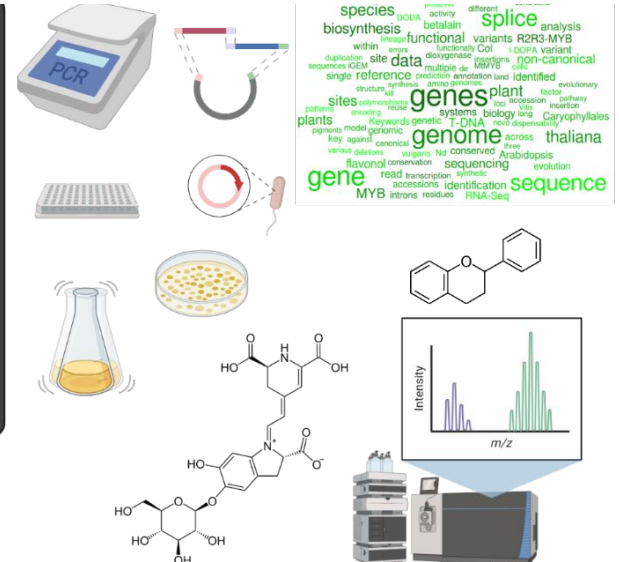
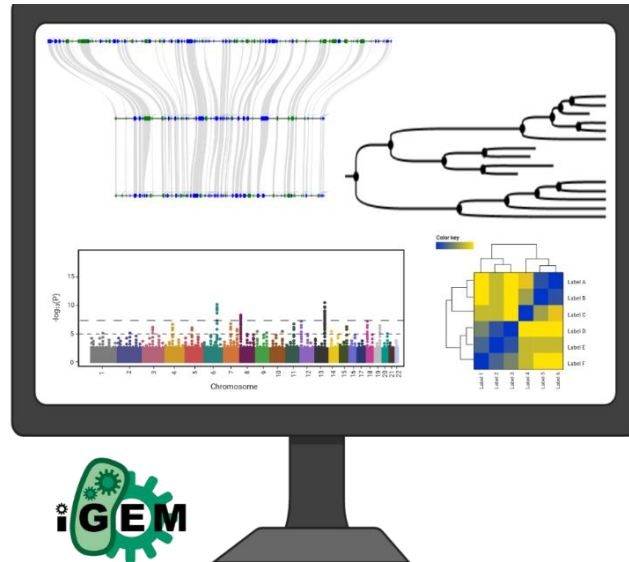




Technische  
Universität  
Braunschweig



# Plant DNA extraction

Prof. Dr. Boas Pucker  
(Plant Biotechnology and Bioinformatics)

# Availability of slides

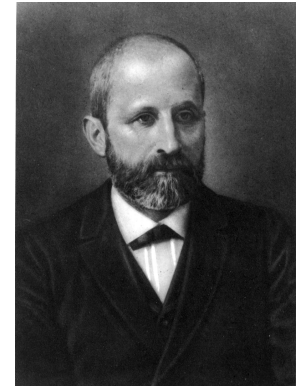
- All materials are freely available (CC BY) - after the lectures:
  - StudIP: **LMChemBSc12**
  - GitHub: <https://github.com/bpucker/teaching>
- Questions: Feel free to ask at any time
- Feedback, comments, or questions: [b.pucker\[a\]tu-bs.de](mailto:b.pucker[a]tu-bs.de)

My figures and content can be re-used in accordance with CC BY 4.0, but this might not apply to all images/logos. Some figure were constructed using bioRender.com.

