

RNA-seq COVID

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D. Blanco-Melo, B. Nilsson-Payant, W.-C. Liu, R. Moeller, M. Panis, D. Sachs, R. Albrecht, B.R. TenOever, SARS-CoV-2 launches a unique transcriptional signature from in vitro, ex vivo, and in vivo systems, BioRxiv. (2020) 2020.03.24.004655. <https://doi.org/10.1101/2020.03.24.004655>.

```
# Clear all objects (from the workspace)
rm(list = ls())

# Suppress Warning messages
options(warn = -1)

# Turn off scientific notation like 1e+06
options(stringsAsFactors = F)

# INSTALL with:

# if (!requireNamespace("BiocManager", quietly = TRUE))
#   install.packages("BiocManager")
# BiocManager::install("GEOquery")
# BiocManager::install("DESeq2")
# BiocManager::install("limma")

# library(devtools)
# install_github("tpq/exprso")

# LOAD Libs
library(dplyr)

##
## Attaching package: 'dplyr'

## The following objects are masked from 'package:stats':
##
##   filter, lag

## The following objects are masked from 'package:base':
##
##   intersect, setdiff, setequal, union

library(tidyverse)

## -- Attaching packages -----
## v ggplot2 3.3.0      v purrr   0.3.3
## v tibble  2.1.3      v stringr 1.4.0
## v tidyr   1.0.2      v forcats 0.5.0
## v readr   1.3.1

## -- Conflicts -----
## x dplyr::filter() masks stats::filter()
## x dplyr::lag()    masks stats::lag()

library(DESeq2)

## Loading required package: S4Vectors
```

```

## Loading required package: stats4
## Loading required package: BiocGenerics
## Loading required package: parallel
##
## Attaching package: 'BiocGenerics'
## The following objects are masked from 'package:parallel':
##
##   clusterApply, clusterApplyLB, clusterCall, clusterEvalQ,
##   clusterExport, clusterMap, parApply, parCapply, parLapply,
##   parLapplyLB, parRapply, parSapply, parSapplyLB
## The following objects are masked from 'package:dplyr':
##
##   combine, intersect, setdiff, union
## The following objects are masked from 'package:stats':
##
##   IQR, mad, sd, var, xtabs
## The following objects are masked from 'package:base':
##
##   anyDuplicated, append, as.data.frame, basename, cbind, colnames,
##   dirname, do.call, duplicated, eval, evalq, Filter, Find, get, grep,
##   grepl, intersect, is.unsorted, lapply, Map, mapply, match, mget,
##   order, paste, pmax, pmax.int, pmin, pmin.int, Position, rank,
##   rbind, Reduce, rownames, sapply, setdiff, sort, table, tapply,
##   union, unique, unsplit, which, which.max, which.min
##
## Attaching package: 'S4Vectors'
## The following object is masked from 'package:tidyr':
##
##   expand
## The following objects are masked from 'package:dplyr':
##
##   first, rename
## The following object is masked from 'package:base':
##
##   expand.grid
## Loading required package: IRanges
##
## Attaching package: 'IRanges'
## The following object is masked from 'package:purrr':
##
##   reduce
## The following objects are masked from 'package:dplyr':
##
##   collapse, desc, slice
## The following object is masked from 'package:grDevices':
##
##   windows
## Loading required package: GenomicRanges
## Loading required package: GenomeInfoDb
## Loading required package: SummarizedExperiment
## Loading required package: Biobase

```

```

## Welcome to Bioconductor
##
## Vignettes contain introductory material; view with
## 'browseVignettes()'. To cite Bioconductor, see
## 'citation("Biobase")', and for packages 'citation("pkgname")'.

## Loading required package: DelayedArray
## Loading required package: matrixStats
##
## Attaching package: 'matrixStats'
## The following objects are masked from 'package:Biobase':
##
## anyMissing, rowMedians
## The following object is masked from 'package:dplyr':
##
## count
## Loading required package: BiocParallel
##
## Attaching package: 'DelayedArray'
## The following objects are masked from 'package:matrixStats':
##
## colMaxs, colMins, colRanges, rowMaxs, rowMins, rowRanges
## The following object is masked from 'package:purrr':
##
## simplify
## The following objects are masked from 'package:base':
##
## aperm, apply, rowsum
library(limma)

##
## Attaching package: 'limma'
## The following object is masked from 'package:DESeq2':
##
## plotMA
## The following object is masked from 'package:BiocGenerics':
##
## plotMA
# LOAD provided functions
source("../script_ejercicios.R")

datos <- load("../GSE147507_datos_covid.Rdata")

names(datos_covid)

## [1] "SARS004_mock_3" "SARS004_mock_2"
## [3] "SARS004_mock_1" "SARS004_CoV2_3"
## [5] "SARS004_CoV2_2" "SARS004_CoV2_1"
## [7] "CoV002.mock3.indexG3" "CoV002.mock2.indexG2"
## [9] "CoV002.mock1.indexG1" "CoV002.CoV2.3.indexG6"
## [11] "CoV002.CoV2.2.indexG5" "CoV002.CoV2.1.indexG4"
## [13] "svRNA184.mock.3.indexF3" "svRNA184.mock.1.indexF1"
## [15] "svRNA184.RSV.3.indexH9" "svRNA184.RSV.1.indexF4"
## [17] "X3_9_mock1_13" "X3_9_mock2_14"
## [19] "X3_9_wt1_15" "X3_9_wt2_16"

```

```
dim(datos_covid)
```

```
## [1] 23710    20
```

```
head(datos_covid)
```

```
##      SARS004_mock_3 SARS004_mock_2 SARS004_mock_1 SARS004_CoV2_3
## DDX11L1           0           0           0           0
## WASH7P           29          24          23          34
## FAM138A           0           0           0           0
## FAM138F           0           0           0           0
## OR4F5            0           0           0           0
## LOC729737        112          119          113          127
##      SARS004_CoV2_2 SARS004_CoV2_1 CoV002.mock3.indexG3
## DDX11L1           0           0           0
## WASH7P           19          44          68
## FAM138A           0           0           0
## FAM138F           0           0           0
## OR4F5            0           0           0
## LOC729737         84          270          11
##      CoV002.mock2.indexG2 CoV002.mock1.indexG1 CoV002.CoV2.3.indexG6
## DDX11L1           0           0           0
## WASH7P           43          33          65
## FAM138A           0           0           0
## FAM138F           0           0           0
## OR4F5            0           0           0
## LOC729737         3           6           8
##      CoV002.CoV2.2.indexG5 CoV002.CoV2.1.indexG4 svRNA184.mock.3.indexF3
## DDX11L1           1           1           1
## WASH7P           79          48          184
## FAM138A           0           0           0
## FAM138F           0           0           0
## OR4F5            0           0           0
## LOC729737        10          10          108
##      svRNA184.mock.1.indexF1 svRNA184.RSV.3.indexH9 svRNA184.RSV.1.indexF4
## DDX11L1           0           0           0
## WASH7P          128          51          43
## FAM138A           0           0           0
## FAM138F           0           0           0
## OR4F5            0           0           0
## LOC729737         95          37          11
##      X3_9_mock1_13 X3_9_mock2_14 X3_9_wt1_15 X3_9_wt2_16
## DDX11L1           0           0           0           0
## WASH7P           15          12           3           3
## FAM138A           0           0           0           0
## FAM138F           0           0           0           0
## OR4F5            0           0           0           0
## LOC729737         1           5           0           2
```

