

# Class 19: Investigating Pertussis Resurgence

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## Investigating Pertussis Cases By Year

B. Pertussis is a severe bacterial lung infection also known as whooping cough.

We will begin by investigating the number of Pertussis cases by year in the US.

This data is available on [this CDC page](#).

We'll use the addin DataPasta to import this data into R. It allows you to paste data you've copied into R as r objects like data frames and tables.

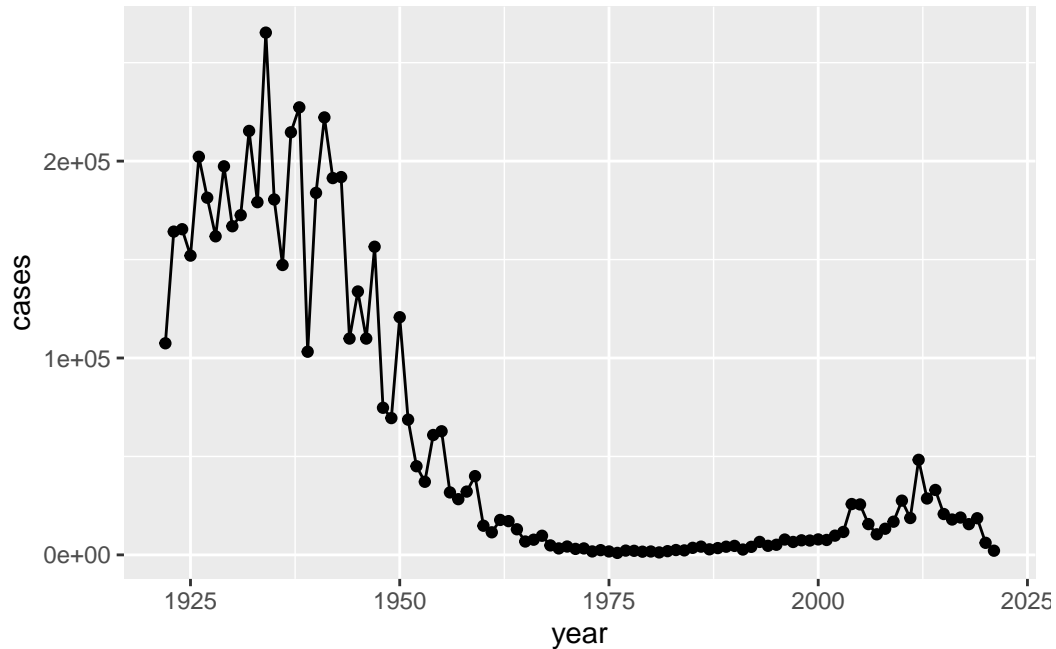
Let's have a quick look at this data.frame:

```
head(cdc)
```

```
  year  cases
1 1922 107473
2 1923 164191
3 1924 165418
4 1925 152003
5 1926 202210
6 1927 181411
```

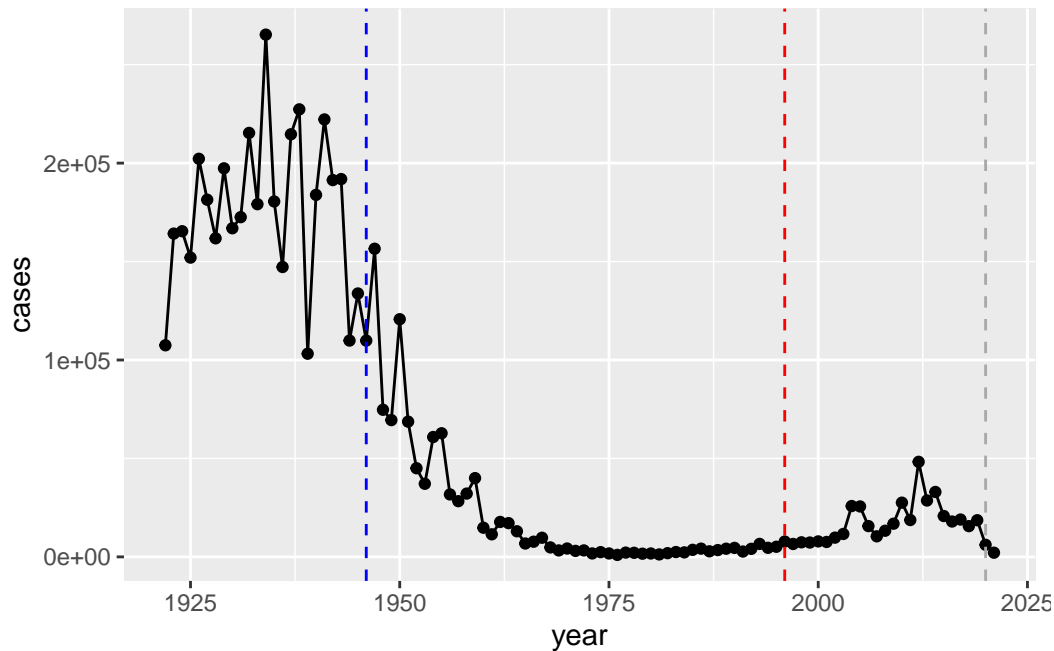
Q1. With the help of the R “addin” package datapasta assign the CDC pertussis case number data to a data frame called cdc and use ggplot to make a plot of cases numbers over time.

```
library(ggplot2)
ggplot(cdc, aes(year, cases)) +
  geom_point() +
  geom_line()
```



