


## ✓ Загрузка библиотек

```
from google.colab import drive
drive.mount('/content/drive')
```

 Mounted at /content/drive

```
import pandas as pd
import warnings
warnings.filterwarnings("ignore")
```

```
import numpy as np
```

```
pip install -U scikit-learn
```

 [Показать скрытые выходные данные](#)

```
import sklearn
```

```
pip install scanpy
```

 [Показать скрытые выходные данные](#)

```
import scanpy as sc
```

```
import scipy as sp
```

```
import pandas as pd
```

```
import matplotlib.pyplot as plt
```

```
import matplotlib.image as mpimg
```

```
import seaborn as sb
```

```
pip install vaex
```

 [Показать скрытые выходные данные](#)

```
pip install loompy
```

 [Показать скрытые выходные данные](#)

```
import loompy
```

## ✓ 1.Загрузка

```
# загрузка файла формата loom    Вариант 1
adata = sc.read_loom('/content/drive/MyDrive/Cell_2/анализ/ПМЖ.loom', sparse=True, cleanup=False,
                    X_name='spliced', obs_names='CellID', var_names='Gene')
```

```
gene_names = pd.read_csv('/content/drive/MyDrive/Cell_2/анализ/ПМЖ гены.csv', index_col = 0)
named_list = adata.var.join(gene_names)
```

```
#загрузка файла формата csv    Вариант 2
data_d = pd.read_csv('/content/drive/MyDrive/Cell_2/анализ/PART_1CSV.csv')
data_d
```

Unnamed: 0															
		PBMC3	PTEN	CDH1	PBMC1	PBMC1.1	BRCA2	BRCA1	ATM	BRCA1.1	...	PBMC1.35	BRCA2.55	BRCA1.47	ATM.4
0	AU565_AAACCAGTTTGG	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	...	0.0	0.0	0.0	0.0
1	AU565_AAACGTGCAGCG	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	...	0.0	0.0	0.0	0.0
2	AU565_AAAGCCACATGC	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	...	0.0	0.0	0.0	0.0
3	AU565_AAAGTCGGCTGG	0.0	0.0	0.0	3.0	0.0	0.0	0.0	0.0	0.0	...	0.0	0.0	0.0	1.0
4	AU565_AAAGTGCCTAAA	1.0	7.0	0.0	0.0	1.0	0.0	0.0	0.0	0.0	...	0.0	0.0	0.0	0.0
...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
583		NaN	1.0	NaN	NaN	NaN	NaN	NaN	NaN	NaN	...	NaN	NaN	NaN	NaN
584		NaN	NaN	NaN	NaN	NaN	NaN	NaN	NaN	NaN	...	NaN	NaN	NaN	NaN
585		NaN	NaN	NaN	NaN	NaN	NaN	NaN	NaN	NaN	...	NaN	NaN	NaN	NaN
586		NaN	NaN	NaN	NaN	NaN	NaN	NaN	NaN	NaN	...	NaN	NaN	NaN	NaN
587		NaN	NaN	NaN	NaN	NaN	NaN	NaN	NaN	NaN	...	NaN	NaN	NaN	NaN

588 rows × 256 columns

```
data_m = pd.read_csv('/content/drive/MyDrive/Cell_2/анализ/breast-cancer (1).csv')
data_m
```

	Gene	Index_Cell	organism	id	Cell_type	radius_mean	texture_mean	perimeter_mean	area_mean
0	ENSG00000006634	AU565_ATAACAGCCACT	Human Breast	89511501	Basal cells	12.20	15.21	78.01	457.9
1	ENSG00000006652	AU565_ATAAGGGCTGAC	Human Breast	8911800	Basal cells	13.59	17.84	86.24	572.3
2	ENSG00000006695	AU565_ATACCGCACACA	Human Breast	924084	Basal cells	12.77	29.43	81.35	507.9
3	ENSG00000006704	AU565_ATACGCGACACA	Human Breast	869218	Basal cells	11.43	17.31	73.66	398.0
4	ENSG00000006712	AU565_ATAGACTCGCAG	Human Breast	902976	Basal cells	13.88	16.16	88.37	596.6
...	...	...	...	...	...	...	...	...	...
565	ENSG00000063438	AU565_TGTGTCTGAGCG	Human Breast	927241	Unknown	20.60	29.33	140.10	1265.0
566	ENSG00000063587	AU565_TGTGTTCTACGT	Human Breast	878796	Unknown	23.29	26.67	158.90	1685.0
567	ENSG00000063601	AU565_TGTTACAATACG	Human Breast	911296202	Unknown	27.42	26.27	186.90	2501.0
568	ENSG00000063660	AU565_TGTTTCACCTGA	Human Breast	8610862	Unknown	20.18	23.97	143.70	1245.0
569	ENSG00000063854	AU565_TTAATGAGGACG	Human Breast	86355	Unknown	22.27	19.67	152.80	1509.0

570 rows × 36 columns

2.1 Анализ данных

Для данных только формата csv

Необходимо данные объединить в один файл

```
data_d.head()
```

	Unnamed: 0	PBMC3	PTEN	CDH1	PBMC1	PBMC1.1	BRCA2	BRCA1	ATM	BRCA1.1	...	PBMC1.35	BRCA2.55	BRCA1.47	ATM.42
0	AU565_AAACCAGTTTGG	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	...	0.0	0.0	0.0	0.0
1	AU565_AAACGTGCAGCG	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	...	0.0	0.0	0.0	0.0
2	AU565_AAAGCCACATGC	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	...	0.0	0.0	0.0	0.0
3	AU565_AAAGTCGGCTGG	0.0	0.0	0.0	3.0	0.0	0.0	0.0	0.0	0.0	...	0.0	0.0	0.0	1.0
4	AU565_AAAGTGCCTAAA	1.0	7.0	0.0	0.0	1.0	0.0	0.0	0.0	0.0	...	0.0	0.0	0.0	0.0

5 rows × 256 columns

Строки в столбцы

```
df = df.T
df.columns = list("ABCD")
```

#Если необходимо поменять местами столбцы и строки в исходных данных  
print(data\_d.T)

	0	1	2	\
Unnamed: 0	AU565_AAACCAGTTTGG	AU565_AAACGTGCAGCG	AU565_AAAGCCACATGC	
PBMC3	0.0	0.0	0.0	
PTEN	0.0	0.0	0.0	
CDH1	0.0	0.0	0.0	
PBMC1	0.0	0.0	0.0	
...	...	...	...	
ATM.43	0.0	0.0	0.0	
ATM.44	0.0	0.0	0.0	
ATM.45	0.0	0.0	0.0	
ATM.46	0.0	0.0	0.0	
ATM.47	0.0	0.0	0.0	

