

Experimental Design and Data Analysis, Lecture 5

Eduard Belitser

VU Amsterdam

Lecture Overview

- 1 two-way ANOVA
- 2 general factorial design
- 3 randomized block design
- 4 repeated measures

two way analysis of variance
(completely randomized design)

Setting

An experiment with:

- a numerical outcome Y ;
- two factors (categorical variables) that can be fixed at I and J levels (categories), respectively.

EXAMPLE Agricultural experiment with outcome total yield from a plot and factors type of fertilizer and crop variety.

EXAMPLE Quality of a genetic algorithm to determine the minimal value of a criterion function with outcome CPU time needed to find true minimum and factors mutation probability and population size.

EXAMPLE Outcome time to develop mold on bread and factors temperature and humidity.

Design

- Select NIJ experimental units randomly from the population of interest.
- Assign combined levels (i, j) of the factors to a random set of N units.
- Independently perform the NIJ experiments.

Randomization in R:

```
> I=4; J=2; N=3
> rbind(rep(1:I,each=N*J),rep(1:J,N*I),sample(1:(N*I*J)))
```

	[,1]	[,2]	[,3]	[,4]	[,5]	[,6]	[,7]	[,8]	[,9]	[,10]	[,11]	[,12]	[,13]
[1,]	1	1	1	1	1	1	2	2	2	2	2	2	3
[2,]	1	2	1	2	1	2	1	2	1	2	1	2	1
[3,]	20	1	3	14	17	24	19	12	22	13	16	15	4

	[,14]	[,15]	[,16]	[,17]	[,18]	[,19]	[,20]	[,21]	[,22]	[,23]	[,24]
[1,]	3	3	3	3	3	4	4	4	4	4	4
[2,]	2	1	2	1	2	1	2	1	2	1	2
[3,]	23	8	10	2	7	21	9	5	6	18	11

For unit 20 use levels (1,1) of (factor 1, factor 2); for unit 1 use levels (1,2);
...; for unit 11 use levels (4,2).

Two-way ANOVA model — data

Data

sample (i, j) : $Y_{ij1}, Y_{ij2}, \dots, Y_{ijn_{ij}}, \quad i = 1, \dots, I; \quad j = 1, \dots, J.$

Assume that these are sampled independently from IJ **normal** populations with (possibly different) **population means** μ_{ij} , and with **equal population variances**.

Commonly, one considers **balanced design**: $n_{ij} = N$ for all subgroups (i, j) .

We want to **test** the following null hypotheses:

- no interaction between the two factors A and B,
- no main effect of the first factor A,
- no main effect of the second factor B.

The **overall** nullhypothesis $H_0 : \mu_{ij} = \mu_{kl}$ for every i, j, k, l is of modest interest.

We also estimate the corresponding parameters.

Two-way ANOVA model — assumptions

The **two-way ANOVA** model is:

$$Y_{ijk} = \mu_{ij} + e_{ijk} = \mu + \alpha_i + \beta_j + \gamma_{ij} + e_{ijk}, \quad i = 1, \dots, I, \quad j = 1, \dots, J, \quad k = 1, \dots, n_{ij}.$$

Assumption: the indep. errors $e_{ijk} \sim N(0, \sigma^2)$, with unknown variance σ^2 .

We decomposed the (i, j) -group means as $\mu_{ij} = \mu + \alpha_i + \beta_j + \gamma_{ij}$, where

- μ is the **overall mean**,
- α_i is the **main effect** of level i of the **first factor A**,
- β_j is the **main effect** of level j of the **second factor B**,
- γ_{ij} is the **interaction effect** of levels i and j of the first and second factors.

Now we can formalize the hypothesis to test:

- $H_{AB} : \gamma_{ij} = 0$ for every (i, j) (no interactions between factor A and B),
- $H_A : \alpha_i = 0$ for every i (no main effect of factor A),
- $H_B : \beta_j = 0$ for every j (no main effect of factor B).