Q2. Using the ggplot `geom_vline()` function add lines to your previous plot for the 1946 introduction of the wP vaccine and the 1996 switch to aP vaccine (see example in the hint below). What do you notice?

```
ggplot(cdc, aes(year, cases)) +
  geom_point() +
  geom_line() +
  geom_vline(xintercept = 1946, linetype = "dashed", col = "blue") +
  geom_vline(xintercept = 1996, linetype = "dashed", col = "red") +
  geom_vline(xintercept = 2020, linetype = "dashed", col = "darkgrey")
```



Q3. Describe what happened after the introduction of the aP vaccine? Do you have a possible explanation for the observed trend?

Cases were low then jumped up in 2004. This could've been caused by different things like the anti-vax movement or a shorter period of immunity when given the aP vs wP vaccine. However none of that is clear so more study needs to be done.

## Exploring CMI-PB Data

Why is this vaccine-preventable disease on the upswing? To answer this question we need to investigate the mechanisms underlying waning protection against pertussis. This requires evaluation of pertussis-specific immune responses over time in wP and aP vaccinated individuals.

This is the goal of the CMI-PB project: <https://www.cmi-pb.org/>

It makes its data available via “API-endpoint” that return JSON format. We will use the ‘jsonlite’ package to access this data.

```
library(jsonlite)
```

```
# Data tables
```

```

subject <- read_json("http://cmi-pb.org/api/subject", simplifyVector = T)
specimen <- read_json("http://cmi-pb.org/api/specimen", simplifyVector = T)
titer <- read_json("http://cmi-pb.org/api/v4/plasma_ab_titer", simplifyVector = T)

```

```
head(subject)
```

	subject_id	infancy_vac	biological_sex	ethnicity	race
1	1	wP	Female Not Hispanic or Latino	White	
2	2	wP	Female Not Hispanic or Latino	White	
3	3	wP	Female	Unknown	White
4	4	wP	Male Not Hispanic or Latino	Asian	
5	5	wP	Male Not Hispanic or Latino	Asian	
6	6	wP	Female Not Hispanic or Latino	White	

	year_of_birth	date_of_boost	dataset
1	1986-01-01	2016-09-12	2020_dataset
2	1968-01-01	2019-01-28	2020_dataset
3	1983-01-01	2016-10-10	2020_dataset
4	1988-01-01	2016-08-29	2020_dataset
5	1991-01-01	2016-08-29	2020_dataset
6	1988-01-01	2016-10-10	2020_dataset

```
head(titer)
```

	specimen_id	isotype	is_antigen_specific	antigen	MFI	MFI_normalised
1	1	IgE	FALSE	Total	1110.21154	2.493425
2	1	IgE	FALSE	Total	2708.91616	2.493425
3	1	IgG	TRUE	PT	68.56614	3.736992
4	1	IgG	TRUE	PRN	332.12718	2.602350
5	1	IgG	TRUE	FHA	1887.12263	34.050956
6	1	IgE	TRUE	ACT	0.10000	1.000000

	unit	lower_limit_of_detection
1	UG/ML	2.096133
2	IU/ML	29.170000
3	IU/ML	0.530000
4	IU/ML	6.205949
5	IU/ML	4.679535
6	IU/ML	2.816431