	3	4	5	\
Unnamed: 0	AU565_AAAGTCGGCTGG	AU565_AAAGTGCCTAAA	AU565_AAATCATCTTAT	
PBMC3	0.0	1.0	2.0	
PTEN	0.0	7.0	0.0	
CDH1	0.0	0.0	0.0	
PBMC1	3.0	0.0	0.0	
...	...	...	...	
ATM.43	0.0	0.0	1.0	
ATM.44	0.0	0.0	0.0	
ATM.45	0.0	0.0	0.0	
ATM.46	0.0	0.0	1.0	
ATM.47	0.0	0.0	0.0	


	6	7	8	\
Unnamed: 0	AU565_AAATCATTGTCT	AU565_AAATTACTTCAT	AU565_AAATTATTAGAT	
PBMC3	0.0	0.0	0.0	
PTEN	0.0	0.0	0.0	
CDH1	0.0	0.0	0.0	
PBMC1	0.0	0.0	0.0	
...	...	...	...	
ATM.43	0.0	0.0	0.0	
ATM.44	0.0	0.0	0.0	
ATM.45	0.0	0.0	0.0	
ATM.46	0.0	0.0	0.0	
ATM.47	0.0	0.0	0.0	

	9	...	578	579	580	581	582	583	584	585	\
Unnamed: 0	AU565_AAATTGATTGT	...	NaN	NaN	NaN	NaN	NaN	NaN	NaN	NaN	
PBMC3	0.0	...	NaN	NaN	NaN	NaN	NaN	1.0	NaN	NaN	
PTEN	1.0	...	NaN	NaN	NaN	NaN	NaN	NaN	NaN	NaN	
CDH1	0.0	...	NaN	NaN	NaN	NaN	NaN	NaN	NaN	NaN	
PBMC1	2.0	...	NaN	NaN	NaN	NaN	NaN	NaN	NaN	NaN	
...	...	...	...	...	...	...	...	...	...	...	
ATM.43	0.0	...	NaN	NaN	NaN	NaN	NaN	NaN	NaN	NaN	
ATM.44	0.0	...	NaN	NaN	NaN	NaN	NaN	NaN	NaN	NaN	
ATM.45	1.0	...	NaN	NaN	NaN	NaN	NaN	NaN	NaN	NaN	
ATM.46	0.0	...	NaN	NaN	NaN	NaN	NaN	NaN	NaN	NaN	
ATM.47	0.0	...	NaN	NaN	NaN	NaN	NaN	NaN	NaN	NaN	

	586	587
Unnamed: 0	NaN	NaN
PBMC3	NaN	NaN
PTEN	NaN	NaN
CDH1	NaN	NaN
PBMC1	NaN	NaN

```
data_d.columns = ['Index_Cell', *data_d.columns[1:]]
```

```
data_d.T.rename(columns=data_d.T.iloc[0]).drop(data_d.T.index[0])
```




	AU565_AAACCAGTTTGG	AU565_AAACGTGCAGCG	AU565_AAAGCCACATGC	AU565_AAAGTCGGCTGG	AU565_AAAGTGCCTAAA	AU565_AAATCATCTTAT	AU
PBMC3	0.0	0.0	0.0	0.0	1.0	2.0	
PTEN	0.0	0.0	0.0	0.0	7.0	0.0	
CDH1	0.0	0.0	0.0	0.0	0.0	0.0	
PBMC1	0.0	0.0	0.0	3.0	0.0	0.0	
PBMC1.1	0.0	0.0	0.0	0.0	1.0	0.0	
...	...	...	...	...	...	...	
ATM.43	0.0	0.0	0.0	0.0	0.0	1.0	
ATM.44	0.0	0.0	0.0	0.0	0.0	0.0	
ATM.45	0.0	0.0	0.0	0.0	0.0	0.0	
ATM.46	0.0	0.0	0.0	0.0	0.0	1.0	
ATM.47	0.0	0.0	0.0	0.0	0.0	0.0	

255 rows × 588 columns

data\_d=data\_d.T

data\_d



	0	1	2	3	4	
Index_Cell	AU565_AAACCAGTTTGG	AU565_AAACGTGCAGCG	AU565_AAAGCCACATGC	AU565_AAAGTCGGCTGG	AU565_AAAGTGCCTAAA	A
PBMC3	0.0	0.0	0.0	0.0	1.0	
PTEN	0.0	0.0	0.0	0.0	7.0	
CDH1	0.0	0.0	0.0	0.0	0.0	
PBMC1	0.0	0.0	0.0	3.0	0.0	
...	...	...	...	...	...	
ATM.43	0.0	0.0	0.0	0.0	0.0	
ATM.44	0.0	0.0	0.0	0.0	0.0	
ATM.45	0.0	0.0	0.0	0.0	0.0	
ATM.46	0.0	0.0	0.0	0.0	0.0	
ATM.47	0.0	0.0	0.0	0.0	0.0	

256 rows × 588 columns

metadata = pd.concat([data\_m], axis=0).drop\_duplicates()  
metadata = data\_m.drop\_duplicates(subset='Index\_Cell', keep="last")  
metadata

	Gene	Index_Cell	organism	id	Cell_type	radius_mean	texture_mean	perimeter_mean	area_mean
0	ENSG00000006634	AU565_ATAACAGCCACT	Human Breast	89511501	Basal cells	12.20	15.21	78.01	457.9
1	ENSG00000006652	AU565_ATAAGGGCTGAC	Human Breast	8911800	Basal cells	13.59	17.84	86.24	572.3
2	ENSG00000006695	AU565_ATACGCACACA	Human Breast	924084	Basal cells	12.77	29.43	81.35	507.9
3	ENSG00000006704	AU565_ATACGCGACACA	Human Breast	869218	Basal cells	11.43	17.31	73.66	398.0
4	ENSG00000006712	AU565_ATAGACTCGCAG	Human Breast	902976	Basal cells	13.88	16.16	88.37	596.6
...	...	...	...	...	...	...	...	...	...
565	ENSG00000063438	AU565_TGTGTCTGAGCG	Human Breast	927241	Unknown	20.60	29.33	140.10	1265.0
566	ENSG00000063587	AU565_TGTGTTCTACGT	Human Breast	878796	Unknown	23.29	26.67	158.90	1685.0
567	ENSG00000063601	AU565_TGTTACAATACG	Human Breast	911296202	Unknown	27.42	26.27	186.90	2501.0
568	ENSG00000063660	AU565_TGTTTCACCTGA	Human Breast	8610862	Unknown	20.18	23.97	143.70	1245.0
569	ENSG00000063854	AU565_TTAATGAGGACG	Human Breast	86355	Unknown	22.27	19.67	152.80	1509.0

