

class19

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```
#1 echo=FALSE

cdc <- data.frame( year = c(1922L,1923L,1924L,1925L,1926L,
                            1927L,1928L,1929L,1930L,1931L,
                            1932L,1933L,1934L,1935L,1936L,1937L,
                            1938L,1939L,1940L,1941L,1942L,
                            1943L,1944L,1945L,1946L,1947L,1948L,
                            1949L,1950L,1951L,1952L,1953L,1954L,
                            1955L,1956L,1957L,1958L,1959L,
                            1960L,1961L,1962L,1963L,1964L,1965L,
                            1966L,1967L,1968L,1969L,1970L,
                            1971L,1972L,1973L,1974L,1975L,1976L,
                            1977L,1978L,1979L,1980L,1981L,
                            1982L,1983L,1984L,1985L,1986L,1987L,
                            1988L,1989L,1990L,1991L,1992L,1993L,
                            1994L,1995L,1996L,1997L,1998L,
                            1999L,2000L,2001L,2002L,2003L,2004L,
                            2005L,2006L,2007L,2008L,2009L,
                            2010L,2011L,2012L,2013L,2014L,2015L,
                            2016L,2017L,2018L,2019L,2020L,
                            2021L), cases = c(107473,164191,165418,152003,
                            202210,181411,161799,197371,166914,
                            172559,215343,179135,265269,180518,
                            147237,214652,227319,103188,183866,
                            222202,191383,191890,109873,133792,
                            109860,156517,74715,69479,120718,
                            68687,45030,37129,60886,62786,
                            31732,28295,32148,40005,14809,11468,
                            17749,17135,13005,6799,7717,9718,
                            4810,3285,4249,3036,3287,1759,
                            2402,1738,1010,2177,2063,1623,1730,
```

```

1248,1895,2463,2276,3589,4195,
2823,3450,4157,4570,2719,4083,6586,
4617,5137,7796,6564,7405,7298,
7867,7580,9771,11647,25827,25616,
15632,10454,13278,16858,27550,18719,
48277,28639,32971,20762,17972,
18975,15609,18617,6124,2116)
)

```

Lets have a wee look at this table

```
head(cdc)
```

```

  year  cases
1 1922 107473
2 1923 164191
3 1924 165418
4 1925 152003
5 1926 202210
6 1927 181411

```

Q1. With the help of the R “addin” package datapasta assign the CDC pertussis case number data to a data frame called cdc and use ggplot to make a plot of cases numbers over time.

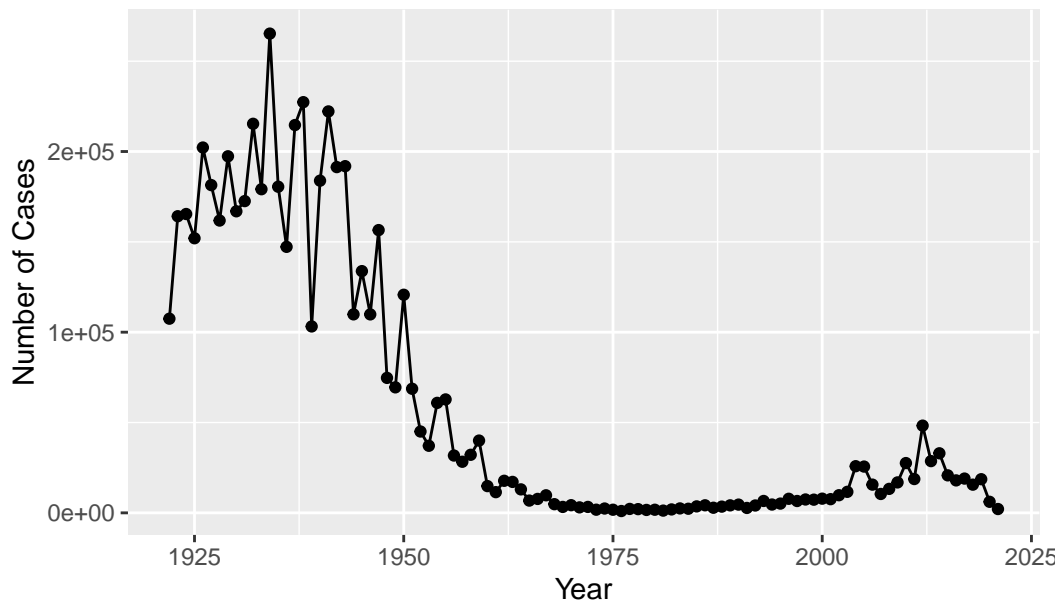
```