Q4. How many aP and wP infancy vaccinated subjects are in the dataset?

```
table(subject$infancy_vac)
```

```
aP wP  
60 58
```

Q5. How many Male and Female subjects/patients are in the dataset?

```
table(subject$biological_sex)
```

```
Female    Male  
    79     39
```

Q6. What is the breakdown of race and biological sex (e.g. number of Asian females, White males etc...)?

```
table(subject$race, subject$biological_sex)
```

	Female	Male
American Indian/Alaska Native	0	1
Asian	21	11
Black or African American	2	0
More Than One Race	9	2
Native Hawaiian or Other Pacific Islander	1	1
Unknown or Not Reported	11	4
White	35	20

The most represented category is white female. For research like this a more diverse study group would be preferable.

### Side-note: Working with Dates

Dates can be super annoying to work with - the format and order is different around the world. Using the lubridate package makes it a LOT easier to calculate dates:

```
library(lubridate)
```

Attaching package: 'lubridate'

The following objects are masked from 'package:base':

date, intersect, setdiff, union

```
today()
```

```
[1] "2023-12-08"
```

```
today() - mdy("10-7-2002")
```

Time difference of 7732 days

```
today() - ymd("2002-10-7")
```

Time difference of 7732 days

```
time_length( today() - ymd("1961-11-9"), "years")
```

```
[1] 62.07803
```

Q7. Using this approach determine (i) the average age of wP individuals, (ii) the average age of aP individuals; and (iii) are they significantly different?

```
library(dplyr)
```

Attaching package: 'dplyr'

The following objects are masked from 'package:stats':

filter, lag

The following objects are masked from 'package:base':

intersect, setdiff, setequal, union

```
# average wP age
subject %>%
  filter(infancy_vac == "wP") %>%
  summarize(meanAge = mean(time_length(today() - ymd(year_of_birth), "years")))

meanAge
1 36.32977

# average aP age
subject %>%
  filter(infancy_vac == "aP") %>%
  summarize(meanAge = mean(time_length(today() - ymd(year_of_birth), "years")))

meanAge
1 26.03304
```

(iii). A ten-year difference between wP and aP vaccination ages seems pretty significant.

Q8. Determine the age of all individuals at time of boost?

```
# Calculating age at time of boost
subject$age <- ymd(subject$date_of_boost) - ymd(subject$year_of_birth)

# Converting time length to years
subject$age_years <- time_length(subject$age, "years")

subject$age_years

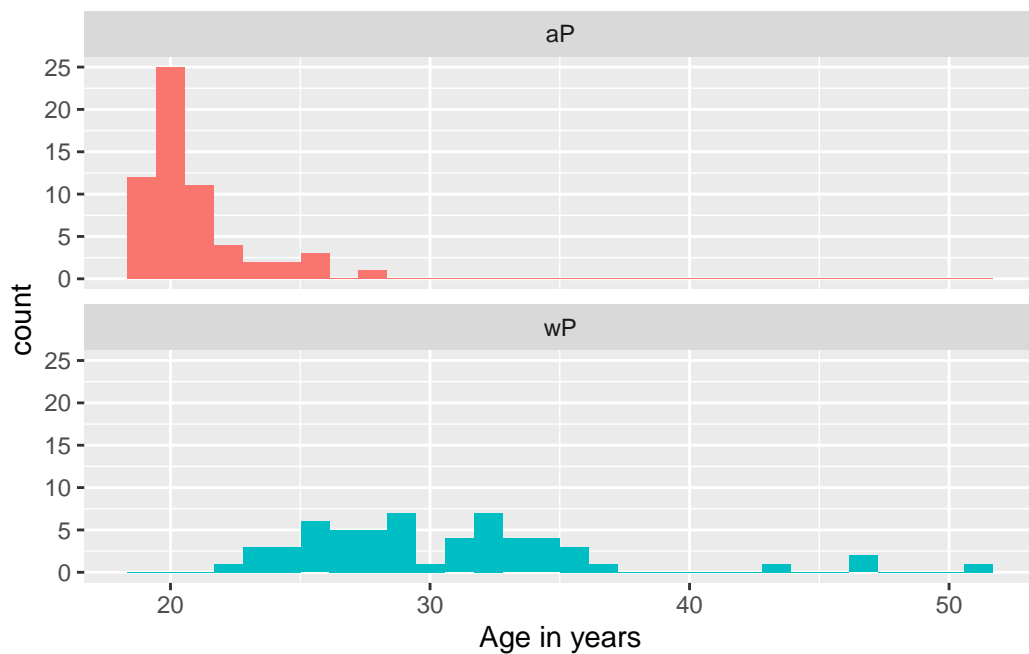
[1] 30.69678 51.07461 33.77413 28.65982 25.65914 28.77481 35.84942 34.14921
[9] 20.56400 34.56263 30.65845 34.56263 19.56194 23.61944 27.61944 29.56331
[17] 36.69815 19.65777 22.73511 35.65777 33.65914 31.65777 25.73580 24.70089
[25] 28.70089 33.73580 19.73443 34.73511 19.73443 28.73648 27.73443 19.81109
[33] 26.77344 33.81246 25.77413 19.81109 18.85010 19.81109 31.81109 22.81177
[41] 31.84942 19.84942 18.85010 18.85010 19.90691 18.85010 20.90897 19.04449
[49] 20.04381 19.90691 19.90691 19.00616 19.00616 20.04381 20.04381 20.07940
[57] 21.08145 20.07940 20.07940 20.07940 32.26557 25.90007 23.90144 25.90007
[65] 28.91992 42.92129 47.07461 47.07461 29.07324 21.07324 21.07324 28.15058
[73] 24.15058 24.15058 21.14990 21.14990 31.20876 26.20671 32.20808 27.20876
[81] 26.20671 21.20739 20.26557 22.26420 19.32375 21.32238 19.32375 19.32375
```

```
[89] 22.41752 20.41889 21.41821 19.47707 23.47707 20.47639 21.47570 19.47707
[97] 35.90965 28.73648 22.68309 20.83231 18.83368 18.83368 27.68241 32.68172
[105] 27.68241 25.68378 23.68241 26.73785 32.73648 24.73648 25.79603 25.79603
[113] 25.79603 31.79466 19.83299 21.91102 27.90965 24.06297
```

