

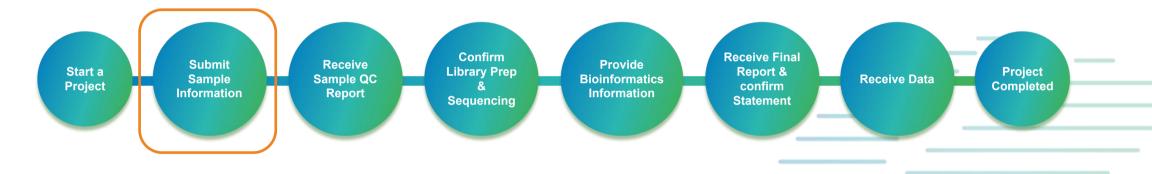
# How to submit Sample Information Form (Premade Libraries)

Novogene Customer Service System (CSS) Instruction Guide
June 2024



### **CSS Overview**

In Novogene, you can manage the sequencing project(s) on CSS through these simple steps.



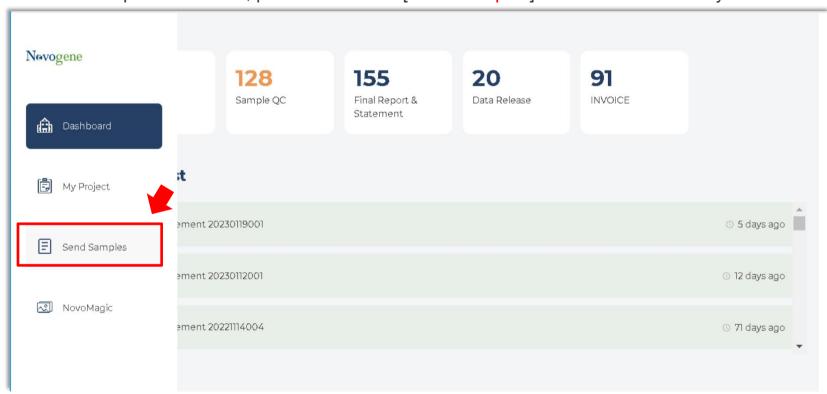
In this guide, we will show you how to submit Sample Information Form for your Premade Libraries using CSS.

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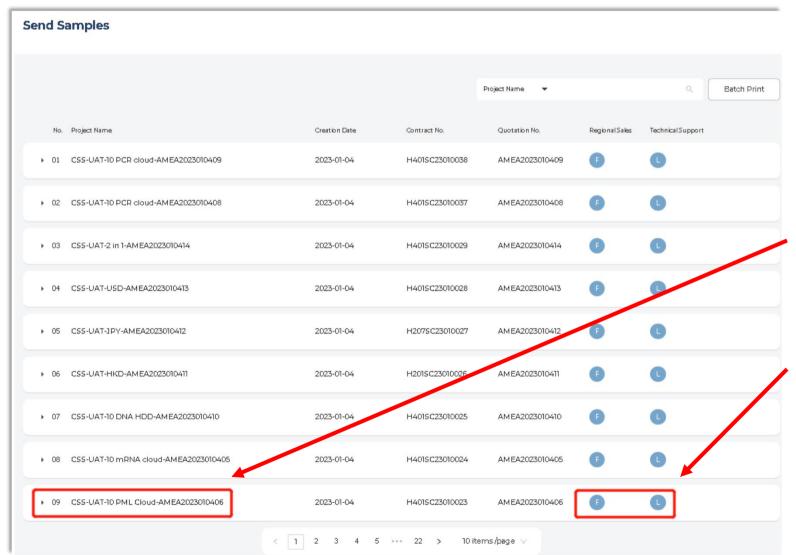
### **Opening the Send Samples menu**

To submit sample information, please locate the [Send Samples] menu on the sidebar your screen.





### **Selecting a Project to Send Samples**



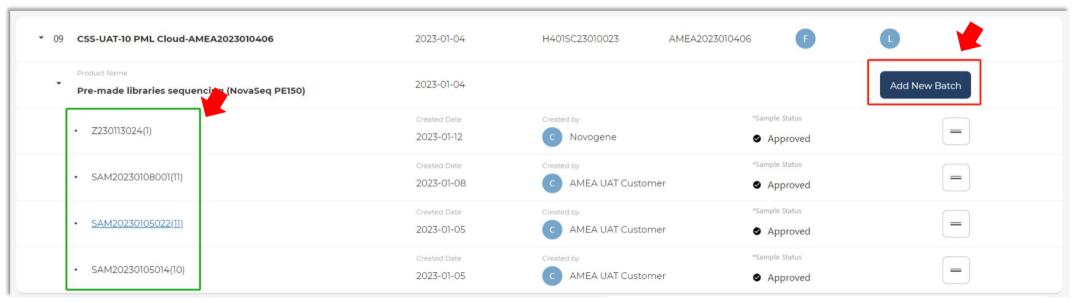
Click on the [Project Name] to expand the project and submit samples.

Hover your cursor over the [name icon] to view the email addresses of the Novogene Representatives for your project. You can contact them if you have any queries.



### **Creating a New Sample Information Form**

Click [Add New Batch] to start a new batch.



Here is a list of Sample Information Forms (SIF) that you have created previously.

The number in the bracket indicates the number of samples you have on the SIF.

