

Critical Oxygen Pressure (P_{CRIT}) and Oxygen Supply Capacity (α) Analysis

Lloyd Trueblood and Kirt Onthank

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1 Loading Libraries

I am reading in the libraries I use for this analysis. Included among these is the “OTools” package, which was written by Kirt Onthank. This can be install from github using the command:

```
install_github('KirtOnthank/OTools')
```

```
library(OTools)
library(respirometry)
library(knitr)
library(nlme)
library(car)
library(emmeans)
```

2 Sorting files for P_{CRIT} analysis

```
files=list.files(recursive=T)
resp.files=grep(".txt",files,value=T)
pcrit.files=grep("pcrit|pcrti",resp.files,value=T,ignore.case=T)
pcrit.files=pcrit.files[!duplicated(basename(pcrit.files))]

pcrit.files.read=pcrit.files[!grepl("ch2.txt|ch3.txt|ch4.txt|\\(1\\).txt",pcrit.files)]
pcrit.files.read
```

```
## [1] "All Pcrits/Gr1 Muus 1000-2 pcrit 7-27-21 B.txt"
## [2] "All Pcrits/Gr1 Muus 1000-2 pcrit 7-27-21.txt"
## [3] "All Pcrits/Gr1 Muus 1800-2 pcrit 25 ml jar 7-29-21 ch2 blank.txt"
## [4] "All Pcrits/Gr1 Muus 1800-2 pcrit 7-28-21.txt"
## [5] "All Pcrits/GR1 Muus 1800-2 pcrit day7 8-3-21.txt"
## [6] "All Pcrits/GR1 Muus1000 7day-7-26-21.txt"
## [7] "All Pcrits/GR1 Muus1000 pcrit 7-21-21.txt"
## [8] "All Pcrits/GR1 Muus1800 7day-pcrit 7-20-21.txt"
## [9] "All Pcrits/GR1 Muus1800 pcrit 7-13-21.txt"
## [10] "All Pcrits/gr2muus1800 7day pcrit 7-20-21.txt"
## [11] "All Pcrits/gr2muus1800-2 pcrit 7-28-21.txt"
## [12] "All Pcrits/gr2muus1800-2 pcrit day7 8-3-21.txt"
## [13] "All Pcrits/gr2muus1800-2 pcrit in 25 ml jar 7-29-21 ch2 is blank.txt"
## [14] "All Pcrits/Gr3 Muus 1000 pcrit 7-21-21.txt"
## [15] "All Pcrits/gr3 muus 1800 7day Pcrit 7-20-21.txt"
## [16] "All Pcrits/gr3 muus 1800 pcrit 7-13-21.txt"
## [17] "All Pcrits/Gr3 Muus 1800-2 pcrit 07-28-21.txt"
## [18] "All Pcrits/Gr3 Muus 1800-2 pcrit 08-03-21.txt"
## [19] "All Pcrits/Gr3 Muus1000-2 7 day pcrit 7-27-21.txt"
## [20] "All Pcrits/GR4MUUS1000-2Pcrit-7-26-21-ch1.txt"
## [21] "All Pcrits/GR4MUUS1000Pcrit-7-21-21-ch1.txt"
## [22] "All Pcrits/GR4MUUS1800-2-7dayPcrit-8-3-21-ch1.txt"
## [23] "All Pcrits/GR4MUUS1800-2Pcrit-7-28-21-ch1.txt"
## [24] "All Pcrits/GR4MUUS1800-7dayPcrit-7-20-21-ch1.txt"
## [25] "All Pcrits/GR4MUUS1800Pcrit-7-13-21-ch1.txt"
## [26] "All Pcrits/tbcto 1000 pcrit tank 1 and 2 day 7 8-19-21.txt"
```

```
## [27] "All Pcrits/tbocto 1000 pcrit tank 3 and 4 8-11-21-ch1.txt"
## [28] "All Pcrits/tbocto 1000 pcrit tank 3 and 4 day 7 8-19-21-ch1.txt"
## [29] "All Pcrits/Tbocto 1000 pcrti tank 1 and 2 8-11-21.txt"
## [30] "Group 2/Pcrit/gr2muus1000 pcrit 7-21-21.txt"
## [31] "Group 2/Pcrit/gr2muus1000-2 pcrit 7-26-21.txt"
## [32] "Trueblood after session/gr2MUUS1800-2pcritday7.8-3-21.txt"
## [33] "Trueblood after session/Muus TB collected data/desktop from presense onthank/tbocto 1800 pcrit
```

3 Reading in log files

Here I am reading in the log files that will provide additional information needed to analyze the raw data files.

```
pcrit.log=read.csv("pcrit_log.csv")
routine=read.csv("RMR_Results.csv")
```

4 Calculating P_{CRIT} from raw data

First I make a empty object where I can place calculated P_{CRITS}

```
pcrits=data.frame(filename=as.character(),
                  spreadsheet_guess=as.character(),
                  octo=as.character(),
                  mass=as.numeric(),
                  pco2=as.numeric(),
                  day=as.numeric(),
                  rmr=as.numeric(),
                  pcrit=as.numeric(),
                  alpha=as.numeric()
                  )
```

Next, I calculate the P_{CRIT} from each data file. Here are a couple of important points on our calculations:

1. We are using the alpha P_{CRIT} method (Seibel et al, 2021) to calculate P_{CRIT} as implemented by the `calc_pcrit()` function from the “respirometry” R package (Birk, 2021).
2. We used the routine metabolic rate that we measured for each individual octopus in this study, unless the maximum metabolic rate measured during the P_{CRIT} run was less than the RMR we had measured. In those cases we used the default MR used by the function, which is “the mean MO_2 value from the oxyregulating portion of the curve (as defined by the broken-stick regression)”.
3. Any run in which O_2 did not drop below 50 mmHgO_2 was dropped from the analysis. All runs, however, including those dropped, are graphed below.
4. Oxygen trace of each run is graphed below with the portion used to calculate P_{CRIT} plotted in red.

```
co=1

for (i in 1:length(pcrit.files.read)){
  filename=pcrit.files.read[i]

  if(length(grep("Group 4|presens|ch\\d\\.txt",basename(filename)))>0){
    pcrit.raw=read.presens(filename)
```

```

}else{
  pcrit.raw=read.pyro(filename)
}

guess=which.min(adist(basename(filename),pcrit.log$filename))
ch=pcrit.log$ch1[guess]
octo=pcrit.log$octo1[guess]
start=pcrit.log[guess,6+ch]
stop=max(pcrit.raw$times)-pcrit.log[guess,10+ch]
mass=mean(routine$mass[routine$octo==octo])
rmr=mean(routine$rmr[routine$octo==octo])

pcrit.working=
  pcrit.raw[
    pcrit.raw$times>start&
    pcrit.raw$times<stop,
  ]
vol=pcrit.log$vol[guess]
drop.time=round((stop-start)/3600,1)
O2.drop=round(diff(range(pcrit.working[,3+ch])),1)
rough.mo2=round((O2.drop*vol)/mass/drop.time,1)

plot(pcrit.raw[,3+ch]~pcrit.raw$times,type="l",main=basename(filename))
points(pcrit.working[,3+ch]~pcrit.working$times,type="l",col="red")
mtext(paste("mass=",mass),side=3,adj=1,line=-1)
mtext(paste0("resp vol=",vol),side=3,adj=1,line=-2)
mtext(paste0("drop time=",drop.time," hr"),side=3,adj=1,line=-3)
mtext(paste0("O2 drop=",O2.drop),side=3,adj=1,line=-4)
mtext(paste0("rough mean MO2=",rough.mo2),side=3,adj=1,line=-5)

pcrit.resp=resp.closed(pcrit.working,volume=pcrit.log$vol[guess],
  weight=mass,smooth="loess",channel=ch,smooth.span = 0.2)

pcrit.bin=aggregate(pcrit.resp$resp~round(pcrit.resp$po2,1),FUN="mean")
colnames(pcrit.bin)=c("po2","resp")

if(max(pcrit.bin$resp)>rmr){
  plot_pcrit(pcrit.bin$po2,pcrit.bin$resp,
    avg_top_n = 3,MR=rmr)
  if (min(pcrit.working[,3+ch])>50){
    mtext("DROPPED FROM ANALYSIS",line=-5,col="red",cex=3)
    mtext("pO2 did not get low enough",line=-7,col="red",cex=2)
  }
}else{
  plot_pcrit(pcrit.bin$po2,pcrit.bin$resp,
    avg_top_n = 3)
  if (min(pcrit.working[,3+ch])>50){
    mtext("DROPPED FROM ANALYSIS",line=-5,col="red",cex=3)
    mtext("pO2 did not get low enough",line=-7,col="red",cex=2)
  }
}
}

```

```

pcrits[co,1]=basename(filename)
pcrits[co,2]=pcrit.log$filename[guess]
pcrits[co,3]=octo
pcrits[co,4]=mass
pcrits[co,5]=pcrit.log$po2[guess]
pcrits[co,6]=pcrit.log$day[guess]
pcrits[co,7]=rmr
if (min(pcrit.working[,3+ch])<50){
  if(max(pcrit.bin$resp)>rmr){
    pcrits[co,8]=as.numeric(calc_pcrit(pcrit.bin$po2,pcrit.bin$resp,
      avg_top_n = 3,MR=rmr)[1])
  }else{
    pcrits[co,8]=as.numeric(calc_pcrit(pcrit.bin$po2,pcrit.bin$resp,
      avg_top_n = 3)[1])
  }
  pcrits[co,9]=calc_alpha(pcrit.bin$po2,pcrit.bin$resp,avg_top_n = 3)$alpha
}else{
  pcrits[co,8]=NA
  pcrits[co,9]=NA
}
co=co+1

if(!is.na(pcrit.log$ch2[guess])){
  ch=pcrit.log$ch2[guess]
  octo=pcrit.log$octo2[guess]
  start=pcrit.log[guess,6+ch]
  stop=max(pcrit.raw$times)-pcrit.log[guess,10+ch]
  mass=mean(routine$mass[routine$octo==octo])
  rmr=mean(routine$rmr[routine$octo==octo])

  pcrit.working=
    pcrit.raw[
      pcrit.raw$times>start&
      pcrit.raw$times<stop,
    ]

  plot(pcrit.raw[,3+ch]~pcrit.raw$times,type="l",main=basename(filename))
  points(pcrit.working[,3+ch]~pcrit.working$times,type="l",col="red")

  pcrit.resp=resp.closed(pcrit.working,volume=pcrit.log$vol[guess],
    weight=mass,smooth="loess",channel=ch,smooth.span = 0.2)
  pcrit.bin=aggregate(pcrit.resp$resp~round(pcrit.resp$po2,1),FUN="mean")
  colnames(pcrit.bin)=c("po2","resp")

  if(max(pcrit.bin$resp)>rmr){
    plot_pcrit(pcrit.bin$po2,pcrit.bin$resp,
      avg_top_n = 3,MR=rmr)
    if (min(pcrit.working[,3+ch])>50){
      mtext("DROPPED FROM ANALYSIS",line=-5,col="red",cex=3)
      mtext("pO2 did not get low enough",line=-7,col="red",cex=2)
    }
  }else{
    plot_pcrit(pcrit.bin$po2,pcrit.bin$resp,

```

```

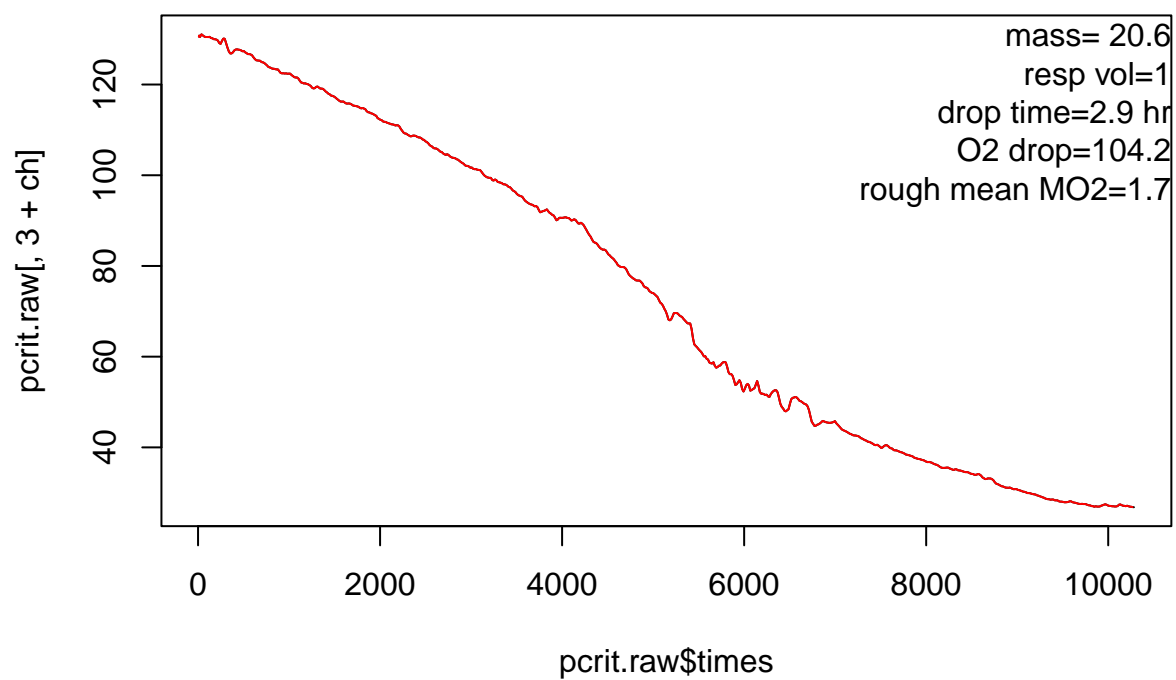
        avg_top_n = 3)
    if (min(pcrit.working[,3+ch])>50){
        mtext("DROPPED FROM ANALYSIS",line=-5,col="red",cex=3)
        mtext("pO2 did not get low enough",line=-7,col="red",cex=2)
    }
}

pcrits[co,1]=basename(filename)
pcrits[co,2]=pcrit.log$filename[guess]
pcrits[co,3]=octo
pcrits[co,4]=mass
pcrits[co,5]=pcrit.log$pco2[guess]
pcrits[co,6]=pcrit.log$day[guess]
pcrits[co,7]=rmr
if (min(pcrit.working[,3+ch])<50){
    if(max(pcrit.bin$resp)>rmr){
        pcrits[co,8]=as.numeric(calc_pcrit(pcrit.bin$po2,pcrit.bin$resp,
            avg_top_n = 3,MR=rmr)[1])
    }else{
        pcrits[co,8]=as.numeric(calc_pcrit(pcrit.bin$po2,pcrit.bin$resp,
            avg_top_n = 3)[1])
    }
    pcrits[co,9]=calc_alpha(pcrit.bin$po2,pcrit.bin$resp,avg_top_n = 3)$alpha
}else{
    pcrits[co,8]=NA
}
co=co+1
}

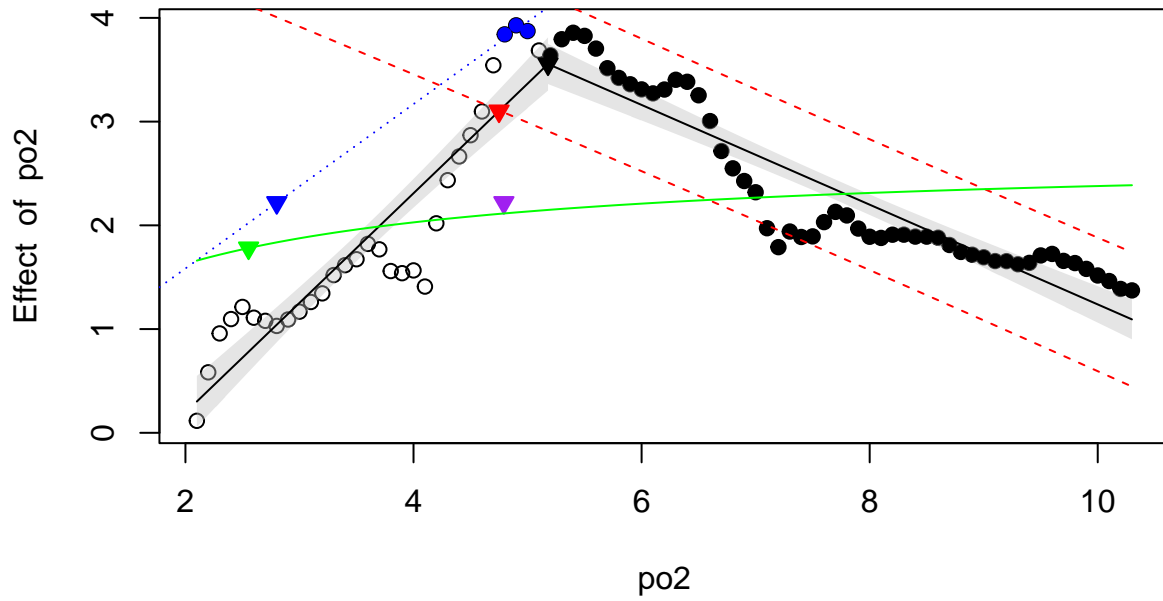
}

