Tutorial: testing microbiome mediation effect using miMediation

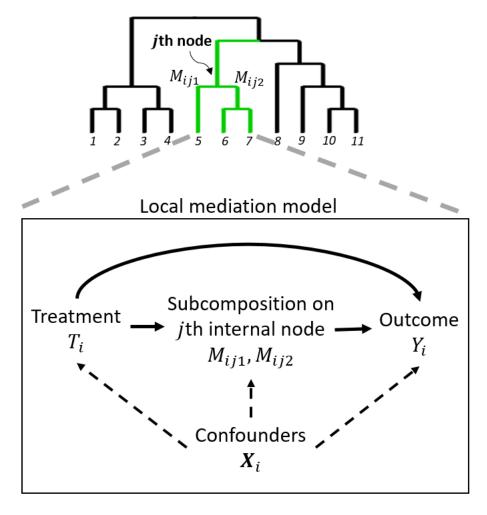
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This is a practical tutorial on the use of miMediation package, which introduces a phylogeny-based mediation test (PhyloMed) for high-dimensional microbial composition mediators. The methodology is described in detail in the Hong, Chen, and Tang (Manuscript).

A brief summary of the PhyloMed

PhyloMed models microbiome mediation effect through a cascade of independent local mediation models of subcompositions on the internal nodes of the phylogenetic tree. Each local model captures the mediation effect of a subcomposition at a given taxonomic resolution. The method improves the power of the mediation test by enriching weak and sparse signals across mediating taxa that tend to cluster on the tree. PhyloMed enables us to test the overall mediation effect of the entire microbial community and pinpoint internal nodes with significant subcomposition mediation effects.



As depicted in the figure above, we propose to construct a local mediation model for the subcomposition at each internal node of the phylogenetic tree. The subcomposition on a given internal node consists of the relative abundance aggregated at its two child nodes. We apply the following robust linear regression model and generalized linear regression model to represent the causal path diagram of the local mediation model at the *j*th internal node

$$E\left\{\log\left(\frac{M_{ij1}}{M_{ij2}}\right)\right\} = \alpha_{jX}^{T}\mathbf{X}_{i} + \alpha_{j}T_{i}$$
$$g\{E(Y_{i})\} = \beta_{jX}^{T}\mathbf{X}_{i} + \beta_{jT}T_{i} + \beta_{j}\log\left(\frac{M_{ij1}}{M_{ij2}}\right)$$

where $g(\cdot)$ is the link function depending on the type of the outcome and we omit the intercept term in both models as it can be absorbed into \mathbf{X}_i .

Under potential outcome framework (VanderWeele (2015)) and assumptions of no unmeasured confounding variables, it leads to the null hypothesis

$$H_0^j = \alpha_j \beta_j = 0$$

, which is equivalent to the union of three disjoint component null hypotheses

$$H_{00}^j: \alpha_j = \beta_j = 0,$$
 (1)

$$H_{10}^j: \alpha_i \neq 0, \beta_i = 0,$$
 (2)

$$H_{01}^j: \alpha_j = 0, \beta_j \neq 0.$$
 (3)

We define the mediation test statistic for H_0^j as

$$P_{\max_i} = \max(P_{\alpha_i}, P_{\beta_i})$$

The P_{α_j} , P_{β_j} could be calculated via asymptotic approach or adaptive permutation approach when sample size is small.

In fact, P_{\max_j} follows a mixture distribution with three components, each of which corresponds to one type of null hypothesis H_{00}^j , H_{10}^j , H_{01}^j . The *p*-value of mediation test in the *j*th local model is given by

$$Pr(P_{\max_{j}} \leq p_{\max_{j}}) = \pi_{00}p_{\max_{j}}^{2} + \pi_{10}p_{\max_{j}}Pr(P_{\alpha_{j}} \leq p_{\max_{j}} \mid \alpha_{j} \neq 0) + \pi_{01}p_{\max_{j}}Pr(P_{\beta_{j}} \leq p_{\max_{j}} \mid \beta_{j} \neq 0)$$

In this formula, we need to estimate three component probabilities: π_{00} , π_{10} , and π_{01} , and two power functions evaluated at p_{\max_j} : $Pr(P_{\alpha_j} \leq p_{\max_j} \mid \alpha_j \neq 0)$ and $Pr(P_{\beta_j} \leq p_{\max_j} \mid \beta_j \neq 0)$. There are two various methods (product, maxp) to estimate three component probabilities. Both are derived from JC's method (Jin and Cai (2007)), which uses the empirical characteristic function and Fourier analysis to estimate $\pi_{0\bullet}$ (the proportion of null $\alpha_j = 0$) and $\pi_{\bullet 0}$ (the proportion of null $\beta_j = 0$).