For the parameters to be identifiable, we need to impose $I + J + 1$ linear restrictions, (done by command **contrasts** in R). The default in R is the **treatment** parametrization:

$\alpha_1 = \beta_1 = \gamma_{1j} = \gamma_{i1} = 0, j = 1, \dots, J, i = 1, \dots, I$. Often one uses the **sum** parametrization: $\sum_i \alpha_i = 0, \sum_j \beta_j = 0, \sum_i \gamma_{ij} = 0$ for all $j = 1, \dots, J$, and $\sum_j \gamma_{ij} = 0$ for all $i = 1, \dots, I$.

Tests in two-way ANOVA

Setting: a two-way ANOVA model: $Y_{ijk} = \mu + \alpha_i + \beta_j + \gamma_{ij} + e_{ijk}$.

Hypotheses: we want to test H_{AB} , H_A , H_B against their negations.

Test statistics: F_{AB} for testing H_{AB} , F_A for testing H_A , and F_B for testing H_B .

Distribution of F 's under H_{AB} , H_A , H_B : $F_{AB} \sim F_{(I-1)(J-1), n-IJ}$, $F_A \sim F_{I-1, n-IJ}$, $F_B \sim F_{J-1, n-IJ}$. $F_{m,k}$ is the **F-distribution** with m and k degrees of freedom.

Test: larger values of $F_{AB} = f_{AB}$ give **more evidence against H_{AB}** , hence we reject H_{AB} if F_{AB} is large. The test is therefore **always right-sided**: compare the p -value $p_{right} = P(F > f_{AB})$ with a significance level α . Similarly for F_A , F_B .

In R: the p -value is in `anova(lm(y~f1*f2))`, with `f1` and `f2` the two factors.

Balanced design: equal group size $n_{ij} = N$ for each i and j , thus $n = NIJ$.

Formula `y~f1*f2` is the same as `y~f1+f2+f1:f2`, meaning that the model includes μ (μ is always included by default), and all α_i 's, β_j 's and γ_{ij} 's.

If H_{AB} is not rejected (i.e., we concluded that all $\gamma_{ij} = 0$), then it is **proper practice** to test for main effects A and B under the **additive model** $\mu_{ij} = \mu + \alpha_i + \beta_j$ (in R: `y~f1+f2`). Otherwise, we proceed to test for main effects using the full model.

F-statistics in two-way ANOVA

The idea of the F -statistics is $F = \frac{\text{explained variance}}{\text{unexplained variance}} = \frac{\text{between-groups SS}}{\text{within-groups SS}}$.

Denote the total mean $\bar{Y}_{...} = \frac{1}{I} \sum_{i=1}^I \frac{1}{J} \sum_{j=1}^J \frac{1}{n_{ij}} \sum_{k=1}^{n_{ij}} Y_{ijk}$, and

$$\bar{Y}_{ij\cdot} = \frac{1}{n_{ij}} \sum_{k=1}^{n_{ij}} Y_{ijk}, \quad \bar{Y}_{i..} = \frac{1}{J} \sum_{j=1}^J \frac{1}{n_{ij}} \sum_{k=1}^{n_{ij}} Y_{ijk}, \quad \bar{Y}_{\cdot j\cdot} = \frac{1}{I} \sum_{i=1}^I \frac{1}{n_{ij}} \sum_{k=1}^{n_{ij}} Y_{ijk}.$$

The test statistics are

$$F_{AB} = \frac{\sum_{i=1}^I \sum_{j=1}^J n_{ij} (\bar{Y}_{ij\cdot} - \bar{Y}_{i..} - \bar{Y}_{\cdot j\cdot} + \bar{Y}_{...})^2 / ((I-1)(J-1))}{\sum_{i=1}^I \sum_{j=1}^J \sum_{k=1}^{n_{ij}} (Y_{ijk} - \bar{Y}_{ij\cdot})^2 / (n - IJ)},$$

$$F_A = \frac{\sum_{i=1}^I \sum_{j=1}^J n_{ij} (\bar{Y}_{i..} - \bar{Y}_{...})^2 / (I-1)}{\sum_{i=1}^I \sum_{j=1}^J \sum_{k=1}^{n_{ij}} (Y_{ijk} - \bar{Y}_{ij\cdot})^2 / (n - IJ)},$$

similarly for F_B .