```

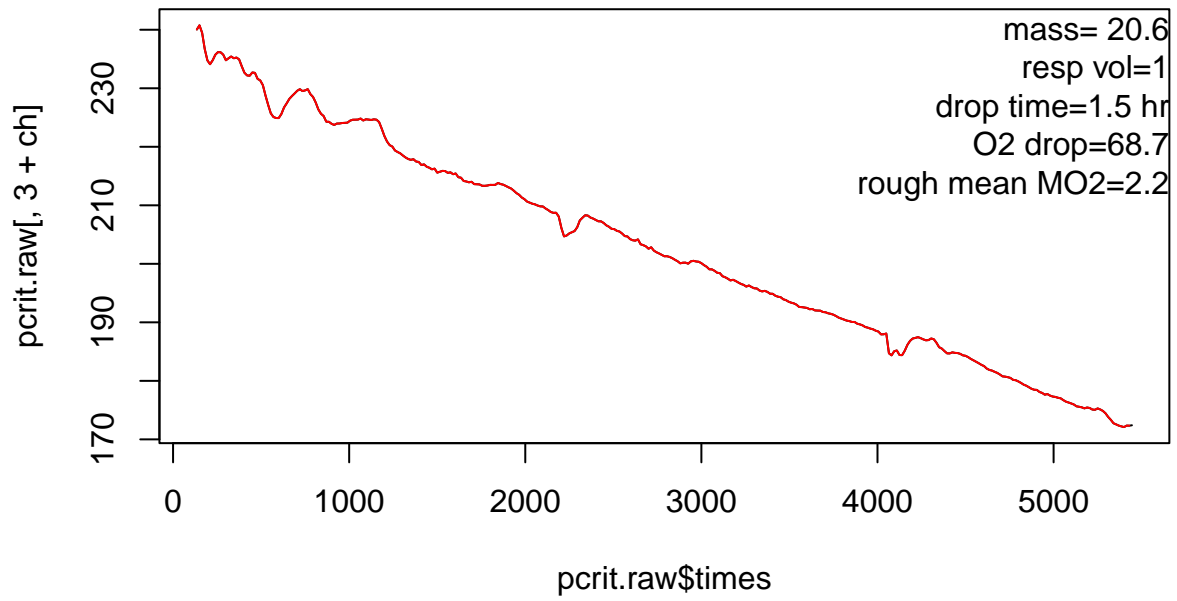
Gr1 Muus 1000-2 pcrit 7-27-21 B.txt



Alpha @ MR of 2.22 = 2.801
 Breakpoint = 5.179
 LLO @ MR of 2.22 = 4.792
 NLR (Michaelis-Menten) = 2.554
 Sub-PI = 4.75



Gr1 Muus 1000-2 pcrit 7-27-21.txt



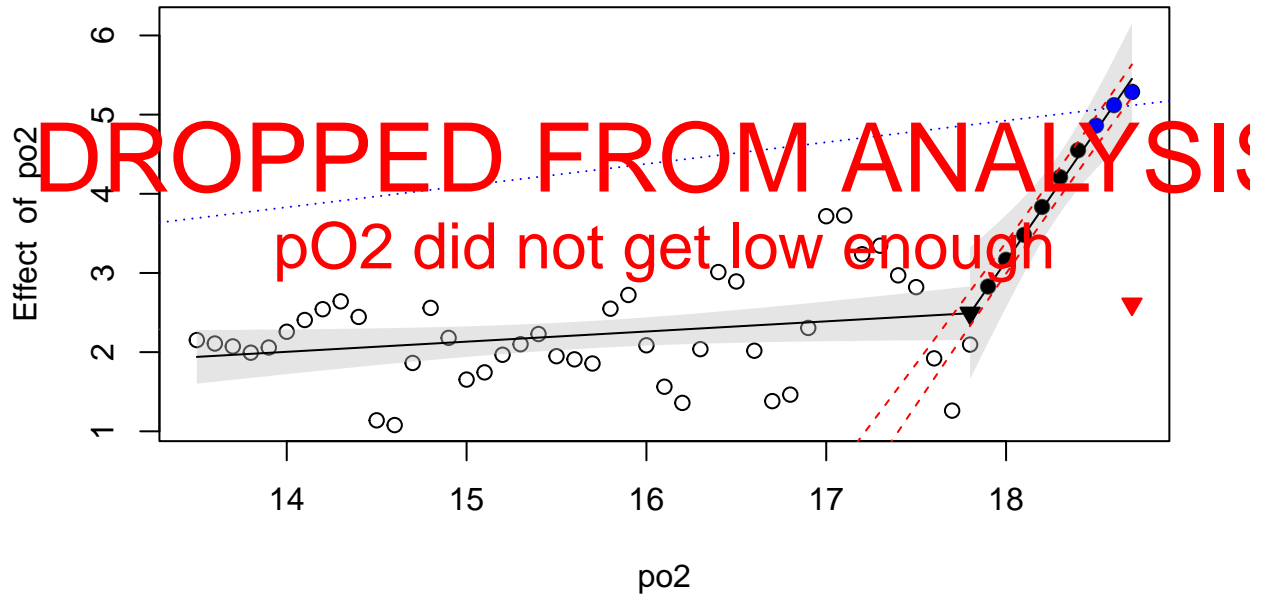
Alpha @ MR of 2.22 = 8.109

Breakpoint = 17.8

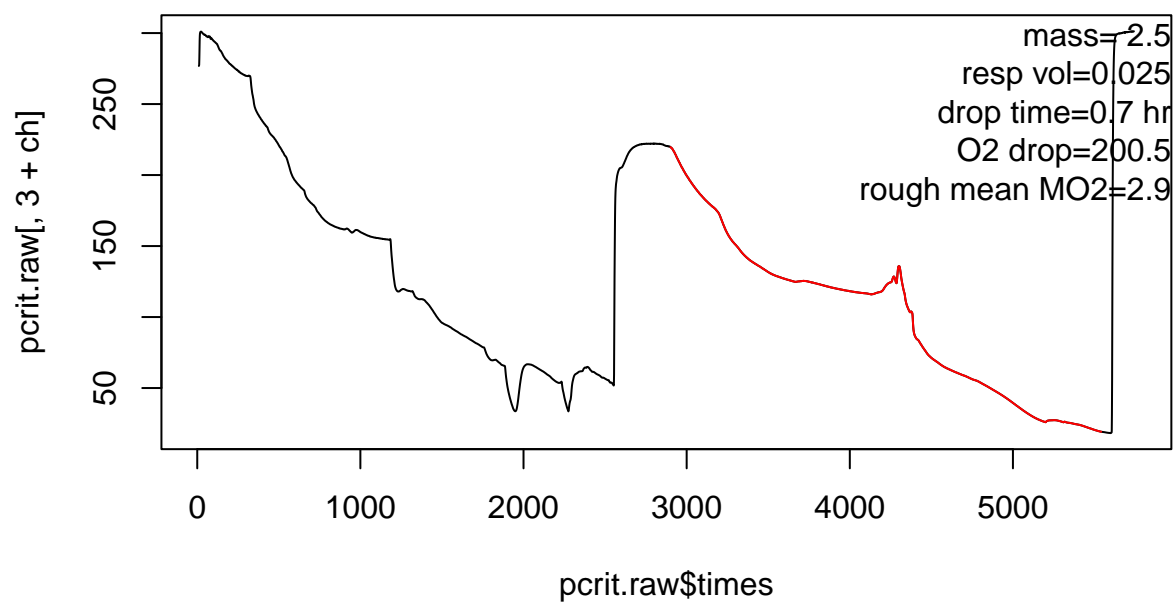
LLO @ MR of 2.22 = 13.235

NLR () = NA

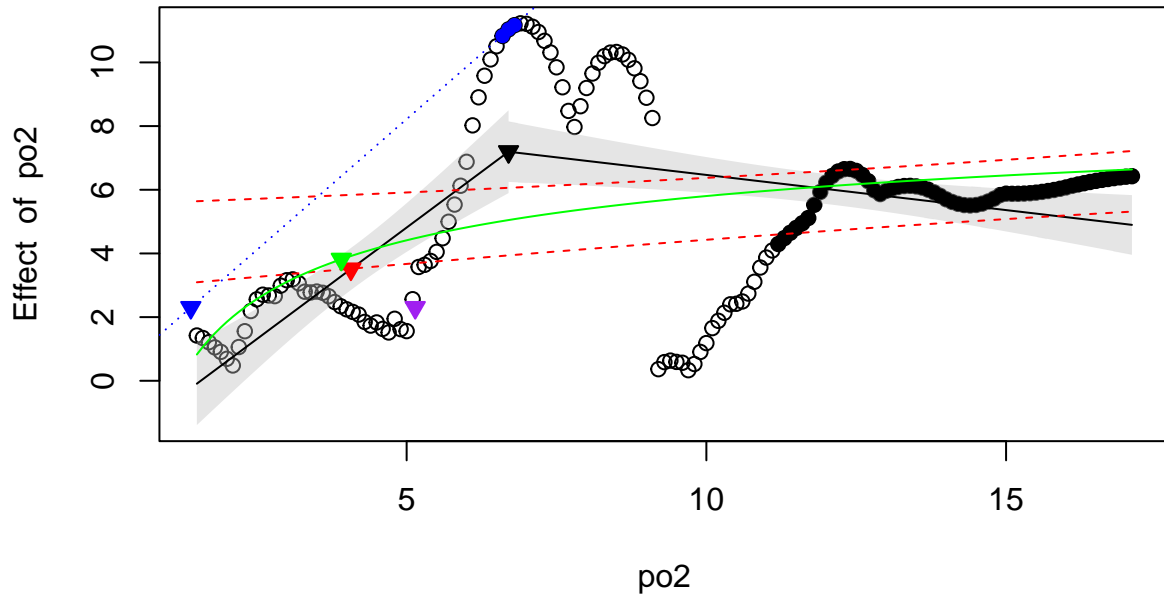
Sub-PI = 18.7



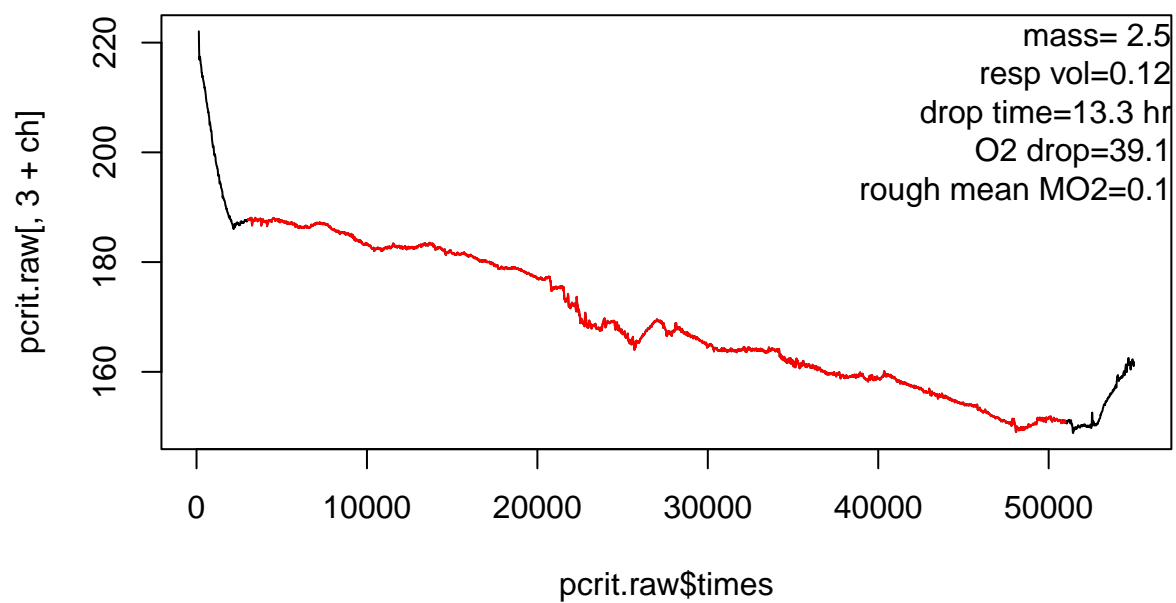
Gr1 Muus 1800-2 pcrit 25 ml jar 7-29-21 ch2 blank.txt



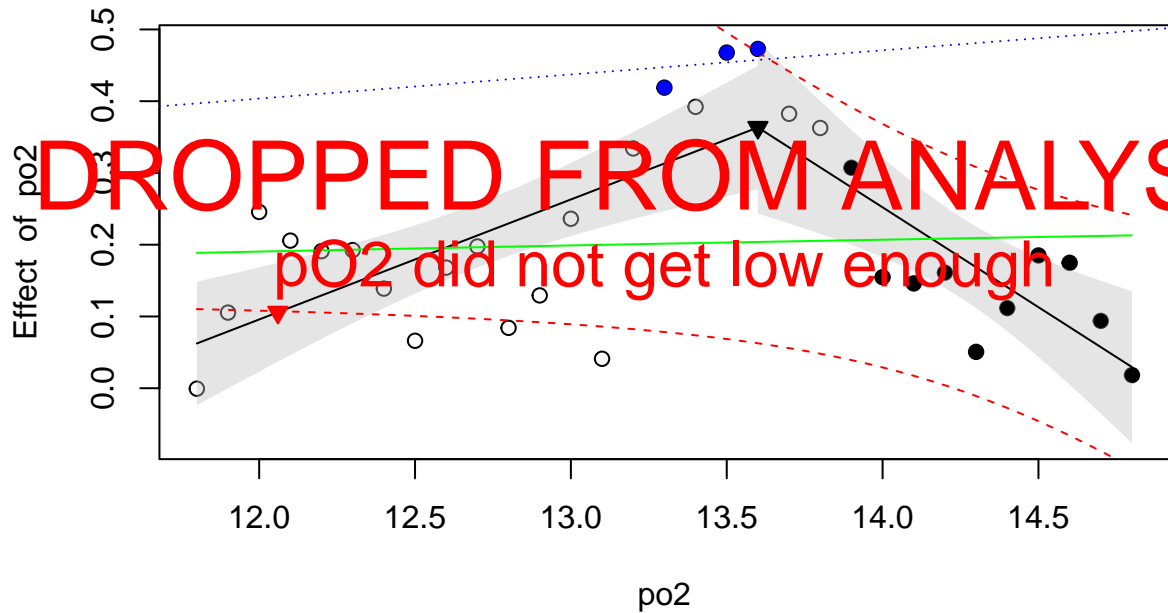
Alpha @ MR of 2.3 = 1.4
Breakpoint = 6.7
LLO @ MR of 2.3 = 5.143
NLR (Pareto) = 3.91
Sub-PI = 4.07