- "product" method: The estimates of π_{00} , π_{10} , π_{01} are $\hat{\pi}_{00} = \hat{\pi}_{0\bullet}\hat{\pi}_{\bullet 0}/\hat{\pi}_{0}$, $\hat{\pi}_{10} = (1 \hat{\pi}_{0\bullet})\hat{\pi}_{\bullet 0}/\hat{\pi}_{0}$, and $\hat{\pi}_{01} = \hat{\pi}_{0\bullet}(1 \hat{\pi}_{\bullet 0})/\hat{\pi}_{0}$, where $\hat{\pi}_{0} = \hat{\pi}_{0\bullet} + \hat{\pi}_{\bullet 0} \hat{\pi}_{0\bullet}\hat{\pi}_{\bullet 0}$.
- "maxp" method: The estimates of π_{00} , π_{10} , π_{01} are $\hat{\pi}_{00} = (\hat{\pi}_{0\bullet} + \hat{\pi}_{\bullet 0} \hat{\pi}_{0})/\hat{\pi}_{0}$, $\hat{\pi}_{10} = (\hat{\pi}_{0} \hat{\pi}_{0\bullet})/\hat{\pi}_{0}$, and $\hat{\pi}_{01} = (\hat{\pi}_{0} \hat{\pi}_{\bullet 0})/\hat{\pi}_{0}$, where $\hat{\pi}_{0}$ is obtained from JC's method by using $p_{\max_{i}}$.

After obtaining the p-values on all internal nodes, we apply Benjamini-Hochberg (BH) false discovery rate procedure (Benjamini and Hochberg (1995)) to identify a collection of nodes on the phylogenetic tree with significant mediation effects. To test the global mediation null hypothesis $H_0: \cap_{j=1}^J H_0^j$, we apply the harmonic mean p-value (HMP) method (Wilson (2019)) to combine local mediation p-values.

Specifically, the weighted harmonic mean of subcomposition mediation test p-values p_1, \ldots, p_J is defined as

$$\mathring{p} = \frac{\sum_{j=1}^{J} w_j}{\sum_{j=1}^{J} w_j p_j},$$

where w_j 's are weights that sum to 1 and we set $w_j = 1/J$ by default. The global test *p*-value can be obtained by calculating the tail probability from the \mathring{p} 's null distribution approximation.

Application with phylognetic information: Cecal data

It is well-known that low dose antibiotics have been used widely to stimulate weight gain in livestock. However, there is growing concern that antibiotic exposure may have long-term consequences. Several studies have shown that antibiotics can have great impact on the abundances of bacteria in the gut community. It is interesting to investigate whether the subtherapeutic antibiotic treatment effect on body weight is mediated through the perturbation of gut microbiome and study the underlying mechanisms.