#### Sample Status:

- [Editing] means that the SIF has not been submitted.
- [Submitted] means that the SIF has been submitted.
- [Approved] means that the SIF has been checked and accepted by the Novogene Representative.



### **Understanding the send samples Policy and Procedure**

#### Start a New Batch

Please read the instructions below before you start the new batch. To avoid any issues/delays, please ensure you follow all the points that are important. If you have any questions, please check out our **FAQ** section in the Help Center.



#### Sample Name

Library/Sub-library Name must begin with a letter of the alphabet and contain a maximum of 17 alphanumeric characters or underscores.



#### Tube Size

Send samples in 1.5 ml or 2.0 ml microcentrifuge tubes. 96-well plates and other tubes will risk processing delays and handlings fees.



#### **Tube Labelling**

Sample names on the tube must match with the Sample Information Form. Please do not include other information on the tube to avoid any discrepancy.



#### Sample Protection

Seal each tube with parafilm before packaging. We highly recommend placing the sample tubes in a container such as a 50ml tube or cryobox to prevent sample tubes from being crushed by dry ice or other packing materials. Cotton or absorbent papers can be used to prevent tubes from being jostled inside the container.



#### Sample Requirement

Please refer to your quotation for the sample requirements. Any sample failing to follow the sample requirements may risk processing delays.



#### Shipment and Delivery

Novogene highly recommends that you choose FedEx overnight or international express shipping with dry ice, and to avoid weekend delivery.



#### **Biosafety Level**

Novogene does not currently accept blood-borne pathogens or purified viral genomes, as well as other agents classified at Biosafety Level 2 or above ("Infectious Sample"). Please notify your Novogene Sales and obtain Novogene's written approval before submitting any such infectious sample(s). Novogene reserves the right to accept or reject the submission of any infectious sample at its sole discretion. Any infectious sample(s) submitted without prior written approval shall be immediately destroyed or returned.

I confirm that I have read, understand and agree to the above policy and procedure.

Cancel

These are useful tips and instructions for sample preparation.

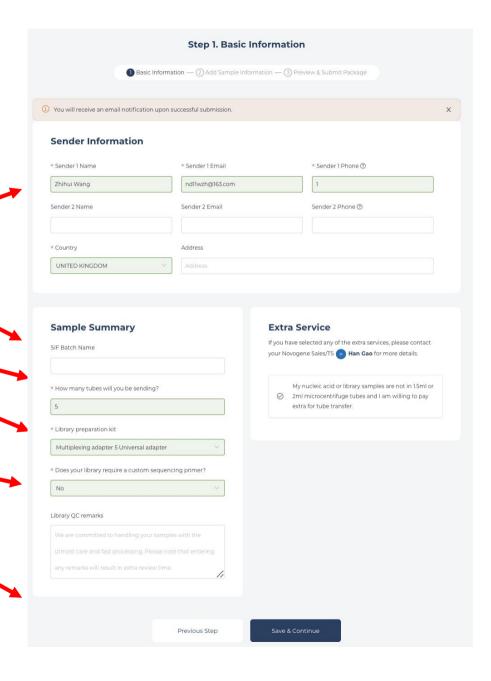
The Library Name has a maximum of 17 alphanumeric characters and MUST begin with an alphabet.

Please **read and follow** the instructions carefully to ensure that there will be no discrepancies when your samples reach our lab.

Click [I confirm] to continue.

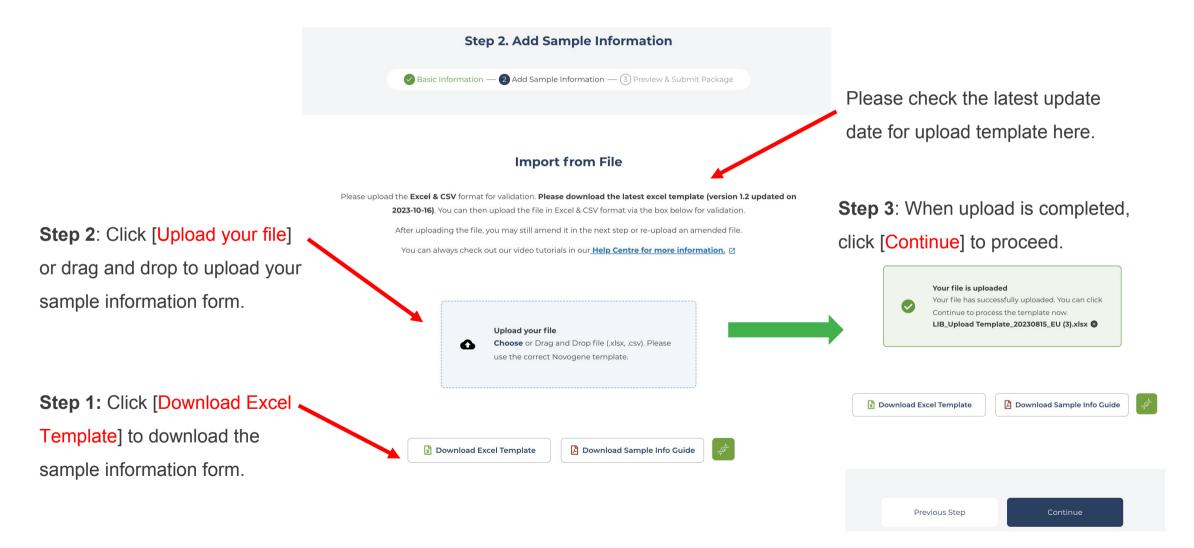
### **Sender Information & Sample Summary**

- Sender Information will be used by our Novogene Representative related to this SIF.
- You can customize the name your batch up (max: 8 characters).
- Please fill in the number of **physical** tubes you will be sending, including the back-up samples.
- Select the library preparation kit you've used from the dropdown list. If you choose Others, please specify it.
- Select "Yes" if you are using customised primers. Please specify your requirments for sequencing.
- You can write note for requirment for library QC stage .
- Click [Save & Continue] to proceed



### **Adding Sample Information**





### Instruction for i5 and i7 Index

Previous Step

Continue



#### **Import from File**



Click on the green icon [ ] to bring up the **premade library instruction** guide.