library(ggplot2)

ggplot(cdc, aes(x = year, y = cases)) +
  geom_point() +
  geom_line() +
  labs(title = "CDC Pertussis Case Numbers Over Time",
       x = "Year",
       y = "Number of Cases")

```

CDC Pertussis Case Numbers Over Time



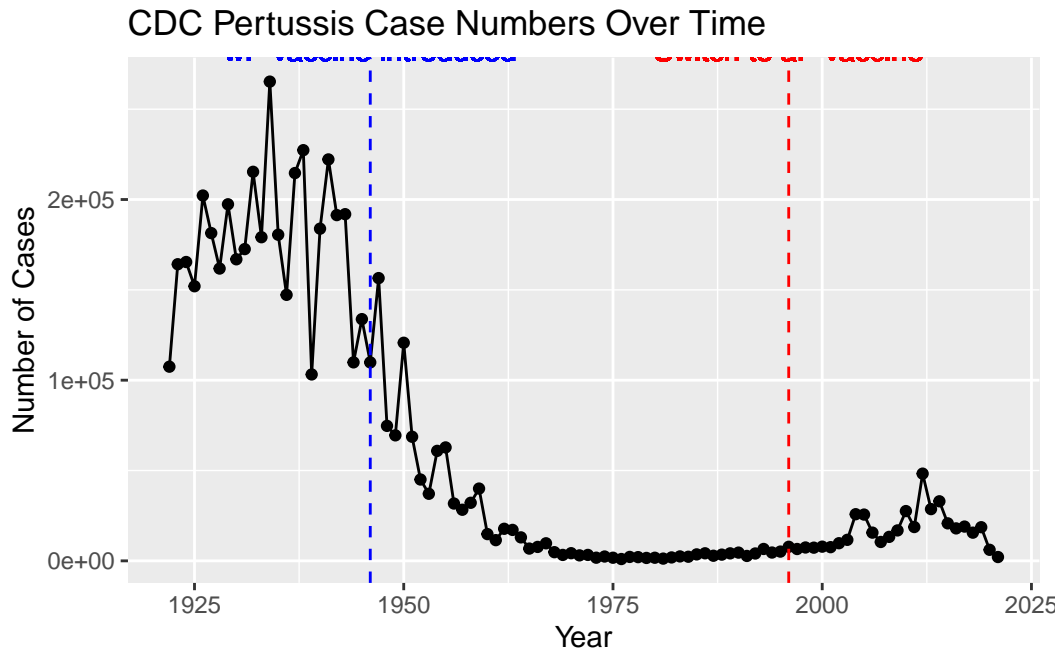
Q2. Using the ggplot `geom_vline()` function add lines to your previous plot for the 1946 introduction of the wP vaccine and the 1996 switch to aP vaccine (see example in the hint below). What do you notice?

