Infrastructure classes for high-throughput SNP data

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This document describes some of the infrastructure classes used for high-throughput genomic data. For the classes used to organize SNP data, we provide examples for initialization and illustrate some of the accessors. We should add a diagram showing the relationships of these classes here.

[Insert diagram of classes here]

1 Feature-level classes

2 Locus-level classes

The examples below are completely simulated and are not meant to convey any biological plausibility.

2.1 SnpSet

2.1.1 Initialization

```
> theCalls <- matrix(sample(1:3, 20, rep = TRUE), nc = 2)
> p <- matrix(runif(20), nc = 2)
> theConfs <- round(-1000 * log2(1 - p))
> obj <- new("SnpSet", call = theCalls, callProbability = theConfs)
```

```
2.1.2 Accessors
> calls(obj)
   1 2
  3 2
  1 3
  2 2
  1 2
  2 3
  3 3
7
  3 2
 3 3
  2 2
10 1 3
> confs(obj)
  0.3188686 0.19022593
  0.4774319 0.18453763
  0.2576987 0.32429589
  0.3892087 0.15464617
  0.6518076 0.88875274
```

```
6 0.5876983 0.64971214
7 0.0924440 0.59464822
8 0.8298373 0.01783897
9 0.1622202 0.28179483
10 0.7553679 0.09516258
2.1.3 Annotating
> if (require("genomewidesnp6Crlmm")) {
      ids <- c("SNP_A-2131660", "SNP_A-1967418", "SNP_A-1969580",
          "SNP_A-4263484", "SNP_A-1978185", "SNP_A-4264431",
          "SNP_A-1980898", "SNP_A-1983139", "SNP_A-4265735",
+
          "SNP_A-1995832")
      rownames(theCalls) <- rownames(p) <- rownames(theConfs) <- ids</pre>
+
      obj <- new("SnpSet", call = theCalls, callProbability = theConfs,</pre>
          annotation = "genomewidesnp6")
      featureData(obj) <- addFeatureAnnotation(obj)</pre>
      fvarLabels(obj)
      isSnp(obj)
      position(obj)
      chromosome(obj)
+ }
 [1] 1 1 1 1 1 1 1 1 1 1
2.2
      CopyNumberSet
2.2.1 Initialization
2.2.2 Accessors
2.2.3 Annotating
2.3
      CNSet
2.3.1 Initialization
> theCalls <- matrix(2, nc = 2, nrow = 10)
> A <- matrix(sample(1:1000, 20), 10, 2)
> B <- matrix(sample(1:1000, 20), 10, 2)
> CA <- matrix(rnorm(20, 1), nrow = 10)
> CB <- matrix(rnorm(20, 1), nrow = 10)
> p <- matrix(runif(20), nc = 2)
> theConfs <- round(-1000 * log2(1 - p))</pre>
> obj <- new("CNSet", alleleA = A, alleleB = B, call = theCalls,
      callProbability = theConfs, CA = CA, CB = CB)
2.3.2 Accessors
> calls(obj)
   1 2
1 2 2
2 2 2
3 2 2
4 2 2
```

5 2 2

```
6 2 2
7 2 2
```

8 2 2

9 2 2

10 2 2

> confs(obj)

1

1 0.65285058 0.20863818

2 0.09425729 0.94128147

3 0.99397598 0.89585801

4 0.85221548 0.86220663

5 0.91791500 0.95056022

6 0.07132831 0.98925162

7 0.57514188 0.22275526

8 0.24346010 0.07780631

9 0.35853458 0.57044264

10 0.07688365 0.67793439

> A(obj)

1 2

1 366 508

2 198 71

3 643 45

4 913 130

5 10 887

6 417 221

7 590 311

8 178 424

9 429 992

10 447 139

> B(obj)

1 2

1 769 908

2 402 394

3 680 358

4 79 970

5 8 696

6 85 582

7 749 929

8 767 435

9 728 21

10 344 205

> CA(obj)

1 2

1 0.09763923 0.7451489

2.16017476 -0.3168747

3 1.76494762 -0.9359526

4 -0.13252238 0.6921249

5 1.01859800 1.8430487

```
-0.12303938 -0.8673966
   2.01012557 1.7466194
7
  -0.38111920 1.1632133
   3.92954265
               0.5046183
10 -0.32755537
               1.6799286
> CB(obj)
                        2
            1
1
  -0.2248659
               2.28865399
2
               2.59149180
   0.4969427
3
   2.3889220
               1.96349414
4
   0.8114921
               0.83451404
5
   2.1217974
               2.57355114
6
   1.1309536
              1.59493715
7
   2.7230521 0.67455462
8
   2.0732825
               0.22707415
9
   1.1630574 -0.09253353
10 1.4193042 1.01571406
```

2.3.3 Annotating

Annotating with chromosome and physical position:

3 Session Information

The version number of R and packages loaded for generating the vignette were:

- R version 2.11.0 (2010-04-22), x86_64-apple-darwin9.8.0
- Locale: en_US.UTF-8/en_US.UTF-8/C/C/en_US.UTF-8/en_US.UTF-8
- Base packages: base, datasets, graphics, grDevices, methods, stats, tools, utils
- Other packages: Biobase 2.8.0, genomewidesnp6Crlmm 1.0.4, oligoClasses 1.11.3
- Loaded via a namespace (and not attached): affyio 1.15.2, Biostrings 2.16.0, IRanges 1.5.74