

BIOC*4540 Enzymology

Winter 2020 Section(s): C01

Department of Molecular and Cellular Biology Credit Weight: 0.75 Version 1.00 - December 17, 2019

1 Course Details

1.1 Calendar Description

This is a laboratory-intensive course where the topics studied include enzyme active sites and the mechanisms of enzyme action; enzyme kinetics and regulation; recombinant proteins and site-directed mutagenesis as tools for understanding enzymes.

Pre-Requisites: BIOC*3560 (may be taken concurrently), BIOC*3570

1.2 Course Description

This is a required Biochemistry fourth-year course on the subject of Enzyme Structure, Function and Mechanism. It features a laboratory component (5 laboratory modules) and an Independent Study where the students research an enzyme of choice and present a Powerpoint seminar with a partner.

1.3 Timetable

- Lectures: Tuesday and Thursday @ 10:00 11:20 in ANNU 156
- Laboratory sections will be held on Mon, Tue, Wed, and Thu from 14:30 17:20 in SSC 3101.

1.4 Final Exam

Exam time and location is subject to change. Please see WebAdvisor for the latest information. The Final Examination will be held on April 16 from 11:30 - 13:30. The location will be determined near the end of the W2020 semester.

2 Instructional Support

2.1 Instructional Support Team

Instructor: Rod Merrill

Email: rmerrill@uoguelph.ca **Telephone:** +1-519-824-4120 x53806

Office: SSC 2250

Office Hours: Tuesday and Thursday @ 1:00 – 2:30 pm or by appointment

Lab Co-ordinator:Catrien Bouwman M.Sc.Email:cbouwman@uoguelph.ca

Office: SSC 3503

2.2 Teaching Assistants

Teaching Assistant: Samantha Wear swear@uoguelph.ca

Office Hours: Samantha will function as the Seminar Coordinator for the

Course. She will also give the PyMOL tutorial and serve as a

resource person for the seminars.

Teaching Assistant: Liam Doyle

Email: Idoyle03@uoguelph.ca

Office Hours: Liam will work as a Laboratory Teaching Assistant for the

course.

Teaching Assistant: Carys Jones

Email: carys@uoguelph.ca

Office Hours: Carys will work as a Laboratory Teaching Assistant for the

course.

3 Learning Resources

3.1 Required Resources

Lab Manual (Lab Manual)

Laboratory manuals must be purchased from the Department of Molecular and Cellular Biology during dates and times to be announced.

The cost of the manual is \$10.00 and please bring correct change.

3.2 Recommended Resources

Principles in Biochemistry (Readings)

 No single textbook is sufficient for the lecture material but Lehninger: Principles in Biochemistry (Lehninger 5th, 6th, or 7th editions) Chapter 6 serves as the basis for basic enzyme understanding and theory and this chapter should be read and carefully studied.

On reserve

3.3 Additional Resources

Structure and Mechanism in Protein Science: A Guide to Enzyme Catalysis and Protein Folding, Series in Structural Biology (Textbook)

Alan Fersht (2017) Structure and Mechanism in Protein Science: A Guide to Enzyme Catalysis and Protein Folding, Series in Structural Biology – vol. 9 (2017), World Scientific Co. Pte. Ltd, Hakensack, New Jersey.

· on reserve

Enzymes: Biochemistry, Biotechnology and Clinical Chemistry (Textbook)

Trevor Palmer (2007) Enzymes: Biochemistry, Biotechnology and Clinical Chemistry, 2nd edition, Albion Press.

· on reserve

Introduction to Protein Structure (Textbook)

Carl Branden & John Tooze (1999) Introduction to Protein Structure, 2nd edition, Garland Publ., New York.

on reserve

Introduction to Proteins: Structure, Function, and Motion (Textbook)

Amit Kessel, Nir Ben-Tal (2018) Introduction to Proteins: Structure, Function, and Motion, 2 nd Edition (Chapman and Hall, CRC; published April 11, 2018)

on reserve

3.4 Note

- A number of related texts have been placed on reserve as resources and to provide background information on the various topics discussed in the course (see Course Subject Outline).
- The Adobe Acrobat (*.pdf) files for each Powerpoint lecture will be available for download from the Courselink website and each lecture will be made available at least 2 weeks before the specified lecture date.