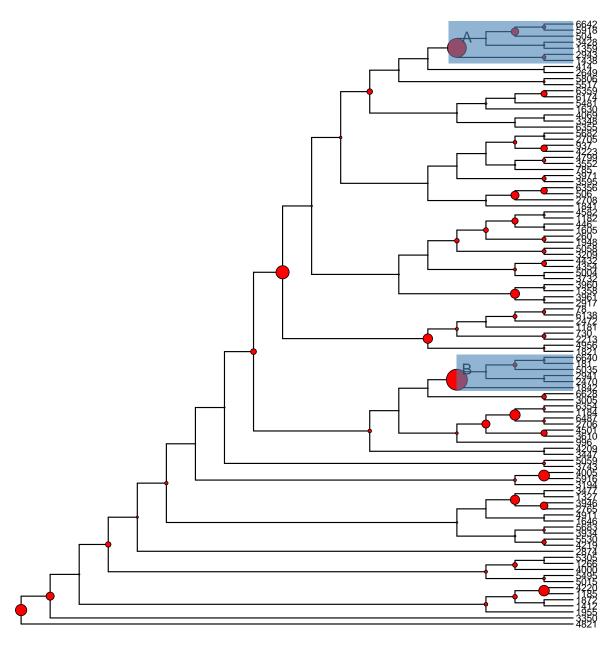
The data here is from an experiment conducted by Cho et al. (2012), in which young mice were treated by different low-dose antibiotic and evaluated changes in body fat and compositions of the microbiome in cecal and fecal samples. The mice in antibiotic group were heavier than those in the control group. We will show how to perform phyloMed function by focusing on cecal samples.

```
> library(miMediation)
> # Load data
> data(data.cecal)
> # Take a look at the data
> Trt <- data.cecal$treatment
> table(Trt) # 0: control 1: antibotics
Trt
0 1
```

```
> M <- data.cecal$mediators
> head(M[,1:6])
          3732 5004 4354 4432 3209 5058
cecal C1
             1
                   2
                       56
                            39
                                  12
                                       13
cecal_C10
             1
                   7
                       60
                            42
                                  34
                                       31
cecal_C2
             9
                   2
                       38
                            40
                                  14
                                        8
cecal_C3
             4
                   4
                       41
                            53
                                  16
                                       18
cecal C4
                   2
                      102
                                       19
             5
                            84
                                  18
cecal_C5
             5
                 13
                       83
                            62
                                  29
                                       29
> Y <- data.cecal$outcome
> summary(Y)
   Min. 1st Qu. Median
                            Mean 3rd Qu.
                                             Max.
  17.20
          20.55
                   21.80
                           22.32
                                    23.38
                                             32.10
> tree <- data.cecal$tree
```

To run phyloMed function, the parameter treatment, mediators, outcome, phylogeny tree and pi.method are required. Other inputs are optional. Note that if n.perm=1e5, the function will output p-value calculated through adaptive permutation procedure as well and it will take ~ 6 minutes to output the result. You can set verbose=TRUE to keep track of the process. Here is an example described in the Hong, Chen, and Tang (Manuscript).

```
> # set random seed here so that you can get the same result every time you run the code
> set.seed(84)
> cecal.rsltlst <- phyloMed(Trt, M, Y, tree, fdr.alpha = 0.1, n.perm = 1e5, graph = TRUE)
> # take a look at phyloseq-class object
> cecal.physeq <- cecal.rsltlst$clean.data
> cecal.physeq
phyloseq-class experiment-level object
              OTU Table:
                                [ 100 taxa and 48 samples ]
otu_table()
sample_data() Sample Data:
                                 [ 48 samples by 2 sample variables ]
              Phylogenetic Tree: [ 100 tips and 99 internal nodes ]
phy_tree()
> cecal.rslt <- cecal.rsltlst$rslt
> # take a look at rslt (PhyloMed.P)
> cecal.rslt$PhyloMed.P
$node.pval
 [1] 0.030249720 0.092335444 0.804658359 0.211165304 0.607828169 0.422528957
 [7] 0.982450877 0.814714539 0.169853475 0.015622998 0.771106344 0.862797494
[13] 0.969594333 0.484577282 0.871291406 0.259366125 0.269493336 0.376023190
[19] 0.269843492 0.176746122 0.748603126 0.634689441 0.328734323 0.075560518
[25] 0.658425404 0.604799852 0.566390083 0.877034873 0.891684145 0.909830655
[31] 0.735645104 0.374101141 0.385234590 0.136706734 0.685643960 0.292098093
[37] 0.775748956 0.091486313 0.154111793 0.197397182 0.862797494 0.603295673
[43] 0.862797494
                          NA 0.001977693 0.372196849 0.785181751 0.710874484
[49] 0.106154293 0.395567605 0.789973394 0.670886620 0.871291406 0.154603991
[55] 0.885773955 0.871291406 0.048660456 0.476565395 0.785181751 0.432264342
[61] 0.431437676 0.350083754 0.697090415 0.352930818 0.785181751 0.925446394
[67] 0.480541455 0.089538035 0.063109834 0.507901457 0.550680818 0.145076919
[73] 0.958103959 0.288488025 0.001168934 0.771106344 0.305193584 0.649771384
[79] 0.782804879 0.495994520 0.486618006 0.034923585 0.699030773 0.674523328
[85] 0.295792162 0.546873927 0.958103959 0.859998027 0.059727124 0.101071936
[91] 0.951414794 0.452994463 0.479541885 0.258716008 0.925446394 0.382561329
[97] 0.311923787 0.877034873 0.039373253
```