# Who discovered DNA and when?

# Discovery of DNA

- Isolation of DNA by Friedrich Miescher in 1869
  - Working in the castle in Tübingen
- WATSON, J., CRICK, F. Molecular Structure of Nucleic Acids: A Structure for Deoxyribose Nucleic Acid. *Nature* **171**, 737–738 (1953). <https://doi.org/10.1038/171737a0>



Friedrich Miescher  
(1844-1895)

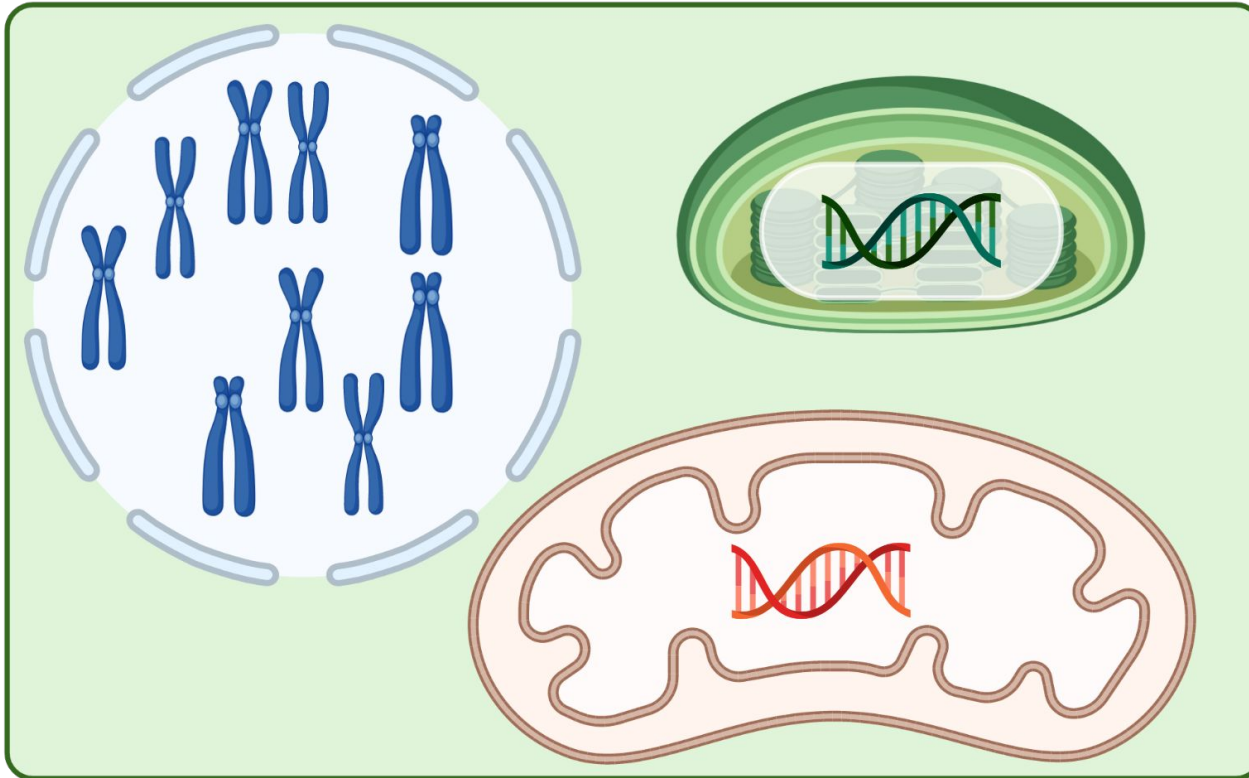
MOLECULAR STRUCTURE OF  
NUCLEIC ACIDS

**A Structure for Deoxyribose Nucleic Acid**

**W**E wish to suggest a structure for the salt of deoxyribose nucleic acid (D.N.A.). This structure has novel features which are of considerable biological interest.



# Plant DNA



- Nucleome: DNA in nucleus (gDNA)
- Plastome: DNA in chloroplast and other plastids (ptDNA)
- Chondrome: DNA in mitochondria (mtDNA)

# What are challenges during DNA extraction?

# DNA extraction challenges

- Cell wall
- Contaminations (proteins, specialized metabolites)
- Other nucleic acids (RNAs)
- DNases
- DNA fragmentation

