Hands-on training session 2

Hui-Walter models for diagnostic test evaluation

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Introduction

Overview

Date/time:

- 19th February 2020
- **1**6.00 17.00

Teachers:

- Matt Denwood (presenter)
- Giles Innocent

Recap

Important points from session $\boldsymbol{1}$

TODO

Session 2a: Hui-Walter models for 2

tests and 1 population

Hui-Walter Model

Background (not necessarily Bayesian)

Rabbits and hats

Model Specification

 $co[1] \sim dho + o(1 1)$

1

1 7

```
model{
      Tally ~ dmulti(prob, TotalTests)
2
      # Test1- Test2-
4
         prob[1] <- (prev * ((1-se[1])*(1-se[2]))) + ((1-prev) *
5
         \hookrightarrow ((sp[1])*(sp[2])))
6
       # Test1+ Test2-
7
         prob[2] <- (prev * ((se[1])*(1-se[2]))) + ((1-prev) *
8
         \hookrightarrow ((1-sp[1])*(sp[2])))
9
       # Test1- Test2+
10
         prob[3] <- (prev * ((1-se[1])*(se[2]))) + ((1-prev) *
11
         \hookrightarrow ((sp[1])*(1-sp[2])))
12
       # Test1+ Test2+
13
         prob[4] <- (prev * ((se[1])*(se[2]))) + ((1-prev) *
14
         \rightarrow ((1-sp[1])*(1-sp[2])))
15
      prev ~ dbeta(1, 1)
16
```

And run it:

Finished running the simulation

```
twoXtwo <- matrix(c(48, 12, 4, 36), ncol=2, nrow=2)
1
    twoXtwo
    ## [,1] [,2]
2 ## [1,] 48 4
3 ## [2,] 12 36
    library('runjags')
1
2
3
    Tally <- as.numeric(twoXtwo)</pre>
    TotalTests <- sum(Tally)</pre>
4
5
    prev <- list(chain1=0.05, chain2=0.95)</pre>
6
    se \leftarrow list(chain1=c(0.5,0.99), chain2=c(0.99,0.5))
7
    sp \leftarrow list(chain1=c(0.5,0.99), chain2=c(0.99,0.5))
8
9
    results <- run.jags('basic_hw.bug', n.chains=2)
10
    ## Loading required namespace: rjags
```

```
##
1
   ## JAGS model summary statistics from 20000 samples (chains = 2;
2
       adapt+burnin = 5000):
   ##
3
                                                  SD
   ## Lower95 Median Upper95 Mean
4
   ## prev 0.30463 0.43905 0.5673 0.43957 0.068498
5
   ## prob[1] 0.36893 0.46065 0.55862 0.46158 0.049011
6
   ## prob[2] 0.072141 0.13274 0.20048 0.13503 0.033383
7
   ## prob[3] 0.01705 0.054976 0.10335 0.058038 0.023171
8
   ## prob[4] 0.25229 0.34432 0.43441 0.34535 0.04675
9
   ## se[1] 0.82443 0.9326 0.99999 0.92407 0.05346
10
11 ## se[2] 0.69339 0.85114 0.99992 0.84739 0.090355
   ## sp[1] 0.7445 0.87349 0.99997 0.87109 0.074509
12
   ## sp[2] 0.85954 0.94725 0.99999 0.94012 0.042404
13
   ##
14
                 Mode MCerr MC%ofSD SSeff AC.10
15
   ##
   ## prev 0.43487 0.0010748 1.6 4062 0.031192
16
17
   ## prob[1] 0.45985 0.00041111 0.8 14213 -0.0059391
18
   ## prob[2] 0.12712 0.00027833
                                   0.8 14386 0.0063839
   ## prob[3] 0.052209 0.00024004 1 9318 -0.011454
19
   ## prob[4]
                0.344 0.00040718
                                   0.9 13182
                                             -0.007306
```

results

20

	Lower95	Median	Upper95	SSeff	psrf
prev	0.305	0.439	0.567	4062	1.000
prob[1]	0.369	0.461	0.559	14213	1.000
prob[2]	0.072	0.133	0.200	14386	1.000
prob[3]	0.017	0.055	0.103	9318	1.000
prob[4]	0.252	0.344	0.434	13182	1.000
se[1]	0.824	0.933	1.000	5875	1.001
se[2]	0.693	0.851	1.000	3455	1.001
sp[1]	0.744	0.873	1.000	3419	1.001
sp[2]	0.860	0.947	1.000	5478	1.000

Note wide confidence intervals

Practicalities

Care with order of combinations in dmultinom

Lots of data needed

And/or strong priors for one of the tests

Convergence can be tricky

Label Switching

sp[1] ~ dbeta(1, 1)

```
How to interpret a test with Se=0\% and Sp=0\%?
The test is perfect - we are just holding it upside down...
. . .
We can force se+sp >= 1:
  se[1] ~ dbeta(1, 1)
  sp[1] \sim dbeta(1, 1)T(1-se[1], )
. . .
Or:
  se[1] \sim dbeta(1, 1)T(1-sp[1], )
```

Simulating data

+ TIOX + TIO

12

Analysing simulated data is useful to check that we can recover parameter values.