Cell Culture: * Normal human bronchial epithelial (NHBE) * Human adenocarcinomic alveolar basal epithelial (A549) cells

Viruses: * SARS-related coronavirus 2 (SARS.CoV.2) * influenza A/Puerto Rico/8/1934 (H1N1) virus (IAV) * human respiratory syncytial virus (RSV)

Filtrar genes con baja expresión y con bajos conteos

```
# FILTRAR genes - conteos
```

```
# Para cada gen, contar el número de muestras con mayor a 5 conteos
```

```
datos_covid_filter <-
```

```
  apply(datos_covid, 1, function(x)
```

```
length(which(x >= 5)))
table(datos_covid_filter)
```

```
## datos_covid_filter
##      0      1      2      3      4      5      6      7      8      9     10     11     12
## 6527   668   380   307   262   238   655   240   221   205   250   240   245
##    13    14    15    16    17    18    19    20
##   273   445   282   261   289   370   575 10777
```

```
# Filtrar genes con menor a 2 muestras con más de 5 conteos
datos_covid <- datos_covid[which(datos_covid_filter >= 2),]
```

```
# Let's take a LOOK
names(datos_covid)
```

```
## [1] "SARS004_mock_3"      "SARS004_mock_2"
## [3] "SARS004_mock_1"      "SARS004_CoV2_3"
## [5] "SARS004_CoV2_2"      "SARS004_CoV2_1"
## [7] "CoV002.mock3.indexG3" "CoV002.mock2.indexG2"
## [9] "CoV002.mock1.indexG1" "CoV002.CoV2.3.indexG6"
## [11] "CoV002.CoV2.2.indexG5" "CoV002.CoV2.1.indexG4"
## [13] "svRNA184.mock.3.indexF3" "svRNA184.mock.1.indexF1"
## [15] "svRNA184.RSV.3.indexH9" "svRNA184.RSV.1.indexF4"
## [17] "X3_9_mock1_13"        "X3_9_mock2_14"
## [19] "X3_9_wt1_15"          "X3_9_wt2_16"
```

```
dim(datos_covid)
```

```
## [1] 16515    20
```

```
head(datos_covid)
```

```
##           SARS004_mock_3 SARS004_mock_2 SARS004_mock_1 SARS004_CoV2_3
## WASH7P                29                24                23                34
## LOC729737             112                119                113                127
## LOC100133331           19                 21                 36                 16
## LOC100288069           13                 17                 27                 19
## LINC00115              9                  12                 26                 12
## LOC643837              93                 74                143                 88
##           SARS004_CoV2_2 SARS004_CoV2_1 CoV002.mock3.indexG3
## WASH7P                19                 44                 68
## LOC729737             84                270                 11
## LOC100133331           23                 54                 54
## LOC100288069           9                  50                 23
## LINC00115              7                  28                 29
## LOC643837             95                301                250
##           CoV002.mock2.indexG2 CoV002.mock1.indexG1 CoV002.CoV2.3.indexG6
## WASH7P                  43                 33                 65
## LOC729737                3                  6                  8
## LOC100133331             23                 20                 45
## LOC100288069             18                  5                 30
## LINC00115                15                  2                 17
## LOC643837              127                 83                243
##           CoV002.CoV2.2.indexG5 CoV002.CoV2.1.indexG4
## WASH7P                  79                 48
## LOC729737               10                 10
## LOC100133331             42                 36
## LOC100288069             17                 14
## LINC00115                13                 16
## LOC643837              197                 136
##           svRNA184.mock.3.indexF3 svRNA184.mock.1.indexF1
## WASH7P                  184                 128
## LOC729737               108                 95
## LOC100133331            127                 93
```

## LOC100288069	7	12	
## LINC00115	38	24	
## LOC643837	145	95	
##	svRNA184.RSV.3.indexH9	svRNA184.RSV.1.indexF4	X3_9_mock1_13
## WASH7P	51	43	15
## LOC729737	37	11	1
## LOC100133331	25	16	16
## LOC100288069	4	3	5
## LINC00115	13	6	5
## LOC643837	41	30	39
##	X3_9_mock2_14	X3_9_wt1_15	X3_9_wt2_16
## WASH7P	12	3	3
## LOC729737	5	0	2
## LOC100133331	14	10	3
## LOC100288069	15	8	3
## LINC00115	11	4	0
## LOC643837	81	77	18

Classify samples

SARS004_mock: "SARS004_mock_3" "SARS004_mock_2" "SARS004_mock_1"

SARS004_CoV2: "SARS004_CoV2_3" "SARS004_CoV2_2" "SARS004_CoV2_1"

CoV002.mock: "CoV002.mock3.indexG3" "CoV002.mock2.indexG2" "CoV002.mock1.indexG1"

CoV002.CoV2: "CoV002.CoV2.3.indexG6" "CoV002.CoV2.2.indexG5" "CoV002.CoV2.1.indexG4"

svRNA184.mock: "svRNA184.mock.3.indexF3" "svRNA184.mock.1.indexF1"

svRNA184.RSV: "svRNA184.RSV.3.indexH9" "svRNA184.RSV.1.indexF4"

X3_9_mock: "X3_9_mock1_13" "X3_9_mock2_14"

X3_9_wt: "X3_9_wt1_15" "X3_9_wt2_16"

```

SARS004_mock = datos_covid[,grep('SARS004_mock', colnames(datos_covid))]
SARS004_CoV2 = datos_covid[,grep('SARS004_CoV2', colnames(datos_covid))]

CoV002.mock = datos_covid[,grep('CoV002.mock', colnames(datos_covid))]
CoV002.CoV2 = datos_covid[,grep('CoV002.CoV2', colnames(datos_covid))]

svRNA184.mock = datos_covid[,grep('svRNA184.mock', colnames(datos_covid))]
svRNA184.RSV = datos_covid[,grep('svRNA184.RSV', colnames(datos_covid))]

X3_9_mock = datos_covid[,grep('X3_9_mock', colnames(datos_covid))]
X3_9_wt = datos_covid[,grep('X3_9_wt', colnames(datos_covid))]

```

Apply DESeq2 to SARS004 (SARS)

```

tbl_merge = merge(SARS004_mock, SARS004_CoV2, by="row.names", all=TRUE)
rownames(tbl_merge) <- tbl_merge[,1] # First column contains the row names
tbl_merge = tbl_merge %>% select(-contains(c("Row.names"))) # DROP extra data
print(head(tbl_merge))