Q9. With the help of a faceted boxplot or histogram (see below), do you think these two groups are significantly different?

```
ggplot(subject) +
  aes(time_length(age, "year"),
       fill=as.factor(infancy_vac)) +
  geom_histogram(show.legend=FALSE) +
  facet_wrap(vars(infancy_vac), nrow=2) +
  xlab("Age in years")
```

`stat\_bin()` using `bins = 30`. Pick better value with `binwidth`.



They are significantly different in age. aP subjects are all around 20 years old while none of the wP subjects are younger than 20.



## Merge or Join Tables

Q9. Complete the code to join specimen and subject tables to make a new merged data frame containing all specimen records along with their associated subject details:

```
meta <- inner_join(specimen, subject)
```

Joining with `by = join\_by(subject\_id)`

```
head(meta)
```

	specimen_id	subject_id	actual_day_relative_to_boost
1	1	1	-3
2	2	1	1
3	3	1	3
4	4	1	7
5	5	1	11
6	6	1	32

	planned_day_relative_to_boost	specimen_type	visit	infancy_vac	biological_sex
1	0	Blood	1	wP	Female
2	1	Blood	2	wP	Female
3	3	Blood	3	wP	Female
4	7	Blood	4	wP	Female
5	14	Blood	5	wP	Female
6	30	Blood	6	wP	Female

	ethnicity	race	year_of_birth	date_of_boost	dataset
1	Not Hispanic or Latino	White	1986-01-01	2016-09-12	2020_dataset
2	Not Hispanic or Latino	White	1986-01-01	2016-09-12	2020_dataset
3	Not Hispanic or Latino	White	1986-01-01	2016-09-12	2020_dataset
4	Not Hispanic or Latino	White	1986-01-01	2016-09-12	2020_dataset
5	Not Hispanic or Latino	White	1986-01-01	2016-09-12	2020_dataset
6	Not Hispanic or Latino	White	1986-01-01	2016-09-12	2020_dataset

	age	age_years
1	11212 days	30.69678
2	11212 days	30.69678
3	11212 days	30.69678
4	11212 days	30.69678
5	11212 days	30.69678
6	11212 days	30.69678