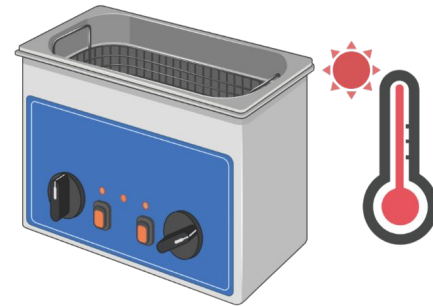
# Breaking the cell wall



Mortar &  
pestle



Ribolyzer



Ultrasonic  
& heat

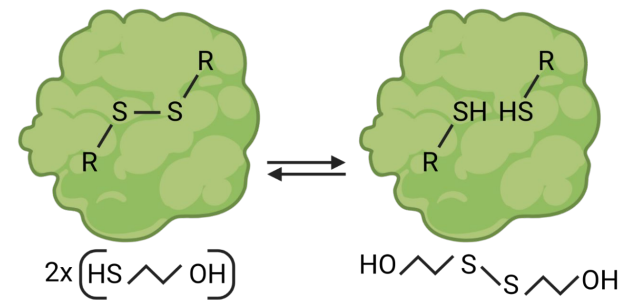
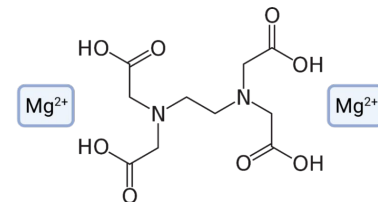


# Plant cell composition - potential contaminants

macromolecule	Percentage of total dry weight	Number of molecules per cell
protein	55	3,000,000
RNA	20	-
DNA	3	-
lipid	9	20,000,000

# How to inactivate DNases?

- Heat
- EDTA
- $\beta$ -mercaptoethanol
- Proteases

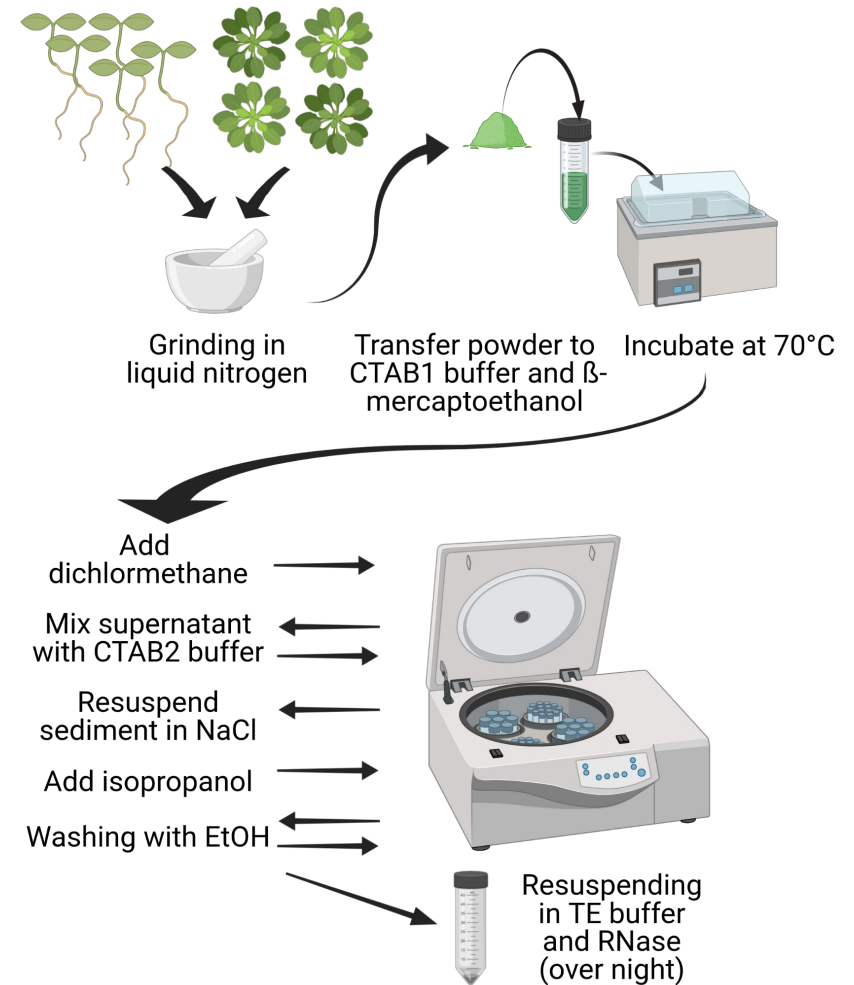


# DNA extraction methods

- CTAB
- Edwards
- Kit-based
- Nucleus isolation
- Leaf lysis methods
- Magnetic beads