Some simultion code:

```
se1 <- 0.9
    sp1 < -0.95
    sp2 < -0.99
    se2 < -0.8
    prevalence <- 0.5
    N < -100
6
7
    truestatus <- rbinom(N, 1, prevalence)</pre>
8
    Test1 <- rbinom(N, 1, (truestatus * se1) + ((1-truestatus) *
    \hookrightarrow (1-sp1)))
    Test2 <- rbinom(N, 1, (truestatus * se2) + ((1-truestatus) *</pre>
10
    11
    twoXtwo <- table(Test1, Test2)</pre>
12
```

Exercise

Modify JAGS code to force tests to be better than useless Simulate data and recover parameters for:

■ N=10, N=100, N=1000

Optional Exercise

Use priors for test1 taken from session 1 and compare the results

Solution

2

4

5

6

7

8

9

11

12

13

14

15

Model definition: model{ Tally ~ dmulti(prob, TotalTests) # Test1- Test2prob[1] <- (prev * ((1-se[1])*(1-se[2]))) + ((1-prev) * \rightarrow ((sp[1])*(sp[2]))) # Test1+ Test2prob[2] <- (prev * ((se[1])*(1-se[2]))) + ((1-prev) * $\hookrightarrow ((1-sp[1])*(sp[2])))$ # Test1- Test2+ prob[3] <- (prev * ((1-se[1])*(se[2]))) + ((1-prev) * $\hookrightarrow ((sp[1])*(1-sp[2])))$ # Test1+ Test2+ prob[4] <- (prev * ((se[1])*(se[2]))) + ((1-prev) * \rightarrow ((1-sp[1])*(1-sp[2])))

Optional Solution

```
HPSe[1,] \leftarrow c(148.43, 16.49)
   HPSp[1,] \leftarrow c(240.03, 12.63)
2
3
   HPSe
1 ## [,1] [,2]
2 ## [1,] 148.43 16.49
3 ## [2,] 1.00 1.00
   HPSp
1 ## [,1] [,2]
2 ## [1,] 240.03 12.63
   ## [2,] 1.00 1.00
   results <- run.jags('basic_hw.bug', n.chains=2)
```

Finished running the simulation

tests and N populations

Session 2b: Hui-Walter models for 2

Independent intercepts for populations

```
model{
2
      for(p in 1:Populations){
        Tally[1:4, p] ~ dmulti(prob[1:4, p], TotalTests[p])
        # Test1- Test2- Pop1
5
6
          prob[1, p] \leftarrow (prev[p] * ((1-se[1])*(1-se[2]))) +
    \rightarrow ((1-prev[p]) * ((sp[1])*(sp[2])))
7
        ## etc ##
8
9
        prev[p] ~ dbeta(1, 1)
10
11
12
      se[1] ~ dbeta(HPSe[1,1], HPSe[1,2])T(1-sp[1], )
13
      sp[1] ~ dbeta(HPSp[1,1], HPSp[1,2])
14
      se[2] ~ dbeta(HPSe[2,1], HPSe[2,2])T(1-sp[2], )
15
      sp[2] ~ dbeta(HPSp[2,1], HPSp[2,2])
16
17
18
      #data# Tally, TotalTests, Populations, HPSe, HPSp
19
      #monitor# prev, prob, se, sp
```

Auto Hui-Walter

hand (aimdata)

We would usually start with individual-level data in a dataframe e.g.:

```
se1 <- 0.9
    sp1 < -0.95
    sp2 < -0.99
3
    se2 <- 0.8
    prevalences <- c(0.1, 0.5, 0.9)
5
    N < -100
6
7
    simdata <- data.frame(Population = sample(seq_along(prevalences),</pre>
8

→ N, replace=TRUE))
    simdata$probability <- prevalences[simdata$Population]</pre>
9
    simdata$truestatus <- rbinom(N, 1, simdata$probability)</pre>
10
    simdata$Test1 <- rbinom(N, 1, (simdata$truestatus * se1) +</pre>
11
    \rightarrow ((1-simdata\$truestatus) * (1-sp1)))
    simdata$Test2 <- rbinom(N, 1, (simdata$truestatus * se2) +</pre>
12
    13
```

The model code and data format for an arbitrary number of populations (and tests) can be determined automatically

There is a function (soon to be included in the runjags package, but for now provided in the GitHub repo) that can do this for us:

- $_1$ $\mbox{\em \##}$ The model and data have been written to autohw.bug in the \hookrightarrow current working directory
- 2 ## You should check and alter priors before running the model