### Filling up a Sample Information Template

<b>A</b>	В	С	D	E	F	G	Н	1	J	K	L
Library Type (Required)	Library Name (name on tube) (Required)	Sub-library Name(for pooled library) (Required. If is not pooled library, just keep it the same as Library Name)	Data Delivery Method (Required)	i7 Index Sequences (Required when data need be multiplexing)	i5 Index Sequences	Insert Size(bp) (Required)	Library Status (Required)	Total Data Amount (Required)	Data Unit (Required)	Concentration(ng/ul)	Volume(ul)
Premade-WES Library	PML-A	PML_Test_1	Partial lane sequencing-With Demultiplexing	TGCAATGTTC	GCTTGTCGAA	300	Others	12	G raw data		
Premade-WES Library	PML-A	PML_Test_2	Partial lane sequencing-With Demultiplexing	TTAATACGCG	CACCTCGGGT	300	Others	12	G raw data		
Premade-WES Library	PML-A	PML_Test_3	Partial lane sequencing-With Demultiplexing	CCTTCTAGAG	AATACAACGA	300	Others	12	G raw data		
Premade-WES Library	PML-A	PML_Test_4	Partial lane sequencing-With Demultiplexing	GCAGTATAGG	TTCCGTGCAC	300	Others	12	G raw data		
Premade-WES Library	PML-A	PML_Test_5	Partial lane sequencing-With Demultiplexing	TGCAATGTTC	GCTTGTCGAA	300	Others	12	G raw data		
Premade-WES Library	PML-B	PML_Test_6	Partial lane sequencing-With Demultiplexing	TTAATACGCG	CACCTCGGGT	300	Others	20	G raw data		
Premade-WES Library	PML-B	PML_Test_7	Partial lane sequencing-With Demultiplexing	TGCAATGTTC	GCTTGTCGAA	300	Others	20	G raw data		
Premade-WES Library	PML-B	PML_Test_8	Partial lane sequencing-With Demultiplexing	TTAATACGCG	CACCTCGGGT	300	Others	20	G raw data		
Premade-WES Library	PML-B	PML_Test_9	Partial lane sequencing-With Demultiplexing	CCTTCTAGAG	AATACAACGA	300	Others	20	G raw data		
Premade-WES Library	PML-B	PML_Test_10	Partial lane sequencing-With Demultiplexing	GCAGTATAGG	TTCCGTGCAC	300	Others	20	G raw data		
Premade-WES Library	PML-B	PML_Test_11	Partial lane sequencing-With Demultiplexing	AATTCCGGTT	AAAAAATTCC	300	Others	20	G raw data		

- A) Library Type: The type of library that you are sending
- **B)** Library Name: This name should match what is written on the tube. Maximum of 17 characters begin with an alphabet. If you are sending pooled library, fill in the pooled library name and on the physical tube.
- C) Sub-library Name: The name of the individual libraries in the pooled library. In this example, there are 2 pooled libraries, PML A and PML B.
- D) Data Delivery Method: This will be the method of sequencing e.g., Lane sequencing with demultiplexing or Partial lane sequencing with demultiplexing.
- E) i7 and i5 Index Sequences: Please select i5 indexes under v1.0 reagent kits, Sample sheet v2 regardless of sequencing platform. \*\*It is recommended to use dual indexes (i5 and i7) as single index libraries will result in longer turnaround times.
- **G) Insert Size**: The size of the insert sequence.
- H) Library Status: Select Others unless your libraries are in dry powder form
- I) Total Data Amount: The amount of data output required. If you are sending a pooled library, the amount of data for each line will be the total data amount in terms of "G raw data" e.g., the data output for PML-A is 12G raw data. For PE150, only unit G is allowed.



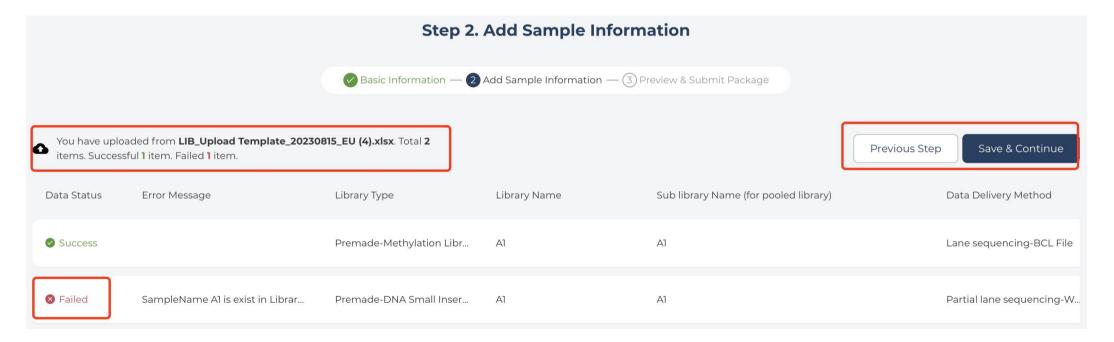
### **Reviewing Uploaded Information**

**Important!** Please ensure all sample information are uploaded successfully.

For [Failed] items, please review the error message and reupload the SIF.