# CTAB: Cetyltrimethylammonium bromide

- High DNA quality (suitable for sequencing)
- Substantial time commitment
- Low material costs



# Edwards

- Good DNA quality (sufficient for PCR)
- Modest time commitment
- Low material costs



leaf sample



leaf disk extraction



grinding



400 $\mu$ L extraction buffer



vortexing



centrifugation



transfer of supernatant and  
addition of 300 $\mu$ L isopropanol



resolve in 100 $\mu$ L TE

# Kit-based DNA extraction

- High DNA purity
- Fragment length and total amount limited
- Modest time commitment
- High material costs



grinding sample



lysis (protease)



RNA digestion (RNase A)



Binding DNA to column



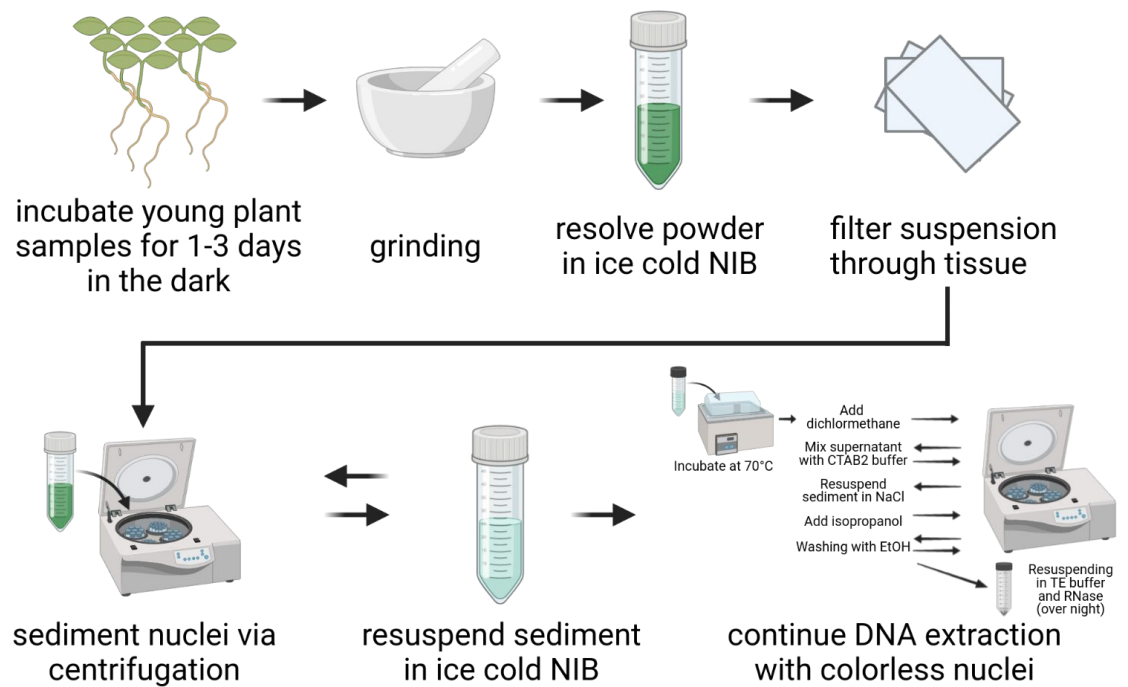
Washing



DNA elution

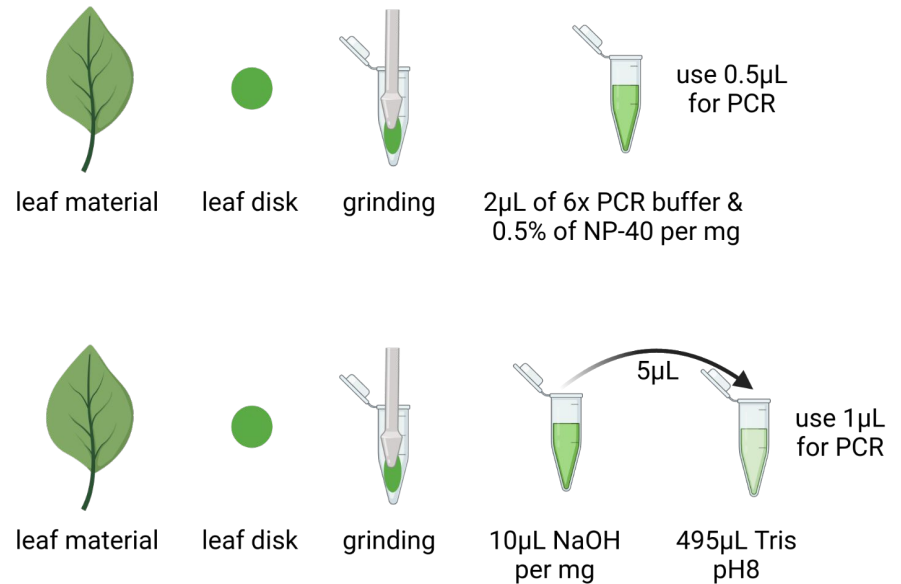
# Nucleus isolation

- Excellent DNA quality
- Very high time commitment
- Modest material costs



# Leaf lysis methods

- Low DNA quality/purity; only suitable for immediate use in PCR
- Extremely fast
- Low costs

