# AnalysisTools user manual

Karel Šindelka k.sindelka@gmail.com April 20, 2020

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### 0. Contents

## 1. Introdution

This software package is a set of utilities aimed to analyse trajectories of coarse-grained simulations. It was initially developed to work with DL\_MESO simulation package. However, it works with vtf trajectory files, so it can be used to analyse any vtf trajectories.

The emphasis is mainly on assembly of molecules, e.g., self-assembly of polymers. It therefore includes utilities to determine if molecules are in aggregates and to calculate various properties of aggregates. Also, throughout the utilities and this manual, the term aggregates refers to any supramolecular structure (such as, e.g., a bilayer).

However, the package also contains some utilities to generate initial configurations (or adapt existing configurations) as well as to create a data file from vtf trajectories for the LAMMPS molecular dynamic simulator.

## 2. Installation

All programs can be compiled on linux using cmake which generates Makefile and subsequently running make. It requires C and FORTRAN compilers. The compilation should be done in a separate directory, such as build.

To create the Makefile, simply run cmake \$ROOT\_DIR in the separate directory, where \$ROOT\_DIR is the AnalysisTools root directory. All utilities can be then compiled by simply running make without any arguments. To compile individual utilities, simply run make <utility name>. The binaries will be in build/bin directory.

Individual utilities can also be compiled without the use of cmake (and without Makefile) by using gcc -O3 -lm \$ROOT\_DIR/AnalysisTools.c \$ROOT\_DIR/Error.c \$ROOT\_DIR/Options.c \$ROOT\_DIR/Utility/program.c.

## 3. Format of input/output files

All utilities read information about the system from vsf/vcf files (formatted as described below) and FIELD file (input file for DL\_MESO simulation package). All system information is read from the vsf structure file (Section 3.1) and from the vcf coordinate file (Section 3.2). One vtf file containing a structure and coordinate sections can also be used. The FIELD file is only used when bead charge and/or mass is missing from the vsf structure file or for generation (or modification) of configurations. The utilities consider only bead types that are present in the vcf coordinate file (i.e., bead types present in the vsf file but not in the vcf file are not seen by the utilities). Chapter 5 briefly describes how the data are read.

Both vsf and vcf files can be generated from the DL\_MESO HISTORY file using a traject utility provided by the DL\_MESO simulation package (traject-v2\_5 and traject-v2\_6 provided here are modified versions from earlier DL\_MESO simulation package versions). Both structure and coordinates can be in one file (with vtf extension) – this file can be used instead of separate vsf and vcf files.

All utilities assume cuboid simulation box with dimensions from 0 to N, where N is the side length of the box (which can be different in all three dimensions).

### 3.1 vsf structure file

The structure file contains all information about all beads and bonds except for their Cartesian coordinates. The utilities are written for a vsf file created by a traject utility (for DL\_MESO versions from 2.5 to 2.7), but other vsf files should work as long as they adhere to the following format.

A vsf file is divided into two parts. The first part contains bead definitions. Each line contains the description of a single bead and follows these rules:

- the line starts with atom (or just a)
- the second string is a bead index number that starts from 0 and increases with every subsequent line (the last bead definition line therefore shows the total number of beads in the simulation)
- the line contains bead name as name <char(8)>
- the first bead definition line may contain **default** instead of the index number; every bead that is not explicitly written in the bead definition lines is of the

default type

- if the bead is in a molecule, the line contains molecule name (resname <char(8)>) and molecule id (resid <int>) that starts from 1
- mass and charge keywords are read if present (otherwise the mass and charge of beads is read from FIELD)
- other keywords are ignored

The following is an example of bead definition lines containing all required data:

```
atom default name Bead_A
atom 0 name Bead_B
atom 3 name Bead_C resname Mol_A 1
atom 4 name Bead_C resname Mol_A 1
atom 6 name Bead_C resname Mol_A 2
atom 7 name Bead_C resname Mol_A 2
atom 8 name Bead_D resname Mol_B 3
atom 9 name Bead_A resname Mol_B 3
```

In this example, there are four bead types (named Bead\_A, Bead\_B, and Bead\_C, and Bead\_D) and 10 beads in all (with indices from 0 to 9). Beads with indices 1, 2, 5, and 9 are of the default type (Bead\_A); notice that if a bead of a default type is in a molecule, it must have its own atom line. There are two molecule types named Mol\_A and Mol\_B with molecule indices 1 to 3. All molecules with the same name must have the same structure, i.e., the same number of beads and the same bond connectivity.

The second part of a **vsf** file contains bond definitions. Each bond definition line follows these rules:

- the line starts with bond (or just b)
- bond between two beads is specified by their indices separated by a colon

The following is an example of bead definition lines that complement the aboveshown bead definition lines:

```
bond 3: 4
bond 6: 7
bond 8: 9
```

In this example, there three bonds between beads with indices 3 and 4, 6 and 7, 8 and 9

Blank lines and comments (lines beginning with #) are allowed in both parts of the vsf file. Instead of a separate vsf file, a vtf file can be used (via -i <name.vtf>command line option).

### 3.2 vcf coordinate file

The coordinate file contains Cartesian coordinates of the beads and the size of the cuboid simulation box. Coordinates are read from a vcf file containing either ordered timesteps (Section 3.2.1) or indexed timesteps (Section 3.2.2).

An ordered vcf file must contain all beads defined in the vsf file, while an indexed vcf file can contain only a subset of defined beads. Both indexed and ordered vcf files contain a line before every timestep specifying the file type — timestep ordered or timestep indexed (the keyword timestep can be omitted). In both ordered and indexed vcf files, the size of the simulation box is given by a line pbc <float> <float> <float>. Because everything up to the first timestep or pbc line is ignored, vtf file can be used instead of a vcf file.

The vcf file may contain comment lines (beginning with #) and blank lines between timesteps, but the coordinate block must be continuous.

#### 3.2.1 Ordered coordinate file

Coordinate lines in ordered vcf file contain only the Cartesian coordinates of the beads in the form <float> <float> <float>. The beads are written in ascending order of their indices as defined in the vsf file. The following is an example of an ordered vcf file:

```
timestep ordered
pbc 10 10 10
0.0 0.0 0.0
0.5 0.5 0.5
...

# comments between timesteps
timestep ordered
# another comment
```

1.0 1.0 1.0 1.5 1.5 1.5

# any number of comments or blank lines

In this example, the simulation box is cubic with side length of 10. Beginnings of two timesteps are represented by coordinates of the first two beads with indices 0 and 1 (as defined in the vsf file).

#### 3.2.2 Indexed coordinate file

Indexed coordinate file contains not only Cartesian coordinates, but also bead indices (preceding the coordinates). Therefore, an indexed timestep does not have to contain all beads in the vsf structure file; however, AnalysisTools utilities work with whole sets of beads, that is, when one bead of a specific type is missing, all beads of that type (i.e., beads with the same name) must be missing as well. Also, the beads do not have to be ordered according to their ascending indices. The following is an example of an ordered vcf file:

```
# any number of comments or blank lines
timestep indexed
pbc 10 10 10
  2 0.5 0.5 0.5
21 0.0 0.0 0.0
...

# comments between timesteps
timestep indexed
# another comment
21 1.0 1.0 1.0
  2 1.5 1.5 1.5
```

This example is similar to that for the ordered vcf file, but two beads have indices 2 and 21 instead of 0 and 1.

## 3.3 Aggregate file (agg)

The aggregate file with agg extension is generated using Aggregates (or Aggregates -NotSameBeads) utility. The file contains information about the number of aggregates in each timestep and which molecules and monomeric (i.e., unbonded) beads belong to which aggregate. It serves as an additional input file for utilities that calculate aggregate properties; agg file is, therefore, linked to the vcf file that was used to generate it.

The agg file is a simple text file. The first line contains the command used to generate it – parts of this command can be necessary for subsequent analysis of aggregates. The second line is blank, and from the third line, the data for individual timesteps are shown. It follows these rules:

- each timestep starts with Step: <int> (only Step keyword is read by the utilities)
- the second line is the number of aggregates in the given timestep and is followed by a blank line
- there are two lines for each aggregate:
  - (1) number of molecules in the aggregate followed by their indices taken from the vsf file (resid indices)
  - (2) number of monomeric beads in the aggregate followed by their indices taken from the vsf file (atom indices)
- no blank or comment lines are allowed inside the aggregate block
- not all molecules present in the vcf file used to generate this file must be present in every timestep

Note that the term aggregate also refers to free chains (i.e., fully dissolve dchains). When the keyword Last is present instead of Step, it signalises the end of the agg file; no utility will read anything beyond this keyword.

Following is an example of an agg file:

Aggregates in.vcf 1 1 out.agg A

```
Step: 1
2
2: 13
3: 10 100 1000
1: 2
1: 20
Step: 2
1
3: 123
4: 10 20 100 2000
Last Step: 2
```

anything from here on is ignored

In this example, command Aggregates in.vcf 1 1 out.agg A was used to generate the file (see Section 4.2 for details about this utility). There are two

timesteps here – the first contains two aggregates (although one of them is a free, dissolved molecule) and the second an aggregate comprised of two molecules (i.e., a dimer). As an example, the aggregate in the second step contains three molecules with indices 1, 2, and 3 (taken from the vsf file) and four monomeric beads (i.e., solvent or counterions) with indices 10, 20, 100, and 2000 (again, taken from the vsf file).

Besides using this file for further analysis by other utilities, the indices can be used in vmd to visualize, e.g., only a specific aggregate by using resid 1 2 3 in the Selected Atoms box in vmd.

## 4. Utilities

All utilities have command line help with short description when -h argument is used. Besides -h, most of the utilities have several standard command line options that are the same. The standard options can be used with any utility unless stated otherwise.

Standard options		
-i <name></name>	use custom structure file instead of traject.vsf (vsf or vtf for-	
	$\operatorname{mat}$ )	
<b>-</b> ∆	verbose output that provides information about all bead and	
	molecule types	
-s	run silently, i.e., without any output at all (overrides -v option)	
script	no progress output (useful if output is routed to a file)	
-h	print this help and exit	
version	print version of the utilities and exit	

## 4.1 AddToSystem

This utility takes an existing system specified by vcf coordinate and vsf structure files and adds new beads into it placing them either randomly or using provided coordinates. Because it is meant to produce configurations for new simulations, it required a coordinate file that contains all beads (i.e., all beads defined in the structure file, must be present in the coordinate). If an incomplete coordinate file, AddToSystem can show undefined behaviour, i.e., it can run without problems, exit with an error, or freeze.