To reupload, click [Previous Step]

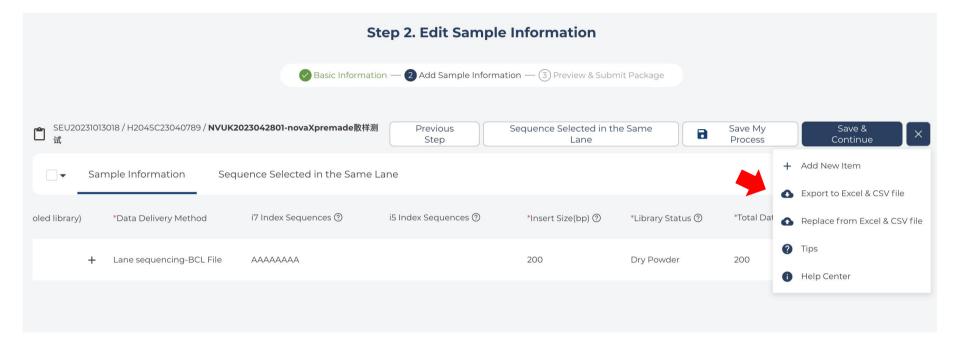
To proceed, click [Save & Continue]



Note: After you click [Save & Continue], we can only save the success items.



### **More information**



You can find **more options** by clicking =

- Add New Item: add one more sample
- Export to Excel & CSV File: Exports current data to an excel file
- Replace from Excel & CSV File: brings you back to the template upload page
- Tips&Help Center: useful tips and FAQs



### **Example 1: Partial Lane Sequencing Non-pooled**

### Library

When you are sending individual libraries.

- Fill in the data amount needed for each individual library.
- Partial lane sequencing-With Demultiplexing: requires
   i5 and i7 index
- Partial lane sequencing-Without Demultiplexing: we don't provide this service anymore.

В	С	I
Library Name (name on tube) (Required)	Sub-library Name(for pooled library) (Required. If is not pooled library, just keep it the same as Library Name)	Total Data Amount (Required)
S1	S1	6
S2	<b>S2</b>	6
S3	<b>S</b> 3	6

### **Example 2: Partial Lane Sequencing Pooled Library**

When you are sending 1 tube of pooled library (please make sure the pooled library is in the correct ratio).

- Fill in the total data amount for the pooled library. E.g., 18G for pooled library S.
- Partial lane sequencing-With Demultiplexing: requires i5 and i7 index.Do not mix libraries with the same index in the same library, otherwise they cannot be distinguished.

В	С	l
Library Name (name on tube) (Required)	Sub-library Name(for pooled library) (Required. If is not pooled library, just keep it the same as Library Name)	Total Data Amount (Required)
S	S1	18
S	<b>S2</b>	18
S	<b>S</b> 3	18



## Example 3: Whole Lane Sequencing Non-pooled Library(without phix)

When you are sending individual library for lane sequencing.

 Fill in the data amount per individual library. The sum of total data amount <u>must not</u> be more than the Total Lane Data Amount. e.g. 800G for one NovaSeq lane for PE150, or 375G for Novaseq Xplus-10B lane, or 1000G for Novaseq Xplus-25B.

В	С	I
Library Name (name on tube) (Required)	Sub-library Name(for pooled library) (Required. If is not pooled library, just keep it the same as Library Name)	Total Data Amount (Required)
S1	S1	40
S2	<b>S2</b>	40
S3	<b>S</b> 3	30

## Example 4: Whole Lane Sequencing Non-pooled Library(with phix)

- Fill in the data amount per individual library with phix. The sum of total data amount <u>must not</u> be more than the Total Lane Data Amount.e.g. 800G for one NovaSeq lane for PE150, or 375G for Novaseq Xplus-10B lane, or 1000G for Novaseq Xplus-25B.
- e.g. If you need 10% phix for the lane, and you want 40G for the sample, then you need to fill in 44G as the total data amount for the sample.

В	С		1
Library Name (name on tube) (Required)	Sub-library Name(for pooled library) (Required. If is not pooled library, just keep it the same as Library Name)	Data amount needed	Total Data Amount (Required)
S1	S1	40	44
S2	S2	40	44
<b>S</b> 3	<b>S</b> 3	40	44



### **Example 5: Whole Lane Sequencing Pooled Library**

When you are sending 1 tube of pooled library for lane sequencing.

- The Total data amount corresponds to the pooled library, not the sub-library.
- Hence, if the pooled Library S needs to be sequenced in 1 NovaSeq lane, S1, S2, S3 needs to indicate as 800G on every row.

В	С	1		
Library Name (name on tube) (Required)	Sub-library Name(for pooled library) (Required. If is not pooled library, just keep it the same as Library Name)	Total Data Amount (Required)		
s	S1	800		
s	S2	800		
S	<b>S</b> 3	800		



**Example 6: 10x Premade Library-(Single index-Non-pooled library)** 

 For certain 10x libraries, if there are four i7 indexes per library, fill in 4 lines of the same library and one i7 index for each library as shown below.