570 rows × 36 columns

```
#Если одинаковые таблицы по наполнению, то их можно объединить
data_d = sc.AnnData(data_d)
data_m.index = metadata.Index_Cell.map(str)
data_m.obs = metadata
adata

AnnData object with n_obs × n_vars = 2544 × 58347
  obs: 'ACCUMULATION_LEVEL', 'ALIGNED_READS', 'AT_DROPOUT', 'Aligned 0 time', 'Aligned 1 time', 'Aligned >1 times', 'Aligned concordantly 1 time', 'Aligned concordantly >1 times', 'Aligned concordantly or discordantly 0 t', 'Aligned discordantly 1 time', 'BAD_CYCLES.FIRST_OF_PAIR', 'BAD_CYCLES.PAIR', 'BAD_CYCLES.SECOND_OF_PAIR', 'CODING_BASES', 'CORRECT_STRAND_READS', 'ESTIMATED_LIBRARY_SIZE', 'GC_DROPOUT', 'GC_NC_0_19', 'GC_NC_20_39', 'GC_NC_40_59', 'GC_NC_60_79', 'GC_NC_80_100', 'IGNORED_READS', 'INCORRECT_STRAND_READS', 'INTERGENIC_BASES', 'INTRONIC_BASES', 'MAX_INSERT_SIZE', 'MEAN_INSERT_SIZE', 'MEAN_READ_LENGTH.FIRST_OF_PAIR', 'MEAN_READ_LENGTH.PAIR', 'MEAN_READ_LENGTH.SECOND_OF_PAIR', 'MEDIAN_3PRIME_BIAS', 'MEDIAN_5PRIME_BIAS', 'MEDIAN_5PRIME_TO_3PRIME_BIAS', 'MEDIAN_ABSOLUTE_DEVIATION', 'MEDIAN_CV_COVERAGE', 'MEDIAN_INSERT_SIZE', 'MIN_INSERT_SIZE', 'NUM_R1_TRANSCRIPT_STRAND_READS', 'NUM_R2_TRANSCRIPT_STRAND_READS', 'NUM_UNEXPLAINED_READS', 'Overall alignment rate', 'PAIR_ORIENTATION', 'PCT_ADAPTER.FIRST_OF_PAIR', 'PCT_ADAPTER.PAIR', 'PCT_ADAPTER.SECOND_OF_PAIR', 'PCT_CHIMERAS.FIRST_OF_PAIR', 'PCT_CHIMERAS.PAIR', 'PCT_CHIMERAS.SECOND_OF_PAIR', 'PCT_CODING_BASES', 'PCT_CORRECT_STRAND_READS', 'PCT_INTERGENIC_BASES', 'PCT_INTRONIC_BASES', 'PCT_MRNA_BASES', 'PCT_PF_READS.FIRST_OF_PAIR', 'PCT_PF_READS.PAIR', 'PCT_PF_READS.SECOND_OF_PAIR', 'PCT_PF_READS_ALIGNED.FIRST_OF_PAIR', 'PCT_PF_READS_ALIGNED.PAIR', 'PCT_PF_READS_ALIGNED.SECOND_OF_PAIR', 'PCT_PF_READS_IMPROPER_PAIRS.FIRST_OF_PA', 'PCT_PF_READS_IMPROPER_PAIRS.PAIR', 'PCT_PF_READS_IMPROPER_PAIRS.SECOND_OF_PA', 'PCT_R1_TRANSCRIPT_STRAND_READS', 'PCT_R2_TRANSCRIPT_STRAND_READS', 'PCT_READS_ALIGNED_IN_PAIRS.FIRST_OF_PAIR', 'PCT_READS_ALIGNED_IN_PAIRS.PAIR', 'PCT_READS_ALIGNED_IN_PAIRS.SECOND_OF_PA', 'PCT_RIBOSOMAL_BASES', 'PCT_USABLE_BASES', 'PCT_UTR_BASES', 'PERCENT_DUPLICATION', 'PF_ALIGNED_BASES', 'PF_ALIGNED_BASES.FIRST_OF_PAIR', 'PF_ALIGNED_BASES.PAIR', 'PF_ALIGNED_BASES.SECOND_OF_PAIR', 'PF_BASES', 'PF_HQ_ALIGNED_BASES.FIRST_OF_PAIR', 'PF_HQ_ALIGNED_BASES.PAIR', 'PF_HQ_ALIGNED_BASES.SECOND_OF_PAIR', 'PF_HQ_ALIGNED_Q20_BASES.FIRST_OF_PAIR', 'PF_HQ_ALIGNED_Q20_BASES.PAIR', 'PF_HQ_ALIGNED_Q20_BASES.SECOND_OF_PAIR', 'PF_HQ_ALIGNED_READS.FIRST_OF_PAIR', 'PF_HQ_ALIGNED_READS.PAIR', 'PF_HQ_ALIGNED_READS.SECOND_OF_PAIR', 'PF_HQ_ERROR_RATE.FIRST_OF_PAIR', 'PF_HQ_ERROR_RATE.PAIR', 'PF_HQ_ERROR_RATE.SECOND_OF_PAIR', 'PF_HQ_MEDIAN_MISMATCHES.FIRST_OF_PAIR', 'PF_HQ_MEDIAN_MISMATCHES.PAIR', 'PF_HQ_MEDIAN_MISMATCHES.SECOND_OF_PAIR', 'PF_INDEL_RATE.FIRST_OF_PAIR', 'PF_INDEL_RATE.PAIR', 'PF_INDEL_RATE.SECOND_OF_PAIR', 'PF_MISMATCH_RATE.FIRST_OF_PAIR', 'PF_MISMATCH_RATE.PAIR', 'PF_MISMATCH_RATE.SECOND_OF_PAIR', 'PF_NOISE_READS.FIRST_OF_PAIR', 'PF_NOISE_READS.PAIR', 'PF_NOISE_READS.SECOND_OF_PAIR', 'PF_READS.FIRST_OF_PAIR', 'PF_READS.PAIR', 'PF_READS.SECOND_OF_PAIR', 'PF_READS_ALIGNED.FIRST_OF_PAIR', 'PF_READS_ALIGNED.PAIR', 'PF_READS_ALIGNED.SECOND_OF_PAIR', 'PF_READS_IMPROPER_PAIRS.FIRST_OF_PAIR', 'PF_READS_IMPROPER_PAIRS.PAIR', 'PF_READS_IMPROPER_PAIRS.SECOND_OF_PAIR', 'READS_ALIGNED_IN_PAIRS.FIRST_OF_PAIR', 'READS_ALIGNED_IN_PAIRS.PAIR', 'READS_ALIGNED_IN_PAIRS.SECOND_OF_PAIR', 'READS_USED', 'READ_PAIRS', 'READ_PAIRS_EXAMINED', 'READ_PAIR_DUPLICATES', 'READ_PAIR_OPTICAL_DUPLICATES', 'RIBOSOMAL_BASES', 'SECONDARY_OR_SUPPLEMENTARY_RDS', 'STANDARD_DEVIATION', 'STRAND_BALANCE.FIRST_OF_PAIR', 'STRAND_BALANCE.PAIR', 'STRAND_BALANCE.SECOND_OF_PAIR', 'TOTAL_CLUSTERS', 'TOTAL_READS.FIRST_OF_PAIR', 'TOTAL_READS.PAIR', 'TOTAL_READS.SECOND_OF_PAIR', 'Total pairs', 'Total unpaired reads', 'UNMAAPPED_READS', 'UNPAIRED_READS_EXAMINED', 'UNPAIRED_READ_DUPLICATES', 'UTR_BASES', 'WIDTH_OF_10_PERCENT', 'WIDTH_OF_20_PERCENT', 'WIDTH_OF_30_PERCENT', 'WIDTH_OF_40_PERCENT', 'WIDTH_OF_50_PERCENT', 'WIDTH_OF_60_PERCENT', 'WIDTH_OF_70_PERCENT', 'WIDTH_OF_80_PERCENT', 'WIDTH_OF_90_PERCENT', 'WIDTH_OF_99_PERCENT', 'WINDOW_SIZE', 'alignable reads', 'filtered reads', 'multiple mapped', 'strand', 'total alignments', 'total reads', 'unalignable reads', 'uncertain reads', 'unique aligned'
```