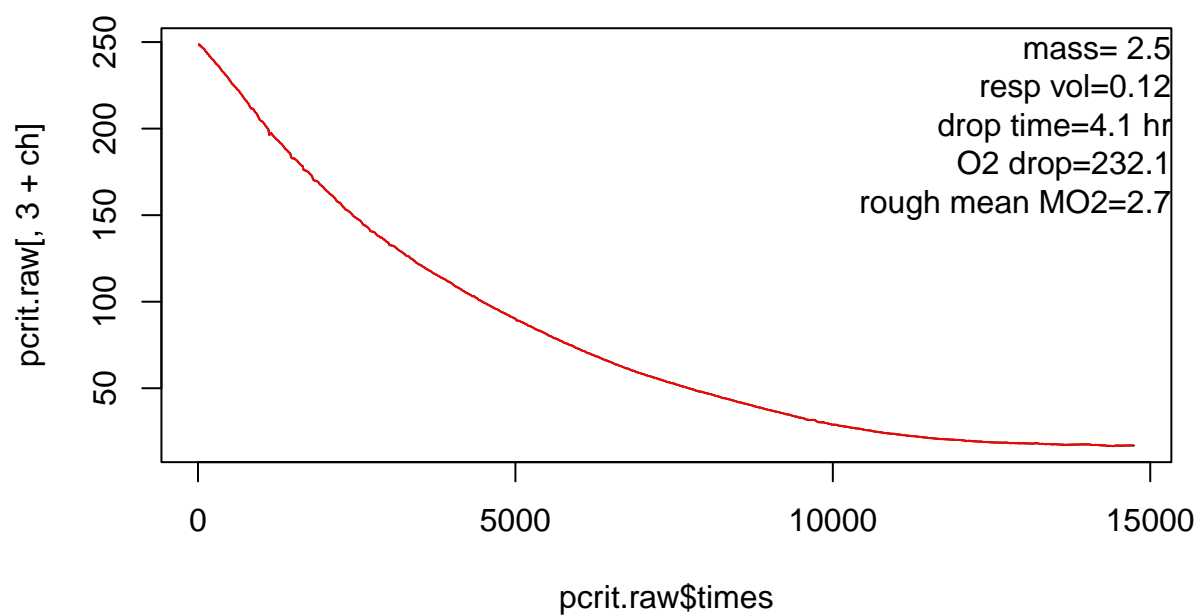
Gr1 Muus 1800-2 pcrit 7-28-21.txt



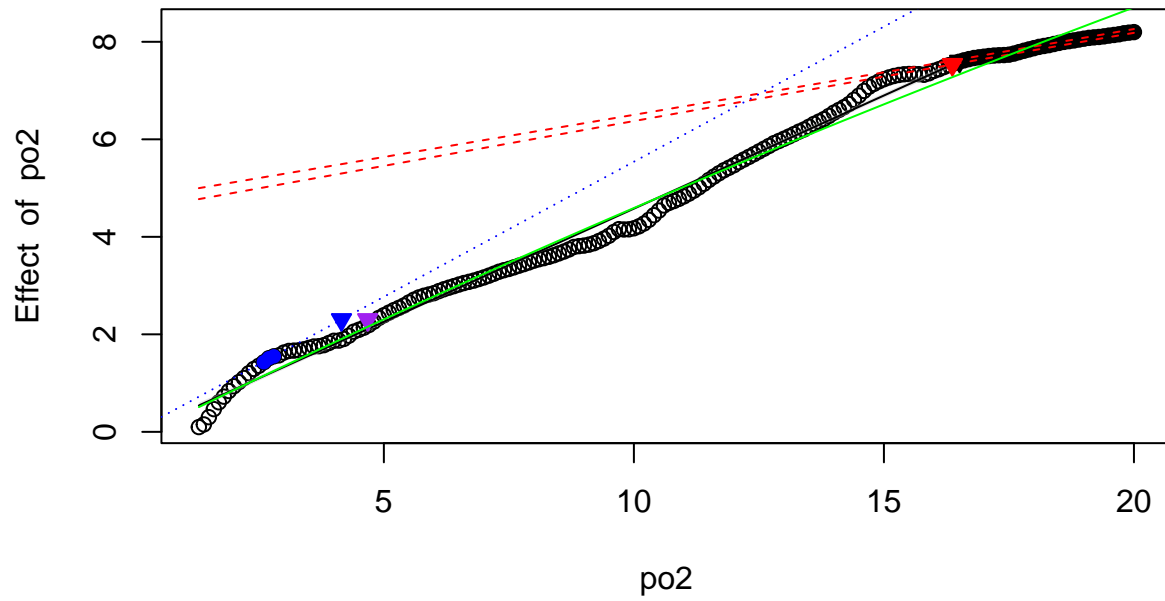
Alpha @ MR of 0.14 = 4.172
Breakpoint = 13.6
LLO @ MR of NA = NA
NLR (Michaelis-Menten) = 0.829
Sub-PI = 12.06



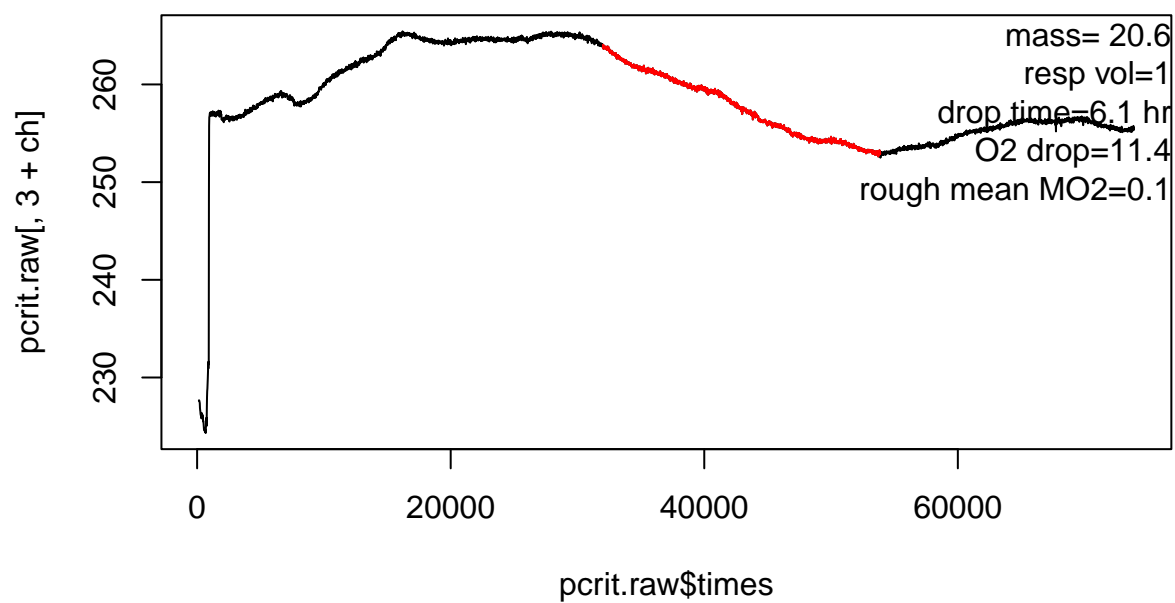
GR1 Muus 1800-2 pcrit day7 8-3-21.txt



Alpha @ MR of 2.3 = 4.153
Breakpoint = 16.514
LLO @ MR of 2.3 = 4.676
NLR (Hyperbola) = -0.483
Sub-PI = 16.37