General form of ANOVA tables

One-way ANOVA results are usually presented in an one-way [ANOVA table](#):

Source	Df	Sum Sq	Mean Sq	F value	p-value
Factor A	$I - 1$	SS_A	$SS_A / (I - 1)$	$F_A = \frac{SS_A / (I - 1)}{RSS / (n - I)}$	$P_A(F_A > f)$
Residuals	$n - I$	RSS	$RSS / (n - I)$		
Total	$n - 1$	SS_T			

Two-way ANOVA results are usually presented in a two-way [ANOVA table](#):

Source	Df	Sum Sq	Mean Sq	F value	p-value
Factor A	$I - 1$	SS_A	$SS_A / (I - 1)$	$F_A = \frac{SS_A / (I - 1)}{RSS / (n - IJ)}$	$P_A(F_A > f)$
Factor B	$J - 1$	SS_B	$SS_B / (J - 1)$		
Interaction	$(I - 1)(J - 1)$	SS_{AB}	$SS_{AB} / ((I - 1)(J - 1))$	$F_{AB} = \frac{SS_{AB} / ((I - 1)(J - 1))}{RSS / (n - IJ)}$	$P_{AB}(F_{AB} > f)$
Residuals	$n - IJ$	RSS	$RSS / (n - IJ)$		
Total	$n - 1$	SS_T			

$$SS_T = SS_A + SS_B + SS_{AB} + RSS = \sum_{i=1}^I \sum_{j=1}^J \sum_{k=1}^{n_{ij}} (y_{ijk} - \bar{y}_{...})^2.$$

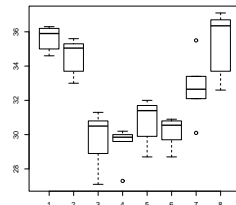
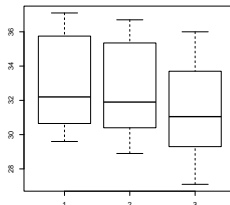
Example — pvc (1)

The following data is from an experiment to study factors affecting the production of the plastic PVC, 3 operators used 8 different devices called resin railcars to produce PVC, two samples for each of the 24 combinations.

```
> pvc=read.table(file="pvc.txt",header=TRUE)
> pvc[1:4,]
  psize operator resin
1  36.2         1     1
2  36.3         1     1
3  35.3         1     2
4  35.0         1     2
```

```
> attach(pvc)
> boxplot(psize~operator)
> boxplot(psize~resin)
```

These pictures give an idea of the main effects of the factors. Interactions are not visible.

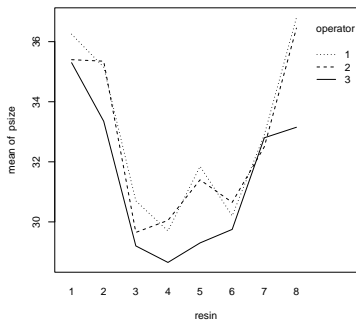
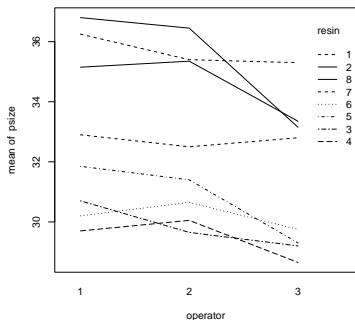


Example — pvc (2)

An **interaction plot** fixes one factor and plots the average outcome (vertical axis) against the levels of the other factor (horizontal axis). Interaction shows up as nonparallel curves.

```
> interaction.plot(operator, resin, psize)
```

```
> interaction.plot(resin, operator, psize)
```



Lines may be unparallel, because of interactions, but also because of noise in the data.

Example — pvc (3)

```
> pvc$operator=as.factor(pvc$operator); pvc$resin=as.factor(pvc$resin)
> pvcaov=lm(psize~operator*resin); anova(pvcaov)
[ some output deleted ]
```

Response: psize

	Df	Sum Sq	Mean Sq	F value	Pr(>F)	
operator	2	20.718	10.359	7.0072	0.00401	**
resin	7	283.946	40.564	27.4388	5.661e-10	***
operator:resin	14	14.335	1.024	0.6926	0.75987	
Residuals	24	35.480	1.478			

The p -value for testing $H_0 : \alpha_i = 0$ for all i is 0.00401; for $H_0 : \beta_j = 0$ for all j is 5.661e-10; for $H_0 : \gamma_{i,j} = 0$ for all (i,j) is 0.75987. So, there is no evidence for interaction (both factors seems to have a main effect but one should not draw conclusions about the factors at this stage).