Удалить NaN

```
data_d3 = data_d.fillna(0)
```

Показать скрытые выходные данные

```
data_m.nunique()
```

```

↗ Gene          570
  Index_Cell    570
    organism      1
      id         570
    Cell_type     7
    radius_mean  455
    texture_mean 479
    perimeter_mean 521
      area_mean  538
    smoothness_mean 473
    compactness_mean 536
    concavity_mean 538
      Cluster     8
    concave points_mean 541
    symmetry_mean  431
    fractal_dimension_mean 499
      radius_se   539
    texture_se     518
    perimeter_se   532
      area_se     527
    smoothness_se  546
    compactness_se 542
    concavity_se   532
    concave points_se 506
    symmetry_se    497
    fractal_dimension_se 544
      radius_worst 456
    texture_worst  510
    perimeter_worst 513
      area_worst   543
    smoothness_worst 410
    compactness_worst 528
    concavity_worst 538
    concave points_worst 492
    symmetry_worst  499
    fractal_dimension_worst 535
dtype: int64

```

```
data_m.dtypes
```

```

↗ Gene          object
  Index_Cell    object
    organism      object
      id         int64
    Cell_type     object
    radius_mean  float64
    texture_mean float64
    perimeter_mean float64
      area_mean  float64
    smoothness_mean float64
    compactness_mean float64
    concavity_mean float64
      Cluster     int64
    concave points_mean float64
    symmetry_mean  float64
    fractal_dimension_mean float64
      radius_se   float64
    texture_se     float64
    perimeter_se   float64
      area_se     float64
    smoothness_se  float64
    compactness_se float64
    concavity_se   float64
    concave points_se float64
    symmetry_se    float64
    fractal_dimension_se float64
      radius_worst float64
    texture_worst  float64
    perimeter_worst float64
      area_worst   float64
    smoothness_worst float64
    compactness_worst float64
    concavity_worst float64
    concave points_worst float64
    symmetry_worst  float64
    fractal_dimension_worst float64
dtype: object

```

```

adata = sc.AnnData(data_m)
adata.obs = data_m
adata

```

```

↗ AnnData object with n_obs × n_vars = 570 × 36
  obs: 'Gene', 'Index_Cell', 'organism', 'id', 'Cell_type', 'radius_mean', 'texture_mean', 'perimeter_mean', 'area_mean',

```

```
'smoothness_mean', 'compactness_mean', 'concavity_mean', 'Cluster', 'concave points_mean', 'symmetry_mean',
'fractal_dimension_mean', 'radius_se', 'texture_se', 'perimeter_se', 'area_se', 'smoothness_se', 'compactness_se', 'concavity_se',
'concave points_se', 'symmetry_se', 'fractal_dimension_se', 'radius_worst', 'texture_worst', 'perimeter_worst', 'area_worst',
'smoothness_worst', 'compactness_worst', 'concavity_worst', 'concave points_worst', 'symmetry_worst', 'fractal_dimension_worst'
```

```
data_m['Cell_type'].replace('PBMC2_1', 'PBMC2', inplace=True)
data_m['Cell_type'].replace('PBMC1_1', 'PBMC1', inplace=True)
data_m['Cell_type'].replace('PBMC3_1', 'PBMC3', inplace=True)
```

```
adata.obs['Cell_type'].value_counts()
```

```
Cell_type
Basal cells      204
Unknown          135
Ductal cells     64
Cholangiocytes   48
malignant        43
Fibroblasts      39
benign           37
Name: count, dtype: int64
```

```
#Выбираем те клетки, которые нам нужны для дальнейшего анализа
adata = adata[adata.obs['Cell_type'].isin(['Basal cells',
'Unknown',
'malignant',
'Ductal cells'
])]
```

 [Показать скрытые выходные данные](#)

```
adata.obs['Cell_type'].value_counts()
```

```
Cell_type
Basal cells      204
Unknown          135
Ductal cells     64
Cholangiocytes   48
malignant        43
Fibroblasts      39
benign           37
Name: count, dtype: int64
```

```
pip install -U scikit-learn
```

```
Requirement already satisfied: scikit-learn in /usr/local/lib/python3.10/dist-packages (1.5.0)
Requirement already satisfied: numpy>=1.19.5 in /usr/local/lib/python3.10/dist-packages (from scikit-learn) (1.25.2)
Requirement already satisfied: scipy>=1.6.0 in /usr/local/lib/python3.10/dist-packages (from scikit-learn) (1.11.4)
Requirement already satisfied: joblib>=1.2.0 in /usr/local/lib/python3.10/dist-packages (from scikit-learn) (1.4.2)
Requirement already satisfied: threadpoolctl>=3.1.0 in /usr/local/lib/python3.10/dist-packages (from scikit-learn) (3.5.0)
```

```
#Нормализация данных
adata.obs['n_counts'] = adata.X.sum(1)
adata.obs['log_counts'] = np.log(adata.X.sum(1))
adata.obs['n_genes'] = (adata.X > 0).sum(1)
#с помощью машинного обучения X_normalized = preprocessing.normalize(X, norm='l2')
```

 [Показать скрытые выходные данные](#)