```

##	SARS004_mock_3	SARS004_mock_2	SARS004_mock_1	SARS004_CoV2_3
## A1BG	8	7	6	7
## A1BG-AS1	23	34	33	28
## A1CF	0	0	0	0
## A2M	2	2	0	1
## A2M-AS1	3	3	0	1
## A2ML1	1487	1267	2156	1259
##	SARS004_CoV2_2	SARS004_CoV2_1		
## A1BG	3	22		

```

## A1BG-AS1          18          68
## A1CF              0          0
## A2M              0          8
## A2M-AS1          0         12
## A2ML1          1280        5239

# Conteos
aux_classes = rep(0, times = ncol(tbl_merge)) # CLASSIFY infected samples as 0s
aux_classes[grep(pattern = "mock", x = colnames(tbl_merge))] = 1 # and mocks as 1s
aux_classes

## [1] 1 1 1 0 0 0

colnames(tbl_merge)

## [1] "SARS004_mock_3" "SARS004_mock_2" "SARS004_mock_1" "SARS004_CoV2_3"
## [5] "SARS004_CoV2_2" "SARS004_CoV2_1"

count_results_SARS004 = DESeq_func(matrix_c = tbl_merge, classes_c = aux_classes)

## estimating size factors
## estimating dispersions
## gene-wise dispersion estimates
## mean-dispersion relationship
## final dispersion estimates
## fitting model and testing

count_results_SARS004 = count_results_SARS004[order(count_results_SARS004$pvalue),]
summary(count_results_SARS004)

##      baseMean      log2FoldChange      lfcSE      stat
## Min.   :    0.00      Min.   : -5.0817      Min.   :0.0771      Min.   : -22.0089
## 1st Qu.:   17.44      1st Qu.: -0.1576      1st Qu.:0.1252      1st Qu.: -0.7192
## Median :   181.43      Median : 0.0063      Median :0.1829      Median :  0.0308
## Mean   :   832.79      Mean   : 0.0146      Mean   :0.4908      Mean   : -0.0437
## 3rd Qu.:   663.88      3rd Qu.: 0.1909      3rd Qu.:0.4475      3rd Qu.:  0.7730
## Max.   :135168.60      Max.   : 4.5330      Max.   :4.0805      Max.   :  8.0613
##      NA's      :561      NA's      :561      NA's      :561
##      pvalue      padj
## Min.   :0.0000      Min.   :0.000
## 1st Qu.:0.1823      1st Qu.:0.569
## Median :0.4558      Median :0.804
## Mean   :0.4615      Mean   :0.712
## 3rd Qu.:0.7277      3rd Qu.:0.916
## Max.   :1.0000      Max.   :1.000
## NA's   :561      NA's   :3957

head(count_results_SARS004)

##      baseMean log2FoldChange      lfcSE      stat      pvalue      padj
## IL8      2374.7575      -2.335177 0.1061013 -22.00894 2.364807e-107 2.969725e-103
## CCL20     412.3543      -3.146069 0.1649441 -19.07355 4.188415e-81 2.629906e-77
## SAA2       576.0496      -2.424539 0.1282341 -18.90713 9.963962e-80 4.170914e-76
## SAA1     3317.5564      -2.223965 0.1396037 -15.93056 3.888422e-57 1.220770e-53
## IL36G      271.4601      -2.734517 0.1761112 -15.52722 2.270245e-54 5.701948e-51
## S100A8    1707.6257      -1.871364 0.1222721 -15.30492 7.089426e-53 1.483817e-49

# HISTOGRAM
pvals = count_results_SARS004["pvalue"]
hist(
  pvals[, 1],
  prob = TRUE,
  col = "black",
  border = "white",

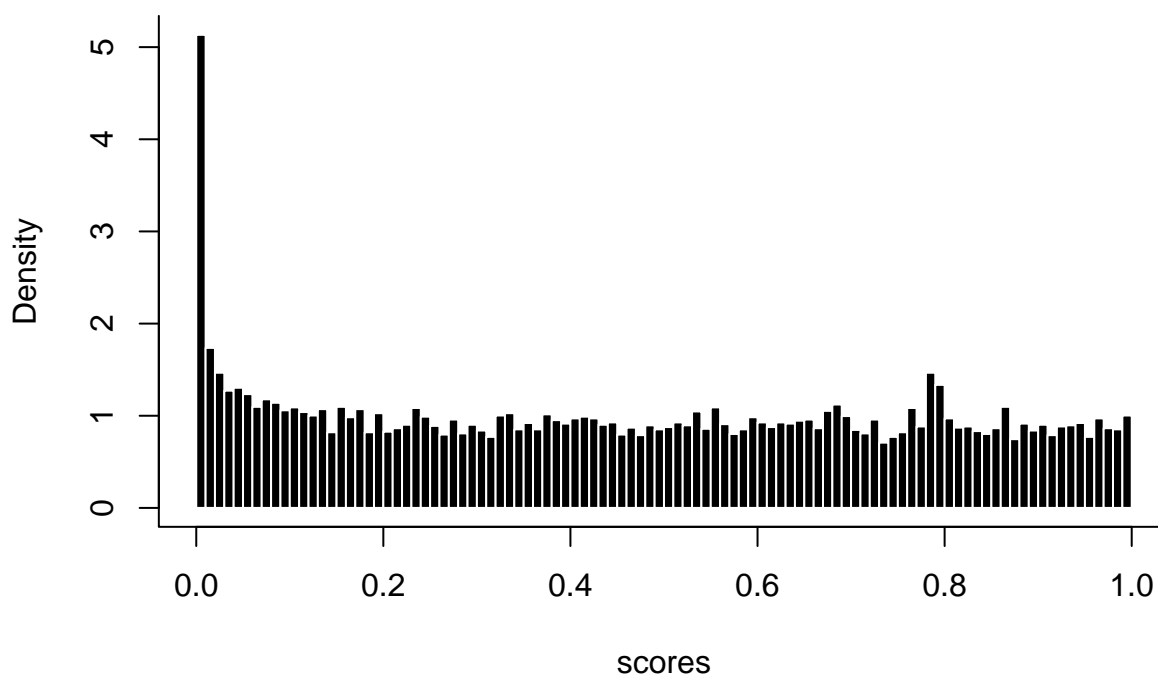
```

```

    xlab = "scores",
    breaks = 100
  )
box(bty = "l")
# Draw density function (assuming normal dist)
score_mean = mean(pvals[, 1])
score_sd = sd(pvals[, 1])
curve(
  dnorm(x, mean = score_mean, sd = score_sd),
  add = TRUE,
  col = "red",
  lwd = 2
)

```

Histogram of pvals[, 1]



```

# Let's take a look to some genes
# count_mat_28v0["ENSG00000128567.16_PODXL",]
# count_mat_28v0["ENSG00000185559.14_DLK1",]

```

Apply DESeq2 to CoV002 (SARS-related coronavirus 2)

```

tbl_merge = merge(CoV002.mock, CoV002.CoV2, by="row.names", all=TRUE)
rownames(tbl_merge) <- tbl_merge[,1] # First column contains the row names
tbl_merge = tbl_merge %>% select(-contains(c("Row.names"))) # DROP extra data
print(head(tbl_merge))

```

```

##          CoV002.mock3.indexG3 CoV002.mock2.indexG2 CoV002.mock1.indexG1
## A1BG                101                47                48
## A1BG-AS1             16                 6                 4
## A1CF                  4                 5                 3
## A2M                   0                 1                 0
## A2M-AS1              37                15                12
## A2ML1                 1                 0                 0
##          CoV002.CoV2.3.indexG6 CoV002.CoV2.2.indexG5 CoV002.CoV2.1.indexG4