GR1 Muus1000 7day-7-26-21.txt



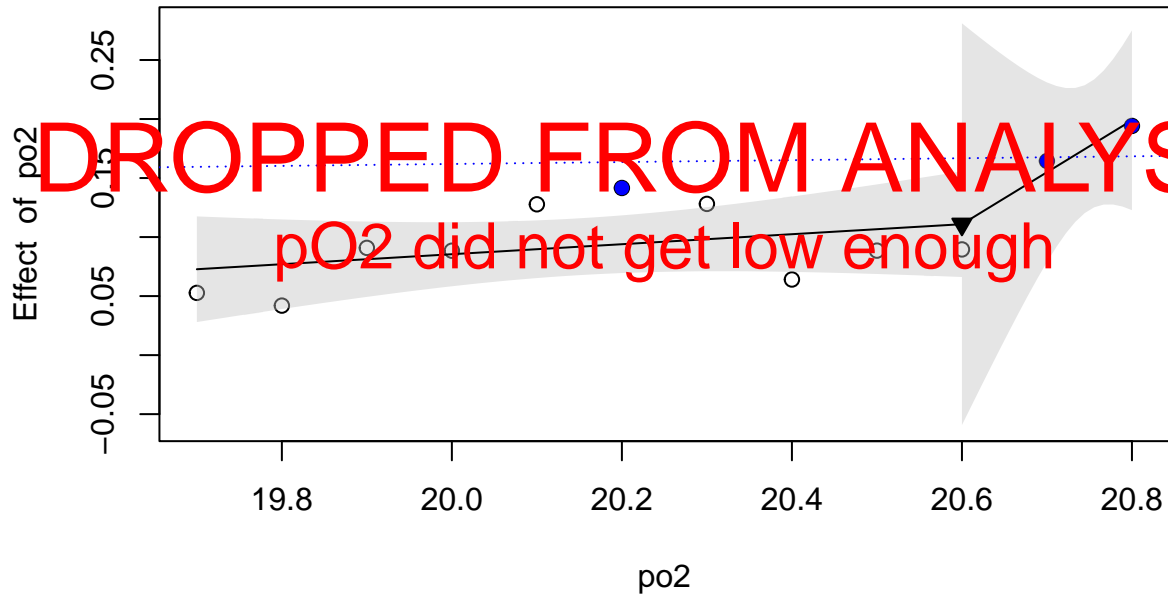
Alpha @ MR of 0.18 = 22.147

Breakpoint = 20.6

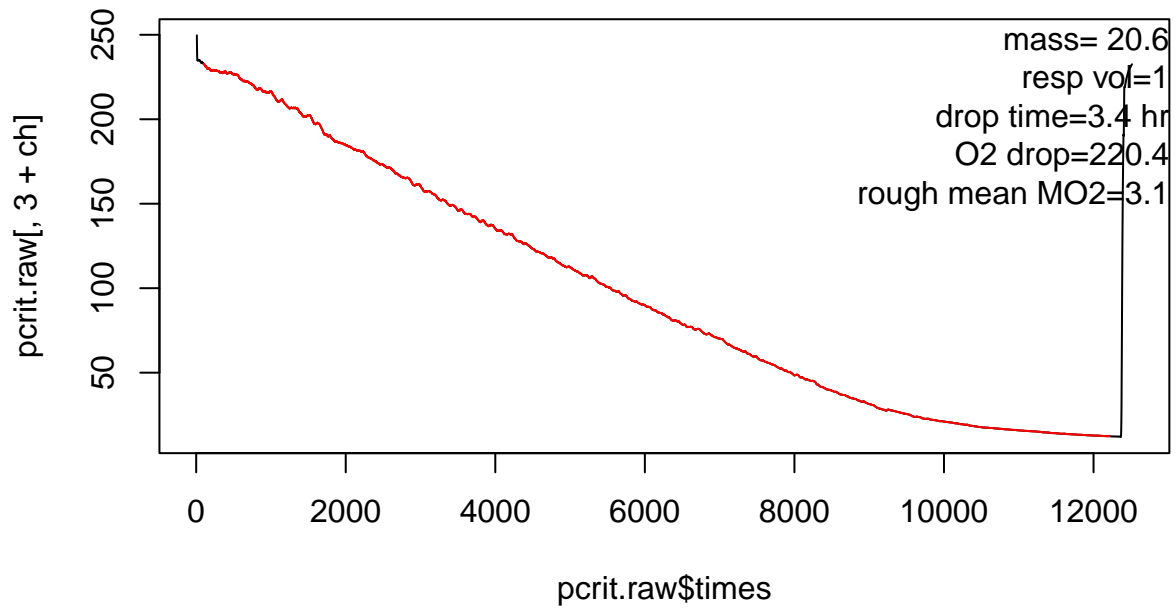
LLO @ MR of NA = NA

NLR () = NA

Sub-PI =



GR1 Muus1000 pcrit 7-21-21.txt



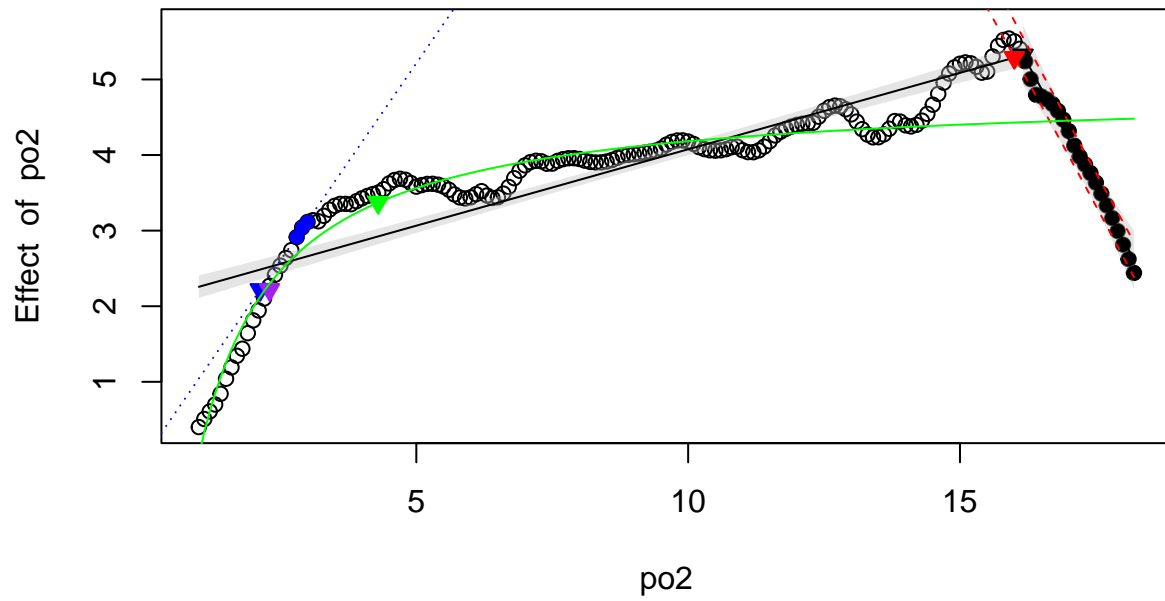
Alpha @ MR of 2.22 = 2.128

Breakpoint = 16.161

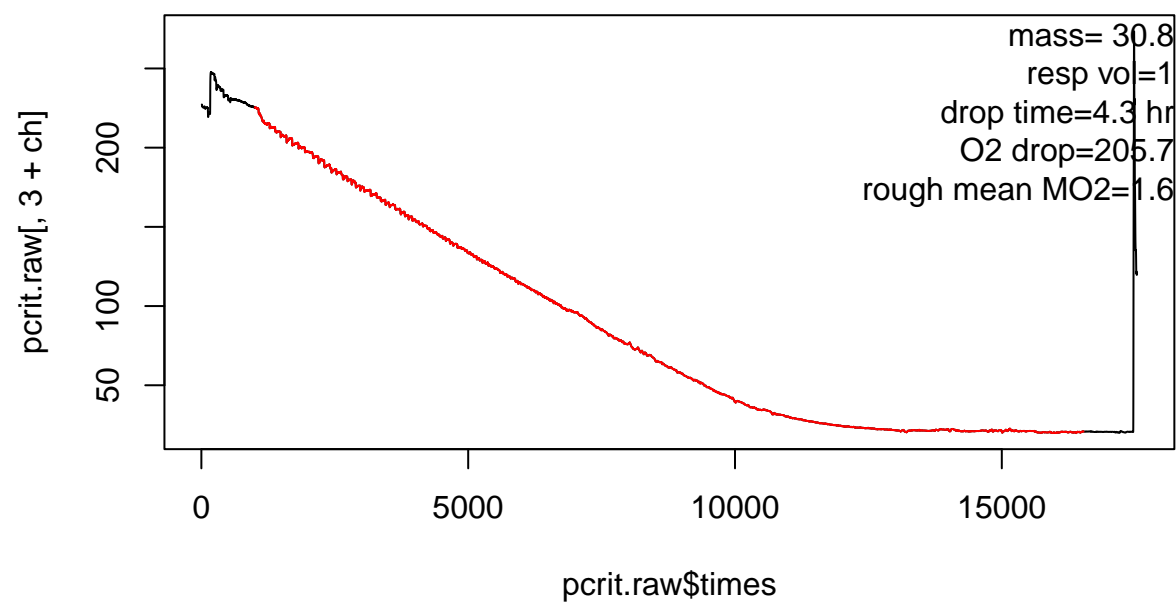
LLO @ MR of 2.22 = 2.3

NLR (Hyperbola) = 4.298

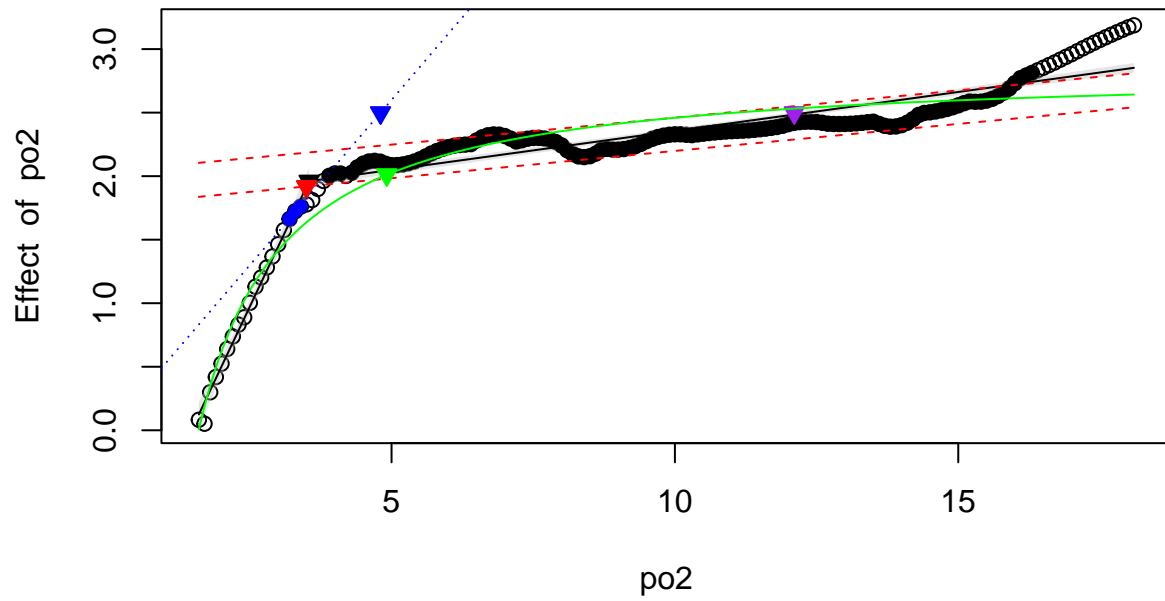
Sub-PI = 16



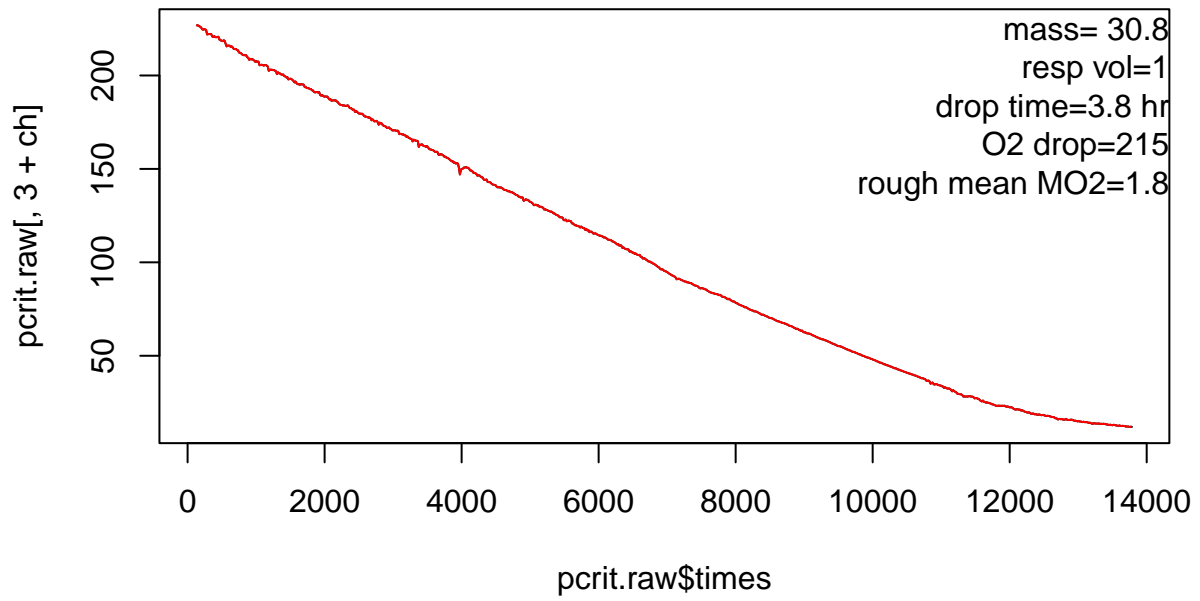
GR1 Muus1800 7day-pcrit 7-20-21.txt



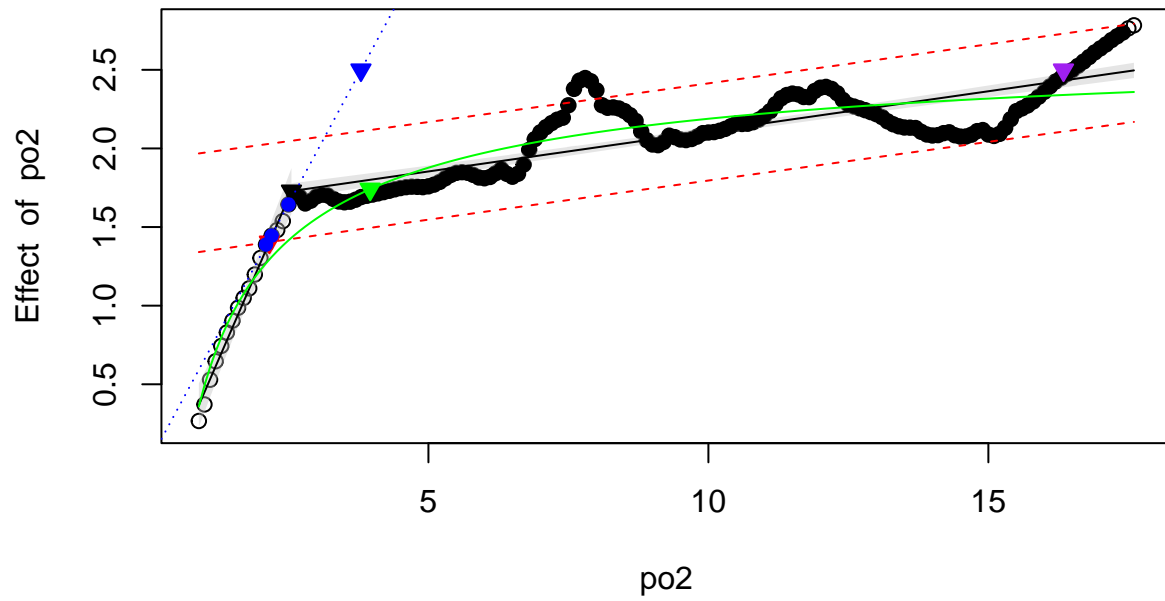
Alpha @ MR of 2.5 = 4.806
Breakpoint = 3.545
LLO @ MR of 2.5 = 12.106
NLR (Pareto) = 4.917
Sub-PI = 3.5



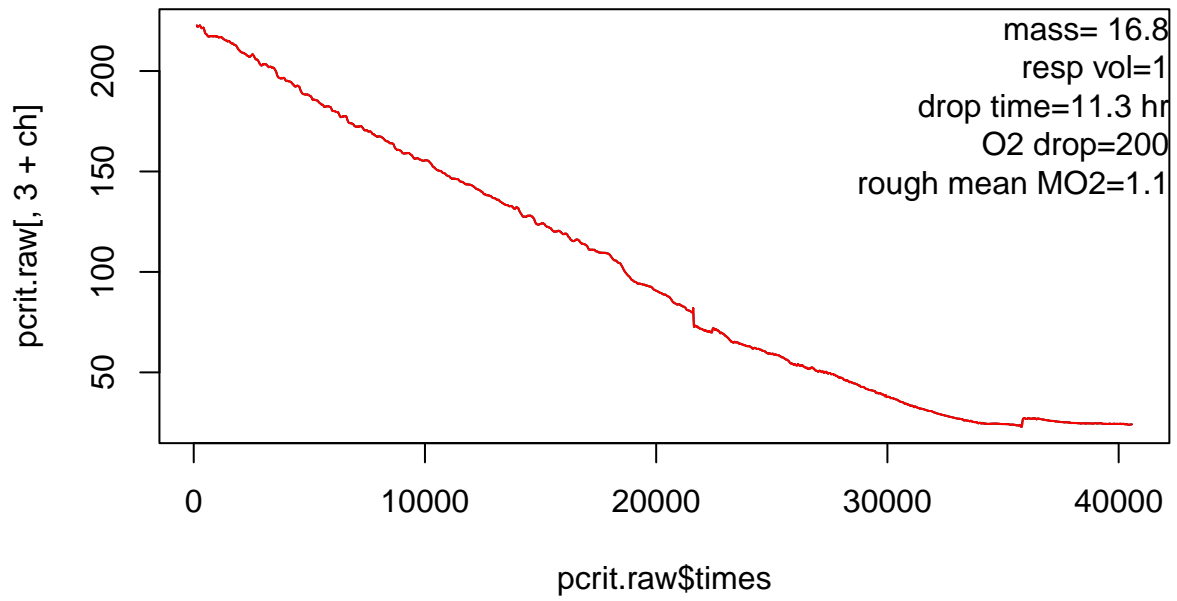
GR1 Muus1800 pcrit 7-13-21.txt



Alpha @ MR of 2.5 = 3.795
Breakpoint = 2.554
LLO @ MR of 2.5 = 16.341
NLR (Weibull with intercept) = 3.96
Sub-PI = 2.16