После данного этапа можно строить различные типа графиков

## ✓ Фильтруем и нормализуем данные

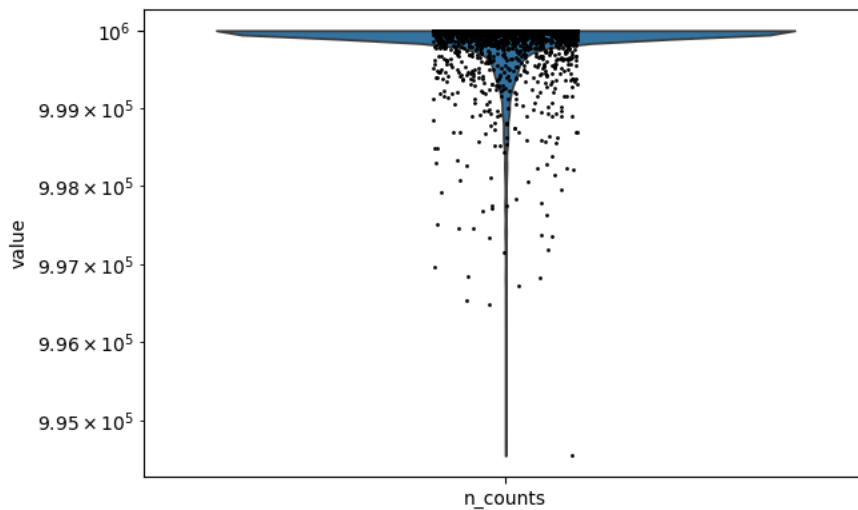
для данных формата csv и loom

```
sc.pp.filter_genes(adata, min_cells=2)
```

```
sc.pp.filter_cells(adata, min_genes=100)
```

```
adata.obs['n_counts'] = np.sum(adata.X, axis=1).A1
```

```
sc.pl.violin(adata, 'n_counts', size=2, log=True, cut=0)
```



```
dat = sc.pp.normalize_per_cell(adata, counts_per_cell_after=1e4, copy=True)
dat
```



```
AnnData object with n_obs x n_vars = 2544 x 37562
obs: 'ACCUMULATION_LEVEL', 'ALIGNED_READS', 'AT_DROPOUT', 'Aligned 0 time', 'Aligned 1 time', 'Aligned >1 times', 'Aligned
concordantly 1 time', 'Aligned concordantly >1 times', 'Aligned concordantly or discordantly 0 t', 'Aligned discordantly 1 time',
'BAD_CYCLES.FIRST_OF_PAIR', 'BAD_CYCLES.PAIR', 'BAD_CYCLES.SECOND_OF_PAIR', 'CODING_BASES', 'CORRECT_STRAND_READS',
'ESTIMATED_LIBRARY_SIZE', 'GC_DROPOUT', 'GC_NC_0_19', 'GC_NC_20_39', 'GC_NC_40_59', 'GC_NC_60_79', 'GC_NC_80_100', 'IGNORED_READS',
'INCORRECT_STRAND_READS', 'INTERGENIC_BASES', 'INTRONIC_BASES', 'MAX_INSERT_SIZE', 'MEAN_INSERT_SIZE',
'MEAN_READ_LENGTH.FIRST_OF_PAIR', 'MEAN_READ_LENGTH.PAIR', 'MEAN_READ_LENGTH.SECOND_OF_PAIR', 'MEDIAN_3PRIME_BIAS',
'MEDIAN_5PRIME_BIAS', 'MEDIAN_5PRIME_TO_3PRIME_BIAS', 'MEDIAN_ABSOLUTE_DEVIATION', 'MEDIAN_CV_COVERAGE', 'MEDIAN_INSERT_SIZE',
'MIN_INSERT_SIZE', 'NUM_R1_TRANSCRIPT_STRAND_READS', 'NUM_R2_TRANSCRIPT_STRAND_READS', 'NUM_UNEXPLAINED_READS', 'Overall alignment
rate', 'PAIR_ORIENTATION', 'PCT_ADAPTER.FIRST_OF_PAIR', 'PCT_ADAPTER.PAIR', 'PCT_ADAPTER.SECOND_OF_PAIR',
'PCT_CHIMERAS.FIRST_OF_PAIR', 'PCT_CHIMERAS.PAIR', 'PCT_CHIMERAS.SECOND_OF_PAIR', 'PCT_CODING_BASES', 'PCT_CORRECT_STRAND_READS',
'PCT_INTERGENIC_BASES', 'PCT_INTRONIC_BASES', 'PCT_MRNA_BASES', 'PCT_PF_READS.FIRST_OF_PAIR', 'PCT_PF_READS.PAIR',
'PCT_PF_READS.SECOND_OF_PAIR', 'PCT_PF_READS_ALIGNED.FIRST_OF_PAIR', 'PCT_PF_READS_ALIGNED.PAIR',
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'PCT_PF_READS_IMPROPER_PAIRS.SECOND_OF_PA', 'PCT_R1_TRANSCRIPT_STRAND_READS', 'PCT_R2_TRANSCRIPT_STRAND_READS',
'PCT_READS_ALIGNED_IN_PAIRS.FIRST_OF_PAIR', 'PCT_READS_ALIGNED_IN_PAIRS.PAIR', 'PCT_READS_ALIGNED_IN_PAIRS.SECOND_OF_PAIR',
'PCT_RIBOSOMAL_BASES', 'PCT_USABLE_BASES', 'PCT_UTR_BASES', 'PERCENT_DUPLICATION', 'PF_ALIGNED_BASES',
'PF_ALIGNED_BASES.FIRST_OF_PAIR', 'PF_ALIGNED_BASES.PAIR', 'PF_ALIGNED_BASES.SECOND_OF_PAIR', 'PF_BASES',
'PF_HQ_ALIGNED_BASES.FIRST_OF_PAIR', 'PF_HQ_ALIGNED_BASES.PAIR', 'PF_HQ_ALIGNED_BASES.SECOND_OF_PAIR',
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'PF_HQ_ALIGNED_READS.FIRST_OF_PAIR', 'PF_HQ_ALIGNED_READS.PAIR', 'PF_HQ_ALIGNED_READS.SECOND_OF_PAIR',
'PF_HQ_ERROR_RATE.FIRST_OF_PAIR', 'PF_HQ_ERROR_RATE.PAIR', 'PF_HQ_ERROR_RATE.SECOND_OF_PAIR',
'PF_HQ_MEDIAN_MISMATCHES.FIRST_OF_PAIR', 'PF_HQ_MEDIAN_MISMATCHES.PAIR', 'PF_HQ_MEDIAN_MISMATCHES.SECOND_OF_PAIR',
'PF_INDEL_RATE.FIRST_OF_PAIR', 'PF_INDEL_RATE.PAIR', 'PF_INDEL_RATE.SECOND_OF_PAIR', 'PF_MISMATCH_RATE.FIRST_OF_PAIR',
'PF_MISMATCH_RATE.PAIR', 'PF_MISMATCH_RATE.SECOND_OF_PAIR', 'PF_NOISE_READS.FIRST_OF_PAIR', 'PF_NOISE_READS.PAIR',
'PF_NOISE_READS.SECOND_OF_PAIR', 'PF_READS.FIRST_OF_PAIR', 'PF_READS.PAIR', 'PF_READS.SECOND_OF_PAIR',
'PF_READS_ALIGNED.FIRST_OF_PAIR', 'PF_READS_ALIGNED.PAIR', 'PF_READS_ALIGNED.SECOND_OF_PAIR',
'PF_READS_IMPROPER_PAIRS.FIRST_OF_PAIR', 'PF_READS_IMPROPER_PAIRS.PAIR', 'PF_READS_IMPROPER_PAIRS.SECOND_OF_PAIR',
'READS_ALIGNED_IN_PAIRS.FIRST_OF_PAIR', 'READS_ALIGNED_IN_PAIRS.PAIR', 'READS_ALIGNED_IN_PAIRS.SECOND_OF_PAIR', 'READS_USED',
'READ_PAIRS', 'READ_PAIRS_EXAMINED', 'READ_PAIR_DUPLICATES', 'READ_PAIR_OPTICAL_DUPLICATES', 'RIBOSOMAL_BASES',
'SECONDARY_OR_SUPPLEMENTARY_RDS', 'STANDARD_DEVIATION', 'STRAND_BALANCE.FIRST_OF_PAIR', 'STRAND_BALANCE.PAIR',
'STRAND_BALANCE.SECOND_OF_PAIR', 'TOTAL_CLUSTERS', 'TOTAL_READS.FIRST_OF_PAIR', 'TOTAL_READS.PAIR', 'TOTAL_READS.SECOND_OF_PAIR',
'Total pairs', 'Total unpaired reads', 'UNMAPPED_READS', 'UNPAIRED_READS_EXAMINED', 'UNPAIRED_READ_DUPLICATES', 'UTR_BASES',
'WIDTH_OF_10_PERCENT', 'WIDTH_OF_20_PERCENT', 'WIDTH_OF_30_PERCENT', 'WIDTH_OF_40_PERCENT', 'WIDTH_OF_50_PERCENT',
'WIDTH_OF_60_PERCENT', 'WIDTH_OF_70_PERCENT', 'WIDTH_OF_80_PERCENT', 'WIDTH_OF_90_PERCENT', 'WIDTH_OF_99_PERCENT', 'WINDOW_SIZE',
'alignable reads', 'filtered reads', 'multiple mapped', 'strand', 'total alignments', 'total reads', 'unalignable reads',
'uncertain reads', 'unique aligned', 'n_genes', 'n_counts'
var: 'n_cells'
```