```



```
## A1BG                133                129                91
## A1BG-AS1            27                 19                 19
## A1CF                8                  9                  5
## A2M                 0                  0                  0
## A2M-AS1            47                 32                 17
## A2ML1               0                  0                  1

# Conteos
aux_classes = rep(0, times = ncol(tbl_merge)) # CLASSIFY infected samples as 0s
aux_classes[grep(pattern = "mock", x = colnames(tbl_merge))] = 1 # and mocks as 1s
aux_classes

## [1] 1 1 1 0 0 0

colnames(tbl_merge)

## [1] "CoV002.mock3.indexG3" "CoV002.mock2.indexG2" "CoV002.mock1.indexG1"
## [4] "CoV002.CoV2.3.indexG6" "CoV002.CoV2.2.indexG5" "CoV002.CoV2.1.indexG4"

count_results_CoV002 = DESeq_func(matrix_c = tbl_merge, classes_c = aux_classes)

## estimating size factors
## estimating dispersions
## gene-wise dispersion estimates
## mean-dispersion relationship
## final dispersion estimates
## fitting model and testing

count_results_CoV002 = count_results_CoV002[order(count_results_CoV002$pvalue),]
summary(count_results_CoV002)

##      baseMean      log2FoldChange      lfcSE      stat
## Min.   : 0.00      Min.   : -6.0860      Min.   : 0.0724      Min.   : -34.3980
## 1st Qu.: 24.97      1st Qu.: -0.1699      1st Qu.: 0.1087      1st Qu.: -0.8638
## Median : 273.65      Median : -0.0065      Median : 0.1458      Median : -0.0330
## Mean   : 946.00      Mean   : -0.0092      Mean   : 0.3873      Mean   : -0.1118
## 3rd Qu.: 844.23      3rd Qu.: 0.1344      3rd Qu.: 0.3426      3rd Qu.: 0.7286
## Max.   :145716.11      Max.   : 4.0619      Max.   : 4.0805      Max.   : 6.5449
##      NA's      NA's      NA's      NA's
##      :370      :370      :370      :370
##      pvalue      padj
## Min.   :0.0000      Min.   :0.0000
## 1st Qu.:0.1715      1st Qu.:0.5664
## Median :0.4314      Median :0.7605
## Mean   :0.4485      Mean   :0.7036
## 3rd Qu.:0.7088      3rd Qu.:0.8972
## Max.   :1.0000      Max.   :1.0000
##      NA's      NA's
##      :370      :2871

head(count_results_CoV002)

##      baseMean log2FoldChange      lfcSE      stat      pvalue      padj
## IFI6  1308.9324      -4.333094 0.1259693 -34.39801 2.700377e-259 3.684394e-255
## IFIT1  590.6098      -4.296126 0.1695301 -25.34138 1.118549e-141 7.630738e-138
## ISG15  486.2893      -3.766134 0.1540151 -24.45302 4.673998e-132 2.125734e-128
## MX1    386.1696      -5.066631 0.2275617 -22.26487 8.095413e-110 2.761345e-106
## IRF9   761.8900      -2.159944 0.1103179 -19.57927 2.323191e-85 6.339523e-82
## IRF7   357.9020      -3.184034 0.1721120 -18.49978 2.073161e-76 4.714369e-73

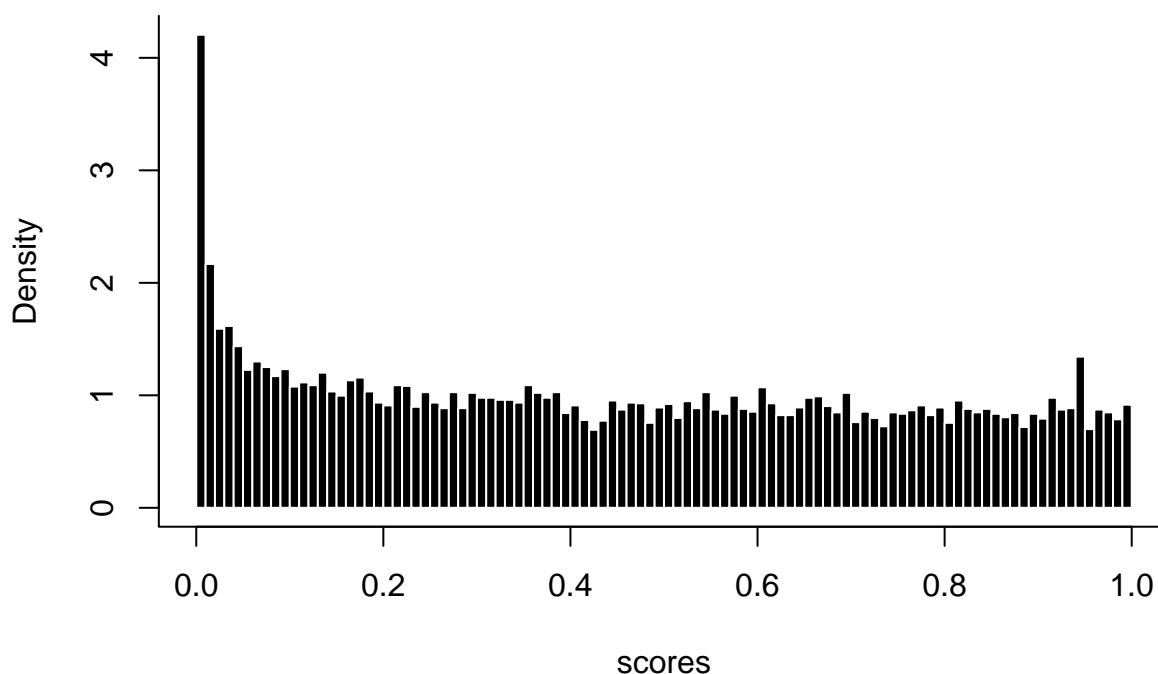
# HISTOGRAM
pvals = count_results_CoV002["pvalue"]
hist(
  pvals[, 1],
  prob = TRUE,
  col = "black",
```

```

border = "white",
xlab = "scores",
breaks = 100
)
box(bty = "l")
# Draw density function (assuming normal dist)
score_mean = mean(pvals[, 1])
score_sd = sd(pvals[, 1])
curve(
  dnorm(x, mean = score_mean, sd = score_sd),
  add = TRUE,
  col = "red",
  lwd = 2
)

```

Histogram of pvals[, 1]



```

# Let's take a look to some genes
# count_mat_28v0["ENSG00000128567.16_PODXL",]
# count_mat_28v0["ENSG00000185559.14_DLK1",]

```

Apply DESeq2 to svRNA184 (human respiratory syncytial virus)

```

tbl_merge = merge(svRNA184.mock, svRNA184.RSV, by="row.names", all=TRUE)
rownames(tbl_merge) <- tbl_merge[,1] # First column contains the row names
tbl_merge = tbl_merge %>% select(-contains(c("Row.names"))) # DROP extra data
print(head(tbl_merge))