The command `as.factor` (or `factor`) is necessary, because the 2nd and 3rd columns of the data matrix were read in as numerical variables (with values 1, 2, 3, 4), but should be treated as factors in the analysis.

Example — pvc (4)

```
> summary(pvcaov) # estimates in the default treatment contrasts
[ some output deleted ]
Coefficients:
```

	Estimate	Std. Error	t value	Pr(> t)
(Intercept)	36.2500	0.8598	42.164	< 2e-16 ***
operator2	-0.8500	1.2159	-0.699	0.491216
operator3	-0.9500	1.2159	-0.781	0.442245
resin2	-1.1000	1.2159	-0.905	0.374615
[some output deleted]				
resin8	0.5500	1.2159	0.452	0.655078
operator2:resin2	1.0500	1.7195	0.611	0.547175
[some output deleted]				
operator3:resin8	-2.7000	1.7195	-1.570	0.129454

The output shows estimates of $\mu, \alpha_2, \alpha_3, \beta_2, \beta_2, \dots, \beta_8, \gamma_{22}, \dots, \gamma_{38}$ in the default **treatment** parametrization: $\alpha_1 = \beta_1 = \gamma_{1j} = \gamma_{i1} = 0, i = 1, 2, 3, j = 1 \dots, 8$. The corresponding estimates $\hat{\alpha}_1 = \hat{\beta}_1 = \hat{\gamma}_{11} = \dots = \hat{\gamma}_{31} = 0$ are not shown. The p -values are for testing the null hypothesis that the coefficient is 0.

Example — pvc (5)

The command `contrasts` overrules the default `treatment` parametrization (e.g., to `sum` parameterization), `lm` and `anova` have to be run again.

```
> contrasts(pvc$operator)=contr.sum; contrasts(pvc$resin)=contr.sum
> pvcaov2=lm(psize~operator*resin,data=pvc); summary(pvcaov2)
[ some output deleted ]
```

Coefficients:

	Estimate	Std. Error	t value	Pr(> t)
(Intercept)	32.35417	0.17550	184.359	< 2e-16 ***
operator1	0.58958	0.24819	2.376	0.025855 *
operator2	0.32708	0.24819	1.318	0.199983
resin1	3.29583	0.46432	7.098	2.45e-07 ***
[some output deleted]				
resin7	0.37917	0.46432	0.817	0.422183
operator1:resin1	0.01042	0.65664	0.016	0.987474
[some output deleted]				
operator2:resin7	-0.56042	0.65664	-0.853	0.401844

The output shows estimates of $\mu, \alpha_1, \alpha_2, \beta_1, \beta_2, \dots, \beta_7, \gamma_{11}, \gamma_{12}, \dots$ in the `sum` parametrization. The estimates of α_3 (for operator 3) and β_8 (for resin 8) are not shown. These can be found from the restrictions $\sum_{i=1}^3 \hat{\alpha}_i = 0$, $\sum_{j=1}^8 \hat{\beta}_j = 0$; similarly for the interactions: $\sum_{i=1}^3 \hat{\gamma}_{ij} = 0$ for $j = 1, \dots, 8$ and $\sum_{j=1}^8 \hat{\gamma}_{ij} = 0$ for $i = 1, 2, 3$.

Example — pvc (6)

As we see, the previous analysis says there are no interactions. Now we remove interaction term from the model and fit the [additive model](#)

$$\mu_{ij} = \mu + \alpha_i + \beta_j, \quad i = 1, \dots, I, \quad j = 1, \dots, J.$$

```
> pvc$operator=as.factor(pvc$operator)
> pvc$resin=as.factor(pvc$resin)
> pvcaov=lm(psize~operator+resin,data=pvc)
> anova(pvcaov)
```

[some output deleted]