gr2muus1800 7day pcrit 7-20-21.txt



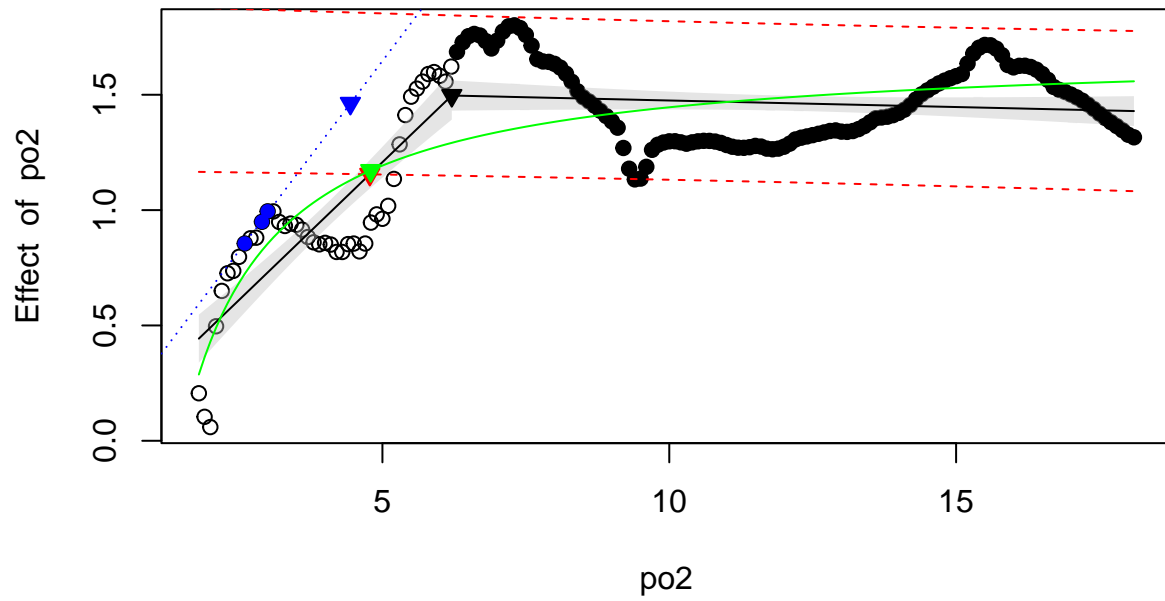
Alpha @ MR of 1.46 = 4.442

Breakpoint = 6.209

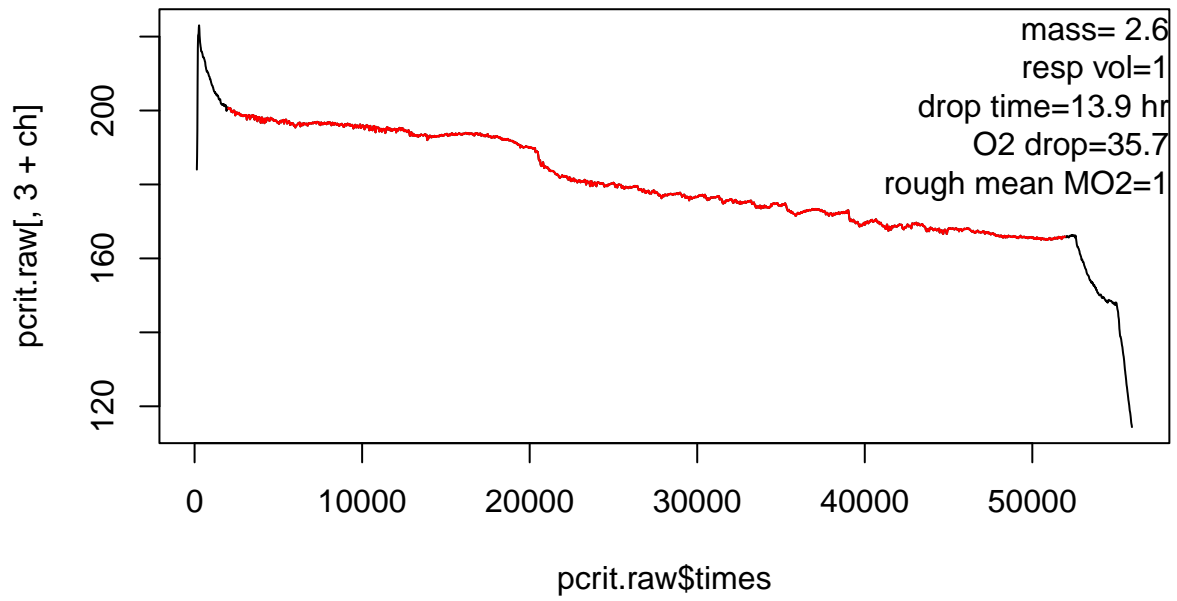
LLO @ MR of NA = NA

NLR (Pareto) = 4.796

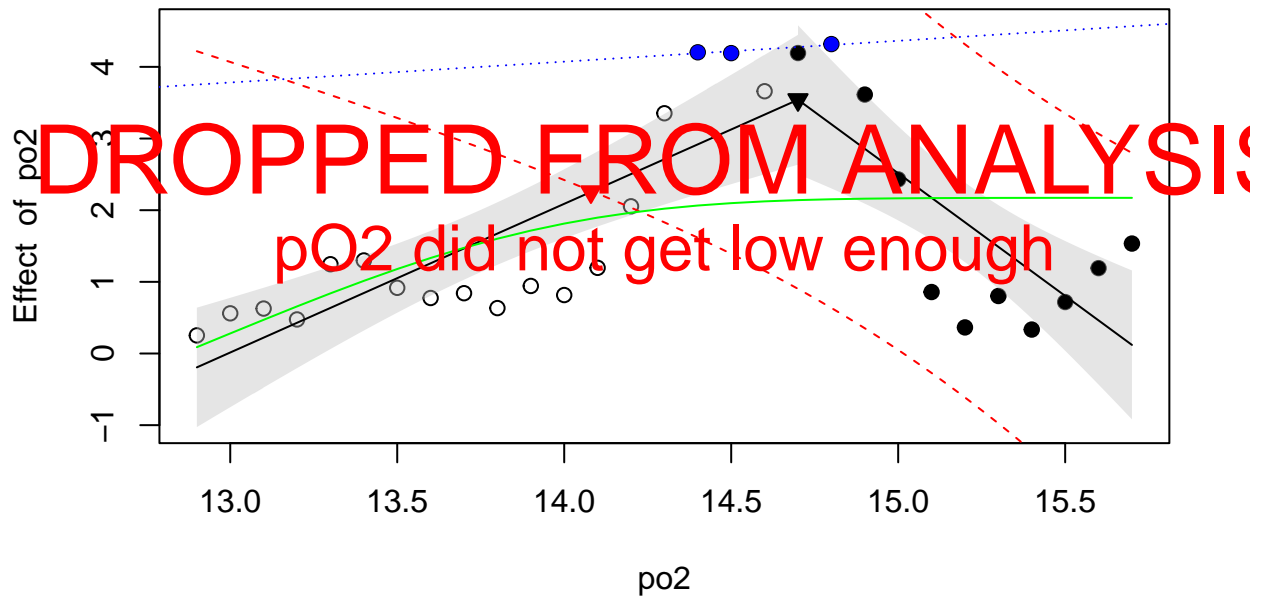
Sub-PI = 4.78



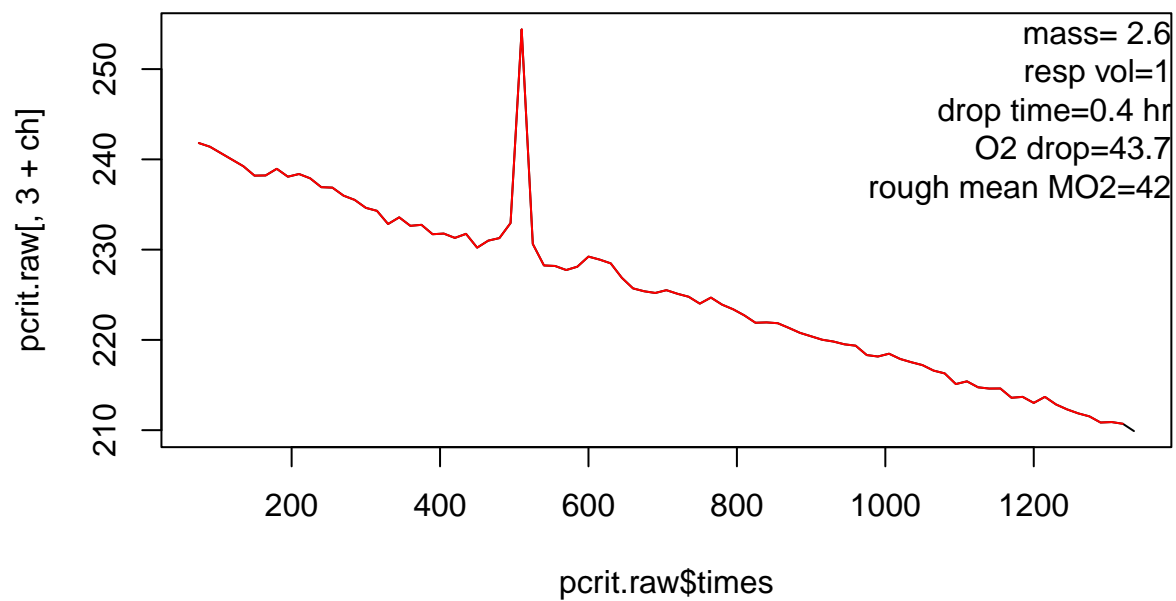
gr2muus1800-2 pcrit 7-28-21.txt



Alpha @ MR of 1.85 = 6.361
Breakpoint = 14.7
LLO @ MR of NA = NA
NLR (Weibull with intercept) = 10.283
Sub-PI = 14.08