3.4 Additional Texts

 All indicated additional texts, papers and treatises are available at the Reserve Desk at the library on two hour loan.

4 Learning Outcomes

Objectives: (i) To integrate the practical aspects of enzymology with the kinetic theories to provide a mechanistic overview of enzyme activity and regulation in cells; and (ii) to prepare students to confidently and competently work with enzyme systems in both Academia and Industry.

4.1 Course Learning Outcomes

By the end of this course, you should be able to:

- 1. Plan and execute an enzyme assay
- 2. Analyse and plot enzyme kinetic data and study the effect of pH on the kinetic activity of the enzyme.
- 3. Learn how to use Bioinformatic tools and software, including PyMOL to study and analyze enzyme structure and function.
- 4. Perform literature research on a specific enzyme topic
- 5. Prepare and deliver a Powerpoint seminar to your peers

5 Teaching and Learning Activities

5.1 Course Subject Outline

- I. ENZYMES AS CATALYSTS (Lehninger Ch 6; Fer Ch 2, 3, 4, 6; Palm Ch 1, 6, 8, 16)
 - · Lect#1: Introduction and History of Enzymes

Historical aspects

Discovery of enzymes

Chemistry of enzymes

Function and importance

Enzymes in biotechnology

Lect#2: Enzyme Purification and Assay

Initial velocity measurements

Assay types

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Enzyme units of activity
        Turnover number and properties
        Purification and assessment
        Methods for measurements
· Lect#3: Michaelis-Menten Kinetics
        Introduction
        Assumptions
        Derivation
        Description of vo versus [So]
        Michaelis constant (K<sub>M</sub>)
        Specificity/substrate constant (SpC)
• Lect#4: Graphical Analysis of Kinetic Data, pH, Temp Dependence and Allosteric
  Enzymes
        Graphical analysis
              Lineweaver-Burk Analysis
              Hanes-Woolf Analysis
              Eadie-Hofstee Analysis
              Direct Linear Plot (Eisenthal/Cornish-Bowden Plot)
              Nonlinear Curve Fitting
        pH-dependence of Michaelis-Menten Enzymes
        Temperature-dependence of enzyme reactions
        Allosteric enzymes
· Lect#5: Enzyme Classification, Characteristics and Properties
        Classification
        Catalytic power and specificity
        Enzymes as catalysts
        Enzyme - substrate interactions
              lock & key model
              induced fit model
              transition state model
              quantum tunnelling model
        Enzymes as proteins
        Enzyme cofactors
· Lect#6: Enzyme Inhibition and Kinetics
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Classification of Inhibitors
Reversible
Irreversible
Iodoacetamide
DIFP

Suicide substrates

Classification of Reversible Inhibitors

Competitive

Uncompetitive

Noncompetitive

Substrate

Lect#7: Single Molecule Enzymology

Movies of single enzymes

Advantages of Single Molecule Studies

Applications of Single Molecule Studies

Following enzymes in real time

ATP Synthase with tethered actin

ATP Synthase mechanism

Myosin-V

Kinesin motor attached to a fluorescent bead

Single Molecule Studies of Cholesterol Oxidase

ß-galactosidase: a model Michaelis-Menten enzyme?