Response: psize

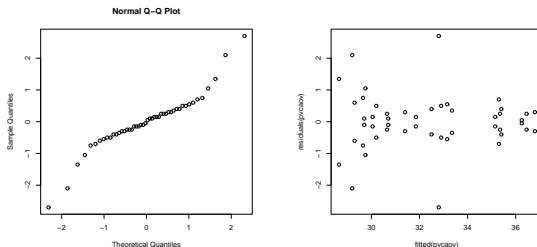
	Df	Sum Sq	Mean Sq	F value	Pr(>F)
operator	2	20.718	10.359	7.902	0.00135 **
resin	7	283.946	40.564	30.943	8.111e-14 ***
Residuals	38	49.815	1.311		

The p -value for testing $H_A : \alpha_i = 0$ for all i is 0.00135; for $H_B : \beta_j = 0$ for all j is $8.111e - 14$. So both factors have a main effect in the additive model as well.

Example — pvc (7)

We check the normality and the assumption of equal variances. The **residuals** $\hat{e}_{ijn} = Y_{ijn} - \hat{\mu} - \hat{\alpha}_i - \hat{\beta}_j - \hat{\gamma}_{ij}$ are the data corrected for the different population means and ought to look normal. The **fitted value** for Y_{ijn} is the estimated mean $\hat{\mu}_{ij} = \hat{\mu} + \hat{\alpha}_i + \hat{\beta}_j + \hat{\gamma}_{ij}$. The spread in the residuals should not change systematically with any variable, in particular not with the fitted values.

```
> qqnorm(residuals(pvcaov2)); plot(fitted(pvcaov2),residuals(pvcaov2))
```



Left plot: normality is doubtful. Right plot: the spread in the residuals seems to be bigger for smaller fitted values. Some data-points also seem extreme. Perhaps transform the data or consider **outliers**.

One observation per cell (1)

The following dataset contains the strength of a thermoplastic composite depending on power of a laser and speed of a tape.

```
> composite=read.table("composite.txt",head=T); composite
```

	strength	laser	tape
1	25.66	40W	slow
2	29.15	50W	slow
3	35.73	60W	slow
4	28.00	40W	medium
5	35.09	50W	medium
6	39.56	60W	medium
7	20.65	40W	fast
8	29.79	50W	fast
9	35.66	60W	fast

Notice that we have only one observation per cell (i.e., per each combination of levels of the two factors laser and tape). o estimate and test interaction effects, it is necessary to have at least 2 observations per combination (i, j) of factor levels.

One observation per cell (2)

R produces a warning message if the data is not sufficient to fit the model, in this case it is impossible to estimate interactions with one observation per cell:

```
> attach(composite); anova(lm(strength~laser*tape))
```

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
laser	2	224.184	112.092		
tape	2	48.919	24.459		
laser:tape	4	10.503	2.626		
Residuals	0	0.000			

Warning message:

```
In anova.lm(lm(strength ~ laser * tape, data = composite)) :
```

ANOVA F-tests on an essentially perfect fit are unreliable

If it can be assumed a priori that all interactions are 0, then it is possible to test and estimate main effects. (Interaction plots may help to justify this assumption.)

```
> anova(lm(strength~laser+tape,data=composite))
```

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
laser	2	224.184	112.092	42.6893	0.002003 **
tape	2	48.919	24.459	9.3151	0.031242 *
Residuals	4	10.503	2.626		

general factorial design

General factorial design

- Everything extends to an arbitrary number of factors.
- A practical difficulty is that the number of combinations of factors increases rapidly, so that many experiments are necessary.
- In the decomposition of the population means this becomes visible through many **interaction parameters**. E.g., given 3 factors there are 3 2nd order and 1 3rd order interactions:

$$\mu_{ijk} = \mu + \alpha_i + \beta_j + \gamma_k + (\alpha\beta)_{ij} + (\alpha\gamma)_{ik} + (\beta\gamma)_{jk} + (\alpha\beta\gamma)_{ijk}.$$

- It is often assumed that higher order interactions are zero. Then lower order interactions can be estimated using fewer experiments (**incomplete designs**).

randomized block design

Setting

An experiment with:

- a **numerical outcome** Y (“dependent variable”),
- a **factor** of interest that can be fixed at I levels (“**treatment**”),
- a **factor** that is *not* of interest that can be fixed at B levels (“**block**”).