```

##	svRNA184.mock.3.indexF3	svRNA184.mock.1.indexF1	svRNA184.RSV.3.indexH9
## A1BG	422	257	158
## A1BG-AS1	43	26	5
## A1CF	127	75	16
## A2M	1	6	3
## A2M-AS1	36	40	6
## A2ML1	0	0	0

```

##          svRNA184.RSV.1.indexF4
## A1BG          82
## A1BG-AS1       3
## A1CF           9
## A2M            1
## A2M-AS1        4
## A2ML1          0

# Conteos
aux_classes = rep(0, times = ncol(tbl_merge)) # CLASSIFY infected samples as 0s
aux_classes[grep(pattern = "mock", x = colnames(tbl_merge))] = 1 # and mocks as 1s
aux_classes

## [1] 1 1 0 0

colnames(tbl_merge)

## [1] "svRNA184.mock.3.indexF3" "svRNA184.mock.1.indexF1"
## [3] "svRNA184.RSV.3.indexH9"   "svRNA184.RSV.1.indexF4"

count_results_svRNA184 = DESeq_func(matrix_c = tbl_merge, classes_c = aux_classes)

## estimating size factors
## estimating dispersions
## gene-wise dispersion estimates
## mean-dispersion relationship
## final dispersion estimates
## fitting model and testing

count_results_svRNA184 = count_results_svRNA184[order(count_results_svRNA184$pvalue),]
summary(count_results_svRNA184)

##      baseMean      log2FoldChange      lfcSE      stat
## Min.   : 0.00      Min.   : -7.7173      Min.   : 0.1631      Min.   : -27.1854
## 1st Qu.: 20.57      1st Qu.: -0.2990      1st Qu.: 0.2291      1st Qu.: -0.8169
## Median : 140.12      Median : 0.0166      Median : 0.3059      Median : 0.0447
## Mean   : 575.65      Mean   : 0.0249      Mean   : 0.6431      Mean   : -0.1005
## 3rd Qu.: 446.26      3rd Qu.: 0.3713      3rd Qu.: 0.6259      3rd Qu.: 0.8506
## Max.   :100749.48      Max.   : 6.6833      Max.   : 4.9920      Max.   : 11.7450
##      NA's      :418      NA's      :418      NA's      :418
##      pvalue      padj
## Min.   :0.0000      Min.   :0.0000
## 1st Qu.:0.1268      1st Qu.:0.3963
## Median :0.4044      Median :0.7240
## Mean   :0.4266      Mean   :0.6226
## 3rd Qu.:0.7035      3rd Qu.:0.8911
## Max.   :0.9999      Max.   :0.9999
##      NA's      :418      NA's      :2600

head(count_results_svRNA184)

##      baseMean log2FoldChange      lfcSE      stat      pvalue      padj
## MX1      1535.804      -5.572993 0.2049993 -27.18543 9.658759e-163 1.344016e-158
## OASL      1095.279      -5.545498 0.2154467 -25.73953 4.221962e-146 2.937430e-142
## IFIT1     4235.123      -5.120843 0.2050420 -24.97460 1.154374e-137 5.354371e-134
## ISG15     5057.195      -4.547792 0.1859176 -24.46133 3.812414e-132 1.326243e-128
## IFIT3     1694.756      -5.012684 0.2065019 -24.27427 3.666006e-130 1.020250e-126
## HELZ2     2747.630      -3.838123 0.1768202 -21.70636 1.786523e-104 4.143245e-101

# HISTOGRAM
pvals = count_results_svRNA184["pvalue"]
hist(
  pvals[, 1],
  prob = TRUE,

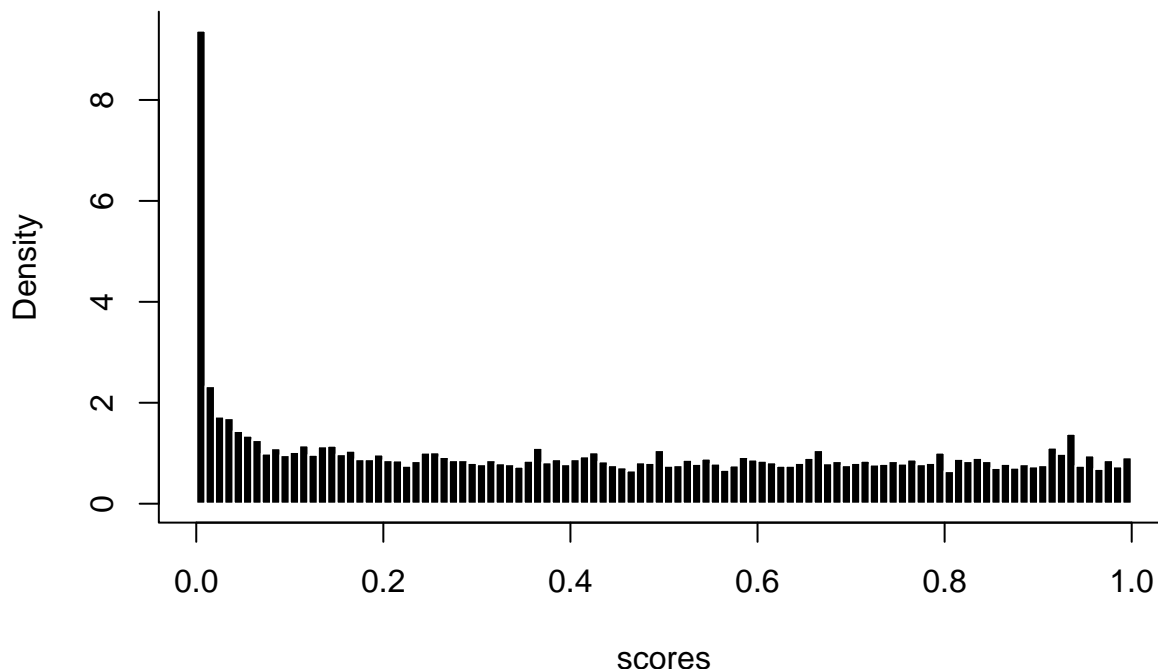
```

```

col = "black",
border = "white",
xlab = "scores",
breaks = 100
)
box(bty = "l")
# Draw density function (assuming normal dist)
score_mean = mean(pvals[, 1])
score_sd = sd(pvals[, 1])
curve(
  dnorm(x, mean = score_mean, sd = score_sd),
  add = TRUE,
  col = "red",
  lwd = 2
)

```

Histogram of pvals[, 1]



```

# Let's take a look to some genes
# count_mat_28v0["ENSG00000128567.16_PODXL",]
# count_mat_28v0["ENSG00000185559.14_DLK1",]

```

Apply DESeq2 to X3_9 (influenza A/Puerto Rico/8/1934)

```

tbl_merge = merge(X3_9_mock, X3_9_wt, by="row.names", all=TRUE)
rownames(tbl_merge) <- tbl_merge[,1] # First column contains the row names
tbl_merge = tbl_merge %>% select(-contains(c("Row.names"))) # DROP extra data
print(head(tbl_merge))

```

```

##           X3_9_mock1_13 X3_9_mock2_14 X3_9_wt1_15 X3_9_wt2_16
## A1BG                21                21             5             8
## A1BG-AS1              0                 6             2             1
## A1CF                  0                 0             0             0
## A2M                   0                 1             0             1
## A2M-AS1              12                26             7             2