This generates self-contained model/data/initial values etc (ignore covse and covsp for now):

```
## ## Auto-generated Hui-Walter model created by script version
    \rightarrow 0.1 on 2020-02-12
    ##
2
    ## model{
3
    ##
4
        ## Observation layer:
    ##
5
    ##
6
    ##
        # Complete observations (N=100):
7
        for(p in 1:Populations){
    ##
8
             Tally_RR[1:4,p] ~ dmulti(prob_RR[1:4,p], N_RR[p])
9
    ##
10
    ##
             prob_RR[1:4,p] <- se_prob[1:4,p] + sp_prob[1:4,p]
    ##
11
    ##
       }
12
13
    ##
14
    ##
15
    ##
        ## Observation probabilities:
    ##
16
    ##
        for(p in 1:Populations){
17
    ##
18
             # Probability of observing Test1- Test2- from a true
    ##
19
```

And can be run directly from R:

```
results <- run.jags('autohw.bug')

## Note: The monitored variables 'covse12' and 'covsp12'

## appear to be non-stochastic; they will not be

## included in the convergence diagnostic

## Finished running the simulation
```

```
results
   ##
   ## JAGS model summary statistics from 20000 samples (chains = 2;
       adapt+burnin = 5000):
   ##
3
   ## Lower95 Median Upper95 Mean SD
                                                     Mode
4
   ## se[1] 0.71517 0.84356 0.96496 0.83979 0.064616 0.85345
5
   ## se[2] 0.65745 0.79572 0.9177 0.79285 0.066441 0.7994
6
   ## sp[1] 0.87605 0.96673 1 0.95596 0.039059 0.98493
   ## sp[2] 0.86925 0.95868 0.99998 0.94921 0.040251 0.98013
8
   ## prev[1] 0.072539 0.18993 0.3269 0.19543 0.066634 0.18175
9
   ## prev[2] 0.26969 0.46757 0.66678 0.4676 0.10224 0.47244
10
   ## prev[3] 0.68481 0.84386 0.98271 0.83654 0.078561 0.85803
11
   ## covse12
                   0
                          0
                                  0
12
   ## covsp12
                                                        0
13
   ##
14
                 MCerr MC%ofSD SSeff AC.10
15
   ##
                                               psrf
   ## se[1] 0.00075224
16
                          1.2 7378 0.0014165 1.0003
17
   ## se[2] 0.0007197 1.1 8522 0.014384 1.0001
18
   ## sp[1] 0.00058798 1.5 4413 0.025856 1.0001
   ## sp[2] 0.00054376 1.4 5479 -0.0065604
                                              1.0002
19
   ## prev[1]
              0.0006472
                          1 10600 0.0049727
                                               1.0003
```

20

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Observation-level model specification

for(n in 2. Pone) {

```
model{
1
 2
      for(i in 1:N){
         Status[i] ~ dcat(prob[i, ])
4
5
           prob[i,1] <- (prev[i] * ((1-se[1])*(1-se[2]))) +
6
                        ((1-prev[i]) * ((sp[1])*(sp[2])))
7
           prob[i,2] \leftarrow (prev[i] * ((se[1])*(1-se[2]))) +
8
                        ((1-prev[i]) * ((1-sp[1])*(sp[2])))
9
           prob[i,3] \leftarrow (prev[i] * ((1-se[1])*(se[2]))) +
10
                        ((1-prev[i]) * ((sp[1])*(1-sp[2])))
11
           prob[i,4] \leftarrow (prev[i] * ((se[1])*(se[2]))) +
12
                        ((1-prev[i]) * ((1-sp[1])*(1-sp[2])))
13
14
           logit(prev[i]) <- intercept +</pre>
15
           → population_effect[Population[i]]
      }
16
17
      intercept ~ dnorm(0, 0.33)
18
      population_effect[1] <- 0</pre>
19
```

Just like in session 1, the main difference is the prior for prevalence (this time in each population)

We also need to give initial values for intercept and population_effect rather than prev, and tell run.jags the data frame from which to extract the data (except N and Pops):

```
intercept <- list(chain1=-1, chain2=1)</pre>
population_effect <- list(chain1=c(NA, 1, -1), chain2=c(NA, -1,
```

1

→ 1)) $se \leftarrow list(chain1=c(0.5,0.99), chain2=c(0.99,0.5))$

```
3
   sp \leftarrow list(chain1=c(0.5,0.99), chain2=c(0.99,0.5))
4
5
   simdata$Status <- with(simdata, factor(interaction(Test1, Test2),</pre>
6
    → levels=c('0.0','1.0','0.1','1.1')))
   N <- nrow(simdata)</pre>
```

Pops <- length(levels(simdata\$Population))</pre> 8 glm_results <- run.jags('glm_hw.bug', n.chains=2, data=simdata)</pre> 9

Note: The monitored variable 'population_effect[1]' ## appears to be non-stochastic; it will not be included

in the convergence diagnostic

Also like in session 1, the estimates for se/sp should be similar, although this model runs more slowly.

Note: this model could be used as the basis for adding covariates

For a handy way to generate a GLM model see

runjags::template.jags

 Look out for integration with autohuiwalter in the near (ish) future...

Practicalities

Need to be very careful with tabulating the data, or use automatically generated code

Works best when populations have very different prevalences

Exercise

Play around with the autohuiwalter function

Notice the model and data and initial values are in a self contained file

Ignore the covse and covsp for now