В	С	E	I
Library Name (name on tube) (Required)	Sub-library Name(for pooled library) (Required. If is not pooled library, just keep it the same as Library Name)	i7 Index Sequences (Required when data need be multiplexing)	Total Data Amount (Required)
S10X	Sublibrary1	CGCTATGT	6
S10X	Sublibrary1	GCTGTCCA	6
S10X	S10X Sublibrary1		6
S10X Sublibrary1		CCTATCCT	6



### **Example 7: 10x Premade Library-(Single index-Pooled library)**

- When you are sending 1 tube of pooled 10X library for sequencing. The Total data amount corresponds to the pooled library, not the sub-library. For example: Sublirary1 need 6G, Sublirary2 need 6G, so you need to fill in 12G for the Total Data Amount.
- If there are four i7 index per sub-library, fill in 4 lines of the same sub-library and one i7 index for each sub-library as shown below.

В	l C	Ε	
Library Name (name on tube) (Required)	Sub-library Name(for pooled library) (Required. If is not pooled library, just keep it the same as Library Name)	i7 Index Sequences (Required when data need be multiplexing)	Total Data Amount (Required)
S10X	S10X Sublibrary1 CGCTATGT		12
\$10X	Sublibrary1	GCTGTCCA	12
\$10X	Sublibrary1	TTGAGATC	12
S10X	Sublibrary1	CCTATCCT	12
S10X	S10X Sublibrary2 CCTTAAGG		12
S10X	Sublibrary2	GGGCTTAA	12
S10X	Sublibrary2	AAGGTTCC	12
S10X	S10X Sublibrary2		12



**Example 8: 10x Premade Library (Dual-index library-Non-pooled library)** 

• For certain 10x libraries, if there is only one i7&i5 index, fill in 1 line for the library will be fine. Usually for dual-index plate, there will be only one unique i7 and one unique i5 sample index per well.

В	С	E		l l
Library Name (name on tube) (Required)	Sub-library Name(for pooled library) (Required. If is not pooled library, just keep it the same as Library Name)	i7 Index Sequences (Required when data need be multiplexing)	i5 Index Sequences	Total Data Amount (Required)
S10X	SS001	TAGGGTCAAA	CTTCTAATGT	6

### **Example 9: 10x Premade Library (Dual-index library-pooled library)**

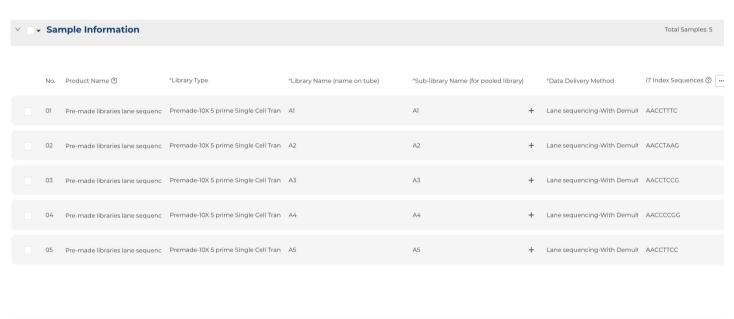
When you are sending 1 tube of pooled 10X library for sequencing. The Total data amount corresponds to the pooled library, not the sub-library.

В	С	E		I
Library Name (name on tube) (Required)	Sub-library Name(for pooled library) (Required. If is not pooled library, just keep it the same as Library Name)	i7 Index Sequences (Required when data need be multiplexing)	i5 Index Sequences	Total Data Amount (Required)
S10X	SS001	TAGGGTCAAA	CTTCTAATGT	12
S10X	SS002	TAGGCAATAA	AGTGCGCACT	12



### **Indicating Lane Sequencing**

- **Step 1:** click [1] +to add a lane. Only lane sequencing project will show this item.
- **Step 2**: click [2] to select samples;

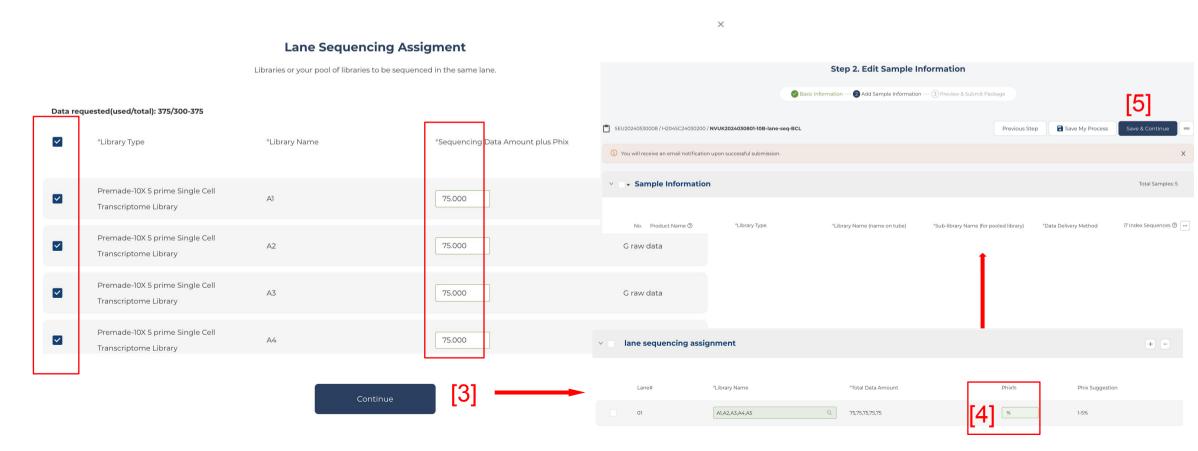






### **Indicating Lane Sequencing**

- **Step 3:** After choosing libraries and filling in data amount in the lane, click [3] 'Continue' to next step.
- **Step 4**:Filling in [4]Phix%;if you don't need any, you can leave it or fill in 0.
- **Step 5**: If everything is OK, Click [5] Save and continue to next step.