```

```
## A2ML1          0          0          0          0
# Conteos
aux_classes = rep(0, times = ncol(tbl_merge)) # CLASSIFY infected samples as 0s
aux_classes[grep(pattern = "mock", x = colnames(tbl_merge))] = 1 # and mocks as 1s
aux_classes

## [1] 1 1 0 0
colnames(tbl_merge)

## [1] "X3_9_mock1_13" "X3_9_mock2_14" "X3_9_wt1_15"  "X3_9_wt2_16"
count_results_X3_9 = DESeq_func(matrix_c = tbl_merge, classes_c = aux_classes)

## estimating size factors
## estimating dispersions
## gene-wise dispersion estimates
## mean-dispersion relationship
## final dispersion estimates
## fitting model and testing
count_results_X3_9 = count_results_X3_9[order(count_results_X3_9$pvalue),]
summary(count_results_X3_9)

##      baseMean      log2FoldChange      lfcSE      stat
## Min.      : 0.00      Min.      :-6.8682      Min.      :0.2386      Min.      : -5.0992
## 1st Qu.: 6.58      1st Qu.: -0.3830      1st Qu.: 0.3352      1st Qu.: -0.7039
## Median : 76.35      Median : 0.0014      Median : 0.4516      Median : 0.0025
## Mean      : 267.67      Mean      : 0.0576      Mean      : 1.0106      Mean      : 0.0420
## 3rd Qu.: 242.78      3rd Qu.: 0.4995      3rd Qu.: 1.0701      3rd Qu.: 0.7245
## Max.      :35035.18      Max.      : 6.0135      Max.      : 4.9730      Max.      : 5.8261
##              NA's      :701      NA's      :701      NA's      :701
##      pvalue      padj
## Min.      :0.0000      Min.      :0.000
## 1st Qu.:0.1929      1st Qu.:0.607
## Median :0.4750      Median :0.809
## Mean      :0.4749      Mean      :0.742
## 3rd Qu.:0.7431      3rd Qu.:0.929
## Max.      :1.0000      Max.      :1.000
## NA's      :701      NA's      :3760
head(count_results_X3_9)

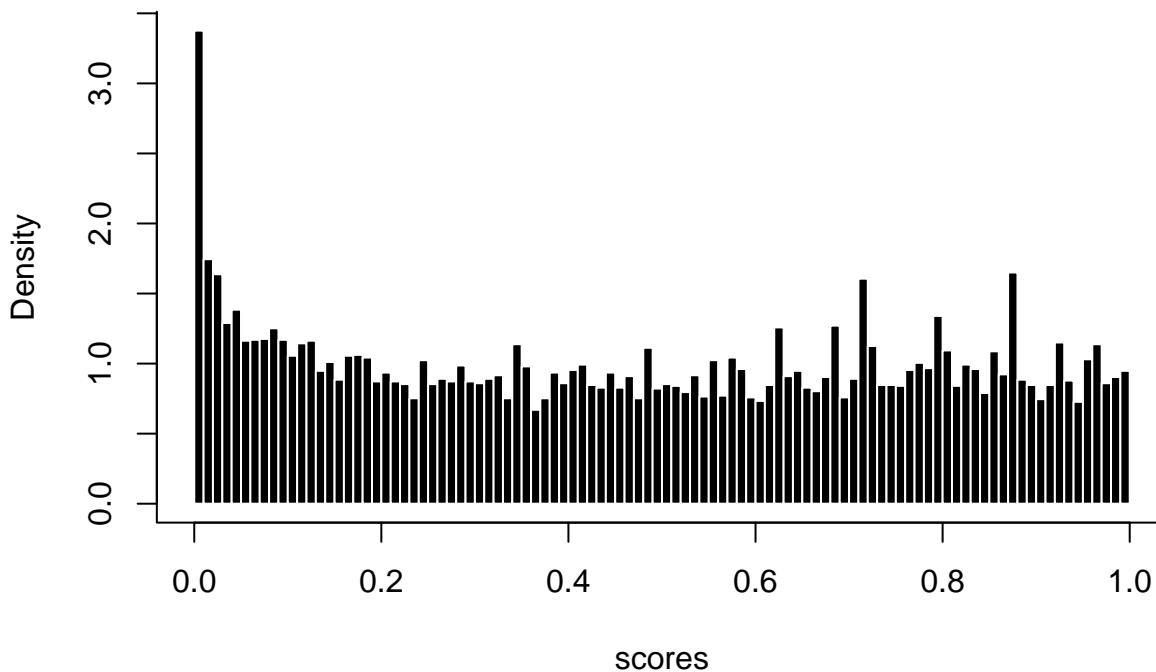
##      baseMean log2FoldChange      lfcSE      stat      pvalue
## RPL39      488.84050      1.734375 0.2976903 5.826106 5.673559e-09
## MAFK       228.13370      2.126322 0.3824744 5.559385 2.707271e-08
## CPLX2      6171.97802      1.384211 0.2553515 5.420807 5.933062e-08
## NEAT1      1145.10887      1.363436 0.2604940 5.234037 1.658470e-07
## ZDHHC24    55.77728      3.486370 0.6770468 5.149378 2.613512e-07
## LINC00641  79.21125      -2.306046 0.4522335 -5.099237 3.410247e-07
##              padj
## RPL39      7.236624e-05
## MAFK       1.726562e-04
## CPLX2      2.522540e-04
## NEAT1      5.288447e-04
## ZDHHC24    6.667068e-04
## LINC00641  7.249616e-04
# HISTOGRAM
pvals = count_results_X3_9["pvalue"]
hist(
  pvals[, 1],
  prob = TRUE,
```

```

col = "black",
border = "white",
xlab = "scores",
breaks = 100
)
box(bty = "l")
# Draw density function (assuming normal dist)
score_mean = mean(pvals[, 1])
score_sd = sd(pvals[, 1])
curve(
  dnorm(x, mean = score_mean, sd = score_sd),
  add = TRUE,
  col = "red",
  lwd = 2
)

```

Histogram of pvals[, 1]



```

# Let's take a look to some genes
# count_mat_28v0["ENSG00000128567.16_PODXL",]
# count_mat_28v0["ENSG00000185559.14_DLK1",]