The new beads replace either neutral unbonded ones with the lowest indices (as ordered in the vsf file) from the original system or option-specified unbonded beads. Right now, molecular beads cannot be exchanged. If molecules are added, AddToSystem places them at the end (for the sake of DL\_MESO which requires molecules to be after unbonded beads). If there are not enough of the beads in the original system replace, AddToSystem exits with an error.

The utility has two modes: by default, it reads what is to be added from a FIELD-like file (see below) and places the new beads and molecules randomly; if, however, -vtf option is used, the utility reads provided vsf and vcf files and uses

these coordinates.

If the new beads and molecules are added randomly, their coordinates are either completely random within the simulation box or ruled by constraining options; either the first bead (default behaviour) or the geometric centre (-gc option) of the new molecules obey these options. The coordinates of the remaining beads in a molecule are governed by the coordinates in the FIELD-like file. Therefore, not all the molecular beads necessarily obey the constraining rules. Molecules are added with a random orientation.

There are two types of constraining options (that can be combined): place new beads either specified distance from other beads or in a specified interval in x, y, or z directions. The first case uses otions -ld and -hd which, if present, must be accompanied by -bt option. Then, the new beads are placed at least -ld <float> and at most -hd <float> distance from beads specified by the -bt option. The second case uses options -cx, -cy, and -cz that basically change the box size for the added beads; for example, -cx 5 9 would generate x coordinates only in the interval (5,9). All these options can be combined, but note that AddToSystem does not perform any sanity checks, that is, if any combination of the provided options is impossible to achieve, the utility will run forever.

As hinted above, the structure and number of added molecules and monomeric beads are read from a FIELD-like file. This file must contain species section followed by molecule section as described in the DL\_MESO simulation package.

The species section contains the number of bead types and their properties:

```
species <int>
<name> <mass> <charge> <number of unbonded beads>
```

The first line must start with **species** keyword followed by the number of bead types. For each bead type, a single line must contain the name of the bead, its mass and charge, and a number of these beads that are not in a molecule (i.e., monomeric or unbonded beads).

The molecule section, which must be behind the species section, contains information about structure and numbers of molecules to be added:

```
molecule <int>
<name>
nummols <int>
beads <int>
<bead name> <float> <float> <float>
...
<bead name> <float> <float> <float> bonds <int>
<bead sint>
<string> <int> <int>
```

Number of types of molecules

Name of the first molecule type

Number of these molecules

Number of beads in these molecules

One line for each of the <int> beads

specifying bead name and its

Cartesian coordinates

Number of bonds in these molecules

Number of bonds in these molecules One line for each of the <int> bonds

```
containing arbitrary string and
indices of connected beads
Anything beyond here is ignored
finish

containing arbitrary string and
indices of connected beads
Anything beyond here is ignored
Description of a molecule is finished
```

The molecule keyword specifies the number of molecule types, that is the number of finish keywords that must be present. The <bed>
bead name<br/>
must be present in the species section. The arbitrary <string<br/>
in the bonds is ignored by AddToSystem (it is a relic from the DL\_MESO simulation package, where the <string<br/>
specifies a type of bond). The indices in bond lines run from 1 to the number of beads in the molecules and are ordered according to the beads part of the section. Because molecule section in the FIELD file from DL\_MESO can also include bond and dihedral angles, anything beyond the last bond line is ignored (until the finish keyword is read).

If no molecules are to be added, the line molecule 0 must be still be present in the file.

The following is an example of the FIELD-like file:

```
species 3
    1.0
        1.0
    1.0 0.0
    1.0 -1.0 30
molecule 2
Dimer
nummols 10
beads 2
A 0.0 0.0 0.0
A 0.5 0.0 0.0
bonds 1
harm 1 2
finish
surfact
nummols 10
beads 3
A 0.0 0.0 0.0
B 0.5 0.0 0.0
B 1.0 0.0 0.0
bonds 2
harm 1 2
harm 2 3
```

angles 1 harm 1 2 3 finish

In this example, 30 unbonded (or monomeric) negatively charged beads called CI are added as well as 20 molecules – 10 molecules called Dimer and 10 molecules called surfact. Dimer molecules contain two A beads and one bond each; surfact molecules contain three beads and two bonds each. The part starting with angles and ending with finish is ignored. All in all, 80 beads are added – 30 CI, 30 A, and 20 B beads.

If -vtf option is used, provided vsf and vcf files are read instead of a FIELD-like file. AddToSystem incorporates the beads and molecules from these files, using the provided coordinates instead of generating new ones. Therefore, above-described constraining options have no effect.

The -vtf option has priority over default behaviour, that is, if it is present, the utility ignores any FIELD-like file (either FIELD itself or a file provided via -f option).

The utility creates the vcf and vsf files with the new system and can also write the coordinates into a xyz file.

Usage:

AddToSystem <input.vcf> <out.vsf> <out.vcf> <options>

Mandatory arguments		
<pre><input.vcf></input.vcf></pre>	input coordinate file (either vcf or vtf format)	
<out.vsf></out.vsf>	output vsf structure file for the new system	
<out.vcf></out.vcf>	output vcf coordinate file for the new system	
Non-standard option	ns	
-f <name></name>	FIELD-like file specifying additions to the system (default: FIELD)	
-vtf <vsf> <vcf></vcf></vsf>	add ready-made system from vtf files instead of randomly placing beads read from the FIELD-like file	
-st <int></int>	<pre><input.vcf> timestep to add new beads to (default: 1)</input.vcf></pre>	
-xyz <name></name>	also save coordinates to xyz file	
-ld <float></float>	lowest distance from beads specified by -bt option	
-hd <float></float>	highest distance from beads specified by -bt option	
-bt <bead names=""></bead>	bead types to use in conjunction with <code>-ld</code> and/or <code>-hd</code> options	
-cx <num> <num2></num2></num>	constrain x coordinate to interval $\langle num, num2 \rangle$	

```
-cy <num> <num2> constrain y coordinate to interval \langle num, num2 \rangle
-cz <num> <num2> constrain z coordinate to interval \langle num, num2 \rangle
-gc use molecule's geometric centre for distance check
```

## 4.2 Aggregates and Aggregates-NotSameBeads

These utilities determine which molecules belong to which aggregates (or a different compact structure) according to a simple criterion: two molecules belong to the same aggregate if they share at least a specified number of contact pairs. A contact pair is a pair of two beads belonging to different molecules which are closer than a specified distance. The information is written in .agg text file described in Section 3.3.

Note that throughout AnalysisTools, an aggregate refers to any structure formed from molecules.

The number of contact pairs, the distance, and bead type(s) to use for aggregate determination are all arguments of the utilities. Any molecule type(s) can be excluded from aggregate determination (-x <mol name(s)> option); they are also excluded from the output agg file). Moreover, any molecules close to specified molecule(s) can be excluded (-xm <mol name(s)> option); here, 'close' means any of the bead types used in aggregate determination is closer than <distance> to any bead of the specified molecule.

Also, periodic boundary conditions can be removed from whole aggregates and the new coordinates saved to an indexed vcf file (-j option). Therefore aggregates will not be split by simulation box boundaries when, for example, visualizing the molecules with vmd.

While the Aggregates utility uses all possible pairs of given bead types, Aggregates-NotSameBeads does not use same-type pairs. For example, if bead types A and B are given, Aggregates will use all three possible bead type pairs (i.e., A-A, A-B, and B-B), but Aggregates-NotSameBeads will use only A-B bead type pairs.

Usage:

Aggregates (or Aggregates-NotSameBeads) <input> <distance> <contacts>

<output.agg> <bead type name(s)> <options>

Mandatory arguments		
<input/>	input coordinate file (either vcf or vtf format)	
<distance></distance>	minimum distance for two beads to be in contact (thus	
	constituting a contact pair)	
<contacts></contacts>	minimum number of contact pairs between two molecules	
	to be in one aggregate	

<pre><output.agg></output.agg></pre>	output agg file (must end with .agg) with aggregate information			
<pre><bead type(s)=""></bead></pre>	bead type name(s) to use for determining contact pairs (at least two for Aggregates-NotSameBeads)			
<pre><options></options></pre>				
Non-standard options	Non-standard options			
-x <mol name(s)=""></mol>	exclude specified molecule type(s) from aggregate determination (and from the output agg file)			
-xm <mol name(s)=""></mol>	exclude molecules that are close to specified molecule type(s)			
-j <output.vcf></output.vcf>	output vcf file with coordinates of joined aggregates (i.e., without periodic boundary conditions)			

## 4.3 AngleMolecules

This utility calculates angles between beads in each molecule of specified molecule type(s). The beads do not have to be connected, so the angle does not have to be between two bonds.

Using  $\neg n$  option, the angle is specified by three bead indices taken from the bead order in the vsf file. These indices go from 1 to N, where N is the number of beads in the molecule type. Generally, the numbering of beads inside a molecule is made according to the first molecule of the given type in vsf file. For example, assume that beads of the first molecule called mol in the vsf file are ordered A (vsf index 123), B (vsf index 124), C (vsf index 200). Then, bead A is 1, bead B is 2, and C is 3.

More than one angle can be specified (i.e., a multiple of three numbers have to be supplied to the -n option.). For example, assuming indices 1 2 3 1 3 2 are specified, two angles will be calculated. The first angle will be between lines defined by beads with indices 1 2 and 2 3; the second one will be between lines defined by beads with indices 1 3 and 3 2. An angle is calculated in degrees and is between 0 and 180°.

The utility calculates distribution of angles for each specified trio of bead indices for each molecule type and prints overall averages at the end of an **<output>** file. If **-a** option is used, it can also write all the angles for all individual molecules in each timestep (i.e., time evolution of the angle for each individual molecule).