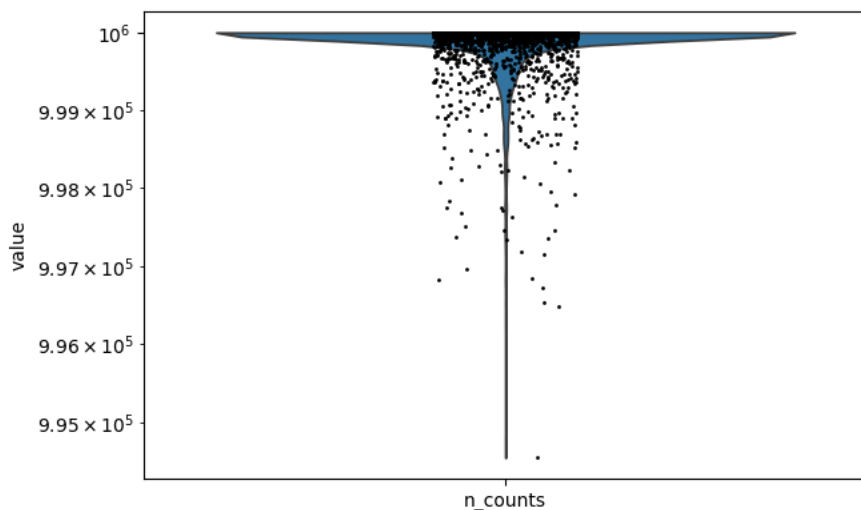
```
#Пример нормализации используя машинное обучение
#X_normalized = preprocessing.normalize(X, norm='l2')
```

```
#Выбираем часть генов в клетке (первую 500 по экспрессии генов )
data_T = []
for i in range(dat.shape[0]):
    indices = np.asarray(np.argsort(dat.X[i,:].todense())[0,:]).flatten()[::-1][0:500]
    data_T = np.union1d(data_T, indices)
```

```
adata = adata[:, data_T.astype('int')]
```

```
sc.pl.violin(adata, 'n_counts', size=2, log=True, cut=0)
```





Подготовка данных к кластеризации

```
sc.pp.pca(adata, svd_solver='arpack')
sc.pp.neighbors(adata)
```

```
sc.tl.tsne(adata)
sc.tl.umap(adata)
```

```
print(adata)
```

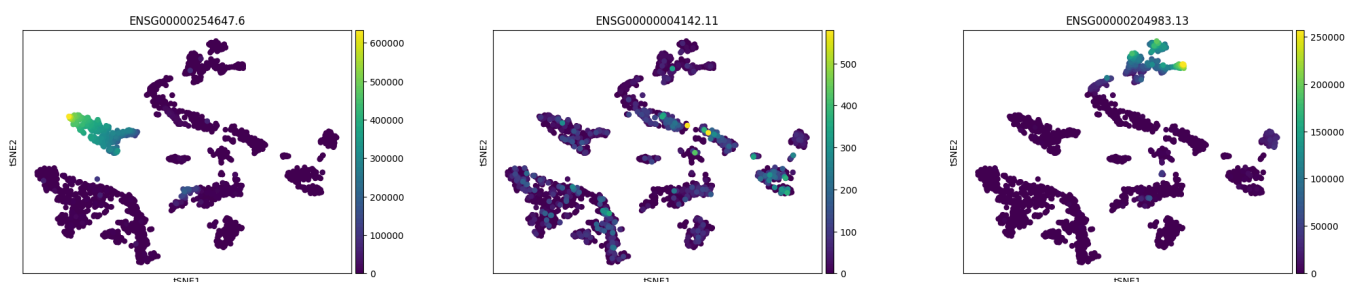
```
AnnData object with n_obs x n_vars = 2544 x 18968
  obs: 'ACCUMULATION_LEVEL', 'ALIGNED_READS', 'AT_DROPOUT', 'Aligned 0 time', 'Aligned 1 time', 'Aligned >1 times', 'Aligned concn'
  var: 'n_cells'
  uns: 'pca', 'neighbors', 'tsne', 'umap'
  obsm: 'X_pca', 'X_tsne', 'X_umap'
  varm: 'PCs'
  obsp: 'distances', 'connectivities'
```