```

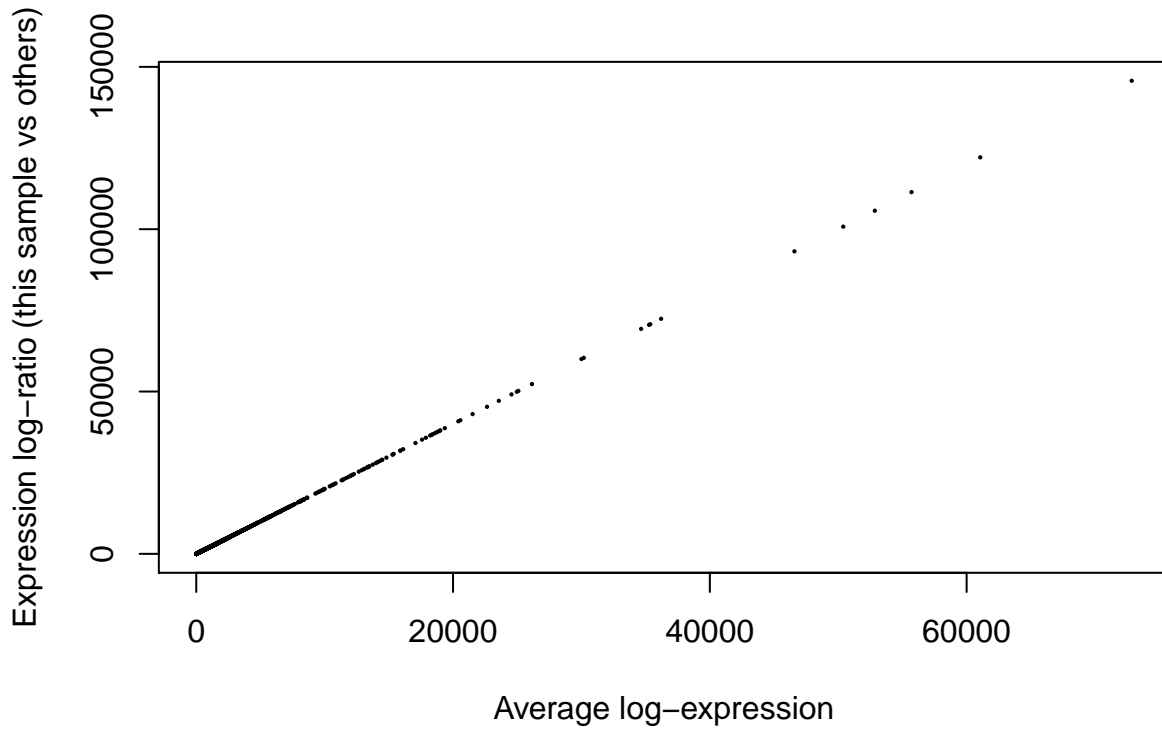
Comparar CoV002 (SARS-related coronavirus 2) vs SARS004 (SARS), svRNA184 (human respiratory syncytial virus) y X3_9 (influenza A/Puerto Rico/8/1934)

```

plotMA(count_results_CoV002, main = "CoV002")

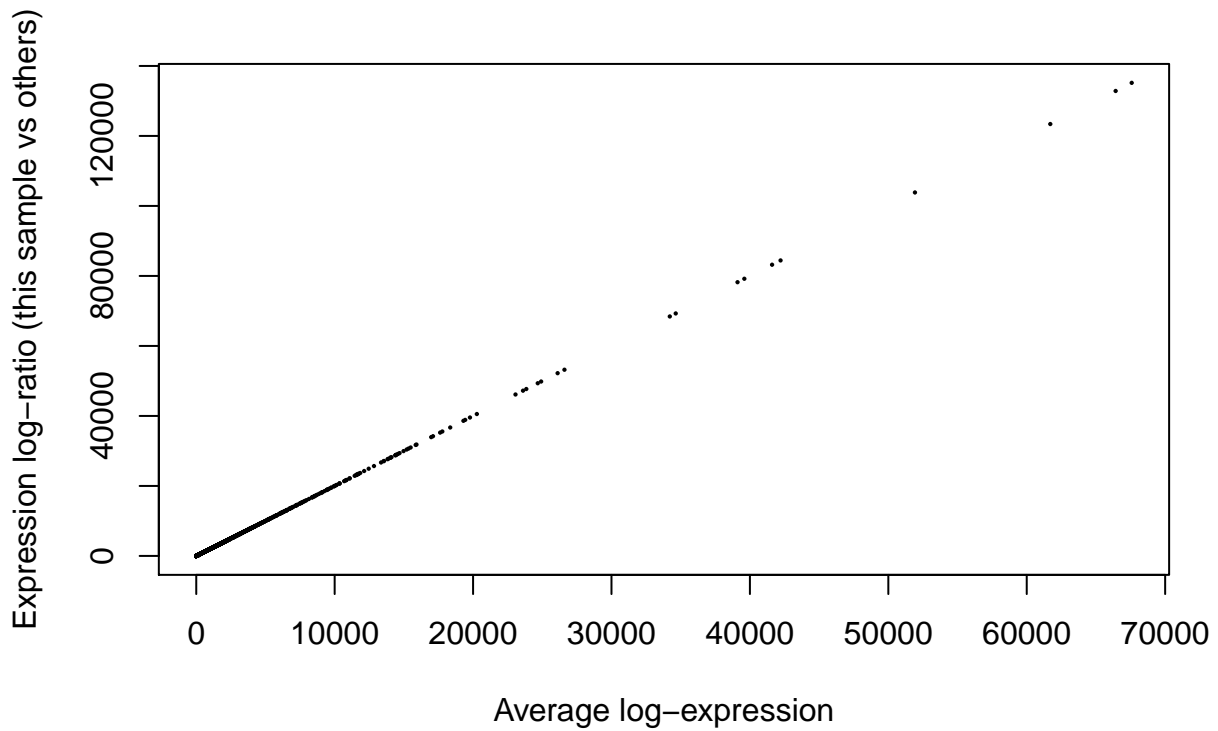
```

CoV002

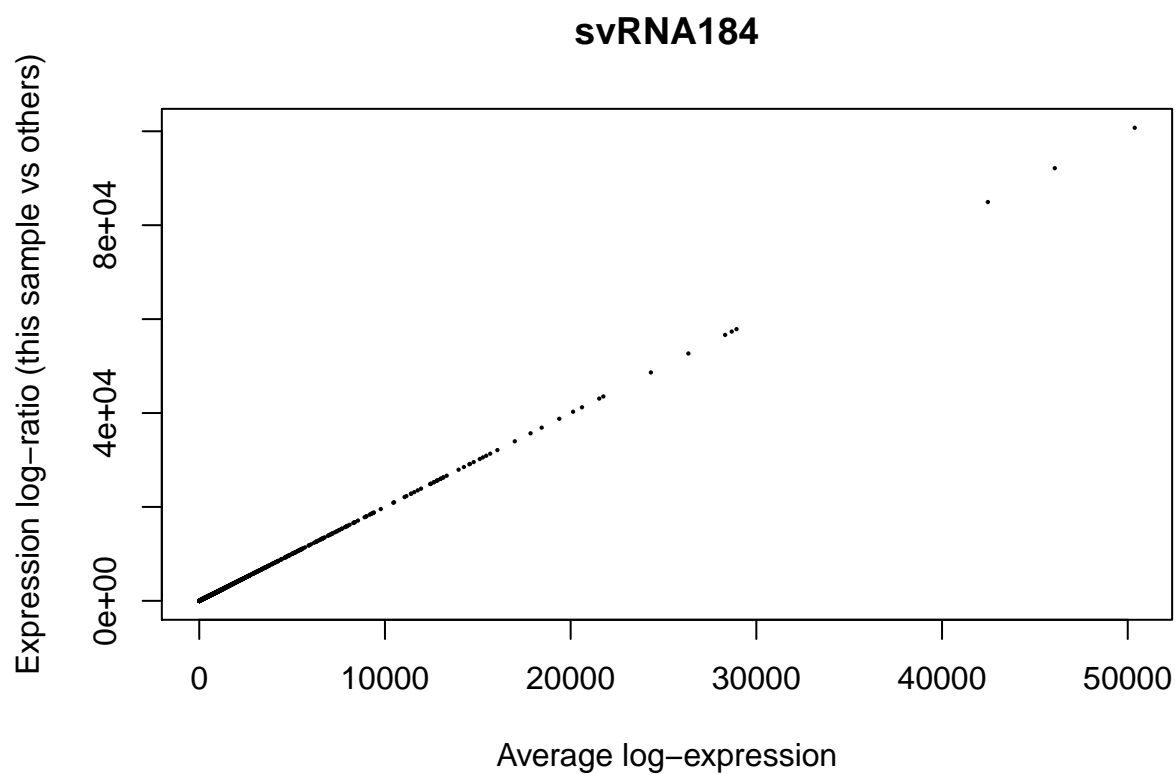


```
plotMA(count_results_SARS004, main = "SARS004")
```