```
library(ggplot2)

# Your existing plot code
ggplot(cdc, aes(x = year, y = cases)) +
  geom_point() +
  geom_line() +
  labs(title = "CDC Pertussis Case Numbers Over Time",
        x = "Year",
        y = "Number of Cases") +

  geom_vline(xintercept = 1946, linetype = "dashed", color = "blue") +
  geom_vline(xintercept = 1996, linetype = "dashed", color = "red") +

  geom_text(aes(x = 1946, y = max(cases), label = "wP vaccine introduced", vjust = -1), color = "blue") +
  geom_text(aes(x = 1996, y = max(cases), label = "Switch to aP vaccine", vjust = -1), color = "red")
```



Q3. Describe what happened after the introduction of the aP vaccine? Do you have a possible explanation for the observed trend?

From the lab hands-on The answer is : It is clear from the CDC data that pertussis cases are once again increasing. For example, we can see that in 2012 the CDC reported 48,277 cases of pertussis in the United States. This is the largest number of cases reported since 1955, when 62,786 cases were reported. The pertussis field has several hypotheses for the resurgence of pertussis including (in no particular order): 1) more sensitive PCR-based testing, 2) vaccination hesitancy 3) bacterial evolution (escape from vaccine immunity), 4) waning of immunity in adolescents originally primed as infants with the newer aP vaccine as compared to the older wP vaccine.

More people get the diseases.

```
library(jsonlite)
subject <- read_json("https://www.cmi-pb.org/api/subject", simplifyVector = TRUE)
head(subject, 3)
```

	subject_id	infancy_vac	biological_sex	ethnicity	race
1	1	wP	Female	Not Hispanic or Latino	White
2	2	wP	Female	Not Hispanic or Latino	White

	3	wP	Female	Unknown White
	year_of_birth	date_of_boost	dataset	
1	1986-01-01	2016-09-12	2020_dataset	
2	1968-01-01	2019-01-28	2020_dataset	
3	1983-01-01	2016-10-10	2020_dataset	

Q4. How many aP and wP infancy vaccinated subjects are in the dataset?

```
table(subject$infancy_vac)
```

```
aP wP
60 58
```

```
library(jsonlite)

# Subject table
subject <- read_json("https://www.cmi-pb.org/api/subject", simplifyVector = TRUE)
specimen <- read_json("http://cmi-pb.org/api/specimen", simplifyVector = TRUE)
titer <- read_json("http://cmi-pb.org/api/v4/plasma_ab_titer", simplifyVector = TRUE )
```

Q5. How many Male and Female subjects/patients are in the dataset?

```
table(subject$biological_sex)
```

```
Female  Male
79      39
```

Q6. What is the breakdown of race and biological sex (e.g. number of Asian females, White males etc...)?

```
table(subject$race, subject$biological_sex)
```

	Female	Male
American Indian/Alaska Native	0	1
Asian	21	11
Black or African American	2	0

More Than One Race	9	2
Native Hawaiian or Other Pacific Islander	1	1
Unknown or Not Reported	11	4
White	35	20

```
library(lubridate)
```

Attaching package: 'lubridate'

The following objects are masked from 'package:base':

date, intersect, setdiff, union

```
today()
```

```
[1] "2023-12-07"
```

```
today() - ymd("2000-01-01")
```

Time difference of 8741 days

```
time_length( today() - ymd("2000-01-01"), "years")
```

```
[1] 23.93155
```

Q7. Using this approach determine (i) the average age of wP individuals, (ii) the average age of aP individuals; and (iii) are they significantly different?

```
library(dplyr)
```

Attaching package: 'dplyr'

The following objects are masked from 'package:stats':

filter, lag

The following objects are masked from 'package:base':

```
intersect, setdiff, setequal, union
```

```
library(lubridate)

subject$age <- ymd(subject$date_of_boost) - ymd(subject$year_of_birth)

subject$age_years <- time_length(subject$age, "years")

ap <- subject %>% filter(infancy_vac == "aP")
round(summary(time_length(ap$age, "years")))
```

Min.	1st Qu.	Median	Mean	3rd Qu.	Max.
19	20	20	21	21	28

```
wp <- subject %>% filter(infancy_vac == "wP")
round(summary(time_length(wp$age, "years")))
```

Min.	1st Qu.	Median	Mean	3rd Qu.	Max.
23	26	29	31	34	51

Q8. Determine the age of all individuals at time of boost?