Usage:

AngleMolecules <input> <width> <output> <mol name(s)> <options>

#### Mandatory arguments

<input/>	input coordinate file (either vcf or vtf format)		
<width></width>	width of each bin of the distribution		
<mol name(s)=""></mol>	molecule name(s) to calculate angles for		
<output></output>	output file for distribution		
Non-standard op	Non-standard options		
joined	specify that <input/> contains joined coordinates (i.e., periodic		
	boundary conditions for molecules do not have to be removed)		
-a <name></name>	write all angles for all molecules in all timesteps to <name></name>		
-n <ints></ints>	multiple of three indices for angle calculation (default: 1 2 3)		
-st <int></int>	starting timestep for calculation (default: 1)		
-e <int></int>	ending timestep for calculation (default: none)		

#### Format of output files:

- (1) <output> distribution of angles
  - first line: command used to generate the file
  - second line: angle-specifying bead indices (the dash-separated numbers correspond to indices inside every molecule and are the same as the arguments to the -n option) with the numbers in brackets corresponding to nth column of data for each molecule type
  - third line: numbering of columns (i.e., column headers)
    - first is the centre of each bin in angles (governed by <width>); i.e., if <width> is 5°, then the centre of bin 0 to 5° is 2.5, centre of bin 5 to 10° is 7.5 and so on
    - the rest are for the calculated data: the range for each molecule type specifies which column numbers correspond to the calculated angles for that particular molecule type and the order of angles is given by the second line
  - last two lines: arithmetic means for each calculated angle (last line) and headers (second to last line) that again give range of columns in the last line for each molecule type
- (2) -a <name> all angles for all molecules in all timesteps
  - first and second lines are the same as for <output>
  - third line: column headers
    - first is simulation timestep
    - the rest are the calculated data: the range for each molecule type corresponds to the number of molecules of the given type times the number of calculated angles; for each molecule the angles are ordered according to the second line

4. Utilities 4.4. Average

## 4.4 Average

This utility uses the binning method to analyse data in a text file. It does not use any of the standard options and prints the result only to standard output (e.i., screen).

Average calculates average value, statistical error, and estimate of autocorrelation time  $\tau$ . Empty lines and comments (lines beginning with #) are skipped. Average prints four numbers to screen:  $n_blocks < average > < std error > < tau estimate >:$ 

<n_blocks></n_blocks>	number of blocks used for the binning analysis
<average></average>	simple arithmetic average
<std error=""></std>	one- $\sigma$ statistical error
<tau estimate=""></tau>	estimate of autocorrelation time $\tau$

The average value of an observable  $\mathcal{O}$  is a simple arithmetic mean:

$$\langle \mathcal{O} \rangle = \frac{1}{N} \sum_{i=1}^{N} \mathcal{O}_i,$$
 (4.1)

where N is the number of measurements and the subscript i denotes individual measurements. If the measurements are independent (i.e., uncorrelated), the statistical error,  $\epsilon$ , is given by:

$$\epsilon^2 = \frac{\sigma_{\mathcal{O}_i}^2}{N},\tag{4.2}$$

where  $\sigma_{\mathcal{O}_i}^2$  is the variance of the individual measurements,

$$\sigma_{\mathcal{O}_i}^2 = \frac{1}{N-1} \sum_{i=1}^N (\mathcal{O}_i - \langle \mathcal{O} \rangle)^2. \tag{4.3}$$

For correlated data, the autocorrelation time,  $\tau$ , representing the number of steps between two uncorrelated measurements must be determined. Every  $\tau$ -th measurement is uncorrelated, so the equation (4.2) can then be used to estimate the error.

A commonly used method to estimate  $\tau$  is the binning (or block) method. In this method, the correlated data are divided into  $N_{\rm B}$  non-overlapping blocks of size k ( $N=kN_{\rm B}$ ) with per-block averages,  $\mathcal{O}_{{\rm B},n}$ , defined as:

$$\mathcal{O}_{B,n} = \frac{1}{k} \sum_{\substack{i=1+\\(n-1)k}}^{kn} \mathcal{O}_i.$$
 (4.4)

If  $k \gg \tau$ , the blocks are assumed to be uncorrelated and equation (4.2) can be used:

$$\epsilon^2 = \frac{\sigma_{\rm B}^2}{N_{\rm B}} = \frac{1}{N_{\rm B}(N_{\rm B} - 1)} \sum_{n=1}^{N_{\rm B}} (\mathcal{O}_{{\rm B},n} - \overline{\mathcal{O}})^2.$$
 (4.5)

An estimate of the autocorrelation time can be obtained using the following formula:

$$\tau_{\mathcal{O}} = \frac{k\sigma_{\mathcal{B}}^2}{2\sigma_{\mathcal{O}_i}^2}.\tag{4.6}$$

A way to quickly get  $\tau$  estimate is to use a wide range of <n\_blocks> values and plot the <tau estimate> values as a function of <n\_blocks>. Because the number of data points in one block should be significantly larger than  $\tau$  (e.g., ten times larger), plotting f(x) =(number of data lines in the file)/(10x) will produce an exponential function that intersects the <tau estimate> line. A value of <tau estimate> near the intersection (but to the left, where the exponential is above <tau average>) can be considered a safe estimate of  $\tau$ .

Usage:

Average <input> <column> <discard> <n\_blocks>

Mandatory arguments		
<input/>	input text file column number in <input/> for data analysis	
<discard></discard>	number of data lines to discard from the beginning of <input/>	
$< n_blocks>$	number of blocks for binning analysis	

## 4.5 BondLength

This utility calculates a distribution of bond lengths in specified molecule type(s). It can also calculate a distribution of distances between any two beads in the molecules.

For each of the specified molecule type(s), BondLength calculates bond lengths between all different types of connected bead pairs. For example, assume two linear molecule types Mol\_1 and Mol\_2, both composed of bead types A and B. Mol\_1 is connected like this: A-A-B; Mol\_2 like this: A-B-B. If both molecule types are used, BondLength calculates for each molecule type distribution of lengths for bonds A-A, A-B, and B-B (separate for each molecule even though the molecules share the same bead types).

To calculate the distribution of distances between specific (possibly unconnected) beads in a molecule, use -d option which takes as arguments pairs of bead indices

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(according to the order of beads in the molecule in vsf — similarly to the —n option in AngleMolecules, i.e., Section 4.3). More than one pair can be specified. These indices are the same for all <mol name(s)>. If an index higher than the number of beads in the molecule is provided, the utility takes the last bead of the molecule (i.e., the highest index). For example, using —d file.txt 1 2 1 9999 would write two distributions for each <mol name(s)> into <file.txt>: distribution of distances between the first and the second bead in each <mol name(s)> and between the first bead and the last one (or the 9999th bead).

In both cases, BondLength appends at the end of the file minimum and maximum bond lengths/distances.

Usage:

BondLength <input> <width> <output> <mol name(s)> <options>

Mandatory arguments			
<input/>	input coordinate file (either vcf or vtf format)		
<width></width>	width of each bin of the distribution		
<output></output>	output file with distribution of bond lengths		
<mol name(s)=""></mol>	molecule name(s) to calculate bond lengths for		
Non-standard options			
-st <int></int>	starting timestep for calculation (default: 1)		
-e <int></int>	ending timestep for calculation (default: none)		
-d <out> <ints></ints></out>	write distribution of distances between specified beads in		
	each specified molecule to <out></out>		
-w <double></double>	warn if a bond length exceeds <double> (default: half a box length)</double>		

#### Format of output files:

- (1) <output> distribution of bond length between all bead pairs
  - first line: command used to generate the file
  - second line: column headers
    - first is the centre of each bin (governed by <width>); i.e., if <width> is 0.1, then the centre of bin 0 to 0.1 is 0.05, centre of bin 0.1 to 0.2 is 0.15, etc.
    - the rest are for the calculated data: for each molecule type, there is a list of column numbers corresponding to all possible bead type pairs in the molecule; if no beads of the given types are connected, the data column contains nan
  - next lines: the calculated data
  - second to last line: column headers for minimal and maximal bond lengths
    - for each molecule, all the possible beads pairs are again listed
    - for each pair, the column number corresponds to the minimum, while the

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next column is always the maximum

• last line: commented out (i.e., the line starts with #) minimal and maximal bond lengths

- (2) -d <output> <ints> distribution of distances between specified beads
  - first line: command used to generate the file
  - second line: order of beads (given only by bead names) in each molecule
  - third line: column headers
    - first is again the centre of every bin
    - the rest are for the calculated data: for each molecule type, there is a list of column numbers corresponding to the given pairs of bead indices in the molecule (and to the -d option's arguments); the numbers also correspond to the order of beads in the previous line
  - next lines: the calculated data
  - second to last line: column headers for minimal and maximal bond lengths
    - for each molecule, all the possible beads pairs are again listed
    - for each pair, the column number corresponds to the minimum, while the next column is always the maximum
  - last line: commented out (i.e., the line starts with #) minimal and maximal bond lengths

## 4.6 Config

This utility creates DL\_MESO CONFIG file. It requires input coordinate file with all beads; otherwise the utility will still run without any error, but it will produce an incomplete CONFIG. Config always prints first unbonded beads and then beads in molecules (required by DL\_MESO).

Usage:

Config <input.vcf> <options>

Mandatory argument			
<input/>	input coordinate file (either vcf or vtf format)		
Non-standard option			
-st <int></int>	timestep for creating CONFIG file from (default: last step)		

There is also utility Config\_from\_xyz which takes a xyz coordinate file and creates the DL\_MESO CONFIG file. Because xyz file does not contain information about the simulation box, the resulting CONFIG file must be modified manually — Config\_from\_xyz prints x, y, and z into the output file where box dimensions should

be.

Usage:

Config\_from\_xyz <input.xyz> <options>

Mandatory argument

<input.xyz> input xyz coordinate file

Non-standard option

-st <int> timestep for creating CONFIG file from (default: last step)

## 4.7 DensityAggregates

This utility calculates radial density profiles (RDPs, or radial number densities) for bead types in an aggregate with specified size (the number of molecules or aggregation number,  $A_{\rm S}$ ) from its centre of mass.

 $\mathrm{RDP}_i(r)$  of bead type i, where r is distance from an aggregate's centre of mass, is the number of these beads in a spherical shell between the distances r and  $r+\mathrm{d}r$  (in DensityAggregates,  $\mathrm{d}r$  is the <width> argument) divided by the volume of this shell. The utility also prints radial number profile (RNP $_i(r)$ ), or the number of beads in each shell without dividing it by its volume. In addition, it prints one- $\sigma$  error for both RDP and RNP.

Composition of an aggregate with given size (i.e., average numbers of different types of molecules in that aggregate) is appended to the output file.

Instead of 'true' aggregate size, a number of molecules of specific type(s) can be used (-m option). For example, an aggregate containg 1 Mol\_A molecule and 2 Mol\_B molecules (i.e., three molecules in all) can be specified in several ways:

- (1) with  $\langle agg size(s) \rangle$  of 3;
- (2) with <agg size(s)> of 3 and -m Mol\_A Mol\_B;
- (3) with <agg size(s)> of 1 and -m Mol\_A; or
- (4) with <agg size(s)> of 2 and -m Mol\_B.