The output consists of four components:

- node.pval: mediation p-values on each internal node of the phylogenetic tree.
- sig.clade: identified mediation nodes with their descendants.
- null.prop: estimated proportion of three disjoint component null hypotheses.
- global.pval: global test *p*-value.

In the figure above, the size of the circle on internal node is proportional to $-\log_{10}(\text{subcompostion mediation p-value } p_j)$, where p_j lives in the node.pval output. The identified mediation node is highlighted by a blue rectangle.

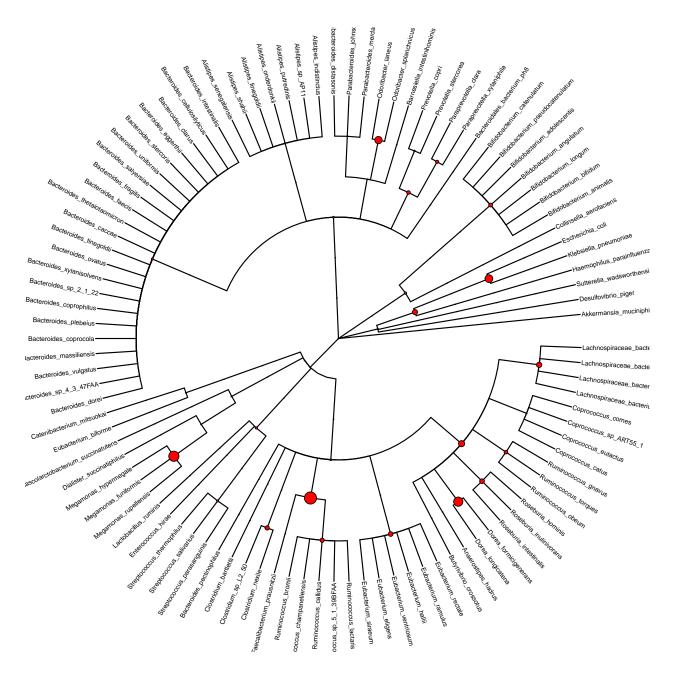
Application with taxonomic information: ZeeviD data

When there is no phylogenetic information available, the phyloMed function could construct taxonomic tree based on the taxonomic information. Here we sampled 200 subjects out of 900 healthy subjects in a real microbiome dataset Zeevi et al. (2015) and divided them into two equal-sized treatment groups. We used the