The purpose is to understand the dependence of Y on the **treatment factor**. The **block variable** is thought (or known) to be of influence. It is used to create homogeneous groups of experimental units, in which the treatment effect is easier to see and not blurred by variation due to the block factor.

EXAMPLE Chemical production process with outcome **total yield**, treatment variable **temperature** fixed at levels low, medium and high and block **blend of raw material**.

EXAMPLE Study of web design with outcome **total time of a user on webpage**, treatment variable **type of design** and block **user skill**. Each user is tested with a single type of web design.

Design

Independently, for $b = 1, 2, \dots, B$:

- select N experimental units randomly from the population of units with block level b ,
- assign level i of the factor to a random set of N units ($i = 1, 2, \dots, I$),
- perform the experiment N times, independently.

Randomization in R.

```
> I=4; B=5; N=1
> for (i in 1:B) print(sample(1:(N*I)))
[1] 3 1 2 4
[1] 4 3 2 1
[1] 1 4 2 3
[1] 3 4 1 2
[1] 2 4 3 1
```

For block 1 assign unit 3 to treatment 1, unit 1 to treatment 2, etc., for block 2 assign unit 4 to treatment 1, unit 3 to treatment 2, etc.

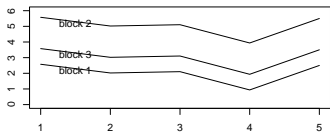
Given many blocks, it is typical to use one replicate per treatment level per block:
 $N = 1$.

Analysis

Data (Y_{ibn}) are assumed to follow the model

$$Y_{ibn} = \mu + \alpha_i + \beta_b + e_{ibn}, \quad i = 1, \dots, I; \quad b = 1, \dots, B; \quad n = 1, \dots, N,$$

where the “errors” (e_{ibn}) are a random sample from a **normal** population.



The pattern ($\alpha_1, \alpha_2, \dots, \alpha_I$) of treatment effects is assumed to be **the same within every block**.

We **test** the null hypothesis $H_0 : \alpha_1 = \alpha_2 = \dots = \alpha_I = 0$.

We also **estimate** the treatment effects $\alpha_1, \alpha_2, \dots, \alpha_I$.

The model is the same as in a two-way factorial experiment, with the block as a second factor, but with zero interactions.

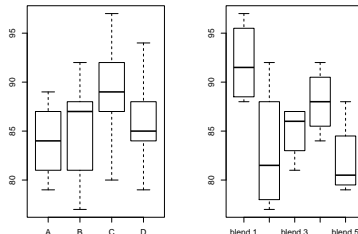
Analysis in R — data input

The following data frame contains the data about penicillin made by production processes A, B, C, D (treatment); with 5 different blends of raw material (blocks), as in a two-way factorial experiment.

```
> penicillin
  treat  blend yield
1      A blend.1   89
2      B blend.1   88
3      C blend.1   97
4      D blend.1   94
5      A blend.2   84
[ some output deleted ]
20     D blend.5   88
> xtabs(yield~treat+blend,data=penicillin)
      blend
treat blend.1 blend.2 blend.3 blend.4 blend.5
  A         89      84      81      87      79
  B         88      77      87      92      81
  C         97      92      87      89      80
  D         94      79      85      84      88
```

Analysis in R — graphics

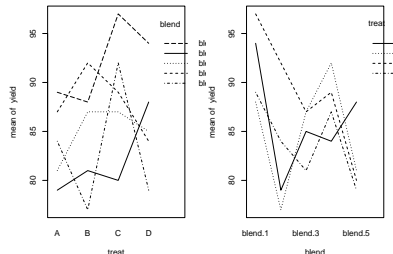
```
> attach(penicillin)
> par(mfrow=c(1,2))
> boxplot(yield~treat)
> boxplot(yield~blend)
```



```
> par(mfrow=c(1,2))
> interaction.plot(treat,blend,yield)
> interaction.plot(blend,treat,yield)
```

The left plot gives estimates of the treatment patterns

$\beta_b + (\alpha_1, \alpha_2, \dots, \alpha_I)$, per block.



Analysis in R — testing and estimation

```
> aovpen=lm(yield~treat+blend)
> anova(aovpen)
Response: yield
```

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
treat	3	70	23.333	1.2389	0.33866
blend	4	264	66.000	3.5044	0.04075 *
Residuals	12	226	18.833		

The treatment effects are not significantly different from 0. The blocks (blend) are, but this was not the research question.