Care must be taken when different molecule types share the same bead type. If one bead type is in more molecule types, the resulting density for that bead type will be the sum of its densities from all molecule types it appears in. The -x option can overcome this – specific molecule types can be excluded from density calculations, i.e., density of beads in the excluded molecule types will not be calculated. For example, assume two molecule types – Mol\_1 and Mol\_2. Mol\_1 contains bead types A and B; Mol\_2 contains bead types A and C. Depending on whether and how the -x option is used, the utility will calculate:

- (1) densities of A, B, and C beads (density of A beads is a sum from both molecules), if no -x is used;
- (2) densities of only A and B beads (with A beads only from Mol\_1), if -x Mol\_2 is used;
- (3) densities of only A and C beads (with A beads only from Mol\_2), if -x Mol\_1 is used; or
- (4) no densities at all if -x Mol\_1 Mol\_2 is used.

Therefore, to be able to plot density of A beads from Mol\_1 and Mol\_2 separately,

(2) and (3) should be used (i.e., DensityAggregates should be run twice). Usage:

DensityAggregates <input> <input.agg> <width> <output> <agg
size(s)> <options>

Mandatory arguments			
<input/>	input coordinate file (either vcf or vtf format)		
<input.agg></input.agg>	input .agg file		
<width></width>	width of a single distribution bin		
<output></output>	output file(s) (one per aggregate size) with automatic #.rho ending (# is aggregate size)		
<agg size(s)=""></agg>	aggregate size(s) (the number of molecules in an aggregate		
	or the aggregation number, $A_{\rm S}$ ) to calculate density for		
Non-standard option	Non-standard options		
joined	specify that <input/> contains joined coordinates (i.e., periodic boundary conditions for aggregates do not have to be removed)		
-n <int></int>	number of bins to average to get smoother density (default: 1)		
-st <int></int>	starting timestep for calculation (default: 1)		
-e <int></int>	ending timestep for calculation (default: none)		
-m <mol name(s)=""></mol>	instead of 'true' aggregate size, use the number of specified molecule type(s) in an aggregate		
-x <mol name(s)=""></mol>	exclude specified molecule type(s) (i.e., do not calculate density for beads in molecules <mol name(s)="">)</mol>		

#### Format of output files:

- (1) <output>#.rho bead densities for one aggregate size
  - first line: command used to generate the file
  - second line: the order of data columns for each bead type rdp is radial density profile, rnp radial number profile and stderr are one- $\sigma$  errors for rdp and rnp

- third line: column headers
  - first is the centre of each bin (governed by <width>); i.e., if <width> is 0.1, then the centre of bin 0 to 0.1 is 0.05, centre of bin 0.1 to 0.2 is 0.15, etc.
  - the rest are for the calculated data: each number specifies the first column with data for the given bead type (i.e., rdp column)
  - last line contains the total number of aggregates the density was calculated for
- second to last line: column headers for average numbers of different molecules in the given aggregate
- last line: average numbers of the molecules

## 4.8 DensityBox

This utility calculates number density for all bead types in the specified direction of the simulation box. The density is calculated from 0 to box length in the given direction, that is, the box is 'sliced' into blocks with width <width> and numbers of different bead types are counted in each of the slices.

The utility does not distinguish between beads with the same name in different molecules, so if one bead type is in more than one molecule type, it's density will be averaged over all molecule types it appears in. In this case, however, the  $-\mathbf{x}$  option can be used to first run the utility and exclude one molecule type and then rerun it, excluding the other molecule type. Thus, two output files are generated, each missing densities from the specified molecule types.

Usage:

DensityBox <input> <width> <output> <axis> <options>

Mandatory arguments	
<input/>	input coordinate file (either vcf or vtf format)
<width></width>	width of each bin of the distribution
<output></output>	output file with automatic <axis>.rho ending</axis>
<axis></axis>	direction in which to calculate density: x, y, or z
Non-standard options	
-n <int></int>	number of bins to average to get smoother density (default: 1)
-st <int></int>	starting timestep for calculation (default: 1)
-x <mol name(s)=""></mol>	exclude specified molecule type(s) (i.e., do not calculate density for beads in molecules <mol name(s)="">)</mol>

Format of output files:

- (1) <output> bead densities
  - first line: command used to generate the file
  - second line: column headers
  - first is the centre of each bin (governed by <width>); i.e., if <width> is 0.1, then the centre of bin 0 to 0.1 is 0.05, centre of bin 0.1 to 0.2 is 0.15, etc.
  - the rest are for the calculated data: each number corresponds to the density of the specified bead type

## 4.9 DensityMolecules

This utility works similarly to DensityAggregates, only instead for whole aggregates, RDPs are calculated for individual molecules. Similarly to DensityAggregates, the output file(s) also contain statistical errors and radial number profiles.

By default, the utility calculates RDPs from the molecule's centre of mass, but any bead in the molecule (with an index according to vsf – similarly to –n option in AngleMolecules, Section 4.3) can be used instead (-c option).

Usage:

DensityMolecules <input> <width> <output> <mol name(s)> <options>

Mandatory arguments	
<input/>	input coordinate file (either vcf or vtf format) width of each bin of the distribution
<pre><output></output></pre>	output file with automatic <mol_name>.rho ending (one file per molecule type is generated)</mol_name>
<mol name(s)=""></mol>	molecule name(s) to calculate density for
Non-standard options	
joined	specify that <input/> contains joined coordinates (i.e., periodic boundary conditions for molecules do not have to be removed)
-n <int></int>	number of bins to average for smoother density (default: 1)
-st <int></int>	starting timestep for calculation (default: 1)
-e <int></int>	ending timestep for calculation (default: none)
-c <name> <int></int></name>	use specified bead in a molecule <name> instead of its centre of mass</name>

Format of output files:

(1) <output><mol\_name>.rho - bead densities for one molecule

- first line: command used to generate the file
- second line: the order of data columns for each bead type rdp is radial density profile, rnp is radial number profile and stderr are one-σ errors for rdp and rnp
- third line: column headers
  - first is the centre of each bin (governed by <width>); i.e., if <width> is 0.1, then the centre of bin 0 to 0.1 is 0.05, centre of bin 0.1 to 0.2 is 0.15, etc.
  - the rest are for the calculated data: each number specifies the first column with data for the given bead type (i.e., rdp column)

#### 4.10 DihedralMolecules

This utility calculates angles between specified planes in each molecule of provided molecule type(s). The planes in a molecule are arbitrary, so they can represent true dihedral angles or improper dihedrals.

The angle is specified by six bead indices (according to the order of beads in the molecule in vsf – similarly to the –n option in AngleMolecules, Section 4.3). The first three indices specify one plane and the next three the other. For example, assuming indices 1 2 3 4 5 6, the first plane is specified by the first three beads in the molecule; second plane by the next three beads (beads 4 5 6). The default indices (i.e., if –n option is not used) are 1 2 3 2 3 4. More than one angle can be specified (i.e., a multiple of six numbers can be supplied to the –n option.).

The utility calculates a distribution for each specified angle for each molecule type and prints overall averages at the end of **<output>** file. If **-a** option is used, it also writes all the angles for all individual molecules in each timestep (i.e., time evolution of the angle for each individual molecule).

Usage:

#### DihedralMolecules <input> <width> <output> <mol name(s)> <options>

Mandatory arguments	
<input/> <width> <output></output></width>	input coordinate file (either vcf or vtf format) width of each bin of the distribution (in degrees) output file for distribution
<mol name(s)=""></mol>	molecule name(s) to calculate angles for
Non-standard options	
joined	specify that <input/> contains joined coordinates (i.e., periodic boundary conditions for molecules do not have to be removed)

-a <file></file>	write all angles for all molecules in all timesteps to <file></file>
-n <ints></ints>	multiple of six indices for angle calculation (default: 1 2 3
	2 3 4)
-st <int></int>	starting timestep for calculation (default: 1)
-e <int></int>	ending timestep for calculation (default: none)

#### Format of output files:

- (1) <output> distribution of angles
  - first line: command used to generate the file
  - second line: calculated angles (the dash-separated numbers correspond to indices inside every molecule and are the same as the arguments to the -n option) with the numbers in brackets corresponding to nth column of data for each molecule type
  - third line: numbering of columns (i.e., column headers)
    - first is the centre of each bin in angles (governed by <width>); i.e., if <width> is 5°, then the centre of bin 0 to 5° is 2.5, centre of bin 5 to 10° is 7.5 and so on
    - the rest are for the calculated data: the range for each molecule type specifies which column numbers correspond to the calculated angles for that particular molecule type and the order of angles is given by the second line
  - last three lines: arithmetic means for each calculated angle (last line), headers (second to last line) that again give range of columns in the last line for each molecule type, and calculated angles (third to last line which is the same as the second line at the file beginning)
- (2) -a <file> all angles for all molecules in all timesteps
  - first and second lines are the same as for <output>
  - third lines: column headers
    - first is simulation timestep
    - the rest are the calculated data (in degrees): the range for each molecule type corresponds to the number of molecules of the given type times the number of calculated angles; for each molecule the angles are ordered according to the second line

## 4.11 DistrAgg

This utility calculates average aggregate mass and aggregation number for each timestep (i.e., time evolution) and the averages over all timesteps from a supplied agg file (see Section 3.3 for its format). It calculates number, weight, and z averages. It also calculates distribution functions of aggregation sizes and volumes.

For a quantity  $\mathcal{O}$ , the number, weight, and z averages,  $\langle \mathcal{O} \rangle_{n}$ ,  $\langle \mathcal{O} \rangle_{w}$ , and  $\langle \mathcal{O} \rangle_{z}$ , respectively, are defined as

$$\langle \mathcal{O} \rangle_{\rm n} = \frac{\sum_{i} N_i \mathcal{O}_i}{N}, \quad \langle \mathcal{O} \rangle_{\rm w} = \frac{\sum_{i} N_i m_i \mathcal{O}_i}{\sum_{i} N_i m_i}, \text{ and } \quad \langle \mathcal{O} \rangle_{\rm z} = \frac{\sum_{i} N_i m_i^2 \mathcal{O}_i}{\sum_{i} N_i m_i^2}, \quad (4.7)$$

where N is the total number of measurements, i.e., the total number of aggregates for per-aggregate averages (or molecules for per-molecule averages);  $N_i$  is the number of measurements with the value  $\mathcal{O}_i$ , and  $m_i$  is mass of an aggregate i (or a molecule i).

Per-timestep averages are written to the <output avg> and overall averages are appended as comments (with commented legend) to both <output avg> and <output distr> files.