top 100 most abundant OTUs and the associated taxonomy table to run phyloMed function.

```
> # Load data
> data(data.zeeviD)
> # Take a look at the data
> Trt <- data.zeeviD$treatment
> table(Trt) # 0: control 1: treatment
  0
100 100
> M <- data.zeeviD$mediators
> dim(M)
[1] 200 100
> Y <- data.zeeviD$outcome
> summary(Y)
   Min. 1st Qu. Median
                           Mean 3rd Qu.
-8.1661 -2.1830 -0.1615 0.4094 2.1486 8.8780
> tree <- data.zeeviD$tree</pre>
> head(tree)
Taxonomy Table:
                    [6 taxa by 7 taxonomic ranks]:
                                       Kingdom
                                                  Phylum
                                       "Bacteria" "Firmicutes" "Negativicutes"
s__Megamonas_hypermegale
s__Megamonas_funiformis
                                       "Bacteria" "Firmicutes" "Negativicutes"
s__Megamonas_rupellensis
                                       "Bacteria" "Firmicutes" "Negativicutes"
s__Phascolarctobacterium_succinatutens "Bacteria" "Firmicutes" "Negativicutes"
                                       "Bacteria" "Firmicutes" "Negativicutes"
s__Dialister_succinatiphilus
                                       "Bacteria" "Firmicutes" "Clostridia"
s__Ruminococcus_bromii
                                                          Family
s__Megamonas_hypermegale
                                       "Selenomonadales" "Veillonellaceae"
                                        "Selenomonadales" "Veillonellaceae"
s__Megamonas_funiformis
s__Megamonas_rupellensis
                                       "Selenomonadales" "Veillonellaceae"
s_Phascolarctobacterium_succinatutens "Selenomonadales" "Acidaminococcaceae"
                                        "Selenomonadales" "Veillonellaceae"
s__Dialister_succinatiphilus
                                                        "Ruminococcaceae"
s__Ruminococcus_bromii
                                       "Clostridiales"
                                       Genus
s__Megamonas_hypermegale
                                       "Megamonas"
s__Megamonas_funiformis
                                        "Megamonas"
s__Megamonas_rupellensis
                                        "Megamonas"
s__Phascolarctobacterium_succinatutens "Phascolarctobacterium"
s__Dialister_succinatiphilus
                                       "Dialister"
s__Ruminococcus_bromii
                                        "Ruminococcus"
                                       Species
s__Megamonas_hypermegale
                                        "Megamonas_hypermegale"
s__Megamonas_funiformis
                                        "Megamonas_funiformis"
s__Megamonas_rupellensis
                                        "Megamonas_rupellensis"
s_Phascolarctobacterium_succinatutens "Phascolarctobacterium_succinatutens"
s__Dialister_succinatiphilus
                                        "Dialister_succinatiphilus"
s__Ruminococcus_bromii
                                        "Ruminococcus_bromii"
> # only show aysmptotic result
> demo.rsltlst <- phyloMed(Trt, M, Y, tree, graph = TRUE)
> # take a look at phyloseq-class object
> demo.physeq <- demo.rsltlst$clean.data</pre>
> demo.physeq
phyloseq-class experiment-level object
```

```
OTU Table:
                                  [ 100 taxa and 200 samples ]
otu_table()
                                  [ 200 samples by 2 sample variables ]
sample_data() Sample Data:
                                  [ 100 taxa by 7 taxonomic ranks ]
tax_table()
              Taxonomy Table:
> demo.rsltlst$rslt$PhyloMed.A
$node.pval
             Genus.Alistipes
                                         Genus.Bacteroides
                  0.77601346
                                                0.63563292
       Genus.Bifidobacterium
                                             Genus.Blautia
                  0.35594013
                                                0.38486173
           Genus.Clostridium
                                         Genus.Coprococcus
                  0.30845559
                                                0.85310122
                 Genus.Dorea
                                         Genus.Eubacterium
                  0.05920649
                                                0.26856782
Genus.Lachnospiraceae_noname
                                           Genus.Megamonas
                  0.21775102
                                                0.04293845
           Genus.Odoribacter
                                     Genus.Parabacteroides
                  0.11264747
                                                0.96369414
                                          Genus.Prevotella
        Genus.Paraprevotella
                  0.44047726
                                                0.73153056
             Genus.Roseburia
                                        Genus.Ruminococcus
                  0.26436600
                                                0.30956110
         Genus.Streptococcus
                                 Family.Enterobacteriaceae
                  0.63083981
                                                0.09572732
  Family.Erysipelotrichaceae
                                    Family.Lachnospiraceae
                  0.87936798
                                                0.16469013
  Family.Porphyromonadaceae
                                     Family.Prevotellaceae
                   1.0000000
                                                0.38652436
      Family.Ruminococcaceae
                                    Family. Veillonellaceae
                  0.02260512
                                                0.89786580
         Order.Bacteroidales
                                       Order.Clostridiales
                  0.69673427
                                                0.67296673
       Order.Lactobacillales
                                     Order.Selenomonadales
                  0.58618988
                                                0.81632370
        Class.Actinobacteria
                                 Class.Gammaproteobacteria
                  0.56921384
                                                0.23213703
           Phylum.Firmicutes
                                     Phylum.Proteobacteria
                  1.0000000
                                                0.80143156
            Kingdom.Bacteria
                  0.92340636
$sig.clade
NULL
$null.prop
      HOO
                          H01
                H10
0.7177680 0.1811051 0.1011268
$global.pval
      HMP
0.5366665
```



References

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