument

<code><output.vsf></code>	output structure file
---------------------------------	-----------------------

5. Computational details

This chapter will contain some information about how things are coded in the utilities.

5.1 Read system data

`ReadStructure()` function reads all system information from `vsf` and `vcf` files. `FIELD` file is used only to get mass and charge of beads if the information is not in `vsf` file. Provided `vsf` file is used to get all information about beads and molecules – names and numbers of bead and molecules, bonds in molecules. The first timestep of the `vcf` file is used to determine numbers and ids of beads in that `vcf` file which means that all timesteps must contain the same beads.

The procedure in `ReadStructure()` is as follows:

- (1) Go through `atom` section of the `vsf` file to identify default bead type (if `atom default` line is present), to find highest bead and molecule ids, and to find bead type names (and charges and masses if present).
- (2) Go again through the `atom` section to read names and ids of beads and molecules as well as numbers of all beads and molecules for each type.
- (3) Go through `bond` section of the `vsf` file to calculate number of bonds in each molecule type.
- (4) Go again through the `bond` section to read bonds for each molecule type.
- (5) Go through the `atom` section (for the third time) to assign bead ids to molecules.
- (6) Go through the first timestep of `vcf` file to find which beads are in that `vcf` file (if no `vcf` file is provided – e.g., for `DistrAgg` utility – assume all beads are used).
- (7) Modify all arrays to accommodate only the beads that are present in the `vcf` file.
- (8) Read charge and mass from the `FIELD` file, if not already known from `vsf` file.