· Lect#8: Multi-substrate Reactions and Substrate Binding Analysis

Multi-substrate reactions

Cleland convention

Ordered and random mechanisms

Sequential and nonsequential mechanisms

Sequential

Nonsequential

Substrate Binding Analysis

Single Binding Site Model

Binding Data Plots

Direct Plot

Reciprocal Plot

Scatchard Plot

Determination of Enzyme-Substrate Dissociation Constants

Kinetics

Equilibrium Dialysis

Equilibrium Gel Filtration

Ultracentrifugation

Spectroscopic Methods

II. MECHANISM OF ENZYME CATALYSIS (Lehninger Ch 6; Fer Ch 2, 9; Palm Ch 10, 11)

Lect#9: Enzyme Mechanisms-I

Reaction Mechanisms and Catalysis

Enzyme-transition state complementarity

Structure-activity correlations

Transition state analogues

Catalytic antibodies

Summary

Preferential transition state binding

Transition state theory

Proximity effect

Acid-base catalysts

· Lect#10: Enzyme Mechanisms II

Covalent catalysis

Metal ion catalysis

Electrostatic catalysis

Low barrier H-bonds

Structural flexibility

• Lect#11: Enzyme Mechanisms-III: Techniques for Drug Discovery

Drug Design

Techniques of Drug Discovery

Complexity of Drug Discovery

SARS and QSARS

Structure-based Drug Design

Combinatorial Chemistry and High-Throughput Screening

Introduction to Pharmacology

Pharmocokinetics

Toxicity and Adverse Reactions Eliminate Most Drug Candidates

Phase I

Phase II

Phase III

Drug Candidate Statistics

Cytochrome P450 Metabolizes Drugs

Many Drugs are Enzyme Inhibitors

Sulfadrugs

Viagara

· Lect#12: Active Site Investigations I

Kinetic Studies

Variation of substrate concentration

Variation of substrate structure

Reversible inhibition

Variation of pH

Pre-steady state kinetics

Detection of Intermediates

X-ray Crystallographic Studies

NMR for Protein Structure Determination

- Lect#13: MID-TERM EXAMINATION (during class time) Tue, Feb 25, 2020
- Lect#14: Active-Site Investigations II

Chemical modifications of active-site side-chains

applications

Super-reactive side-chains

Suicide substrates

Interpretation of chemical modification experiments

Criteria for establishment of side-chain involvement in catalysis

Lect#15: Enzyme Engineering and Design

Substitution

Insertion

Hybrid Proteins

Genes for Novel Enzymes

Enviropig

Engineering More Stable Enzymes

Incorporation of Non-natural Amino Acids into Enzymes

Protein Engineering by Combinatorial Methods

DNA Shuffling

III. CASE STUDY ENZYMES/INDEPENDENT PROJECT/SEMINAR (Starting Tue, Mar 3, 2020)

- Lect#16: Student Presentations (n = 3 presentations, groups of 2 students)
- Lect#17: Student Presentations (n = 3 presentations, groups of 2 students)
- Lect#18: Student Presentations (n = 3 presentations, groups of 2 students)
- Lect#19: Student Presentations (n = 3 presentations, groups of 2 students)
- Lect#20: Student Presentations (n = 3 presentations, groups of 2 students)
- Lect#21: Student Presentations (n = 3 presentations, groups of 2 students)
- Lect#22: Student Presentations (n = 3 presentations, groups of 2 students)
- Lect#23: Student Presentations (n = 3 presentations, groups of 2 students)
- Lect#24: Course Review

Fer = Ferst; Lehninger = Lehninger 7th ed., Palm = Palmer

6 Assessments

6.1 Marking Schemes & Distributions

Name	Scheme A (%)
Mid-term	20
Laboratory SSC 3101	25
Independent Study/Seminar	20
Final Examination	30
Participation	5
Total	100

6.2 Assessment Details

Midterm (20%)

Date: Tue, Feb 25, 10:00 AM, In class

- There will be a Mid-term Examination (80 min, in class-time) involving multiple choice, short answer and problem questions.
- There are no alternate exams offered since the Mid-term will be given in class time.
- The final grade can be based entirely on the Final Exam (and other components) only if reasons for missing the Mid-term exam are adequately documented.
- Both exams are required!
- Students who score a significantly higher grade on the Final Exam, compared with the Mid-term Exam, may receive a higher weighting of the Final Exam (Mid-term: 10%, Final: 40%), at my discretion. A significantly higher grade is one that is 25 percentage points higher.
- Three problem sets will be assigned, which will assist you in understanding and learning the lecture material (quantitative aspects) and which will serve as prototypes for some of the questions on the Mid-term and the Final Examinations.

Laboratory Component (25%)

Date: SSC 3101

The Laboratory Component grade will be composed of a laboratory report and a lab notebook.

Lab periods will begin the week of January 6th during your scheduled day of the week.