Antibody measurements are stored in the titer. However, it doesn't include any of the meta-data:

```
head(titer)
```

	specimen_id	isotype	is_antigen_specific	antigen	MFI	MFI_normalised
1	1	IgE	FALSE	Total	1110.21154	2.493425
2	1	IgE	FALSE	Total	2708.91616	2.493425
3	1	IgG	TRUE	PT	68.56614	3.736992
4	1	IgG	TRUE	PRN	332.12718	2.602350
5	1	IgG	TRUE	FHA	1887.12263	34.050956
6	1	IgE	TRUE	ACT	0.10000	1.000000

	unit	lower_limit_of_detection
1	UG/ML	2.096133
2	IU/ML	29.170000
3	IU/ML	0.530000
4	IU/ML	6.205949
5	IU/ML	4.679535
6	IU/ML	2.816431

Q10. Now using the same procedure join meta with titer data so we can further analyze this data in terms of time of visit aP/wP, male/female etc.

```
abdata <- inner_join(titer, meta)
```

Joining with `by = join\_by(specimen\_id)`

```
dim(abdata)
```

```
[1] 41810    22
```

Q11. How many specimens (i.e. entries in abdata) do we have for each isotype?

```
table(abdata$isotype)
```

```

IgE  IgG IgG1 IgG2 IgG3 IgG4
6698 3240 7968 7968 7968 7968

```

Q12. What are the different \$dataset values in abdata and what do you notice about the number of rows for the most “recent” dataset?

Let’s focus on one of these - IgG. We’re gonna use dplyr to filter only this data.

```
igg <- abdata %>% filter(isotype == "IgG")
head(igg)
```

	specimen_id	isotype	is_antigen_specific	antigen	MFI	MFI_normalised
1	1	IgG	TRUE	PT	68.56614	3.736992
2	1	IgG	TRUE	PRN	332.12718	2.602350
3	1	IgG	TRUE	FHA	1887.12263	34.050956
4	19	IgG	TRUE	PT	20.11607	1.096366
5	19	IgG	TRUE	PRN	976.67419	7.652635
6	19	IgG	TRUE	FHA	60.76626	1.096457

	unit	lower_limit_of_detection	subject_id	actual_day_relative_to_boost
1	IU/ML	0.530000	1	-3
2	IU/ML	6.205949	1	-3
3	IU/ML	4.679535	1	-3
4	IU/ML	0.530000	3	-3
5	IU/ML	6.205949	3	-3
6	IU/ML	4.679535	3	-3

	planned_day_relative_to_boost	specimen_type	visit	infancy_vac	biological_sex
1	0	Blood	1	wP	Female
2	0	Blood	1	wP	Female
3	0	Blood	1	wP	Female
4	0	Blood	1	wP	Female
5	0	Blood	1	wP	Female
6	0	Blood	1	wP	Female

	ethnicity	race	year_of_birth	date_of_boost	dataset
1	Not Hispanic or Latino	White	1986-01-01	2016-09-12	2020_dataset
2	Not Hispanic or Latino	White	1986-01-01	2016-09-12	2020_dataset
3	Not Hispanic or Latino	White	1986-01-01	2016-09-12	2020_dataset
4	Unknown	White	1983-01-01	2016-10-10	2020_dataset
5	Unknown	White	1983-01-01	2016-10-10	2020_dataset
6	Unknown	White	1983-01-01	2016-10-10	2020_dataset

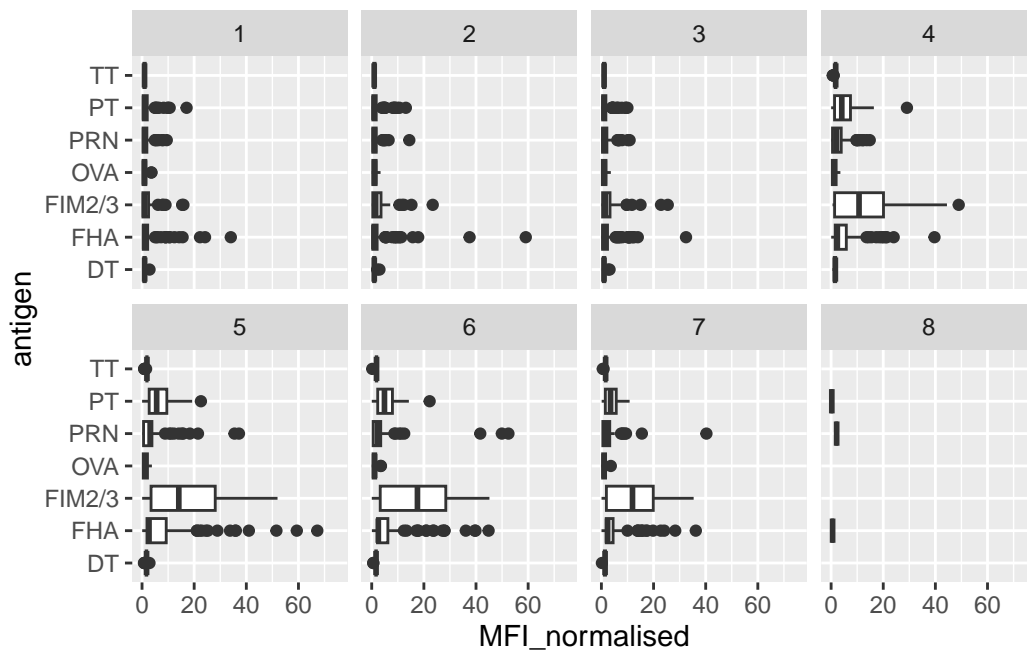
	age	age_years
1	11212 days	30.69678
2	11212 days	30.69678
3	11212 days	30.69678
4	12336 days	33.77413
5	12336 days	33.77413