gr2muus1800-2 pcrit day7 8-3-21.txt



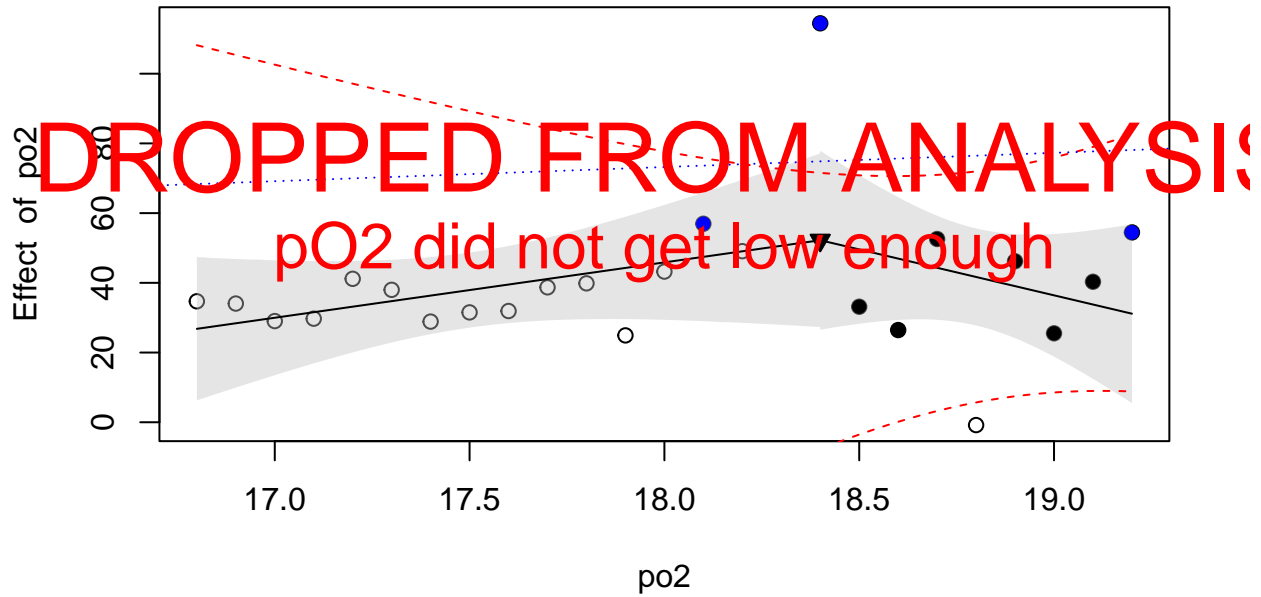
Alpha @ MR of 22.09 = 5.43

Breakpoint = 18.4

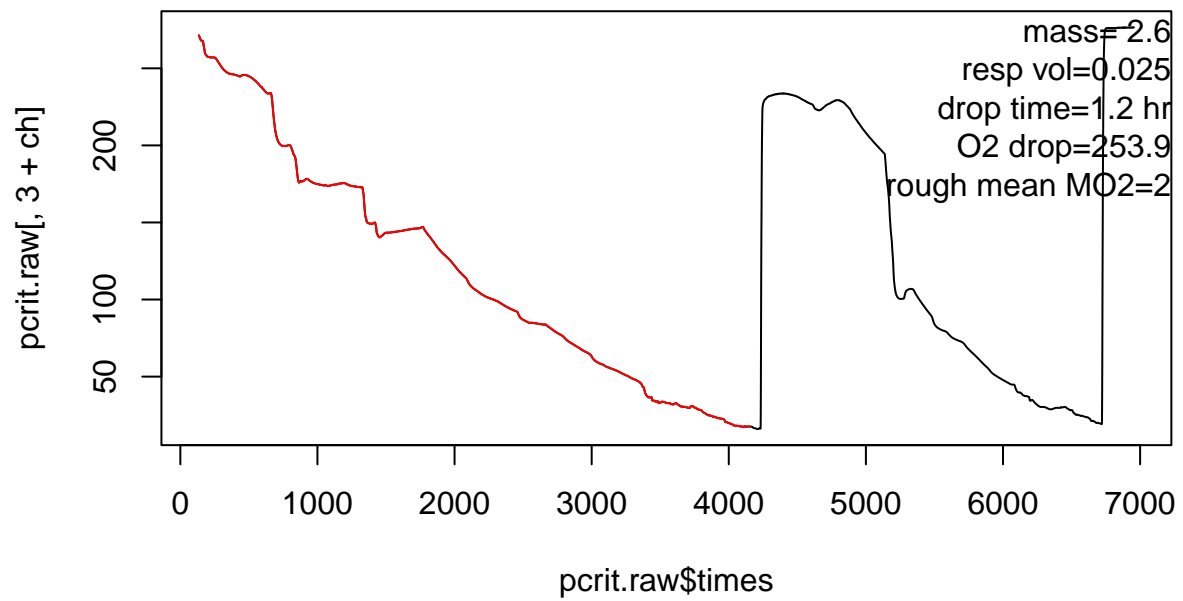
LLO @ MR of 22.09 = NA

NLR () = NA

Sub-PI =



gr2muus1800-2 pcrit in 25 ml jar 7-29-21 ch2 is blank.txt



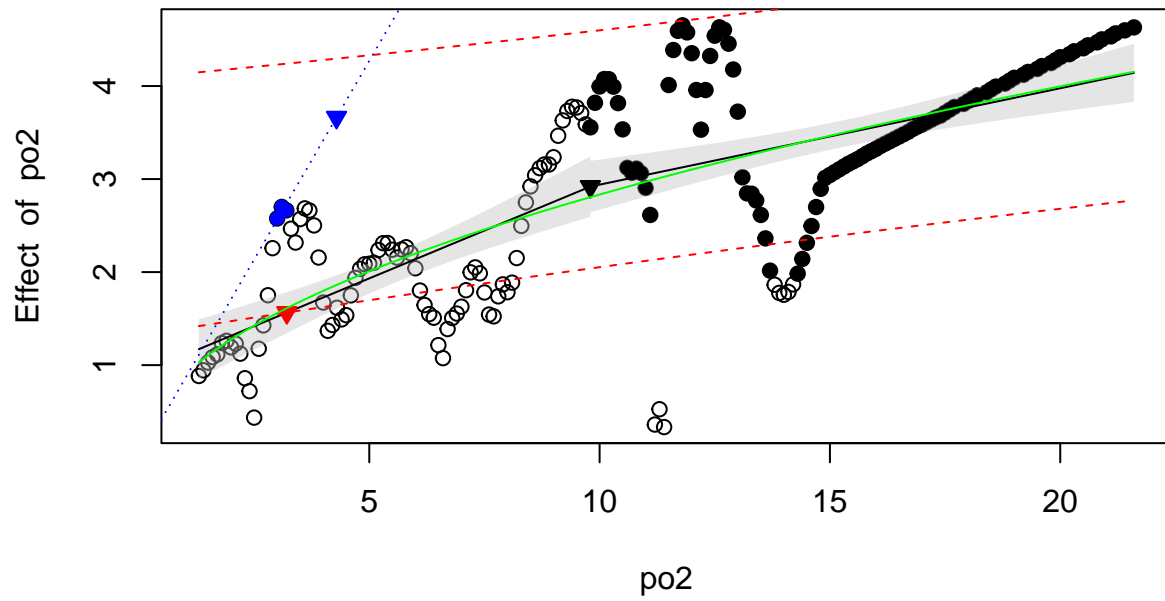
Alpha @ MR of 3.66 = 4.289

Breakpoint = 9.8

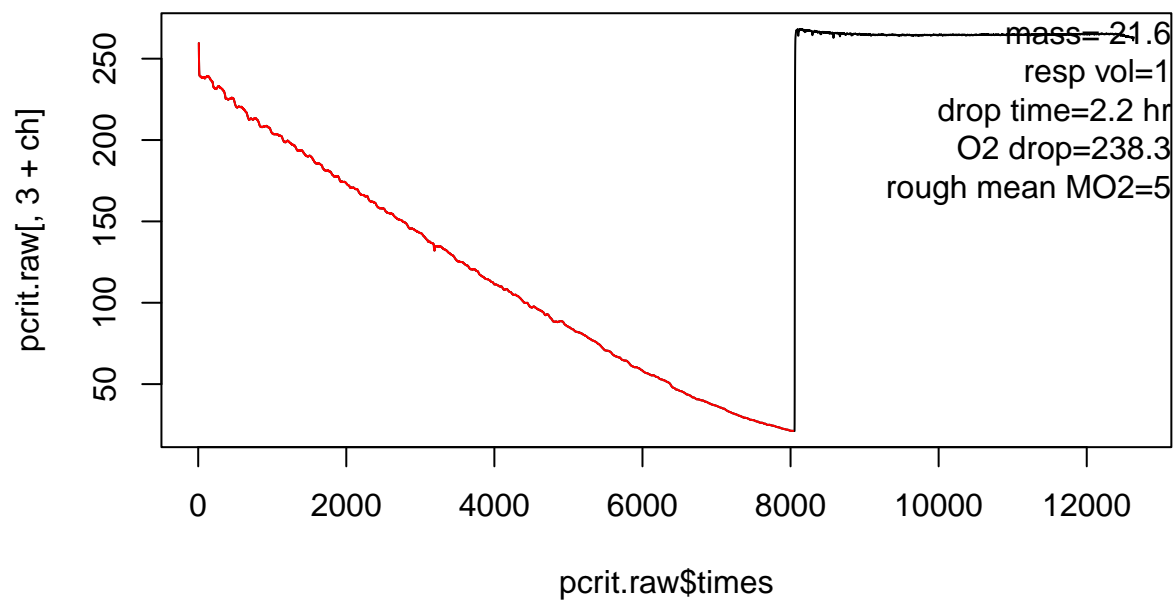
LLO @ MR of NA = NA

NLR (Power) = 40.873

Sub-PI = 3.21



Gr3 Muus 1000 pcrit 7-21-21.txt



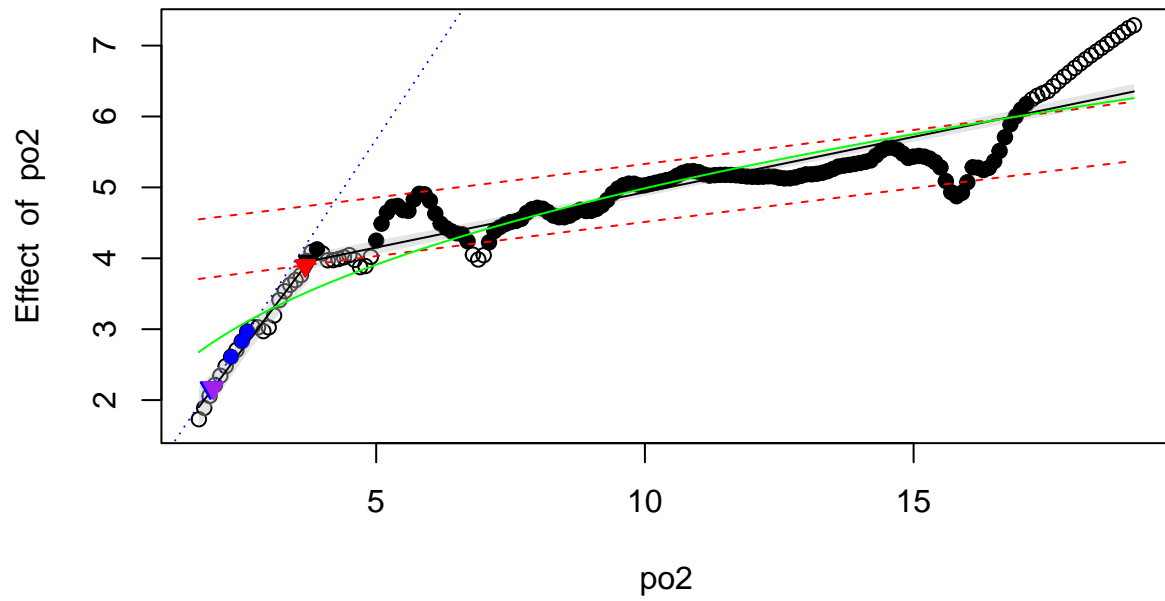
Alpha @ MR of 2.17 = 1.913

Breakpoint = 3.74

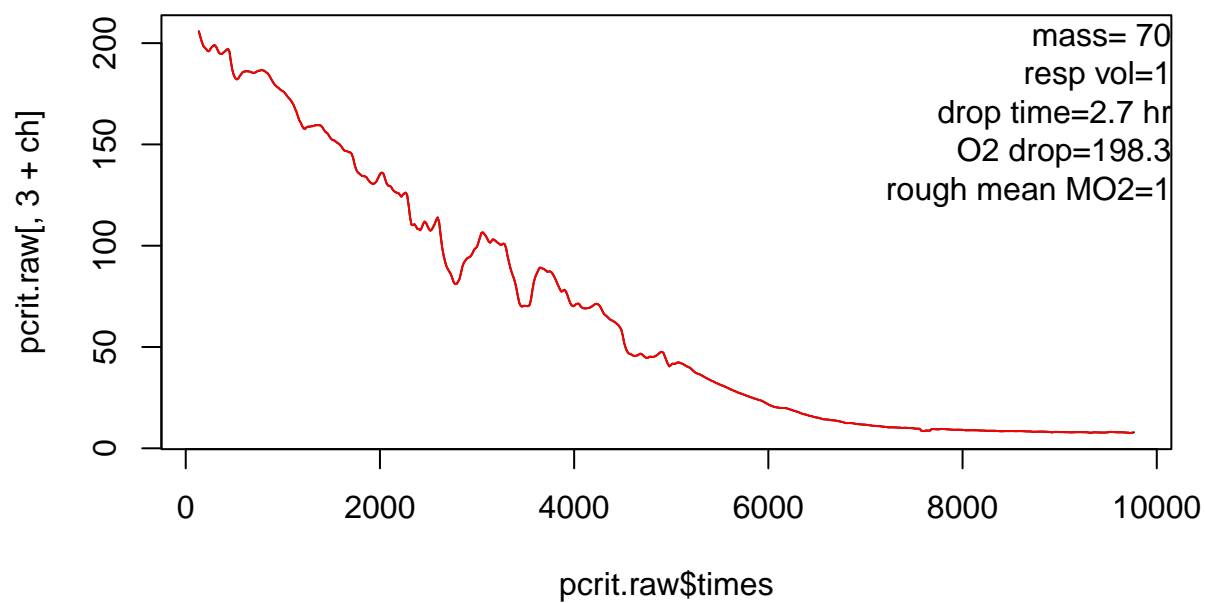
LLO @ MR of 2.17 = 1.97

NLR (Power) = 67.77

Sub-PI = 3.68



gr3 muus 1800 7day Pcrit 7-20-21.txt



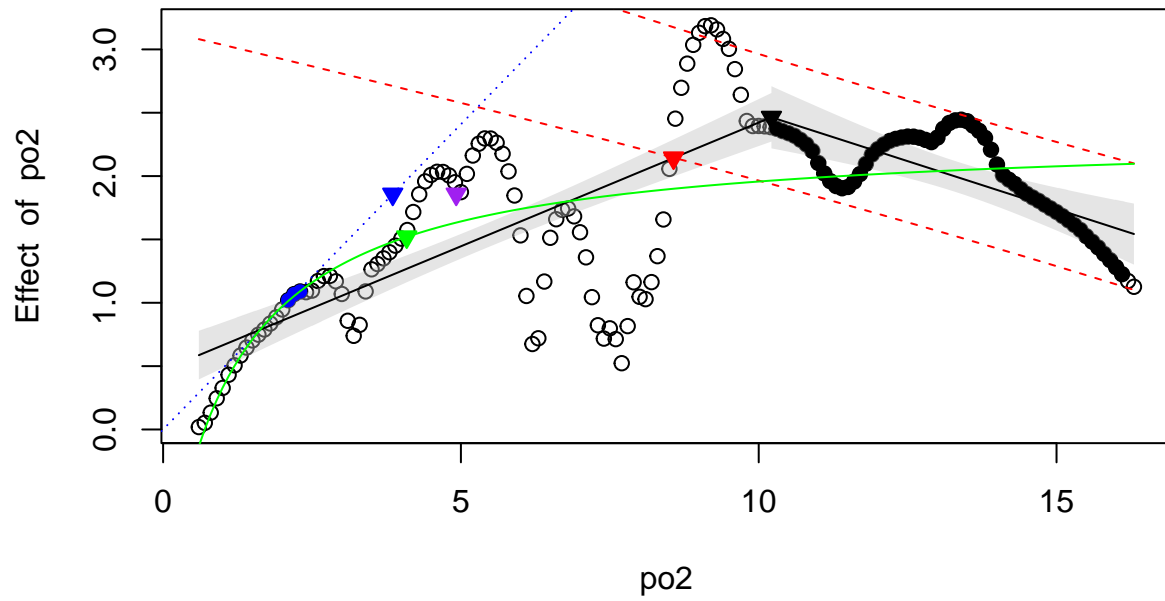
Alpha @ MR of 1.86 = 3.855

Breakpoint = 10.214

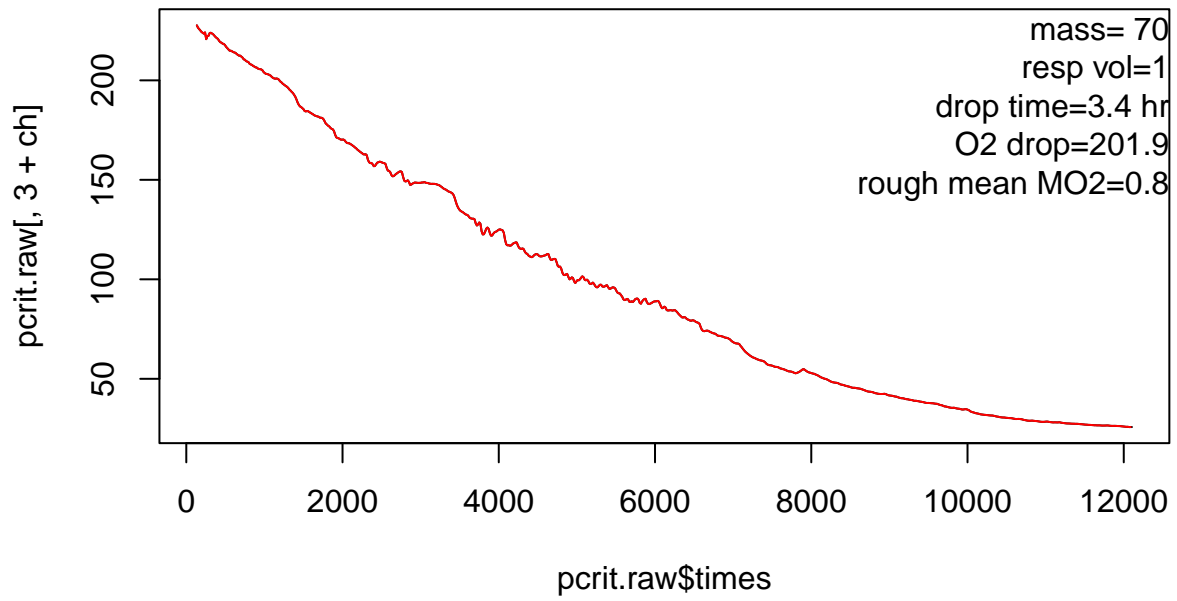
LLO @ MR of 1.86 = 4.921

NLR (Hyperbola) = 4.09

Sub-PI = 8.57



gr3 muus 1800 pcrit 7-13-21.txt



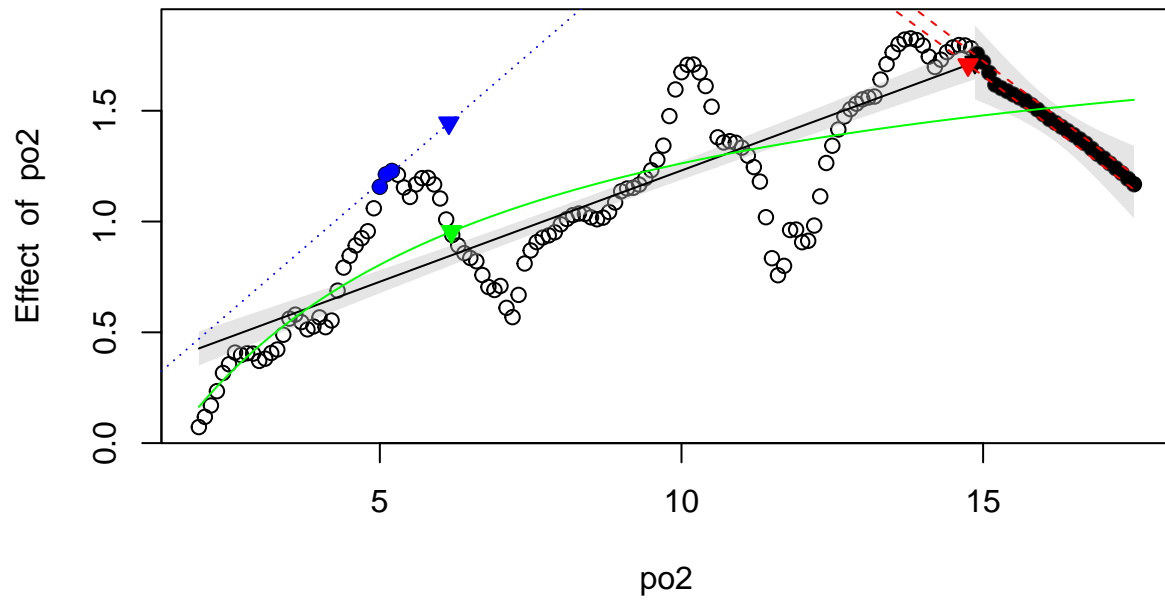
Alpha @ MR of 1.44 = 6.145

Breakpoint = 14.864

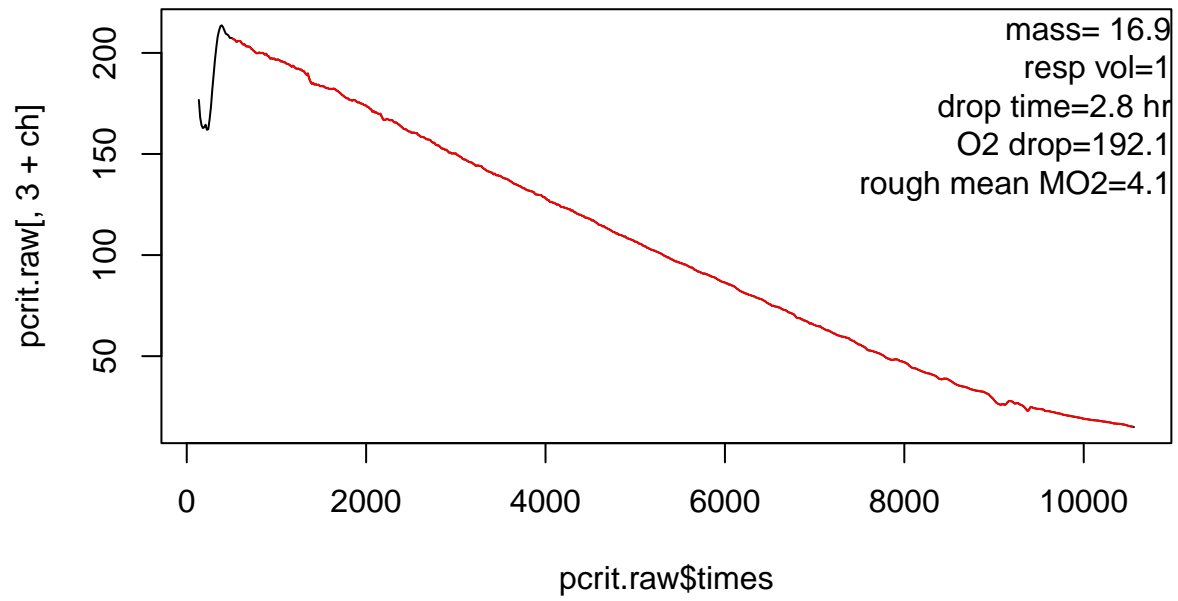
LLO @ MR of NA = NA

NLR (Hyperbola) = 6.188

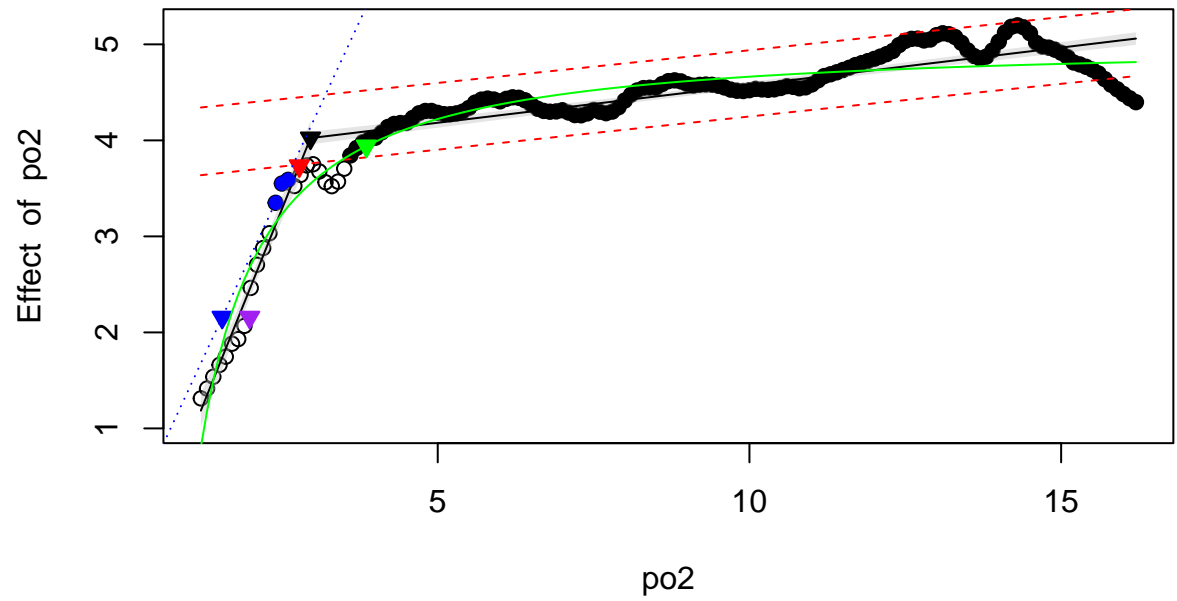
Sub-PI = 14.75



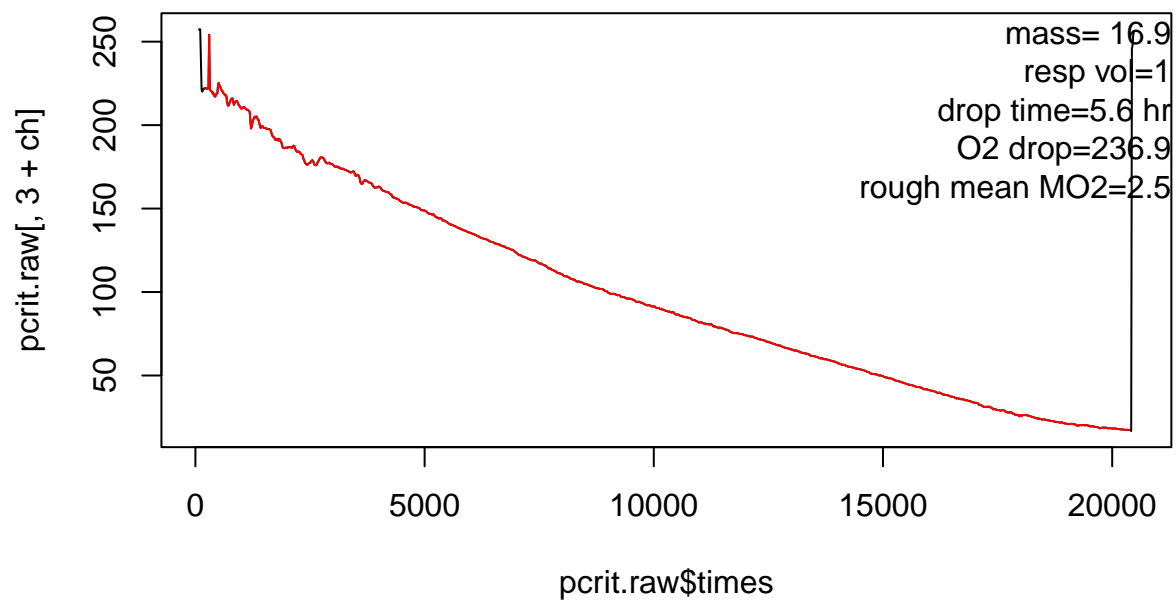
Gr3 Muus 1800-2 pcrit 07-28-21.txt



Alpha @ MR of 2.16 = 1.542
Breakpoint = 2.959
LLO @ MR of 2.16 = 1.983
NLR (Pareto) = 3.857
Sub-PI = 2.78