```
int <- ymd(subject$date_of_boost) - ymd(subject$year_of_birth)
age_at_boost <- time_length(int, "year")
head(age_at_boost)
```

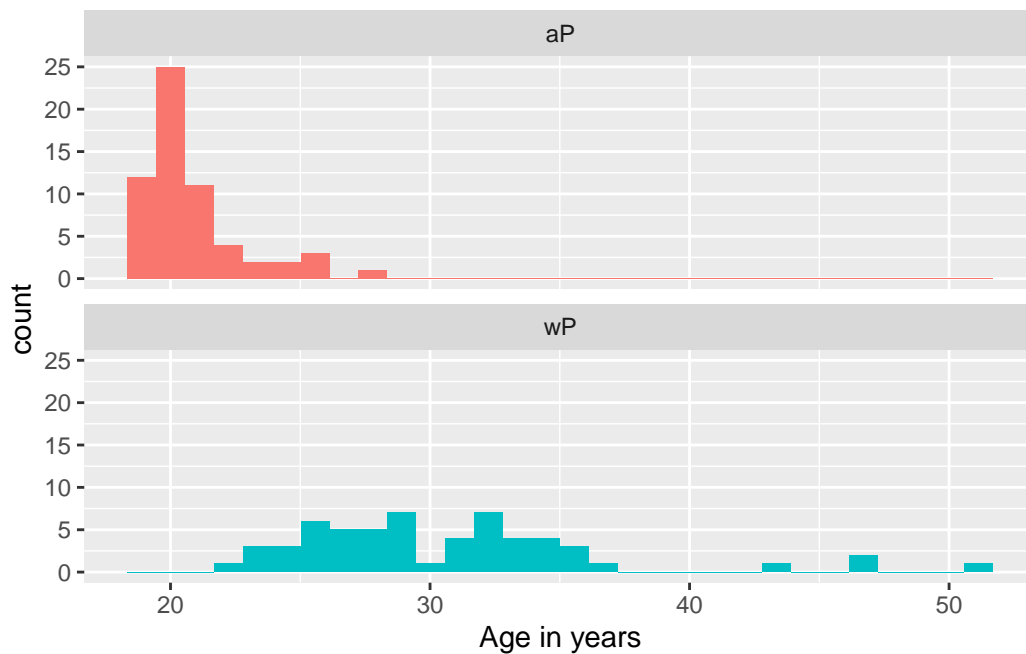
```
[1] 30.69678 51.07461 33.77413 28.65982 25.65914 28.77481
```

Q9. With the help of a faceted boxplot or histogram (see below), do you think these two groups are significantly different?

```
library(ggplot2)
ggplot(subject) +
  aes(time_length(age, "year"),
       fill=as.factor(infancy_vac)) +
  geom_histogram(show.legend=FALSE) +
```

```
facet_wrap(vars(infancy_vac), nrow=2) +
xlab("Age in years")
```

`stat_bin()` using `bins = 30`. Pick better value with `binwidth`.



```
# Or use wilcox.test()
x <- t.test(time_length( wp$age, "years" ),
            time_length( ap$age, "years" ))

x$p.value
```

[1] 9.121472e-19

Q9. Complete the code to join specimen and subject tables to make a new merged data frame containing all specimen records along with their associated subject details:

```
meta <- inner_join(specimen,subject)
```

Joining with `by = join_by(subject_id)`


```
dim(meta)
```

```
[1] 939 15
```

```
head(meta)
```

```
specimen_id subject_id actual_day_relative_to_boost
1           1           1                      -3
2           2           1                       1
3           3           1                       3
4           4           1                       7
5           5           1                      11
6           6           1                      32

planned_day_relative_to_boost specimen_type visit infancy_vac biological_sex
1                           0         Blood     1          wP         Female
2                           1         Blood     2          wP         Female
3                           3         Blood     3          wP         Female
4                           7         Blood     4          wP         Female
5                          14         Blood     5          wP         Female
6                          30         Blood     6          wP         Female