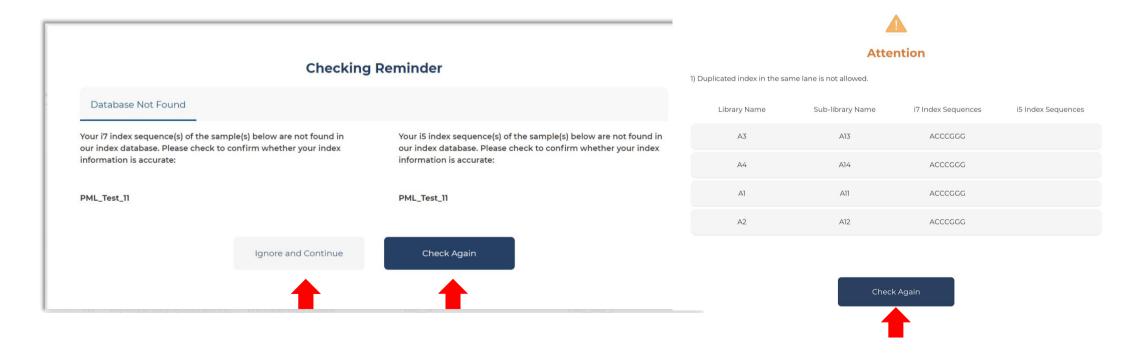
### **System check reminders**

The system will check for indexes information, and you may see a prompt to double check the index information.

If you are sure that the index sequence information is correct, please click [Ignore and Continue]

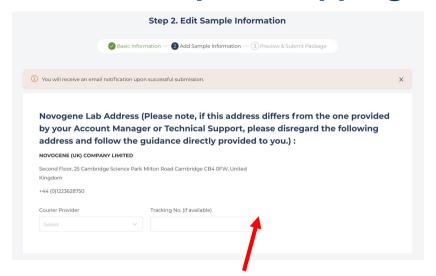
If you would like to re-check the information, click [Check Again] to go back to the previous page.

Duplicated index are not allowed in the same library or the same lane. You have to [Check Again] if it is in this situation.



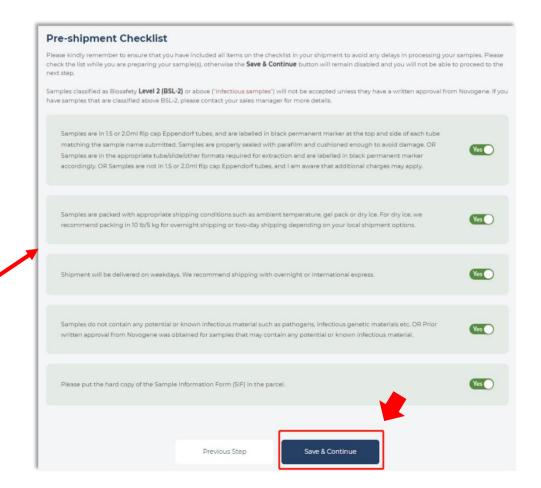


### **About the Samples Shipping Information**



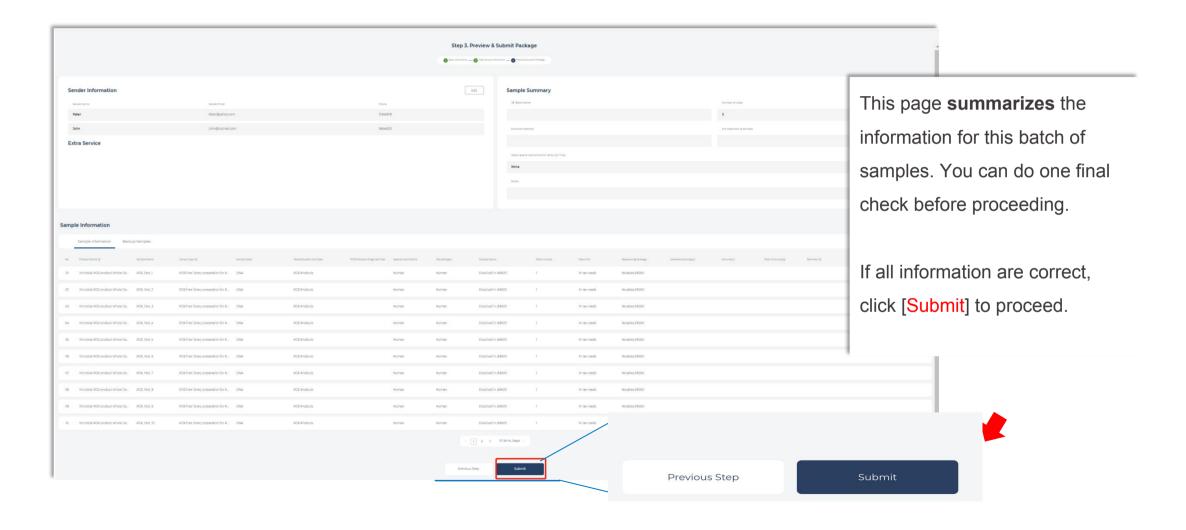
If you already have the tracking number, please help fill in this part. Which could help us login in your samples.