## Графики

plot UMAP

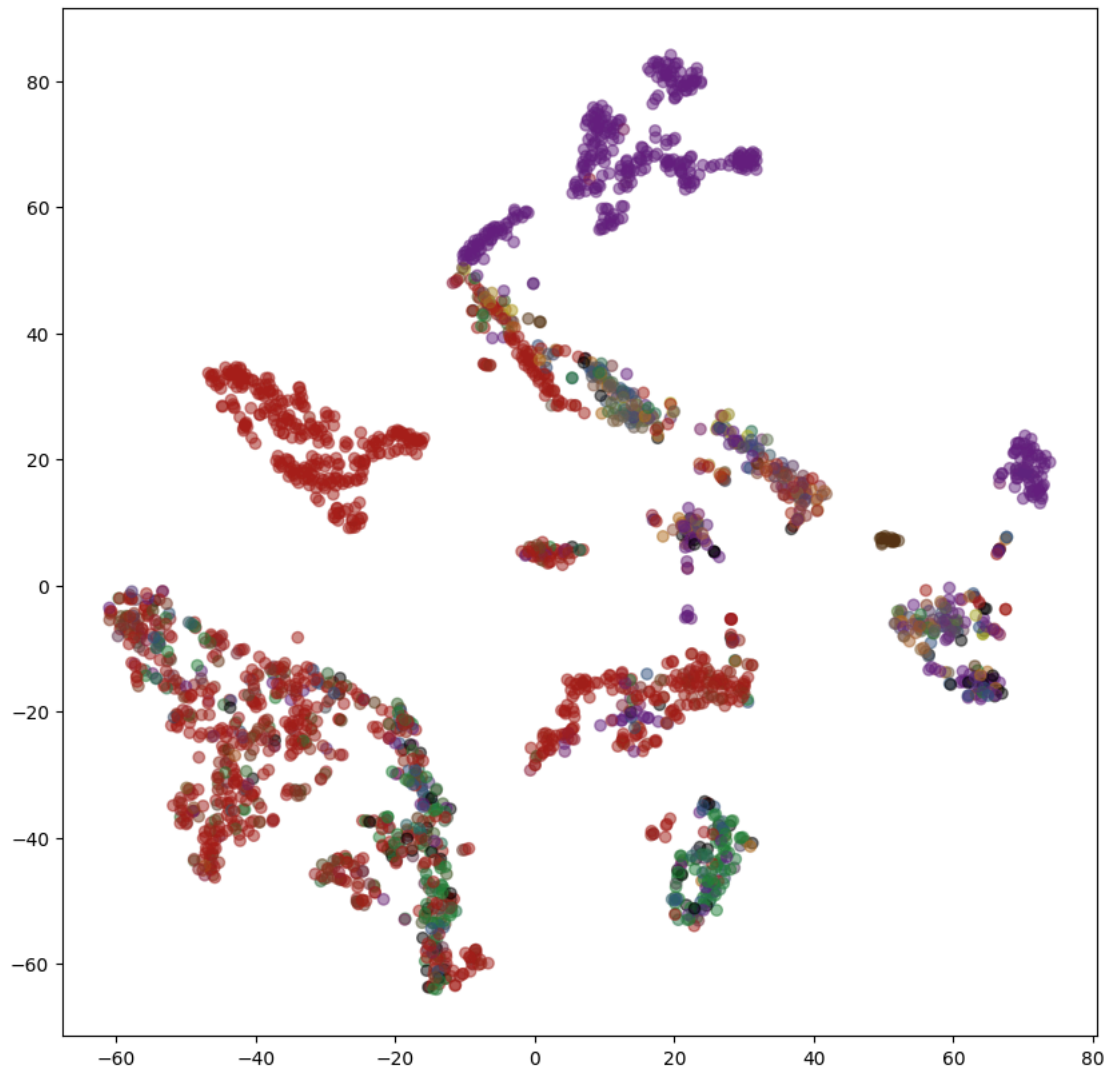
```
genes = ['ENSG00000254647.6', 'ENSG0000004142.11', 'ENSG0000003147.17', 'ENSG00000204983.13', '#TP53 #POLDIP2 #ICA1 #BRCA2'
         'ENSG0000007062.11', 'ENSG00000154096.13', 'ENSG00000108849.7'] #CHEK2 #THY1 #PTEN
colors = np.asarray([[226,43,34], [67,114,175], [48,178,78],
                    [139,47,175], [244,152,46], [249,246,49], [119,73,27]])
weights = adata[:,genes].X.todense()
weights = weights / (weights.sum(axis=1) + 0.0001)
c = np.dot(weights, colors)
```

```
sc.pl.tsne(adata, color=['ENSG00000254647.6', 'ENSG0000004142.11', 'ENSG00000204983.13'],
          wspace=0.3, size=150, linewidths=1, edgecolors='black');
```



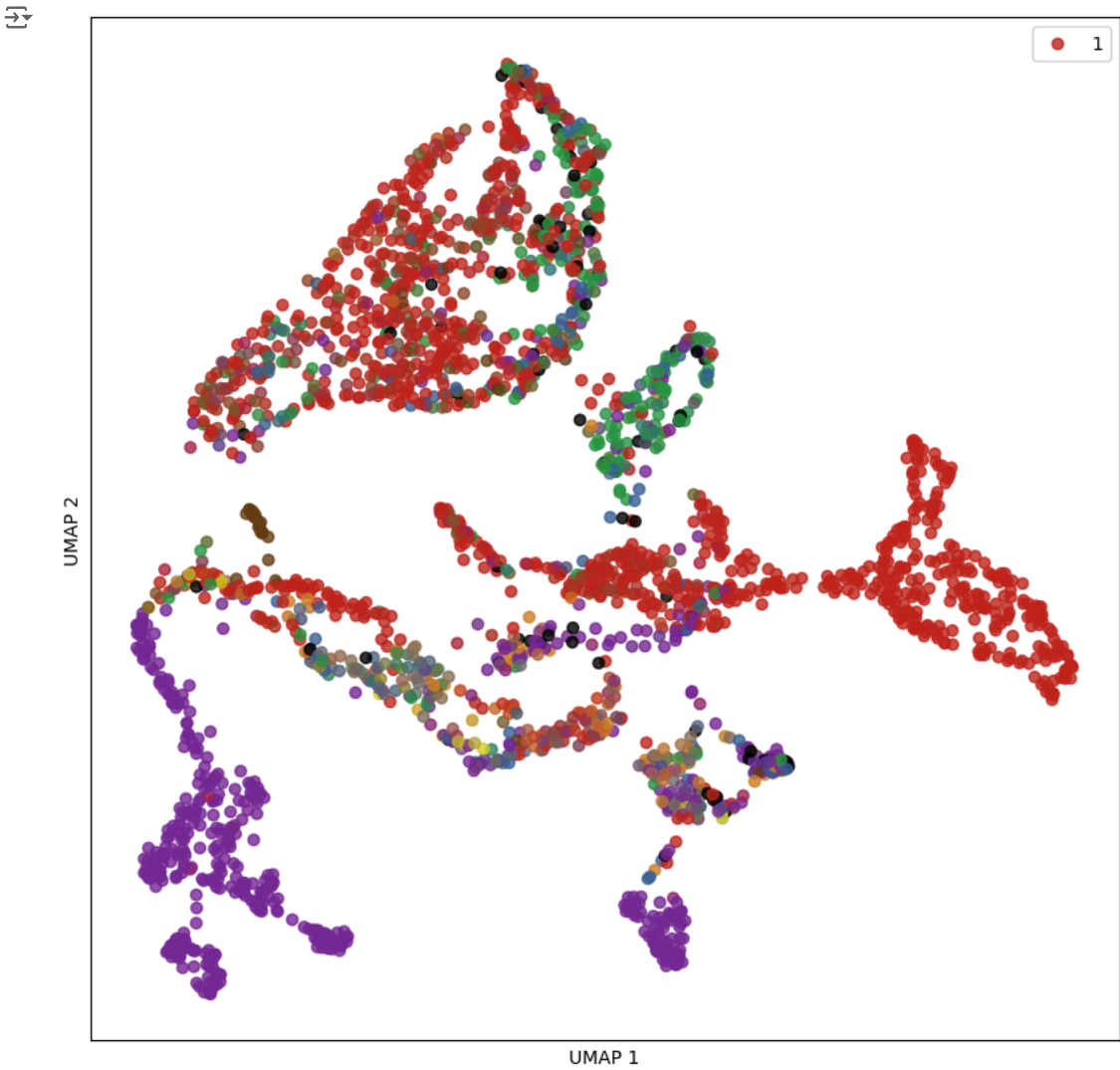
```
plt.figure(figsize=[10, 10])
```

```
plt.scatter(adata.obs['X_tsne'][:,0], adata.obs['X_tsne'][:,1], c=c/350, alpha=0.5);
#Клетки окрашены за счет экспрессии избранных генов
```



```
plt.figure(figsize=[10, 10])
plt.scatter(adata.obs['X_umap'][:,0], adata.obs['X_umap'][:,1], c=c/300, alpha=0.8);
plt.grid(False)
plt.yticks([])
plt.xticks([])
plt.xlabel('UMAP 1')
plt.ylabel('UMAP 2');

plt.legend(['CHEK2'])
plt.show()
```