Gr3 Muus 1800-2 pcrit 08-03-21.txt



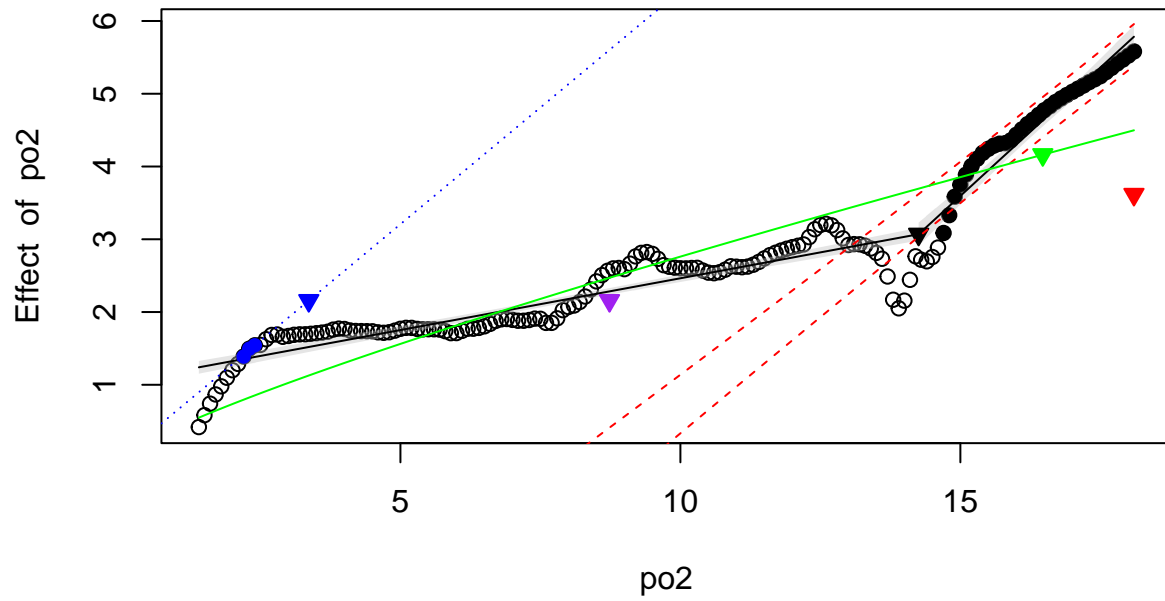
Alpha @ MR of 2.16 = 3.363

Breakpoint = 14.254

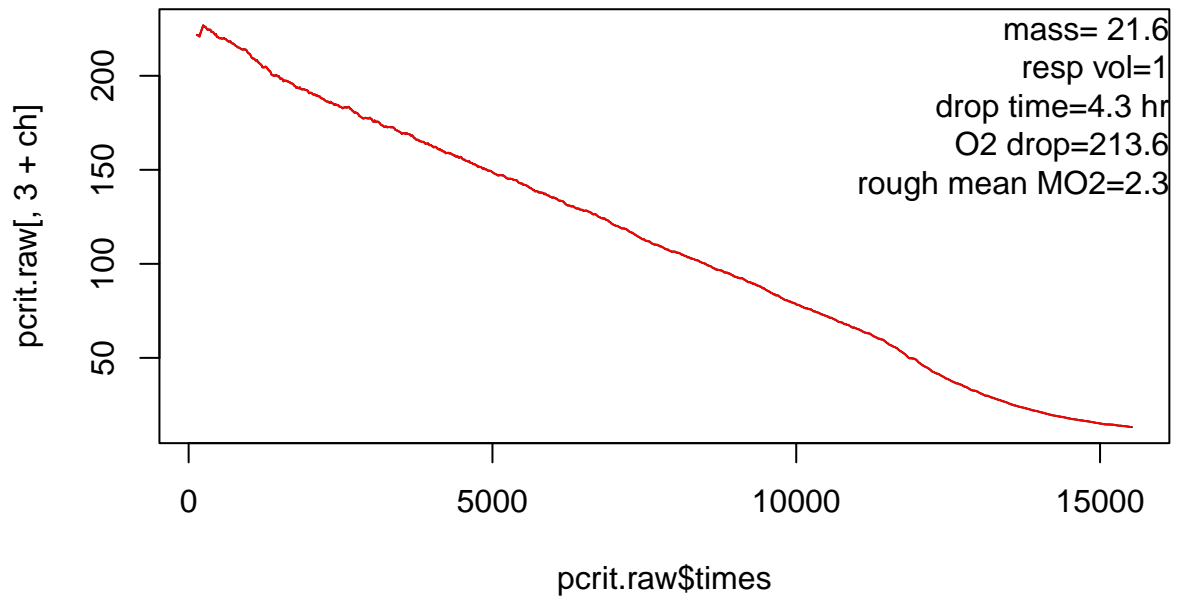
LLO @ MR of 2.16 = 8.734

NLR (Power) = 16.472

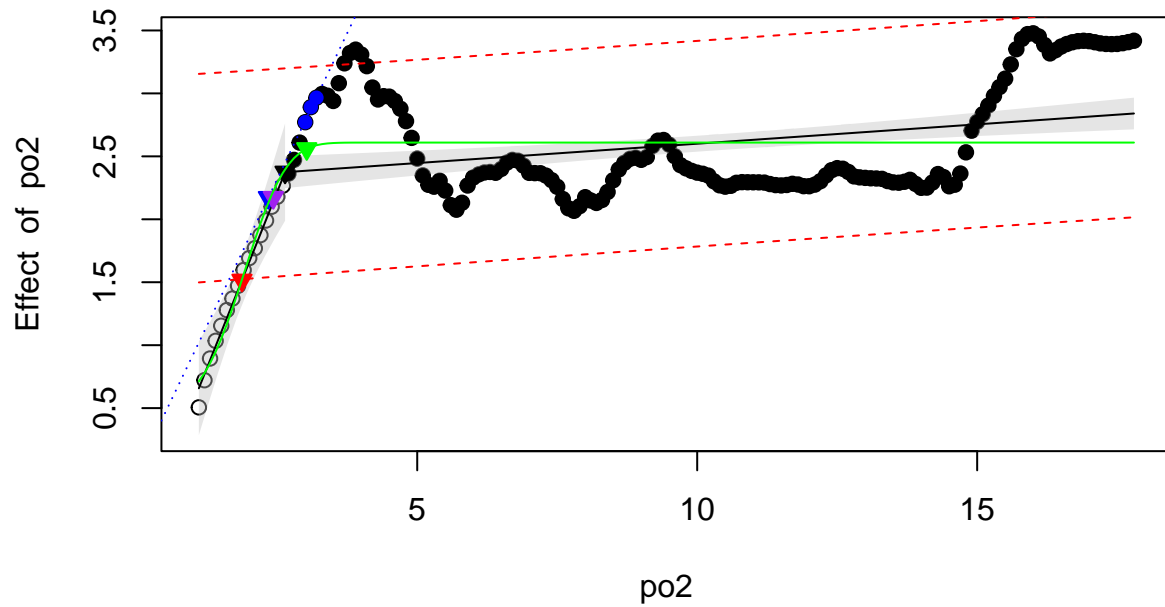
Sub-PI = 18.1



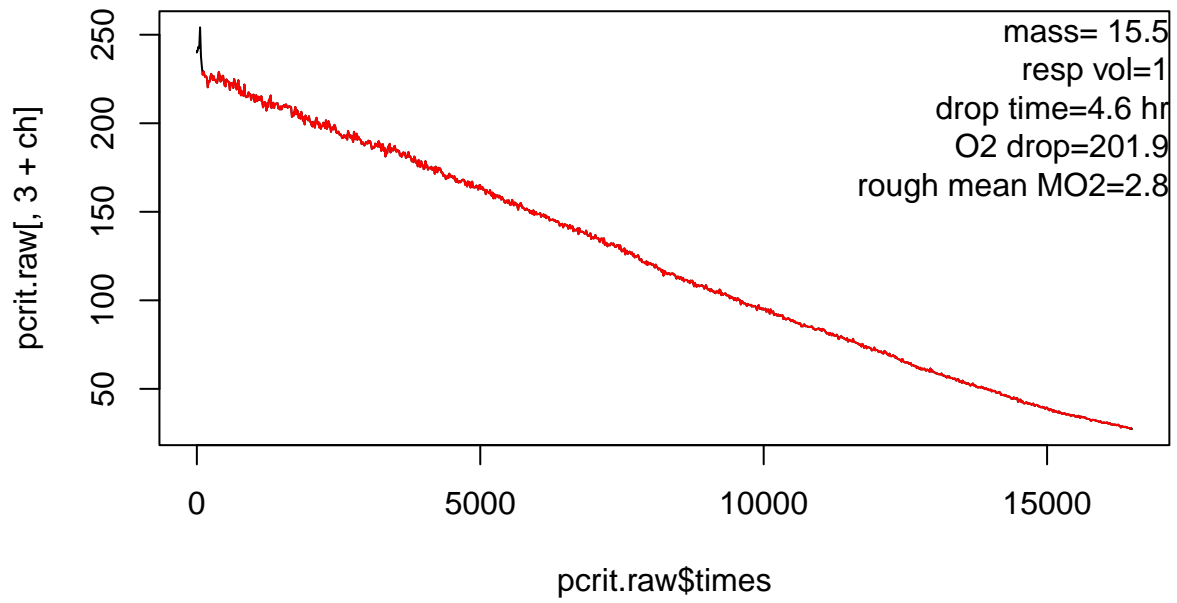
Gr3 Muus1000-2 7 day pcrit 7-27-21.txt



Alpha @ MR of 2.17 = 2.342
Breakpoint = 2.64
LLO @ MR of 2.17 = 2.43
NLR (Weibull with intercept) = 3.027
Sub-PI = 1.87