On the **Pre-shipment Checklist**, these items correspond to the send samples policy and instructions. All 5 items **must be followed and selected "Yes"** to proceed to the next step.





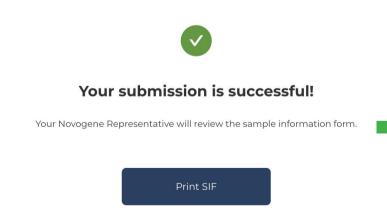
### **Preview and Submit Package**





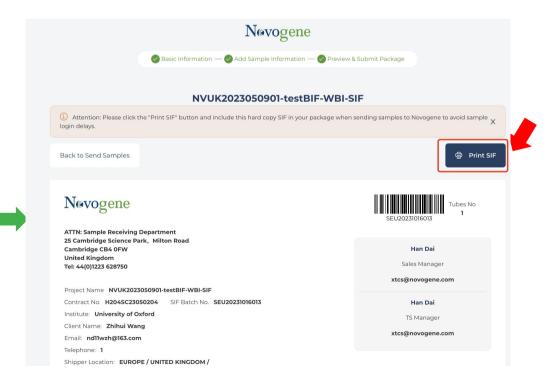


With this, your sample information form is complete! Click [Print SIF] to proceed.



Please **place a hardcopy** of this form with your samples and before sending out your package.

Click [Print SIF].





### **Print Sample Information Form**

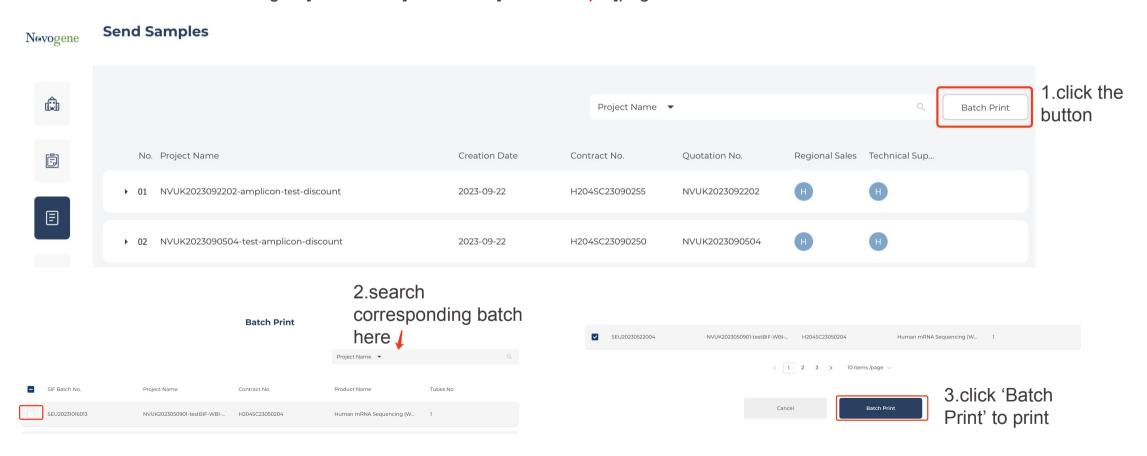
You can also Print SIF accoding to ic Print] on [Send Sample]page.

<b>▼</b> 02	NVUK2023032205-UKI-Nucleome-Lane-F	PE150-2111	2023-03-22	H204SC230307	13 NVUK20230322	205 H	Review
*	Product Name  Mouse Whole Exome Sequencing (WBI)		2023-03-22				Copy 由 Print
	• Z230403003(3)	Submitted By  Novogene		Created Date 2023-04-03	Sample Status  Approved	Submitted Date 2023-04-03	$\left[ \times \right]$
	• Z230403002(3)	Submitted By  Novogene		Created Date 2023-04-03	Sample Status  Approved	Submitted Date 2023-04-03	



### **Print Sample Information Form**

You can also Print SIF according to [Batch Print] button on [Send Sample] page.







### Nevogene

#### NOVOGENE (UK) COMPANY LIMITED

Second Floor, 25 Cambridge Science Park Milton Road Cambridge CB4 0FW, United Kingdom

+44 (0)1223628750

Project Name HU\_SOB\_PE150\_laneseq\_WOBI

Contract No. H204SC21091223 SIF Batch No. SEU20210909004

Institute: SegOmics Biotechnológia Ltd

Client Name: István Nagy

Email: nagyi@seqomics.hu失效失效

Telephone: 36304276152

Shipper Location: EUROPE / HUNGARY / Tracking No. (if available): 774707311011 Transportation Condition: Ice Pack



#### **Tibor Szekere**

Sales Manager

xtcs@novogene.com

Li Li

TS Manager xtcs@novogene.com

#### Sample Information

No.	Library Name (name on tube)	Library Type	Library Status	Remark
01	MEOESZ1	Premade-DNA Small Insert Size Library (Animal or Plant)	Others	
02	MEOESZ2	Premade-DNA Small Insert Size Library (Animal or Plant)	Others	

This is the SIF style we printed out and needs to be shipped to us along with the package.