ethnicity race year_of_birth date_of_boost dataset
1 Not Hispanic or Latino White 1986-01-01 2016-09-12 2020_dataset
2 Not Hispanic or Latino White 1986-01-01 2016-09-12 2020_dataset
3 Not Hispanic or Latino White 1986-01-01 2016-09-12 2020_dataset
4 Not Hispanic or Latino White 1986-01-01 2016-09-12 2020_dataset
5 Not Hispanic or Latino White 1986-01-01 2016-09-12 2020_dataset
6 Not Hispanic or Latino White 1986-01-01 2016-09-12 2020_dataset

age age_years
1 11212 days 30.69678
2 11212 days 30.69678
3 11212 days 30.69678
4 11212 days 30.69678
5 11212 days 30.69678
6 11212 days 30.69678
```

Q10. Now using the same procedure join meta with titer data so we can further analyze this data in terms of time of visit aP/wP, male/female etc.

```
abdata <- inner_join(titer, meta)
```

Joining with `by = join_by(specimen_id)`

```
dim(abdata)
```

```
[1] 41810    22
```

Q11. How many specimens (i.e. entries in abdata) do we have for each isotype?

```
table(abdata$isotype)
```

```
 IgE  IgG IgG1 IgG2 IgG3 IgG4
6698 3240 7968 7968 7968 7968
```

Q12. What are the different \$dataset values in abdata and what do you notice about the number of rows for the most “recent” dataset?

```
table(abdata$dataset)
```

```
2020_dataset 2021_dataset 2022_dataset
        31520         8085         2205
```

```
igg <- abdata %>% filter(isotype == "IgG")
head(igg)
```

	specimen_id	isotype	is_antigen_specific	antigen	MFI	MFI_normalised
1	1	IgG	TRUE	PT	68.56614	3.736992
2	1	IgG	TRUE	PRN	332.12718	2.602350
3	1	IgG	TRUE	FHA	1887.12263	34.050956
4	19	IgG	TRUE	PT	20.11607	1.096366
5	19	IgG	TRUE	PRN	976.67419	7.652635
6	19	IgG	TRUE	FHA	60.76626	1.096457

	unit	lower_limit_of_detection	subject_id	actual_day_relative_to_boost
1	IU/ML	0.530000	1	-3
2	IU/ML	6.205949	1	-3
3	IU/ML	4.679535	1	-3
4	IU/ML	0.530000	3	-3
5	IU/ML	6.205949	3	-3

6	IU/ML	4.679535	3	-3	
	planned_day_relative_to_boost	specimen_type	visit	infancy_vac	biological_sex
1	0	Blood	1	wP	Female
2	0	Blood	1	wP	Female
3	0	Blood	1	wP	Female
4	0	Blood	1	wP	Female
5	0	Blood	1	wP	Female
6	0	Blood	1	wP	Female

	ethnicity	race	year_of_birth	date_of_boost	dataset
1	Not Hispanic or Latino	White	1986-01-01	2016-09-12	2020_dataset
2	Not Hispanic or Latino	White	1986-01-01	2016-09-12	2020_dataset
3	Not Hispanic or Latino	White	1986-01-01	2016-09-12	2020_dataset
4	Unknown	White	1983-01-01	2016-10-10	2020_dataset
5	Unknown	White	1983-01-01	2016-10-10	2020_dataset
6	Unknown	White	1983-01-01	2016-10-10	2020_dataset

	age	age_years
1	11212 days	30.69678
2	11212 days	30.69678
3	11212 days	30.69678
4	12336 days	33.77413
5	12336 days	33.77413
6	12336 days	33.77413

Q13. Complete the following code to make a summary boxplot of Ab titer levels (MFI) for all antigens:

```
ggplot(igg)+
  aes(MFI_normalised ,
      antigen) +
  geom_boxplot() +
  xlim(0,75) +
  facet_wrap(vars(visit),nrow = 2)
```

Warning: Removed 5 rows containing non-finite values (`stat_boxplot()`).