Number, weight, and z distribution functions of aggregate sizes,  $F_n(A_S)$ ,  $F_w(A_S)$ , and  $F_z(A_S)$ , respectively, are defined as

$$F_{\rm n} = \frac{N_{A_{\rm S}}}{\sum_{A_{\rm S}} N_{i}} = \frac{N_{A_{\rm S}}}{N},$$

$$F_{\rm w} = \frac{N_{A_{\rm S}} m_{A_{\rm S}}}{\sum_{A_{\rm S}} N_{i} m_{i}} = \frac{N_{A_{\rm S}} m_{A_{\rm S}}}{\sum_{i=1}^{N} m_{i}} = \frac{N_{A_{\rm S}} m_{A_{\rm S}}}{M}, \text{ and}$$

$$F_{\rm z} = \frac{N_{A_{\rm S}} m_{A_{\rm S}}^{2}}{\sum_{A_{\rm S}} N_{i} m_{i}^{2}} = \frac{N_{A_{\rm S}} m_{A_{\rm S}}^{2}}{\sum_{i=1}^{N} m_{i}^{2}},$$

$$(4.8)$$

where  $N_{A_{\rm S}}$  and  $m_{A_{\rm S}}$  stand for the number and mass, respectively, of aggregates with aggregate size  $A_{\rm S}$ ; M is the total mass of all aggregates. The equations are normalised so that  $\sum F_x(A_{\rm S}) = 1$ .

Distribution of volume fractions of aggregates,  $\phi(A_{\rm S})$ , is defined (assuming all beads have the same volume) as

$$\phi(A_{\rm S}) = \frac{N_{A_{\rm S}} m_{A_{\rm S}}}{\sum_{i=1}^{N} n_i} = \frac{N_{A_{\rm S}} m_{A_{\rm S}}}{n},\tag{4.9}$$

where  $n_i$  is the number of beads in aggregate i and n is the total number of beads in all aggregates. If all beads have unit mass (as is often the case in dissipative particle dynamics), the volume distribution,  $\phi(A_S)$ , is equal to the number distribution,  $F_n(A_S)$ . All distributions are written into the **<output distr>** file.

Lastly, DistrAgg can calculate number distribution of composition for aggregates with specified size(s) (-c option); that is, the distribution of ratios of two molecule types in aggregates with specified size,  $\xi = N_{\text{Mol-A}}/N_{\text{Mol-B}}$ , where  $N_{\text{Mol-A}}$  and  $N_{\text{Mol-B}}$  are the numbers of Mol\_A and Mol\_B molecules in the aggregate with size  $A_{\text{S}} = N_{\text{Mol-A}}/N_{\text{Mol-B}}$ 

 $N_{\text{Mol\_A}} + N_{\text{Mol\_B}}$ . The composition distribution is defined as

$$F_{\rm n}(\xi) = \frac{N_{\xi, A_{\rm S}}}{N_{A_{\rm S}}},$$
 (4.10)

where  $N_{\xi,A_{\rm S}}$  is the number of aggregates with aggregate size  $A_{\rm S}$  and ratio  $\xi$ ;  $N_{A_{\rm S}}$  is the total number of aggregates with aggregate size  $A_{\rm S}$ . If there are more than two molecule types in the simulation, -nc option must be used to specify which molecules should be used in the composition calculation.

The -nc option has no effect if used without the -c.

The <avg file> contains averages for all timesteps regardless of -st and -e options; that is, the starting step (-st option) and ending step (-e option) are ignored.

The definition of aggregate size is flexible. If none of -m, -x, or --only options is used, aggregate size is the 'true' aggregation number,  $A_S$ , i.e., the number of all molecules in the aggregate; if -m is used, aggregate size is the sum of only specified molecule type(s); if -x is used, aggregates containing only specified molecule type(s) are disregarded; if --only is used, only aggregates composed of the specified molecule type(s) are taken into account. For example, consider a system containing three aggregates composed of various numbers of three different molecule types:

Molecule types	Aggregate composition
Mol_A	Agg_1: 1 Mol_A +2 Mol_B +3 Mol_C = 6 molecules
${\tt Mol\_B}$	Agg_2: 1 Mol_A +2 Mol_B = 3 molecules
${\tt Mol\_C}$	Agg_3: 1 Mol_A = 1 molecule

Here is a list of some of the possibilities depending on the option(s) used:

- (1) if none of -m, -x, --only is used, all three aggregates are counted and their sizes are their aggregation numbers, i.e.,  $A_S = 6$ , 3, and 1
- (2) if -m Mol\_A Mol\_B is used, all three aggregates are counted, but their size is the sum of only Mol\_A and Mol\_B molecules: Agg\_1 3; Agg\_2 3; Agg\_3 1
- (3) if -m Mol\_B Mol\_C is used, Agg\_3 is not counted, because its size would be zero; DistrAgg would detect only two aggregates with sizes: Agg\_1 5; Agg\_2 2
- (4) if -x Mol\_A Mol\_B is used, Agg\_2 and Agg\_3 are not counted, because neither contains anything else than Mol\_A and/or Mol\_B; DistrAgg would detect only one aggregate with size: Agg\_1 - 6
- (5) if -x Mol\_A Mol\_B is combined with -m Mol\_A Mol\_B, DistrAgg would again detect only Agg\_1, but its size would be 3
- (6) if --only Mol\_A Mol\_B is used, Agg\_1 is not counted, because it contains a molecule not specified by --only; DistrAgg would detect two aggregates with

- sizes:  $Agg_2 3$ ;  $Agg_3 1$
- (7) if --only Mol\_A Mol\_B is combined with -m Mol\_A, the two detected aggregates have sizes: Agg\_2 1; Agg\_3 1
- (8) if --only Mol\_A Mol\_B is combined with -x Mol\_A, only Agg\_2 is detected as it is the only one composed of only Mol\_A and Mol\_B molecules and its size would be 3
- (9) if --only Mol\_A Mol\_B and -x Mol\_A are combined with -m Mol\_A, the size of the aggregate would be 1

Moreover, only a specified range of aggregate sizes can be taken into account (-n < int > option). These sizes are defined by the -m, -x, and --only options as well.

Usage:

DistrAgg <input.agg> <distr file> <avg file> <options>

Mandatory arguments	
<pre><input.agg> <distr file=""> <avg file=""></avg></distr></input.agg></pre>	input agg file output file with distribution of aggregate sizes output file with per-timestep averages
Non-standard options	
-st <int> -e <int> -n <int> <int> -m <mol name(s)=""> -x <mol name(s)=""></mol></mol></int></int></int></int>	starting timestep for calculation (default: 1) ending timestep for calculation (default: none) use aggregate sizes in a given range use number of specified molecule(s) as aggregate size exclude aggregates containing only specified molecule(s)
<pre>only <mol name(s)=""> -c <output> <int(s)></int(s)></output></mol></pre>	use only aggregates composed of specified molecule(s) save composition distribution for specified aggregate size(s) to <output> file</output>
-nc <name> <name></name></name>	molecule names to use for composition calculation

#### Format of output files:

- (1) <output distr> distributions of aggregate sizes
  - first line: command used to generate the file
  - second line: column headers
    - first is the aggregate size,  ${\tt As}$  either true aggregation number, or the size specified by options
    - F\_n(As), F\_w(As), and F\_z(As) are number, weight, and z distribution of aggregate sizes (Equation (4.8))
    - <volume distribution is distribution according to Equation (4.9)

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- next is the total number of aggregates with specified size
- the remaining columns (<mol name>\_n) show average numbers of every molecule type in an aggregate with the specified size
- second to last line: column headers for overall averages written in the last line
  - <M>\_n, <M>\_w, and <M>\_z are number, weight, and z averages, respectively, of aggregate masses (the averages are defined in Equation (4.7))
  - other column headers are the same as at the file beginning
- (2) <output avg> per-timestep averages
  - first line: command used to generate the file
  - second lines: column headers
    - first is simulation timestep
    - the rest are for the calculated data: number, weight, and z average aggregate mass (<M>\_n, <M>\_w, and <M>\_z, respectively) and aggregate size (<As>\_n, <As>\_w, and <As>\_z, respectively) and the last column is the number of aggregates in the given step
  - the last two lines are the same as in <output distr>
- (3) -c <name> composition distribution
  - first line: command used to generate the file
  - second lines: column headers
    - first is ratio of the two molecule types (i.e., 0 to 1)
    - the rest are aggregate sizes
    - in the data, only ratios that are non-zero for at least one aggregate size are written; in case of more than one aggregate size specified by -a option, if the ratio does not exist for some aggregate size(s), '?' is displayed instead of zero

## 4.12 GenLayers

This utility generates two monolayers composed of molecules specified in a FIELD-like file. For now, only one type of molecule is used (the first one in the FIELD-like file; other molecule types are ignored). The two layers are mirror images of each other, that is, molecules in both layers are grown from box edge to box middle. The layers are placed in z direction (that is, on xy planes of the simulation box).

The first beads of the molecules are arranged on a square lattice defined either by given spacing in x and y directions (-s option) or by the number of molecules per layer (-nm option). The rest of the beads of each molecule are placed based on the coordinates in the FIELD-like file. By default, GenLayers places the two mirror layers at the edges of a simulation box; using -g option, a gap from box edge can be introduced. Therefore, this utility can generate, for example, polymer brushes at box edges or a double layer (such as a biological membrane) in the middle of the box.

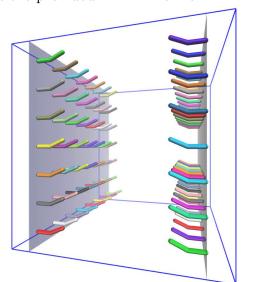
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By default, the total number of beads in the generated system is equal to three times the box volume, that is, the typical number of beads in dissipative particle dynamics simulations. Beads that are not in the molecules, are put at the beginning of the output files (i.e., they have lower indices) with coordinates of the box centre, and name None. The idea is that once the layers are generated, AddToSystem utility (Section 4.1) can be used to exchange these excess beads for different species. The -n option changes the total number of beads. If the number is lower than the total number of beads needed to construct the two layers of molecules, it is adjusted to exactly that number.

The input FIELD-like file must contain species and molecule sections (although, only the first molecule is considered and nummols is disregarded), but the interaction section is ignored (see DL\_MESO manual for details on the FIELD file or below for a simple example). The first line of the FIELD-like file, that is ignored by DL\_MESO, must start with box dimensions, i.e., with three numbers.

This utility does not have error checking for the provided FIELD-like file. If the file is not correct, GenLayers will exhibit undefined behaviour, that is, it will either freeze, crash, or run without errors, producing unexpected output files.