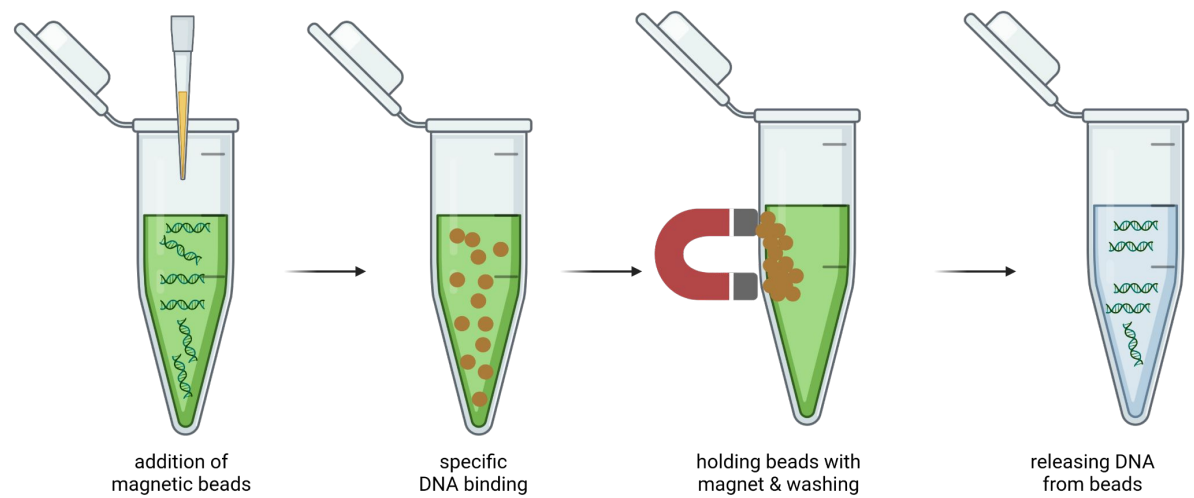


Wang et al., 1993: 10.1093/nar/21.17.4153



# Magnetic beads

- High DNA quality
- Modest time commitment
- High material costs

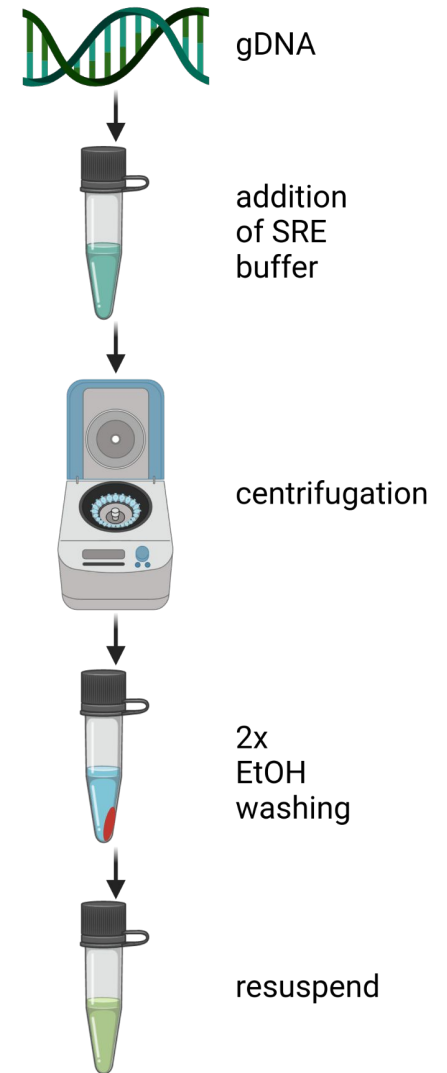


# DNA precipitation

- Cation necessary to neutralize negative charge of DNA
- Low temperature facilitates flocculation of nucleic acids
- DNA concentration needs to be high enough
- DNA precipitation requires:
  - 35% isopropanol + 0.5M salt
  - 75% ethanol + 0.5M salt

# Short Read Eliminator (SRE) kit

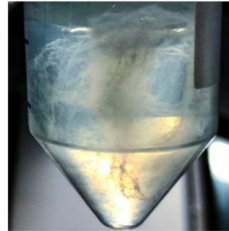
- Proprietary salt mixture allows DNA precipitation