У UMAP Вариант 2

```
warnings.filterwarnings("ignore", message=r"Passing", category=FutureWarning)
```

```
from sklearn.preprocessing import LabelEncoder
le = LabelEncoder()
```

```
X, y = data_d.drop("Index_Cell", axis=1), data_d[["Index_Cell"]].values.flatten()
X.head().sample(10, axis=1)
```

	CDH1.6	PBMC3.14	BRCA1.47	BRCA2.20	CDH1.4	CDH1.14	BRCA2.21	BRCA1.20	BRCA1.27	BRCA2.24
0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
1	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
2	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
3	0.0	0.0	0.0	3.0	0.0	0.0	0.0	0.0	0.0	0.0
4	0.0	7.0	0.0	0.0	1.0	0.0	0.0	0.0	0.0	0.0

```
X.shape
(588, 255)
```

```
y= le.fit_transform(y.astype(str))
```

```
y
Показать скрытые выходные данные
```

```
np.unique(y)
```

[Показать скрытые выходные данные](#)

```
import umap
from sklearn.preprocessing import PowerTransformer
from sklearn.pipeline import make_pipeline
```

```
# Scale
#pipe = make_pipeline(PowerTransformer())
#X = pipe.fit_transform(X.copy())
```

```
# Encode the target to numeric
y_encoded = pd.factorize(y)[0]
```

```
X = np.nan_to_num(X)
```

```
y_encoded = np.nan_to_num(y_encoded)
```

```
manifold = umap.UMAP().fit(X, y_encoded)
```

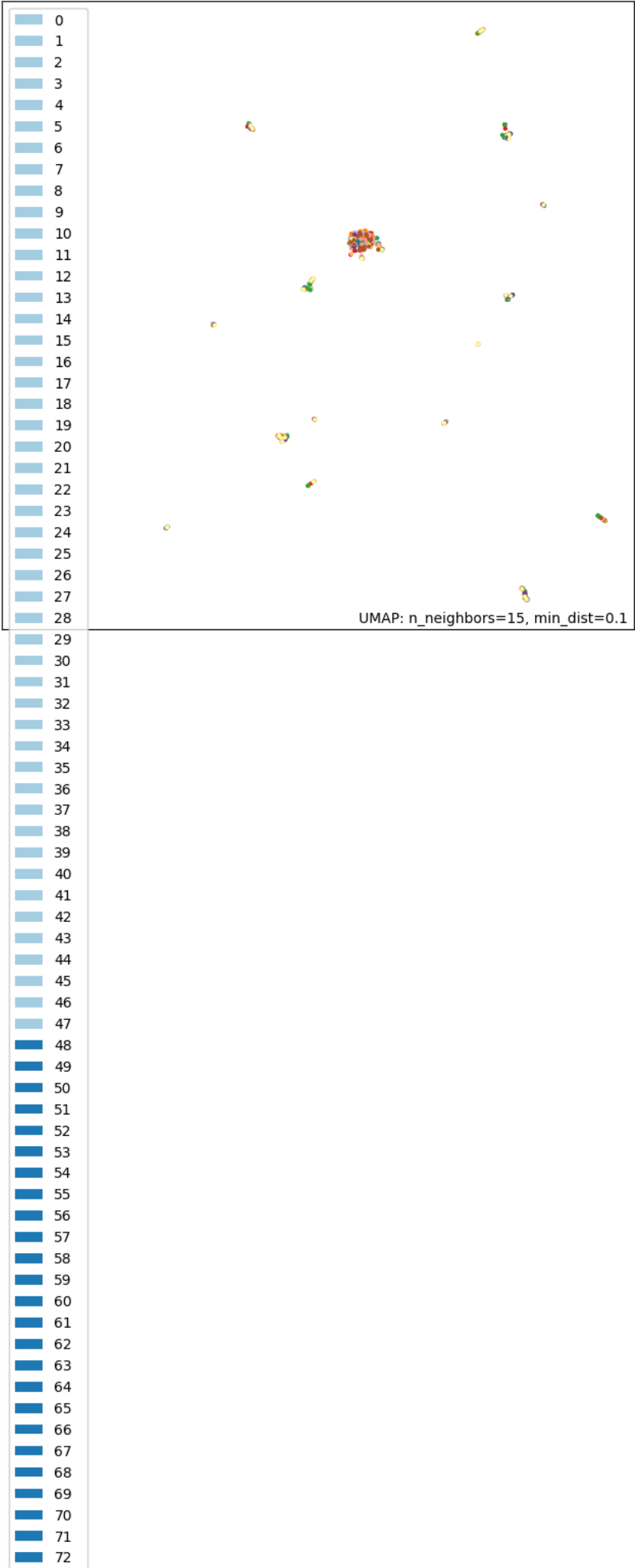
```
pip install umap-learn[plot]
```

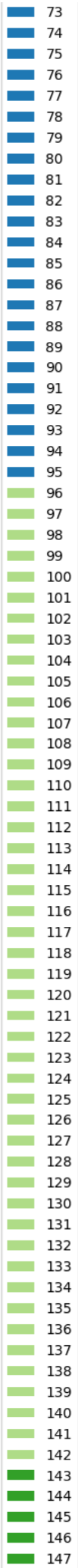
[Показать скрытые выходные данные](#)

```
import umap.plot
```

```
umap.plot.points(manifold, labels=y, color_key_cmap='Paired', background='white')
```

<Axes: >





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