Lab Reports will be monitored with anti-plagiarism software (Turnitin)

Independent Study/Seminar (20%)

- Commencing with Lecture#16 (March 5, 2020), we will have three Powerpoint presentations per lecture period with each seminar being a group effort (two students per team). The presentations will be 15 min in length followed by a 5-min question period. Each team member will receive an identical mark for the presentation, including the ability to answer questions-this activity is meant to be a team-effort and not two individuals presenting separately. Therefore, choose your partner wisely since you must work well with your partner to make the best teambased presentation possible! The independent study/seminar is worth 25% (includes 5% for participation) of the course grade. Therefore, it is important to help each other and to work as a team! It is paramount to remember that anything that you include or say during your presentation is open to questions from the audience and, so you should ensure that you fully understand it. A seminar rubric will be used for evaluation of the seminars. Dr. Merrill, Catrien and Samantha will complete evaluation forms on each presentation. The comments/feedback will be given, but not the marks until all the presentations have been completed. A grade (5% of your seminar mark) will also be given for seminar preparation (following the time-line, not last minute) and participation in the question period (details to follow later).
- Students must form a two-person team by Jan 17th, 2020 and the team must decide upon a case-study enzyme for their presentation and clear the topic with Dr. Merrill by Jan 24th, 2020 (4 pm). The lecture/seminar dates are: Mar 5, 10, 12, 17, 19, 24, 26, and 31. Catrien/Samantha will schedule the presentation dates for all the teams through Courselink. Some research and preparation time will be given during the Enzymology lab sessions (see Catrien/Samantha for details). If you are unable to present your seminar on the scheduled date, a separate written project/assignment will be given.

Participation (5%)

Final Examination (30%)

Date: TBA

- The examination component of the Final Grade can be based entirely on the Final Exam only if reasons for missing the Mid-term exam are adequately documented.
- Both exams are required!
- The Final Exam is cumulative and will cover all lectures, including the research seminars.
- Students who score a significantly higher grade on the Final Exam, compared with the Mid-term Exam, may receive a higher weighting of the Final Exam (Mid-term: 10%, Final: 40%), at my discretion. A significantly higher grade is one that is 25 percentage

- points higher.
- Three problem sets will be assigned but will not be graded. The problem sets will
 assist you in understanding and learning the lecture material (quantitative aspects)
 and will serve as prototypes for some of the questions on the Mid-term and the Final
 Examinations.

7 Course Statements

7.1 Instructor & Course Evaluation

As part of the evaluation process in the Department of Molecular and Cellular Biology, written comments on the Course and/or the Instructors' teaching performance may be sent to the Chair, Department of Molecular and Cellular Biology, at any time. Such letters must be signed. Departmental Evaluations will also be conducted near the end of the semester. Copies of evaluations will be made available to the Instructor after submission of the final grade.

7.2 Course Add and Drop

Notification is not needed for dropping the course before the DROP deadline (last day of class, Apr 3, 2020). Program approval is only needed for drops and adds if your category is "Special" or "Provisional".

8 Department of Molecular and Cellular Biology Statements

8.1 Academic Advisors

If you are concerned about any aspect of your academic program:

Make an appointment with a program counsellor in your degree program. <u>B.Sc.</u>
 Academic Advising or Program Counsellors

8.2 Academic Support

If you are struggling to succeed academically:

 Learning Commons: There are numerous academic resources offered by the Learning Commons including, Supported Learning Groups for a variety of courses, workshops related to time management, taking multiple choice exams, and general study skills.
 You can also set up individualized appointments with a learning specialist.

- http://www.learningcommons.uoguelph.ca/
- Science Commons: Located in the library, the Science Commons provides support for physics, mathematic/statistics, and chemistry. Details on their hours of operations can be found at: http://www.lib.uoguelph.ca/get-assistance/studying/chemistry-physicshelp and http://www.lib.uoguelph.ca/get-assistance/studying/math-stats-help

8.3 Wellness

If you are struggling with personal or health issues:

- Counselling services offers individualized appointments to help students work through personal struggles that may be impacting their academic performance. https://www.uoguelph.ca/counselling/
- Student Health Services is located on campus and is available to provide medical attention. https://www.uoguelph.ca/studenthealthservices/clinic
- For support related to stress and anxiety, besides Health Services and Counselling Services, Kathy Somers runs training workshops and one-on-one sessions related to stress management and high performance situations. http://www.selfregulationskills.ca/

9 University Statements

9.1 Email Communication

As per university regulations, all students are required to check their e-mail account regularly: e-mail is the official route of communication between the University and its students.