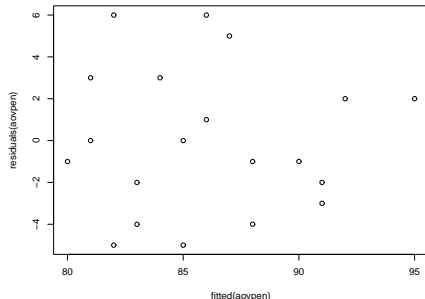
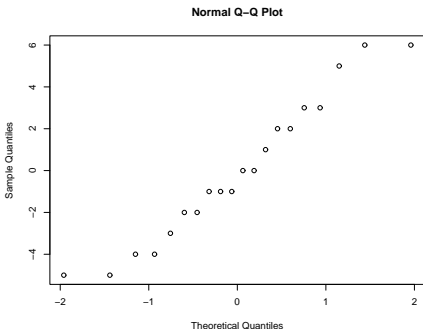
```
> summary(aovpen)
[ some output deleted ]
Coefficients:
```

	Estimate	Std. Error	t value	Pr(> t)
(Intercept)	90.000	2.745	32.791	4.1e-13 ***
treatB	1.000	2.745	0.364	0.72194
treatC	5.000	2.745	1.822	0.09351 .
treatD	2.000	2.745	0.729	0.48018
blendblend.2	-9.000	3.069	-2.933	0.01254 *
blendblend.3	-7.000	3.069	-2.281	0.04159 *
blendblend.4	-4.000	3.069	-1.304	0.21686
blendblend.5	-10.000	3.069	-3.259	0.00684 **

The yield of treatment C is estimated 5 higher than that of treatment A, etc.

Analysis in R — diagnostics

```
> qqnorm(residuals(aovpen))  
> plot(fitted(aovpen),residuals(aovpen))
```



Look OK; perhaps a slight curve in the qq-plot. The interaction plots (see some slides back) can also be considered diagnostic.

Discussion

- The **advantage** of the block design is that more precise conclusions can be obtained by removing variation, present due to block factor. The units must be **similar within the blocks**, and **dissimilar between the blocks**.
- Assuming that *the pattern of treatment effects is the same for each block* means assuming the absence of **interaction** between block and treatment. Without replications ($N = 1$), this cannot be tested. Given $N > 1$ measurements (**replicates**) per combination of block and treatment, the absence of **interaction** between block and treatment can be tested.
- If treatment and blocks do interact, the interpretation of the results of a factorial analysis is more subtle.
- **Multiple treatment factors**: a multi-way factorial experiment can be done within every block (rather than a one factor experiment).
- **Multiple block factors**: all combinations of levels of the block factors can be viewed as a new, single block factor, to which the block design applies.

repeated measures

Setting and design

Setting: an experiment with

- a **numerical outcome** Y (“dependent variable”),
- a **factor** of interest that can be fixed at I levels, (“treatment”).
- **experimental units** that are measured at **every** treatment level.

The purpose is to understand the dependence of Y on the **treatment factor**.

The same **experimental units** are used for every treatment, because this is thought to reduce “extraneous variation”: the units serve as blocks.

For $I = 2$ treatments, this is simply the paired sample design.

EXAMPLE Study of web design with outcome **total time on webpage**, treatment variable **type of design**. Each **user** is tested with every type of design.

EXAMPLE The **velocity** of a ball is measured for **different types of tennis rackets** for a number of **players**, where every player uses all types of rackets.

Design:

- Select B experimental units randomly from a population of units.
- Measure each unit at every treatment level, if possible in random order.

Analysis — exchangeable case

Data vectors $(Y_{1b}, Y_{2b}, \dots, Y_{Ib})$ for B units are assumed to follow the model

$$Y_{ib} = \mu + \alpha_i + \beta_b + e_{ib}, \quad i = 1, \dots, I; \quad b = 1, \dots, B,$$

- the “error vectors” (e_{1b}, \dots, e_{Ib}) for the B units are a random sample from a (multivariate) normal distribution.
- the “errors” e_{1b}, \dots, e_{Ib} within a single unit are **exchangeable** (the ordering is irrelevant, in a way, generalizing the paired samples).
- the effects β_1, \dots, β_B of the units may be considered fixed or random.