On this SIF, we can see the batch NO., Tube NO. and sample detail information. All of the information are important for us to claim your samples. So please make sure to print this file instead of excel upload template.



#### Sample Preparation Instruction

#### A. Sample Preparation

#### ☐ 1. Sample Name

Sample name must begin with alphabet and in maximum of 8 characters. Naming in alphanumeric and underscore are acceptable. Non-standard names will be amended following this naming requirements.



#### 2. Sample Marking

Write sample names at the top and the side of each tube with black permanent marker. Sample names on the tube must match with sample information form. Please do not include other information on the tube to avoid discrepancy.

#### 3. Sample Tube

Send samples in 1.5 ml or 2.0 ml microcentrifuge tubes. 96-well plates and PCR tubestrips cannot be accepted at this time. Incorrect tubes may risk processing delays and handlings fees.

#### 4. Sample Protection

Seal each tube with parafilm before packaging. To avoid crushing, we highly recommend placing the sample tubes in a container such as a 50ml tube or cryobox. Cotton or absorbent papers can be used to prevent tubes from moving inside the container.



#### ☐ 5. Sample Requirements

Please refer to your quotation for the sample requirements. Insufficient sample amount may risk processing delay.



#### ☐ 6. Samples Restriction

In principal, we do not accept blood-borne pathogens, purified viral genomes, and infectious samples classified at biosafety level 2 or above. Please contact us before shipping infectious samples. If you are sending samples derived from animals and plants other than human specimens and experimental animals, you will need to include a confirmation sheet in advance. Details will be provided separately.

#### B. Packing and Shipping Samples

#### ☐ 1. Check

Pack the samples according to the Sample Information Form. If you are sending multiple projects, please pack them separately.

#### 2. Print

Attach a hard copy of Sample Information Form in the sample bag.

#### 3. Notify

Please email Sample Information Form and the courier tracking number to our Technical Support Representative in-charge prior to shipping samples. <u>Samples shipped without Sample Information will be delayed in processing.</u>

#### 4. Ship

Ship samples in appropriate shipping condition such as ambient, gel pack or dry ice. For dry ice, we recommend to pack in 10kg for two-day shipping depends on your shipping location. <u>Please avoid weekend shipment or delivery.</u>



### **Special note**

- 1.Due to inaccuracies or omissions in the provided index sequences, any re-sequencing costs incurred as a result will be the responsibility of the customer. We kindly request that you verify the correctness of all index information for your sublibraries, ensuring that the correct index sequences are used. In the event of contamination or suboptimal output caused by index-related issues, additional re-sequencing fees may apply.
- 2. For partial lane sequencing, we will pool different libraries in one lane, so usually there is no need to add phix.

### **FAQ**



1.If I get a error to reminder me the number of tubes I have filled are not matched with the samples uploaded, how to treat?

Check if you have uploaded all the samples successfully, if yes, then you can check if you have filled in the wrong tube numbers on previous step.



### **Attention**

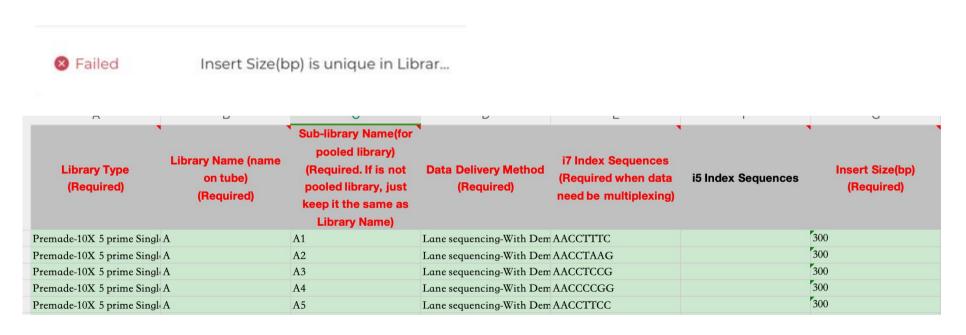
The number of tubes you have filled are not matched with the samples uploaded. Please modify the tube numbers or revise the SIF to make sure these two numbers are the same, so that you can submit it.

Check Again



### 2.If I get a error to reminder me the intersize is unique in the library, what should I do?

Please make sure to fill in the same number on SIF for all the sub-libraries of the same library.







#### 3.If I get an error to reminder me the index have issues, what should I do?

This is just a warning to double check the indexes. If these are correctly written, you can ignore this message.

#### **Checking Reminder**

Mixed Complementary Sequence

Please ensure that only the forward i7 index sequence is provided. According to our index database, we have found both forward and reverse complementary index sequences of the following samples: Mixed Complementary Sequence

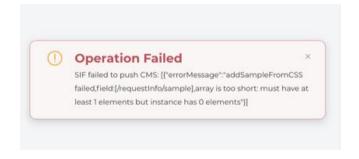
Please ensure that only the forward iS index sequence is provided. According to our index database, we have found both forward and reverse complementary index sequences of the following samples:





### 4.If I get an error to reminder that the SIF can't push to CMS or Failed to upload, waht should I do?

You may used the wrong excel file. Please download the updated excel template in the step 2.



# Thanks!