

Package ‘SmCCNet’

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Type Package

Title Sparse multiple canonical correlation network analysis tool

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Description

A canonical correlation based framework for integrating multiple omics data types and a quantitative phenotype of interest, and constructing phenotype-specific multi-omics networks.

URL <https://github.com/KechrisLab/SmCCNet>

BugReports <https://github.com/KechrisLab/SmCCNet/issues>

Depends R (>= 3.4)

Imports PMA, Matrix, pbapply, igraph

Suggests BiocStyle, knitr, rmarkdown

VignetteBuilder knitr

License GPL-2

Encoding UTF-8

LazyData true

RoxygenNote 6.0.1

NeedsCompilation no

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getAbar	<i>Compute the similarity matrix based on the outer products of absolute canonical weights.</i>
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Description

Compute the similarity matrix based on the outer products of absolute canonical weights.

Usage

```
getAbar(Ws, FeatureLabel = NULL)
```

Arguments

Ws	A canonical weight vector or matrix. If Ws is a matrix, then each column corresponds to one weight vector.
FeatureLabel	A 1×p vector indicating feature names. If FeatureLabel = NULL (default), the feature names will be indices 1 through p, where p is the total number of omics features.

Value

A p×p symmetric non-negative matrix.

Examples

```
w <- matrix(rnorm(6), nrow = 3)
Ws <- apply(w, 2, function(x) return(x/sqrt(sum(x^2))))
abar <- getAbar(Ws)
```

getMultiOmicsModules	<i>Apply a hierarchical tree cutting to the similarity matrix and extract modules that contain both omics data types.</i>
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Description

Apply a hierarchical tree cutting to the similarity matrix and extract modules that contain both omics data types.

Usage

```
getMultiOmicsModules(Abar, P1, CutHeight = 1 - 0.1^10, PlotTree = TRUE)
```

Arguments

Abar	A similarity matrix for both omics data types.
P1	Total number of features for the first omics data type.
CutHeight	Height threshold for the hierarchical tree. Default is 1-0.1 ¹⁰ .
PlotTree	Logical. Whether to create the hierarchical tree plot.

Value

A list of multi-omics modules.

Examples

```
set.seed(123)
w <- rnorm(5)
w <- w/sqrt(sum(w^2))
abar <- getAbar(w)
modules <- getMultiOmicsModules(abar, P1 = 2, CutHeight = 0.5)
```

getRobustPseudoWeights

Compute canonical weights based on sparse multiple canonical correlations (SmCCA), sparse supervised canonical correlations (SsCCA), or sparse canonical correlations (SCCA).

Description

SmCCA and SsCCA take into account of a phenotype/trait. SmCCA maximizes the total (weighted or unweighted) pairwise canonical weights between two omics data type and the trait. It requires the trait to be quantitative. SsCCA prioritizes omics features based on the trait, and assigns non-zero canonical weights to features that are more correlated to the trait. SCCA does not use any trait information for computing the canonical weights. All of these three methods are included in the function, along with an omic feature subsampling scheme.

Usage

```
getRobustPseudoWeights(X1, X2, Trait, Lambda1, Lambda2, s1, s2, NoTrait,
  FilterByTrait, Bipartite, SubsamplingNum, PartitionNum, CCcoef = NULL,
  trace = FALSE)
```

Arguments

X1	An $n \times p_1$ data matrix (e.g. mRNA) with p_1 features and n subjects.
X2	An $n \times p_2$ data matrix (e.g. miRNA) with p_2 features and n subjects.
Trait	An $n \times 1$ trait data matrix for the same n subjects.
Lambda1	LASSO penalty parameter for X1, need to be between 0 and 1.
Lambda2	LASSO penalty parameter for X2, need to be between 0 and 1.
s1	Proportion of mRNA features to be included.
s2	Proportion of miRNA features to be included.
NoTrait	Logical. Whether trait information is provided.
FilterByTrait	Logical. Whether only the top 80 correlation to the trait will be assigned nonzero weights.
Bipartite	Logical. Whether to include random partition.
SubsamplingNum	Number of feature subsamples. Larger number leads to more accurate results, but at a higher cost. We recommend to subsample at least 1000 times.

PartitionNum	Number of random partitions for each set of subsamples. This is only used if Bipartite = FALSE.
CCcoef	Optional coefficients for the pairwise canonical correlations (CC). If CCcoef = NULL (default), then the objective function is the total sum of all pairwise CC. It can also be a coefficient vector that follows the column order of <code>combn(K, 2)</code> .
trace	Logical. Whether to display CCA algorithm trace.

Value

A canonical weight matrix with $p_1 + p_2$ rows. Each column is the canonical weights based on sub-sampled X_1 and X_2 features. The column number equals to `SubsamplineNum`.