- Removal of DNA fragments <10kb
- Depletion of DNA fragments <20kb



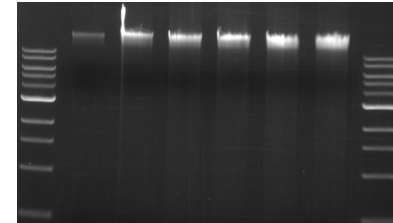
# How to assess the DNA quality?

# DNA quality check

- Optical inspection
- Agarose gel
- NanoDrop
- Qubit
- qPCR (next lecture)

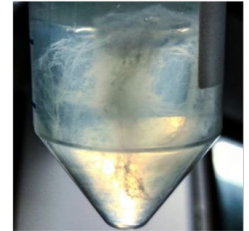


(photo credit: Hanna Schilbert)



# Optical check

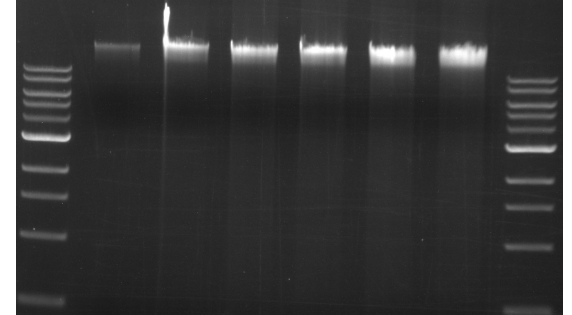
- DNA can be seen in precipitation steps
- Presence of sediment can be checked after centrifugation steps
- White color is often caused by contaminants
- Perfectly clean DNA is almost not visible



(photo credit: Hanna Schilbert)