The following vmd-generated snapshot (the transparent squares are just a visual aid) is an example output using command GenLayers out.vsf out.vcf -n 1 -nm 50 -g 1 and the provided FIELD-like file:



FIELD-like file
10 10 10
species 1
A 1.0 0.0 0
molecules 1
A3
nummols 1
beads 3
A 0 0.0 0.0
A 0 0.0 0.7
A 0 0.3 1.4
bonds 2
harm 1 2 100.0 0.7
harm 2 3 100.0 0.7
finish

The utility generates vsf structure and vcf coordinate files. Usage (GenLayers does not use standard options):

GenLayers <out.vsf> <out.vcf> <options>

Mandatory arguments

<out.vsf></out.vsf>	output vsf structure file
<pre><out.vcf></out.vcf></pre>	output vcf coordinate file
Options	
-s <x> <y></y></x>	spacing of molecules in $x$ and $y$ directions (default: 1 1)
-n < int >	total number of beads (default: three times the box volume)
-nm < int>	number of molecules in each layer (rewrites -s option)
-g <float></float>	gap between box edges and the molecules (default: 0)
-f <name></name>	FIELD-like file (default: FIELD)
-A	verbose output that provides information about all bead and
	molecule types
-h	print this help and exit

## 4.13 GenSystem

This utility was not extensively tested and is not a good generator of initial configuration.

This simple utility uses modified FIELD file to create vsf structure file and to generate coordinates that could be used as an initial configuration. The utility assumes linear chains (no matter the connectivity in the provided FIELD file; it looks at exactly the first n-1 bonds to get bond lengths, where n is the number of beads in a molecule). For each molecule type, it uses equilibrium bond length (or 0.7 if the bond length is 0) to construct a prototype molecule that is fully stretched in z-direction. The utility then creates layers of molecules that are separated by layers of unbonded beads (if there are any). The utility should fill the whole box with given beads.

The input FIELD file must contain species and molecule sections, but the interaction section is ignored (see DL\_MESO manual for details on the FIELD file). The first line of FIELD that is ignored by DL\_MESO must start with box dimensions, i.e., with three numbers.

This utility does not have error checking for the provided FIELD file. If the file is not correct, GenSystem will exhibit undefined behaviour, that is, it will either freeze, crash, or run without errors, producing bad output files.

Usage (GenSystem does not use standard options):

#### GenSystem <out.vsf> <out.vcf> <options>

Mandatory arguments	
<out.vsf></out.vsf>	output vsf structure file
<out.vcf></out.vcf>	output vcf coordinate file

Options	
-f <name></name>	FIELD-like file (default: FIELD)
-Λ	verbose output that provides information about all bead and
	molecule types
-h	print help and exit

## 4.14 GyrationAggregates

This utility calculates the gyration tensor and its eigenvalues (or the roots of the tensor's characteristic polynomial) for all aggregates. Using these eigenvalues, the utility determines shape descriptors: radius of gyration, asphericity, acylindricity, and relative shape anisotropy.

The eigenvalues,  $\lambda_x^2$ ,  $\lambda_y^2$ , and  $\lambda_z^2$ , (sorted so that  $\lambda_x^2 \leq \lambda_y^2 \leq \lambda_z^2$ ) are also written to output file(s), because their square roots represent half-axes of an equivalent ellipsoid.

The radius of gyration,  $R_{\rm G}$ , is defined as

$$R_{\rm G}^2 = \lambda_x^2 + \lambda_y^2 + \lambda_z^2. \tag{4.11}$$

The asphericity, b, and the acylindricity, c, are defined, respectively, as

$$b = \lambda_z^2 - \frac{1}{2}(\lambda_x^2 + \lambda_y^2) = \frac{3}{2}\lambda_z^2 - \frac{R_G^2}{2}$$
 and  $c = \lambda_y^2 - \lambda_x^2$ . (4.12)

The relative shape anisotropy,  $\kappa$ , is defined in terms of the other descriptors as

$$\kappa^2 = \frac{b^2 + 0.75c^2}{R_C^4} \tag{4.13}$$

Number average of all properties and, additionally, weight and z averages for the radius of gyration are calculated (see Equation (4.7) in Section 4.11 for general definitions of averages). Per-timestep averages (i.e., time evolution) are written to the <output> file. Per-size averages can be saved using the -ps option.

The gyration tensor is by default calculated for whole aggregates, but -bt option can be used to specify which bead types to consider.

Similarly to DistrAgg, the definition of aggregate size is flexible – see Section 4.11 for explanations of the -m and -x options.

The starting step (-st option) and ending step (-e option) are used only for averages (both overall averages and per-size averages with -ps option). Per-timestep averages in the <output> file disregard -st and -e options.

Usage:

GyrationAggregates <input> <input.agg> <output> <options>

Mandatory arguments	
<input/> <input.agg> <output></output></input.agg>	input coordinate file (either vcf or vtf format) input agg file output file with per-timestep averages
Non-standard options	
joined	specify that <input/> contains joined coordinates (i.e., periodic boundary conditions for aggregates do not have to be removed)
-bt <bead name(s)=""></bead>	bead type(s) used for calculation
-ps <name></name>	output file with per-size averages
-m <mol name(s)=""></mol>	instead of 'true' aggregate size, use the number of speci-
	fied molecule type(s) in an aggregate
-x <mol name(s)=""></mol>	exclude aggregates containing only specified molecule type(s)
-n <int> <int></int></int>	use only aggregate sizes in given range
-st <int></int>	starting timestep for calculation (default: 1)
-e <int></int>	ending timestep for calculation (default: none)

#### (1) <output> – per-timestep averages

- first line: command used to generate the file
- second line: column headers
  - first is timestep
  - rest are for calculated data: number, weight, and z averages (denoted by \_n, \_w, and \_z, respectively) of radius of gyration and square of radius of gyration (Rg and Rg^2, respectively Equation (4.11)); number averages of relative shape anisotropy (Anis Equation (4.13)), acylindricity, and asphericity (Acyl and Aspher, respectively Equation (4.12)), and all three eigenvalues (eigen.x, eigen.y, and eigen.z  $\lambda_x^2$ ,  $\lambda_y^2$ , and  $\lambda_z^2$ )
- second to last line: column headers for overall averages written in the last line
  - <M>\_n and <M>\_w are number and weight, respectively, averages of aggregate masses (the averages are defined in Equation (4.7)); aggregate mass here is the mass of all beads of the chosen type(s) in the aggregate
  - average numbers of molecules of each type in an aggregate are shown in columns denoted <mol names>
  - the remaining column represent overall averages of the per-timestep quantities described above

#### (2) -ps <name> - per-size averages

- first line: command used to generate the file
- second line: column headers

- first is aggregate size
- last column is the total number of aggregates of the given size
- the columns in between are for the calculated data: simple averages of the numbers of molecules of each type in the aggregate, of radius of gyration and its square, of relative shape anisotropy, acylindricity, and asphericity, and of all three eigenvalues (all are denoted by the above-described symbols)

# 4.15 GyrationMolecules

This utility calculates the gyration tensor and from it the shape descriptors similarly to GyrationAggregates (Section 4.14) but for individual molecules instead of for whole aggregates.

GyrationMolecules <input> <output> <mol name(s)> <options>

1	
Mandatory arguments	
<input/>	input coordinate file (either vcf or vtf format)
<output></output>	output file(s) (one per molecule type) with automatic - <mol_name>.txt ending</mol_name>
<mol name(s)=""></mol>	molecule name(s) to calculcate shape descriptors for
Non-standard options	
joined	specify that <input/> contains joined coordinates (i.e., periodic boundary conditions for molecules do not have to be removed)
-bt <bead name(s)=""></bead>	bead type(s) to be used for calculation
-st <int></int>	starting timestep for calculation (default: 1)
-e <int></int>	ending timestep for calculation (default: none)

- (1) <output> per-timestep averages (one file per molecule type)
  - first line: command used to generate the file
  - second line: name of molecule type
  - third line: column headers
    - first is timestep
    - rest are for calculated data: number, weight, and z averages (denoted by \_n, \_w, and \_z respectively) of radius of gyration (Rg Equation (4.11)); simple averages of relative shape anisotropy (Anis Equation (4.13)), of acylindricity and asphericity (Acyl and Aspher, respectively Equation (4.12)), and of the three eigenvalues (eigen.x, eigen.y, and eigen.z or  $\lambda_x^2$ ,  $\lambda_y^2$ , and  $\lambda_z^2$ )

- last three lines show simple averages over the whole simulation
- third to last line: molecule name
- second to last line: column headers for radius of gyration and shape descriptors
- last line: the averages

## 4.16 Info

This simple utility just prints information about the system read from the provided structure file. If -c option is used, only beads in the provided coordinate file are printed. The information is the same as when the verbose option (-v) is used with any other utility. Here, the -v option prints detailed informations about every bead and every molecule - this information also contains indices as implemented inside the AnalysisTools utilities (i.e., not necessarily consistent with the structure file).

Usage (Info does not use standard options):

Info <input> <options>

Mandatory argument		
<input/> Options	input structure file (either .vsf or .vtf format)	
-c	input coordinate file in either .vcf or .vtf format (default: None)	
-h	print this help and exit	

# 4.17 JoinAggregates

This utility is meant for cases when non-standard option -j is omitted from Aggregates (or Aggregates-NotSameBeads) command. JoinAggregates uses the provided vcf and agg files to join aggregates, i.e., to remove their periodic boundary conditions and save the new coordinates into a vcf file. The utility reads Aggregates command from the agg file to determine distance and number of contact pairs for aggregate check (see Section 4.2 for details on Aggregates utility).

Be warned that if -sk, -st, or -e options are used, <output.vcf> will not be in sync with <input.agg> and, therefore, these two files cannot be used in tandem for further analysis via AnalysisTools.

Usage:

JoinAggregates <input> <input.agg> <output.vcf> <options>

4. Utilities 4.18. JoinRuns

Mandatory arguments		
<input/> <input.agg> <output.vcf></output.vcf></input.agg>	input coordinate file (either vcf or vtf format) input agg file output vcf coordinate file with indexed coordinates	
Non-standard opt	ions	
-sk <int> -st <int> -e <int></int></int></int>	number of steps skip per one used (default: 0) starting timestep for calculation (default: 1) ending timestep for calculation (default: none)	

## 4.18 JoinRuns

This utility joins two independent simulation runs of the same system. That is, the two systems contain identical beads and molecules, but these beads and molecules are numbered differently in the vsf and vcf files from different simulations. The two input vcf files must contain the same timestep type (i.e., both indexed or both ordered) and the same number of beads (i.e., if one bead type is absent from one vcf file, it must be absent from the second one as well).

The output is a vcf coordinate file with beads indexed according to the first vsf structure file (i.e., traject.vsf or provided by -i option). Only specified bead types are saved; using -r switch, the reverse is true, i.e., the specified bead types are excluded from the output file.