The pattern $(\alpha_1, \dots, \alpha_I)$ of treatments is assumed to be **the same for each unit**.

We want to **test** the null hypothesis $H_0 : \alpha_1 = \alpha_2 = \dots = \alpha_I = 0$. We also **estimate** the treatment effects $\alpha_1, \alpha_2, \dots, \alpha_I$.

The model is the same as in a randomized block experiment, with the units as blocks, except for the assumption on the errors. These are allowed to be **dependent** within the units, even though still “exchangeable”.

Analysis in R — data input

Data input is as in a block design, with columns for outcome, treatment level, and block level (=identification of unit).

```
> ashinalong
  pain id order treatment
1  -167  1    pa         a
2  -102  1    pa         p
3  -127  2    pa         a
4   -39  2    pa         p
5   -58  3    pa         a
6   32  3    pa         p
7  -103  4    pa         a
8   28  4    pa         p
[ some output deleted ]
31  -72 16    ap         a
32  -36 16    ap         p
```

The data frame `ashinalong` contains the same data as `ashina`, but every individual is represented by two lines, one for the treatment with the active drug, the other for the placebo. The extra column `id` shows the pairing of the measurements.

Analysis in R — exchangeable case

Analysis is as for a randomized block design, with every unit being a block.

```
> ashinalong$id=factor(ashinalong$id)
> aovashina=lm(pain~treatment+id,data=ashinalong); anova(aovashina)
```

Analysis of Variance Table

Response: pain

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
treatment	1	14706	14706.1	10.413	0.005644 **
id	15	51137	3409.2	2.414	0.049184 *
Residuals	15	21184	1412.3		

Compare to the two sample *t*-test:

```
> t.test(ashina[,1],ashina[,2],paired=TRUE)
```

Paired t-test

data: ashina[, 1] and ashina[, 2]

t = -3.2269, df = 15, p-value = 0.005644

The p-value for treatment is identical to the one of the paired-sample t-test found previously (the order of the treatments was ignored). The p-value for id is not interesting. Note that R had to be told to treat id as labels, not as numbers.

Lack of exchangeability

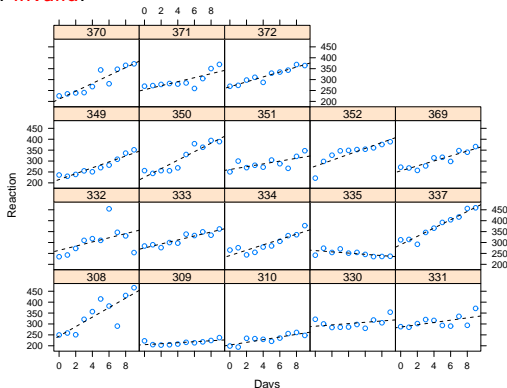
Repeated measures may **not** be exchangeable.

- Time effect: growth, increasing or decreasing variation.
- Learning effect: subject becomes better or bored at tasks (cf. [crossover design](#)).
- Dissimilar subjects: the pattern of response to treatment varies too much.

A block design analysis is then **invalid**.

Example Truck drivers were deprived of sleep and their reaction time was measured daily over a period of 10 days. Each panel gives the reaction times of one truck driver with best fitting line overlaid.

Drivers follow **different** lines, which contradicts the model assumptions.



Discussion

- Taking repeated measures is attractive, because fewer experimental units are needed and “extraneous” variation between units is reduced.
- However, it introduces “dependencies” in the data, which may not be easy to model correctly and may lead to [artifacts](#) if not accounted for.
- The [repeated measures design](#) discussed here is a simple one. It corrects for some dependencies.
- However, in many studies, in particular most “longitudinal studies”, where individuals are followed over time, the assumption of “exchangeability” fails. More complicated models are then necessary.
- Models with [random effects](#) (also called [mixed effects models](#)) are a possibility.

To finish

Today we discussed:

- 1 2-way ANOVA
- 2 general factorial design
- 3 randomized block design
- 4 repeated measures

Next time: Friedman test, random effects, more block designs.