6 12336 days 33.77413

Q13. Complete the following code to make a summary boxplot of Ab titer levels (MFI) for all antigens:

```
ggplot(igg) +  
  aes(MFI_normalised, antigen) +  
  geom_boxplot() +  
  xlim(0,75) +  
  facet_wrap(vars(visit), nrow=2)
```

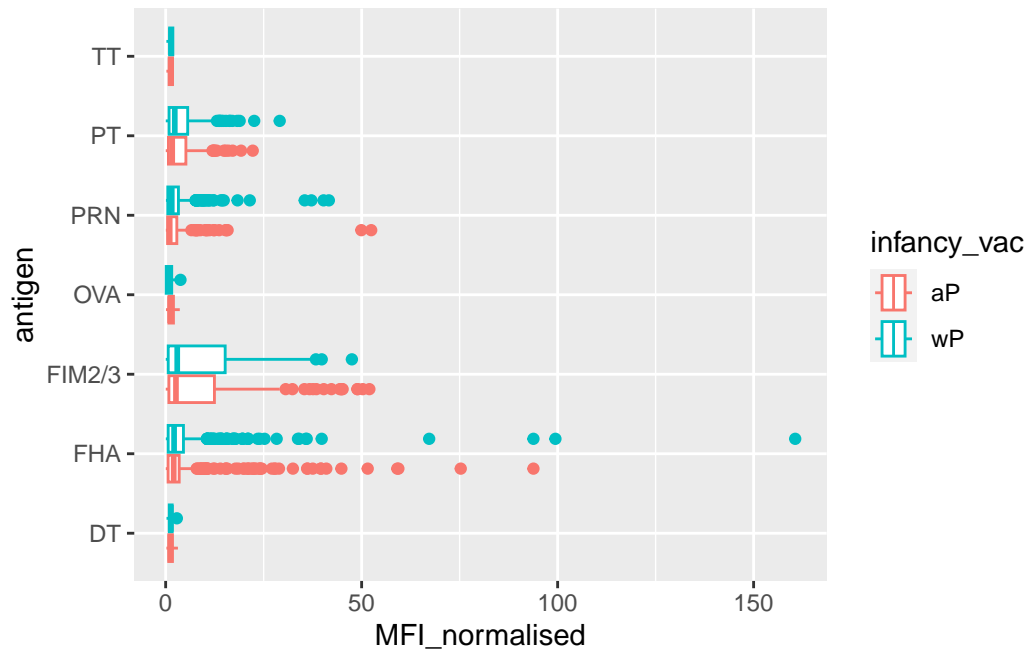
Warning: Removed 5 rows containing non-finite values (`stat\_boxplot()`).



Other box plots we did in class:

```
ggplot(igg) +  
  aes(MFI_normalised, antigen) +  
  geom_boxplot()
```





Focus on IgG to the Pertussis Toxin (PT) antigen in the 2021 dataset:

```
igg.pt <- igg %>% filter(antigen == "PT", dataset == "2021_dataset")
```

```
ggplot(igg.pt) +
  aes(planned_day_relative_to_boost,
      MFI_normalised,
      col = infancy_vac,
      group = subject_id) +
  geom_point() +
  geom_line() +
  geom_vline(xintercept = 0, linetype = "dashed") +
  geom_vline(xintercept = 14, linetype = "dashed")
```