Usage:

JoinRuns <1st input> <2nd input> <2nd vsf> <output> <bead type(s)>
<options>

Mandatory arguments		
<1st input>	input coordinate file from the first simulation (either vcf or vtf format)	
<2nd input>	input coordinate file from the second simulation (either vcf or vtf format)	
<2nd vsf>	input structure file from the second simulation (structure file from the first simulation is traject.vsf; changeable via -i option)	
<output></output>	output vcf coordinate file with indexed coordinates	
<pre><bead type(s)=""></bead></pre>	bead type names to save (can be omitted if -r is used)	

4. Utilities 4.19. lmp\_data

#### Non-standard options

-r	reverse function, i.e., exclude <bead type(s)=""> instead of including them; if no <bead type(s)=""> are specified, all bead types are used (requires input coordinate files with all types)</bead></bead>
join	join molecules by removing periodic boundary conditions
-st1 <int></int>	starting timestep for first run (default: 1)
-st2 <int></int>	starting timestep for second run (default: 1)
-e1 <int></int>	ending timestep for first run (default: none)
-e2 <int></int>	ending timestep for second run (default: none)
-sk1 <int></int>	number of steps to skip per one used for first run (default: 0)
-sk2 <int></int>	number of steps to skip per one used for second run (default: 0)

# 4.19 lmp\_data

This simple utility generates data file for the lammps simulation package (see lammps manual page for details on the data file format).

This utility reads system composition from the provided vsf structure file, structure of molecules from DL\_MESO FIELD file (numbers of bond and angle types and angles – other informations are read from the vsf file), and coordinates from vcf coordinate file. The utility ignores the interactions part from FIELD. For now, the utility also ignores dihedrals.

Option --srp can be used to add one more bead type into the output file (i.e., increment number of atom types and add one more line for Masses section with a mass of 1.0). This is intended for cases when the segmental repulsive potential (srp) is used in lammps (srp prevents bond crossing when soft interparticle potential is used in a simulation).

lmp\_data <input> <out.data> <options>

Mandatory arguments		
<pre><input/> <out.data></out.data></pre>	input vcf coordinate file output data file	
Options		
-f <name>srp -st <int></int></name>	FIELD file (default: FIELD) add one bead type for srp coordinate timestep for creating the data file	

4. Utilities 4.20. lmp\_vtf

# 4.20 lmp\_vtf

This utility generates vsf and vcf files from lammps data file (see lammps manual page for details on its format). Note that while lammps uses the word 'atom', in this manual, 'bead' is used instead.

lmp\_vtf reads the data file header (atoms, bonds, atom types, xlo xhi, ylo
yhi, and zlo zhi keywords) and Masses, Atoms, and Bonds sections; everything
else is ignored. The utility requires lines in the Atoms section to have the following
format: <bed index> <molecule index> <bed type> <charge> <x> <y> <z>
(i.e., atom\_style full in lammps terminology).

If any line of the Masses section ends with a comment, its first string is taken as the name of the bead type. Otherwise, the bead types are called beadN, where N is their type number in the data file.

Charge of every bead type is taken as the charge of the last bead of this type in the Atoms section.

Molecule types are determined according to the order of bead types in molecules as well as according to bead connectivity. Bead order in every molecule is considered from the lowest to the highest bead index in the data file. If <molecule index> in a bead line is 0, this bead is unbonded (i.e., not part of any molecule). If there's only one bead in a molecule, this bead is considered unbonded as well. The types of molecules are called molN, where N goes from 1 to the total number of molecule types.

Following is an example of the data file:

```
7 atoms
2 bonds
3 atom types
0 10 xlo xhi
0 10 ylo yhi
0 10 zlo zhi

Masses
1 1.0
```

2 1.1 # some comment

#### Atoms

3 1.2

1	0	1	0.0	5.0	5.1	5.2
2	0	1	0.0	5.1	5.2	5.3
3	1	2	0.0	5.2	5.3	5.4
4	1	3	0.0	5.3	5.5	5.6

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5	2	3	0.0	5.3	5.5	5.6
6	2	2	0.0	5.3	5.5	5.6
7	3	1	0.0	5.3	5.5	5.6

#### Bonds

1 1 3 4 2 1 5 6

In this example, there are 3 bead types called bead1, some, and bead3. There are 7 beads in all, 3 are unbonded (indices 1, 2, and 7; all called bead3), 4 are in two different molecules. The molecules are considered as different types even though they have the same numbers bonds and the same connectivity; while the first molecule has the order of beads some, bead3 (according to the beads' ascending indices), the second one has the order reversed. These molecule types are called mol1 and mol2.

Note that lines in both Atoms and Bonds sections do not have to be ordered in any way.

Usage (this utility does not use standard options):

lmp\_vtf <input> <out.vsf> <out.vcf> <options>

Mandatory arguments		
<input/> <out.vsf> <out.vcf></out.vcf></out.vsf>	input lammps data file output vsf structure file output vcf coordinate file	
Options		
-v -h	verbose output print this help and exit	

## 4.21 Orientation

This utility calculates orientation order parameter for specified molecules. It determines distribution of the values as well as average values.

Orientation order parameter  $s_{ij}$  indicates how stretched molecules in, e.g., membranes are. It is defined as

$$s_{ij} = \frac{1}{2} (3\cos\theta - 1),$$
 (4.14)

where

$$\cos \theta = \frac{\mathbf{r_{ij}} \cdot \mathbf{n}}{|\mathbf{r_{ij}}||\mathbf{n}|}.\tag{4.15}$$

4. Utilities 4.21. Orientation

That is, the angle  $\theta$  is between a normal vector to a plane,  $\mathbf{n}$ , and a vector connecting beads i and j,  $\mathbf{r_{ij}}$ . The value of s is between -0.5 (parallel to that plane) and 1 (perpendicular to that plane).

The default normal is z-axis, but using -a option, a different axis can be used. The -a option takes as an argument x, y, or z.

The two beads specifying the second vector can be defined via the -a option (similarly to the -n option in AngleMolecules, Section 4.3). The default is the first and second bead of the molecule, i.e., -a 1 2. A multiple of two indices can be provided to calculate different  $s_{ij}$ . All provided index pairs are used for all molecule types, unless both indices in a pair are higher than the number of beads in the molecule type. If only one index is higher, then this index is reduced to the highest index for the given molecule. For example, consider two molecule types to be used: mol1 with four beads and mol2 with two beads. Should -a 1 2 1 3 3 4 be specified, Orientation would calculate  $s_{1,2}$ ,  $s_{1,3}$ , and  $s_{3,4}$  for mol1 and  $s_{1,2}$  and  $s_{1,2}$  for mol2 (it would literally calculate and print the same result twice). The pair 3 4 would be ignored for mol2.

The **<output>** file contains distributions of the orientation order parameter with averages of the parameter appended at the end.

Usage:

#### Orientation <input> <width> <output> <mol name(s)> <options>

Mandatory arguments		
<input/> <width> <output> <mol name(s)=""></mol></output></width>	width of each bin of the distribution output file with distribution of the orientation order parameters.	
Non-standard options		
-st <int> -e <int> -a <axis> -n <ints></ints></axis></int></int>	starting timestep for calculation (default: 1) ending timestep for calculation (default: none) axis to use as the normal vector (default: z) bead indices in the molecules (multiple of two; default: 1 2)	

#### Format of output files:

- (1)  $\langle \text{output} \rangle$  distributions of orientation order parameters,  $s_{ij}$ 
  - first line: command used to generate the file
  - next lines: column headers (on line per specified molecule type)
    - first column is the centre of each bin (governed by width); i.e., if <width> is 0.1, then the centre of the first bin is -0.45 (because  $s_{ij} \in \langle 0.5, 1 \rangle$ ), centre of the second bin -0.35, etc.

4. Utilities 4.22. PairCorrel

- next, each line shows the bead i and j indices in the specified molecule (according to the provided vsf file) to be used for calculation of  $s_{ij}$
- next lines: calculated data
- last line: commented out (i.e., the line starts with #) overall averages
- commented out lines right above the last line: column headers for the averages (same as at the beginning of the file)

## 4.22 PairCorrel

This utility calculates pair correlation function (pcf), or the radial distribution function, between specified bead types. All bead type pairs are used – if A and B are the specified bead types, A-A, A-B, and B-B bead pairs are used.

The utility does not differentiate beads of the same type that are in different molecules, so a pcf will be a sum of the beads from different molecule types (similar the DensityAggregates utility – see Section 4.7).

Usage:

PairCorrel <input> <width> <output> <bead name(s)> <options>

Mandatory arguments		
<input/>	input coordinate file (either vcf or vtf format)	
<width></width>	width of each bin of the pair correlation functions	
<output></output>	output file with pair correlation functions	
<pre><bead name(s)=""></bead></pre>	bead type(s) used for calculation	
Non-standard options		
-n <int></int>	number of bins to average to get smoother pair correlation	
	function (default: 1)	
-st <int></int>	starting timestep for calculation (default: 1)	
-e <int></int>	ending timestep for calculation (default: none)	

#### Format of output files:

- (1) **<output>** pair correlation functions between all bead types
  - first line: command used to generate the file
  - second line: column headers
  - first is the centre of each bin (governed by <width>); i.e., if <width> is 0.1, then the centre of bin 0 to 0.1 is 0.05, centre of bin 0.1 to 0.2 is 0.15, etc.
  - the rest are for the calculated data: each column corresponds to one bead types pair

# 4.23 Potential Aggregates

This utility should be working, but it needs more testing.

Potential Aggregates calculates electrostatic potential for aggregates of specified size(s) as a function of distance from their centre of mass. It places a virtual particle with charge q=1 at several places on the surface of an ever increasing sphere and calculates electrostatic potential acting on that virtual particle.

At long range, the potential is calculated using Coulomb potential,

$$U_{ij}^{\text{long}} = \frac{l_{\text{B}} q_i q_j}{r_{ij}},\tag{4.16}$$

where  $l_{\rm B}$  is the Bjerrum length,  $q_i$  and  $q_j$  are charges of particles i and j, and  $r_{ij}$  is interparticle distance. At short range, the potential is for now calculated using potential between two charges with exponentially decreasing charge density,

$$U_{ij}^{\text{short}} = U_{ij}^{\text{long}} \left[ 1 - (1 + \beta r_{ij}) e^{-2\beta r_{ij}} \right],$$
 (4.17)

where  $\beta = \frac{5r_c}{8\lambda}$  ( $r_c$  is cut-off distance and  $\lambda$  is smearing constant). The utility takes into account periodic images of the simulation box.