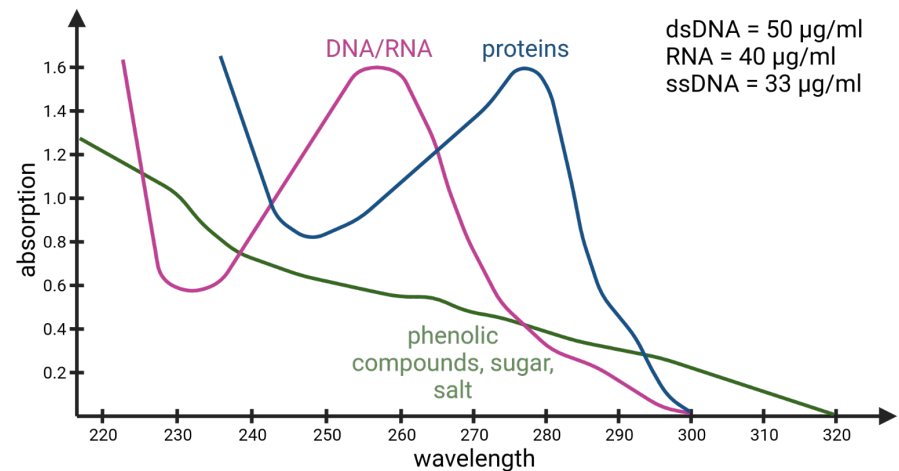
# Agarose gel

- Agarose is a special polysaccharide
- DNA moves through the gel from minus (cathode) to plus (anode)
- DNA is separated by size



# NanoDrop

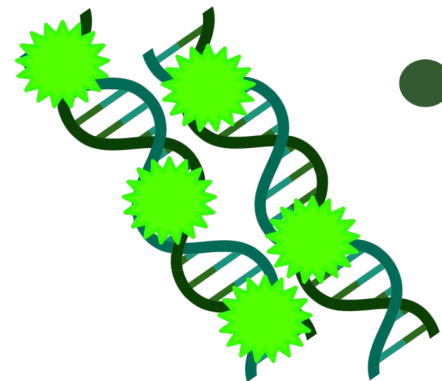
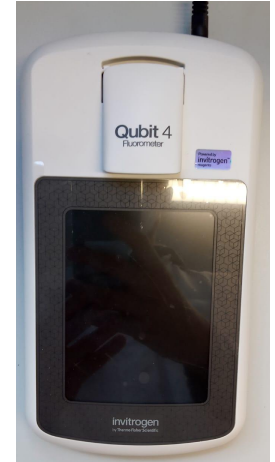
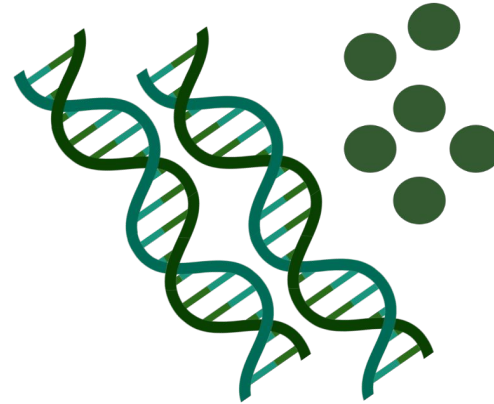
- Photometric analysis of nucleic acids
- Nucleic acid absorbance peak at 260nm
- Protein peak absorbance at 280nm (aromatic amino acids)
- $OD_{260}/OD_{230}$  = small fragment/phenolic compound contamination
- $OD_{260}/OD_{280}$  = protein contamination





# Qubit

- Fluorescence-based (PicoGreen) measurement of nucleic acid concentration
- dsDNA HS Assay: 10pg/ $\mu$ L - 100ng/ $\mu$ L
- dsDNA BR Assay: 50pg/ $\mu$ L - 200ng/ $\mu$ L



# How to store DNA?

# DNA storage

- EDTA in buffer prevents degradation by DNases
- Store at 4°C for up to a week
- Store at -20°C for long term
- High molecular weight DNA should not be frozen to avoid breaks

# Time for questions!

# Questions

1. Who discovered DNA?
2. What is a plant genome?
3. What are challenges during the DNA extraction?
4. Which different DNA extraction methods exist?
5. What are the important steps of a DNA extraction method?
6. How can the DNA quality be checked?
7. What is the concept of a DNA quality check method?