9.2 When You Cannot Meet a Course Requirement

When you find yourself unable to meet an in-course requirement because of illness or compassionate reasons please advise the course instructor (or designated person, such as a teaching assistant) in writing, with your name, id#, and e-mail contact. The grounds for Academic Consideration are detailed in the Undergraduate and Graduate Calendars.

Undergraduate Calendar - Academic Consideration and Appeals https://www.uoguelph.ca/registrar/calendars/undergraduate/current/c08/c08-ac.shtml

Graduate Calendar - Grounds for Academic Consideration https://www.uoguelph.ca/registrar/calendars/graduate/current/genreg/index.shtml

Associate Diploma Calendar - Academic Consideration, Appeals and Petitions https://www.uoguelph.ca/registrar/calendars/diploma/current/index.shtml

9.3 Drop Date

Students will have until the last day of classes to drop courses without academic penalty. The deadline to drop two-semester courses will be the last day of classes in the second semester. This applies to all students (undergraduate, graduate and diploma) except for Doctor of Veterinary Medicine and Associate Diploma in Veterinary Technology (conventional and alternative delivery) students. The regulations and procedures for course registration are available in their respective Academic Calendars.

Undergraduate Calendar - Dropping Courses https://www.uoguelph.ca/registrar/calendars/undergraduate/current/c08/c08-drop.shtml

Graduate Calendar - Registration Changes https://www.uoguelph.ca/registrar/calendars/graduate/current/genreg/genreg-reg-regchg.shtml

Associate Diploma Calendar - Dropping Courses https://www.uoguelph.ca/registrar/calendars/diploma/current/c08/c08-drop.shtml

9.4 Copies of Out-of-class Assignments

Keep paper and/or other reliable back-up copies of all out-of-class assignments: you may be asked to resubmit work at any time.

9.5 Accessibility

The University promotes the full participation of students who experience disabilities in their academic programs. To that end, the provision of academic accommodation is a shared responsibility between the University and the student.

When accommodations are needed, the student is required to first register with Student Accessibility Services (SAS). Documentation to substantiate the existence of a disability is required; however, interim accommodations may be possible while that process is underway.

Accommodations are available for both permanent and temporary disabilities. It should be noted that common illnesses such as a cold or the flu do not constitute a disability.

Use of the SAS Exam Centre requires students to book their exams at least 7 days in advance and not later than the 40th Class Day.

For Guelph students, information can be found on the SAS website https://www.uoguelph.ca/sas

For Ridgetown students, information can be found on the Ridgetown SAS website https://www.ridgetownc.com/services/accessibilityservices.cfm

9.6 Academic Integrity

The University of Guelph is committed to upholding the highest standards of academic integrity, and it is the responsibility of all members of the University community-faculty, staff, and students-to be aware of what constitutes academic misconduct and to do as much as

possible to prevent academic offences from occurring. University of Guelph students have the responsibility of abiding by the University's policy on academic misconduct regardless of their location of study; faculty, staff, and students have the responsibility of supporting an environment that encourages academic integrity. Students need to remain aware that instructors have access to and the right to use electronic and other means of detection.

Please note: Whether or not a student intended to commit academic misconduct is not relevant for a finding of guilt. Hurried or careless submission of assignments does not excuse students from responsibility for verifying the academic integrity of their work before submitting it. Students who are in any doubt as to whether an action on their part could be construed as an academic offence should consult with a faculty member or faculty advisor.

Undergraduate Calendar - Academic Misconduct https://www.uoguelph.ca/registrar/calendars/undergraduate/current/c08/c08-amisconduct.shtml

Graduate Calendar - Academic Misconduct https://www.uoguelph.ca/registrar/calendars/graduate/current/genreg/index.shtml

9.7 Recording of Materials

Presentations that are made in relation to course work - including lectures - cannot be recorded or copied without the permission of the presenter, whether the instructor, a student, or guest lecturer. Material recorded with permission is restricted to use for that course unless further permission is granted.

9.8 Resources

The Academic Calendars are the source of information about the University of Guelph's procedures, policies, and regulations that apply to undergraduate, graduate, and diploma programs.

Academic Calendars https://www.uoguelph.ca/academics/calendars