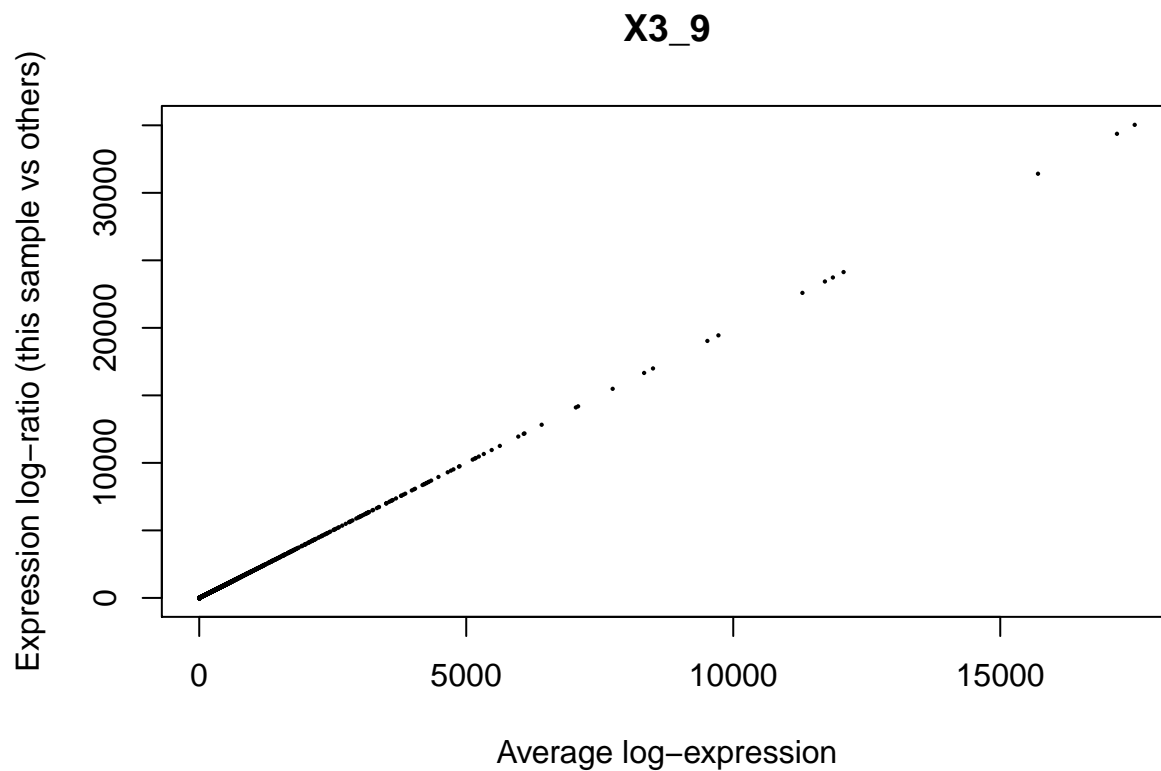
SARS004



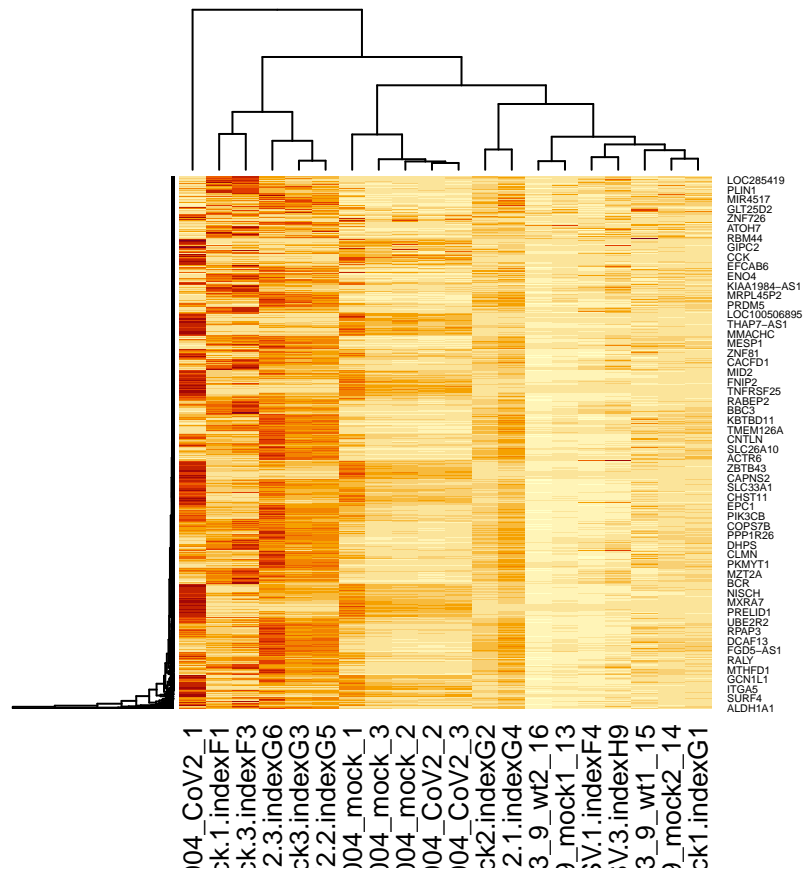
```
plotMA(count_results_svRNA184, main = "svRNA184")
```



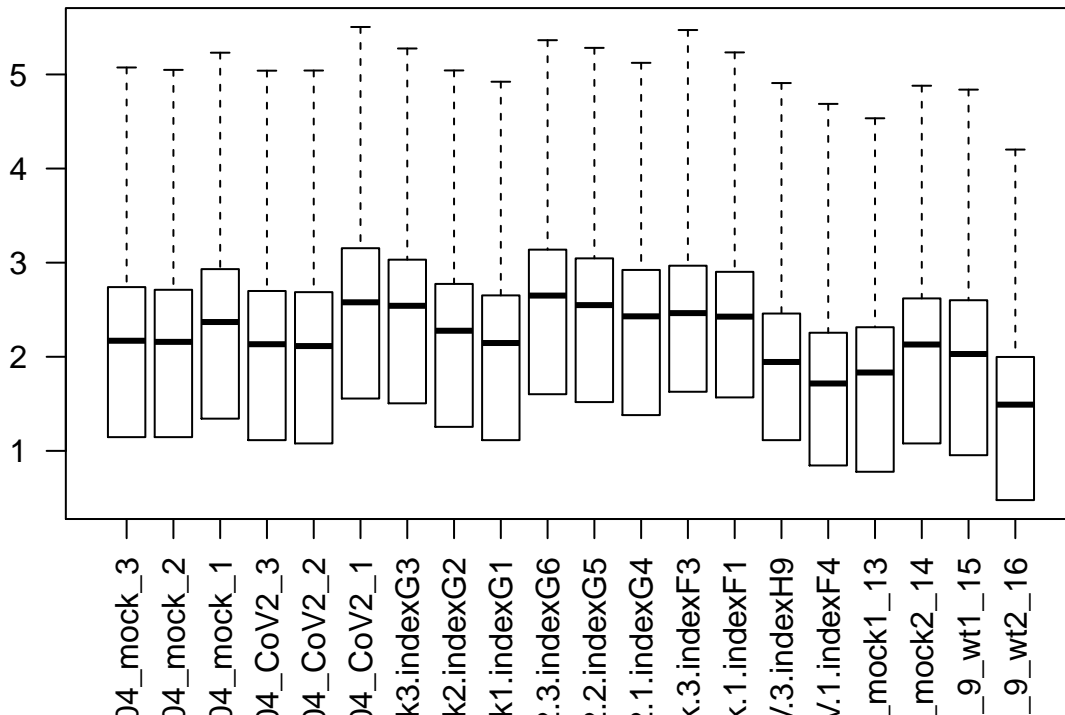
```
plotMA(count_results_X3_9, main = "X3_9")
```



```
heatmap(as.matrix(datos_covid))
```

```
boxplot(log10(datos_covid), range=0, las=2)
```



TNFSF15 y EDN1 son los genes que están presentes al comparar SARS-CoV-2 con otros virus. DESeq2 muestra que los datos proporcionados sugieren que las drogas que aumentan la respuesta antiviral pueden ser una opción efectiva en el tratamiento de COVID-19.