Examples

```
## For illustration, we only subsample 5 times.
set.seed(123)

# SmCCA
W1 <- getRobustPseudoWeights(X1, X2, Trait = Y, Lambda1 = 0.05,
  Lambda2 = 0.05, s1 = 0.7, s2 = 0.9, NoTrait = FALSE, FilterByTrait = FALSE,
  Bipartite = FALSE, SubsamplingNum = 100, PartitionNum = NULL,
  CCcoef = NULL, trace = FALSE)

# SsCCA
W2 <- getRobustPseudoWeights(X1, X2, Trait = Y, Lambda1 = .05, Lambda2 = 0.5,
  s1 = 0.7, s2 = 0.9, NoTrait = FALSE, FilterByTrait = TRUE,
  Bipartite = FALSE, SubsamplingNum = 100, PartitionNum = NULL,
  CCcoef = NULL, trace = FALSE)

# SCCA
W3 <- getRobustPseudoWeights(X1, X2, Trait = Y, Lambda1 = 0.05,
  Lambda2 = 0.05, s1 = 0.7, s2 = 0.9, NoTrait = TRUE, Bipartite = FALSE,
  SubsamplingNum = 100, PartitionNum = 5, CCcoef = NULL, trace = FALSE)
```

`plotMultiOmicsNetwork` *Plot multi-omics modules based on similarity matrix derived from pseudo canonical weights and pairwise feature correlations.*

Description

Plot multi-omics modules based on similarity matrix derived from pseudo canonical weights and pairwise feature correlations.

Usage

```
plotMultiOmicsNetwork(Abar, CorrMatrix, multiOmicsModule, ModuleIdx, P1,
  EdgeCut, FeatureLabel = NULL, AddCorrSign = TRUE, SaveFile = NULL,
  ShowType1Label = TRUE, ShowType2Label = TRUE, PlotTitle = "",
  NetLayout = "lg1", ShowNodes = TRUE, VertexLabelCex = 1,
  VertexSize = 1)
```

Arguments

Abar	A $p \times p$ similiary matrix for both omics data types based on pseudo canonical weights. All entries are non-negative.
CorrMatrix	A $p \times p$ correlation matrix that provides sign information for the network.
multiOmicsModule	A list of multi-omics modules.
ModuleIdx	Index for the module to be plotted. It can not exceed the length of multiOmicsModule.
P1	Total number of features for the first omics data type.
EdgeCut	A numerical value between 0 and 1, indicating an edge threshold for the network. Any features (network nodes) without any edge strength that passes the threshold are excluded from the figure.
FeatureLabel	A $1 \times p$ vector indicating feature names. If FeatureLabel = NULL (default), the feature names will be indices 1 through p, where p is the total number of omics features.
AddCorrSign	Logical. Whether to add a positive or negative sign to each network edge based on pairwise feature correlations.
SaveFile	A pdf file name for the figure output. If SaveFile = NULL (default), the figure will not be saved.
ShowType1Label	Logical. Whether to label the network nodes for the first omics data type.
ShowType2Label	Logical. Whether to label the network nodes for the second omics data type.
PlotTitle	A title for the figure. Default is without any title.
NetLayout	Graphical layout for the network. Possible options are "circle", "sphere" for 3D sphere, "fr" for Fruchterman-Reinhold, and "lgl" for the LGL algorithm. Refer to igraph manual for more details on the layout options.
ShowNodes	Logical. Whether to show network nodes.
VertexLabelCex	Scaling factor for the vertex labels.
VertexSize	Size of the vertices.

Value

A multi-omics network figure.

Examples

```

set.seed(123)
w <- rnorm(5)
w <- w/sqrt(sum(w^2))
abar <- getAbar(w)
modules <- getMultiOmicsModules(abar, P1 = 2, CutHeight = 0.5)
x <- cbind(X1[,1:2], X2[, 1:3])
corr <- cor(x)

plotMultiOmicsNetwork(abar, corr, modules, ModuleIdx = 1, P1 = 2,
  EdgeCut = 0)

```

X1	<i>Two simulated omics datasets and a quantitative phenotype for the same 358 subjects.</i>
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Description

Two simulated omics datasets and a quantitative phenotype for the same 358 subjects.

Usage

X1

Format

A mRNA matrix with 358 rows and 500 columns

X2	<i>A miRNA matrix with 358 rows and 100 columns</i>
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Description

A miRNA matrix with 358 rows and 100 columns

Usage

X2

Format

An object of class `matrix` with 358 rows and 100 columns.

Y	<i>A phenotype matrix with 358 rows and 1 column</i>
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Description

A phenotype matrix with 358 rows and 1 column

Usage

Y

Format

An object of class `matrix` with 358 rows and 1 columns.

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