For now, parameters for the potential are hard coded in the source code: the Bjerrum length is bjerrum=1.1 (aqueous solution), cut-off is r\_c=3, charge smearing constant lambda=0.2, and number of periodic images of the simulation box is images=5. The parameters can be changed in the source code (around line 20), and the utility can then be recompiled.

The aggregate size can be modified using -m option similarly to Density-Aggregates (Section 4.7).

Be aware that the utility does not use any Ewald sum-based algorithm, but simple 'brute force' approach, so the calculation is extremely slow (depending on the number of charged particles in the system).

PotentialAggregates <input> <input.agg> <width> <output> <agg size(s)> <options>

Mandatory argume	ents
<input/>	input coordinate file (either vcf or vtf format)
<input.agg></input.agg>	input agg file
<width></width>	width of each bin of the distribution
<output></output>	output file with electrostatic potential
<agg size(s)=""></agg>	aggregate size(s) for calculation of electrostatic potential

4. Utilities 4.24. SelectedVcf

joined	specify that <input/> contains joined coordinates (i.e., periodic boundary conditions for aggregates do not have to
	be removed)
-st <int></int>	starting timestep for calculation (default: 1)
-e <int></int>	ending timestep for calculation (default: none)
-m <mol name(s)=""></mol>	instead of 'true' aggregate size, use the number of specified
	molecule type(s) in an aggregate

### 4.24 SelectedVcf

This utility creates a new vcf coordinate file containing only beads of specified types. The selected bead types are printed as comments at the beginning of the output vcf file. An xyz coordinate file can also be created from the selected bead type(s).

Using -r switch, the bead types can also be specified in reverse, i.e., bead types to be excluded from the output file(s).

Which timesteps are saved can be controlled using -st option for the first timestep to save and -e option for the last timestep to save; using -sk <int>, <int> steps are ignored per one step saved. If the option to save only specified timesteps (-n) is used, -sk is ignored. A maximum of 100 steps can be explicitly specified using the -n option.

There is also an option to remove periodic boundary conditions (i.e., to join molecules). Conversely, the simulation box can be wrapped (i.e., the periodic boundary conditions applied). If both --join and -w options are used, the simulation box is first wrapped and then the molecules are joined.

Also, specified molecules can be excluded which is useful when the same bead type is shared between more molecule types. However, no utilities can read a vcf file that does not contain all beads of a given type, so the -x option is useful only for visualization, e.g., using vmd.

Lastly, the --last saves to output file(s) only the last step from the input coordinate file; it is useful, when only the last timestep is required, but the total number of timesteps in the input file is unknown.

Usage:

SelectedVcf <input> <output> <bead type(s)> <options>

Mandatory arguments

4. Utilities 4.25. Surface

input coordinate file (either vcf or vtf format)

<input/>	input coordinate me (either vci or vti iormat)	
<output.vcf></output.vcf>	output vcf coordinate file with indexed coordinates	
<pre><bead type(s)=""></bead></pre>	bead type names to save (can be omitted if -r is used)	
Non-standard options		
-r	reverse function, i.e., exclude <bead type(s)=""> instead of in-</bead>	
	cluding them; if no <bead type(s)=""> are specified, all bead</bead>	
	types are used (requires <input/> with all bead types)	
join	join molecules by removing periodic boundary conditions	
<b>-</b> ₩	wrap simulation box (i.e., apply periodic boundary condi-	
	tions)	
-st <int></int>	starting timestep for calculation (default: 1)	
-e <int></int>	ending timestep for calculation (default: none)	
-sk <int></int>	number of steps skip per one used (default: 0)	
-n < int(s) >	save only specified timesteps	
-x < name(s) >	exclude molecules of specified name(s) – do not use if	
	<pre><output.vcf> is further analysed</output.vcf></pre>	
-xyz <name></name>	save coordinates to $xyz$ file – does not take into account $-x$	
	option	
last	save only the last step	

## 4.25 Surface

<innut>

Surface utility determines the average surface of structures such as membranes or or polymer brushes.

The utility cuts the plane perpendicular to the chosen axis into squares of size  $\langle width \rangle *< width \rangle$  and then finds the beads with the highest and lowest coordinates along the chosen axis whose other two coordinates fall into the square for each square on the plane. By default, it searches the chosen axis for the beads with the highest coordinate in the interval  $\langle 0, (boxlength)/2 \rangle$  and with the lowest coordinate in the interval  $\langle (box\ length)/2; (box\ length) \rangle$ , i.e., it assumes something such as a polymer brush on each wall. If  $\neg$ in option is used, it searches for a layer structure inside the box, i.e., it searches for the bead with the lowest and highest coordinates in the whole box,  $\langle 0; (box\ length) \rangle$  (this mode finds surfaces for something like a bilayer).

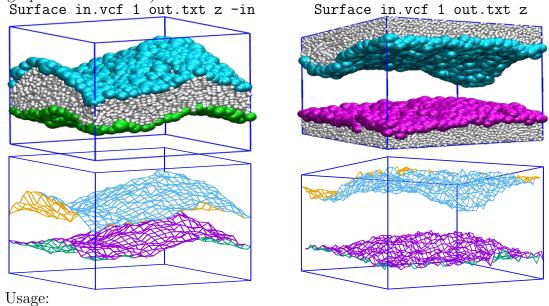
Note that the utility does not determines any structures for now, therefore any molecules outside the brush/membrane can be recognised as a part of the surface (-bt and -m can be sometimes used to eliminate this problem).

By default, all bead types and all molecule types are used, but using -bt and -m options, only specified bead and/or molecule types can be used. This is particularly

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useful when, e.g., other molecules solubilise inside the brush/membrane, leaving some molecules in the solution.

Following are two examples of usage with and without -in option (on top are snapshots with coloured balls representing the detected surface and on the bottom are graphs of the surface):



Surface <input> <width> <output> <axis> <options>

Mandatory arguments		
<input/>	input coordinate file (either vcf or vtf format)	
<width></width>	side length of each square	
<output></output>	output file	
<axis></axis>	direction in which to determine the surface: x, y, or z	
Non-standard options		
-in	start from the box centre instead of from its edges	
-m < name(s) >	molecule type(s) to use (default: all)	
-bt <name(s)></name(s)>	bead type(s) to use (default: all)	
-st <int></int>	starting timestep for calculation (default: 1)	
-e <int></int>	ending timestep for calculation (default: none)	

#### Format of output files:

- (1) <output> 3D coordinates
  - first line: command used to generate the file
  - second line: column headers

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- first two numbers represent the centre of each square (being the coordinates on the sliced up plane); i.e., if <width> is 1, then the centre of the first square is 0.5 0.5, the centre of the second one is 0.5 1.5, etc.

the second two numbers are coordinates of the surface in the third dimension,
 i.e., along the chosen axis

## 4.26 traject

This utility is from the DL\_MESO simulation package and comes in three versions for 2.5, 2.6, and 2.7 versions of DL\_MESO. For the latest DL\_MESO version, the utility is unmodified and therefore is not included here. The utilities traject-v2\_5 and traject-v2\_6 shipped with DL\_MESO 2.5 and 2.6 were modified to generate separate vsf and vcf file (the traject.exe for DL\_MESO 2.7 does this natively with -sc command line option).

Usage of 2.5 and 2.6 versions (see DL\_MESO manual for the description of latest version):

traject-v2\_5 <int> or traject-v2\_6 <int>

#### Mandatory argument

<int> number of computer cores used for the simulation run (equals the number of HISTORY files)

### 4.27 TransformVsf

This utility reads FIELD and vsf files to create a new structure file. Generally, this is useful only if traject-v2\_5 or traject-v2\_6 were used to generate the d1\_meso.vsf file. This file does not contain mass and charge of particles, whereas the one generated with TransformVsf (or with traject.exe from the dl\_meso version 2.7) does and therefore the original FIELD file is not needed for further analysis.

Usage (TransformVsf does not use standard options):

TransformVsf <output.vsf> <options>

1	/Ian	datory	argum	ent

<output.vsf>
Options

output structure file

-i <name></name>	use custom structure file instead of traject.vsf (vsf or vtf
	format)
$-\Lambda$	verbose output
-h	print this help and exit

# 4.28 VisualizeAgg

This utility prints aggregates of given size to output vcf files (one file per aggregate size). The output files contain one aggregate per timestep regardless of how many aggregates of the given size are in the initial coordinate file.

The definition of aggregate size is fairly flexible (similarly to DensityAggregates utility – Section 4.7).

As the output file(s) do not necessarily contain all beads of a bead type, these vcf files cannot be used for further analysis via AnalysisTools.

VisualizeAgg <input> <input.agg> <output> <agg
size(s)> <options>

Mandatory arguments		
<input/>	input coordinate file (either vcf or vtf format)	
<input.agg></input.agg>	input .agg file	
<output></output>	output file(s) (one per aggregate size) with automatic #.vcf	
	ending (# is aggregate size)	
<agg size(s)=""></agg>	aggregate size(s) to save	
Non-standard options		
joined	specify that <input/> contains joined coordinates (i.e., periodic boundary conditions for aggregates do not have to be removed)	
-st <int></int>	starting timestep for calculation (default: 1)	
-e <int></int>	ending timestep for calculation (default: none)	
-m <mol name(s)=""></mol>	instead of 'true' aggregate size, use the number of specified molecule type(s) in an aggregate	
-x <mol name(s)=""></mol>	exclude aggregates containing only specified molecule $\operatorname{type}(s)$	

# 5. Computational details

This chapter will contain some information about how things are coded in the utilities.

## 5.1 Read system data

ReadStructure() function reads all system information from vsf and vcf files. FIELD file is used only to get mass and charge of beads if the information is not in vsf file. Provided vsf file is used to get all information about beads and molecules – names and numbers of bead and molecules, bonds in molecules. The first timestep of the vcf file is used to determine numbers and ids of beads in that vcf file which means that all timesteps must contain the same beads.

The procedure in ReadStructure() is as follows:

- (1) Go through atom section of the vsf file to identify default bead type (if atom default line is present), to find highest bead and molecule ids, and to find bead type names (and charges and masses if present).
- (2) Go again through the atom section to read names and ids of beads and molecules as well as numbers of all beads and molecules for each type.
- (3) Go through bond section of the vsf file to calculate number of bonds in each molecule type.
- (4) Go again through the bond section to read bonds for each molecule type.
- (5) Go through the atom section (for the third time) to assign bead ids to molecules.
- (6) Go through the first timestep of vcf file to find which beads are in that vcf file (if no vcf file is provided e.g., for DistrAgg utility assume all beads are used).
- (7) Modify all arrays to accommodate only the beads that are present in the vcf file.
- (8) Read charge and mass from the FIELD file, if not already known from vsf file.