GR4MUUS1000-2Pcrit-7-26-21-ch1.txt



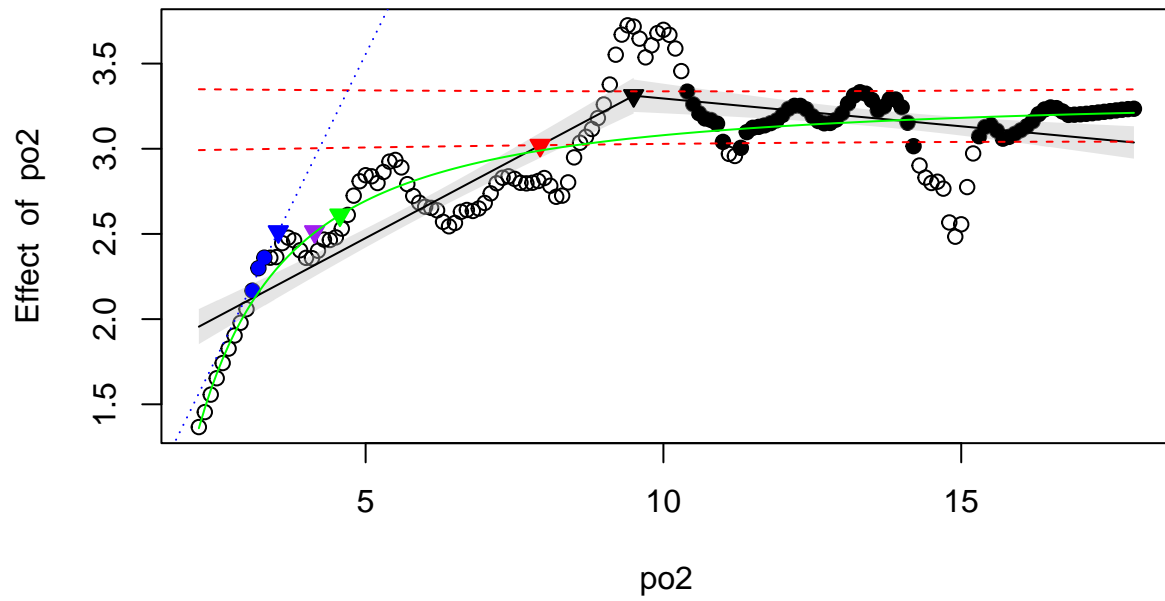
Alpha @ MR of 2.51 = 3.535

Breakpoint = 9.5

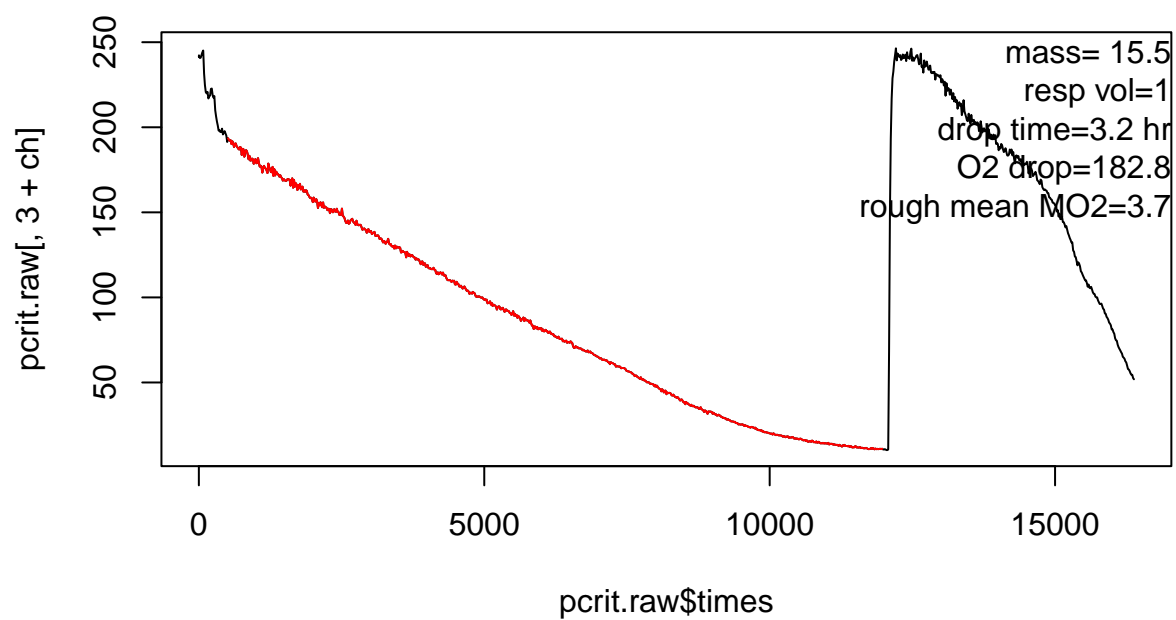
LLO @ MR of 2.51 = 4.144

NLR (Pareto) = 4.569

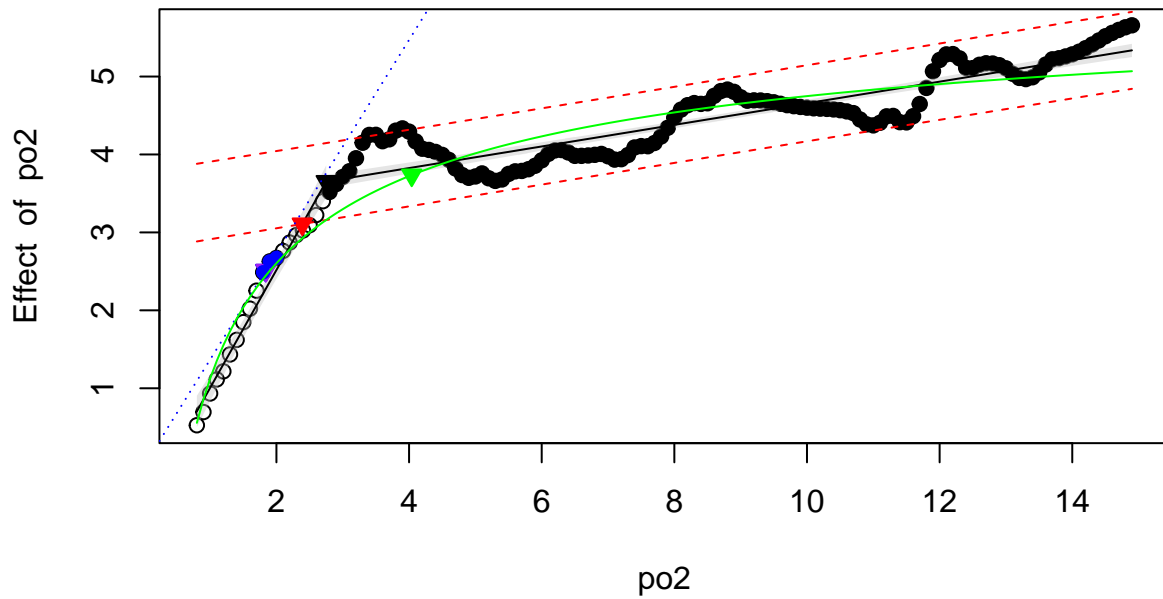
Sub-PI = 7.93



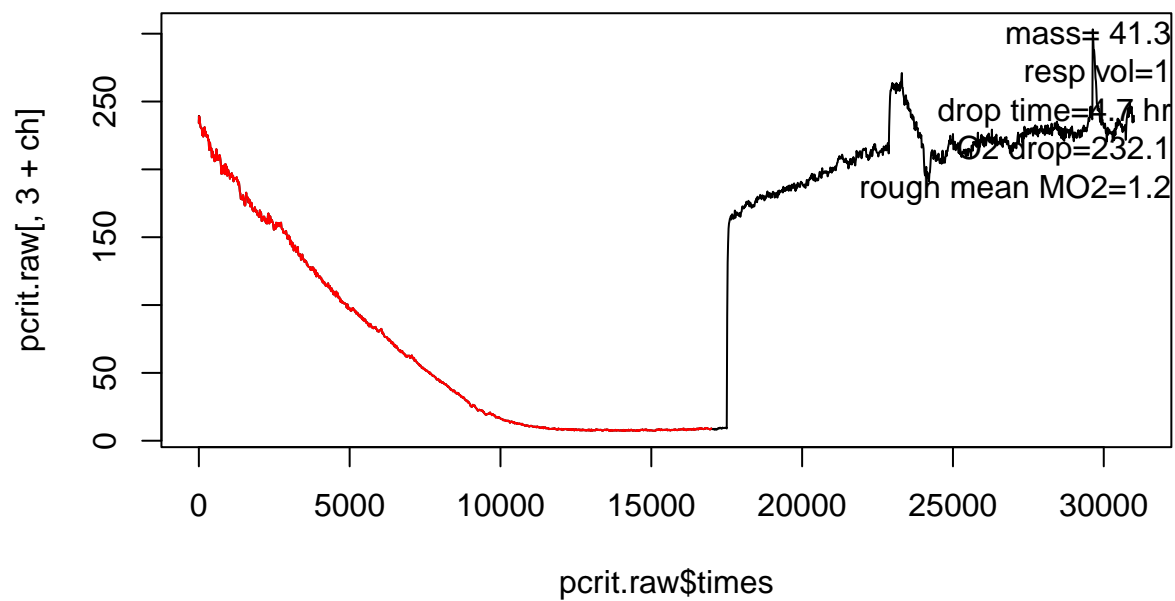
GR4MUUS1000Pcrit-7-21-21-ch1.txt



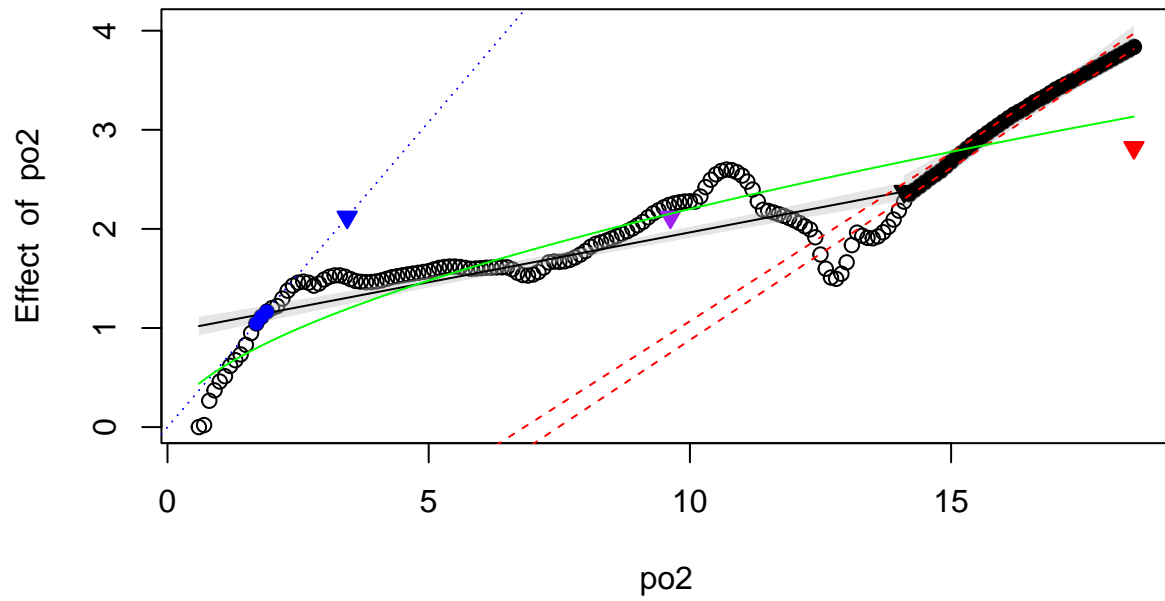
Alpha @ MR of 2.51 = 1.838
Breakpoint = 2.75
LLO @ MR of 2.51 = 1.843
NLR (Weibull with intercept) = 4.04
Sub-PI = 2.39



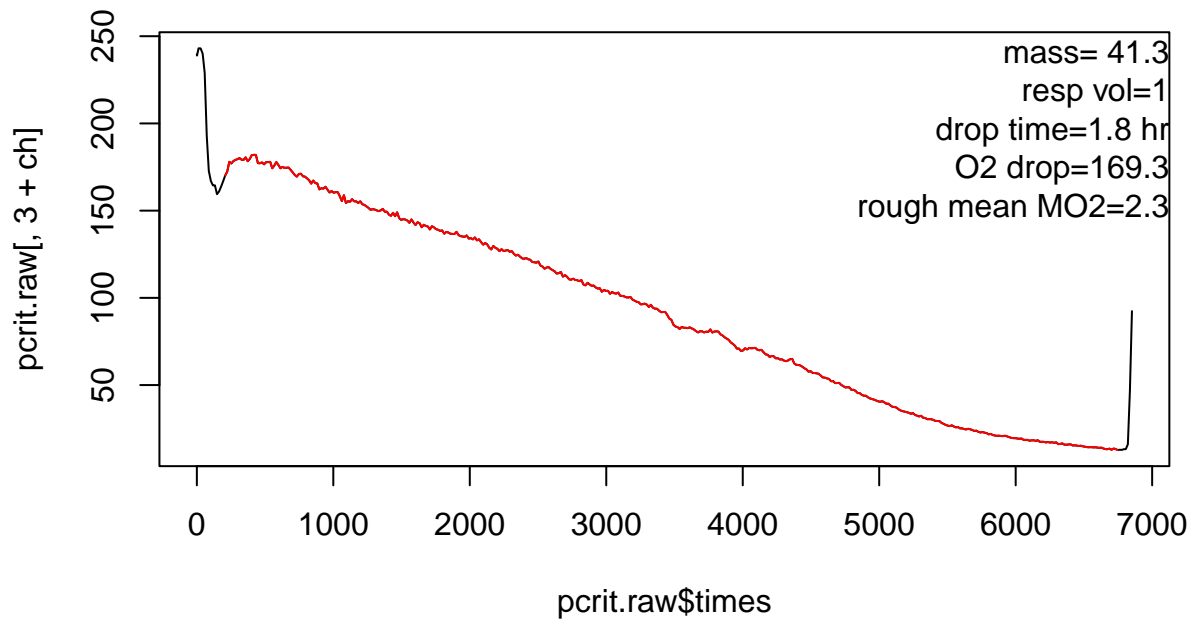
GR4MUUS1800-2-7dayPcrit-8-3-21-ch1.txt



Alpha @ MR of 2.12 = 3.44
Breakpoint = 14.1
LLO @ MR of 2.12 = 9.627
NLR (Power) = 38.75
Sub-PI = 18.5



GR4MUUS1800-2Pcrit-7-28-21-ch1.txt



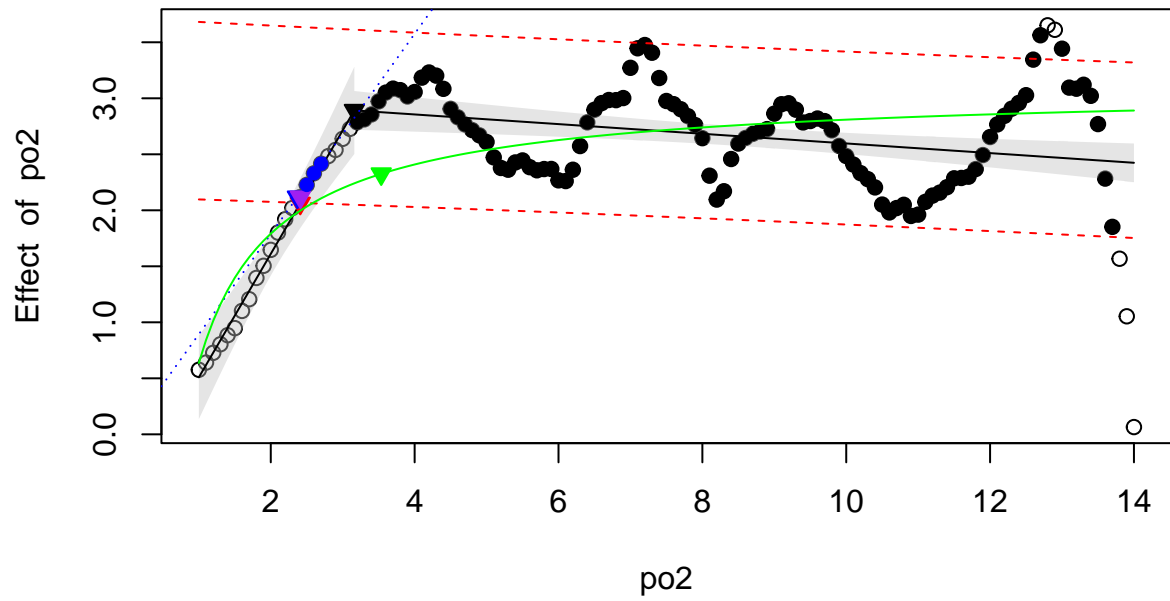
Alpha @ MR of 2.12 = 2.37

Breakpoint = 3.162

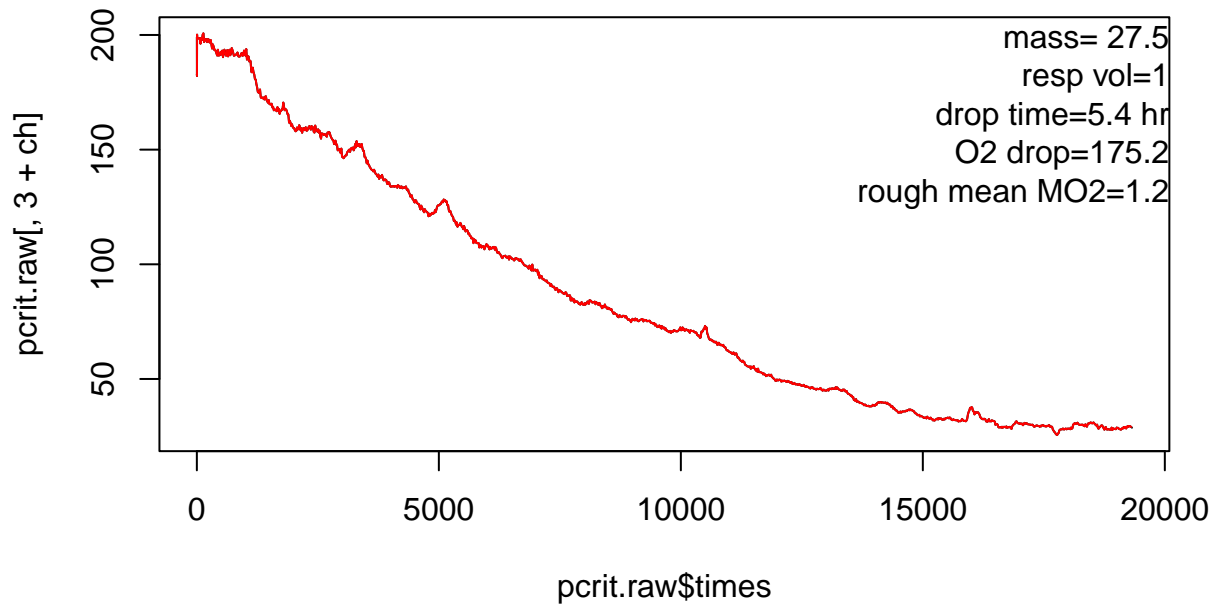
LLO @ MR of 2.12 = 2.413

NLR (Pareto) = 3.535

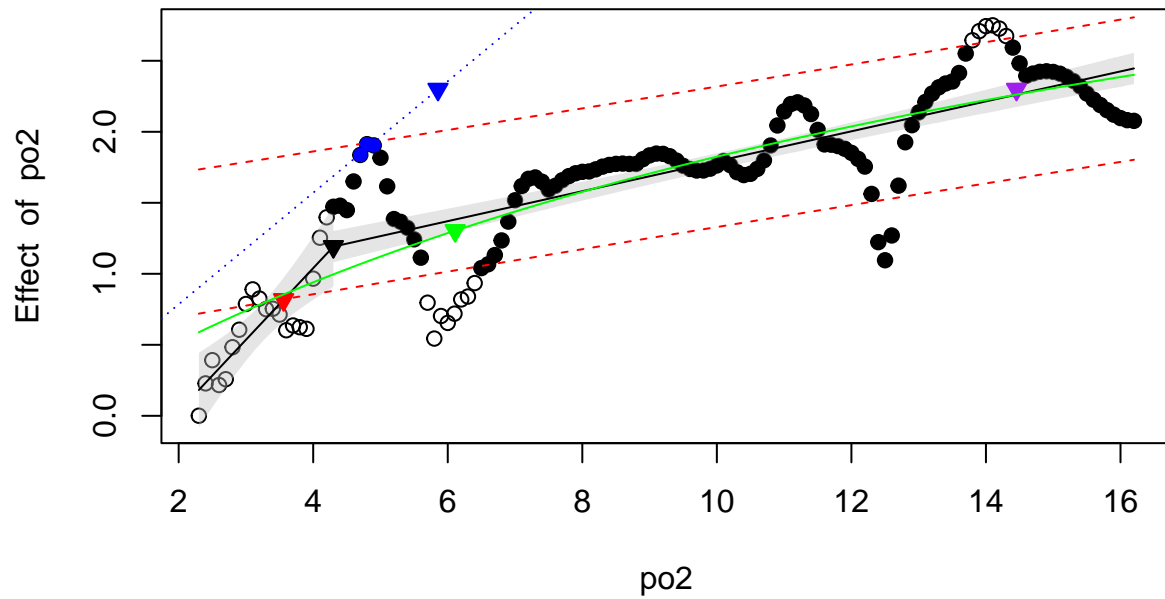
Sub-PI = 2.41



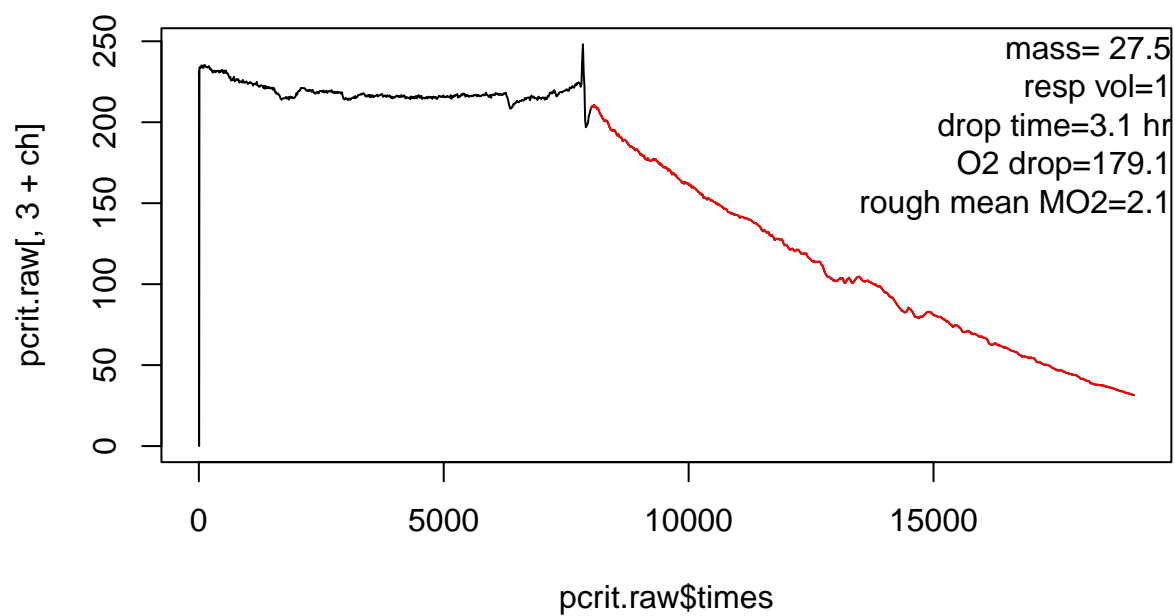
GR4MUUS1800-7dayPcrit-7-20-21-ch1.txt



Alpha @ MR of 2.3 = 5.855
Breakpoint = 4.3
LLO @ MR of 2.3 = 14.454
NLR (Michaelis-Menten) = 6.11
Sub-PI = 3.56



GR4MUUS1800Pcrit-7-13-21-ch1.txt



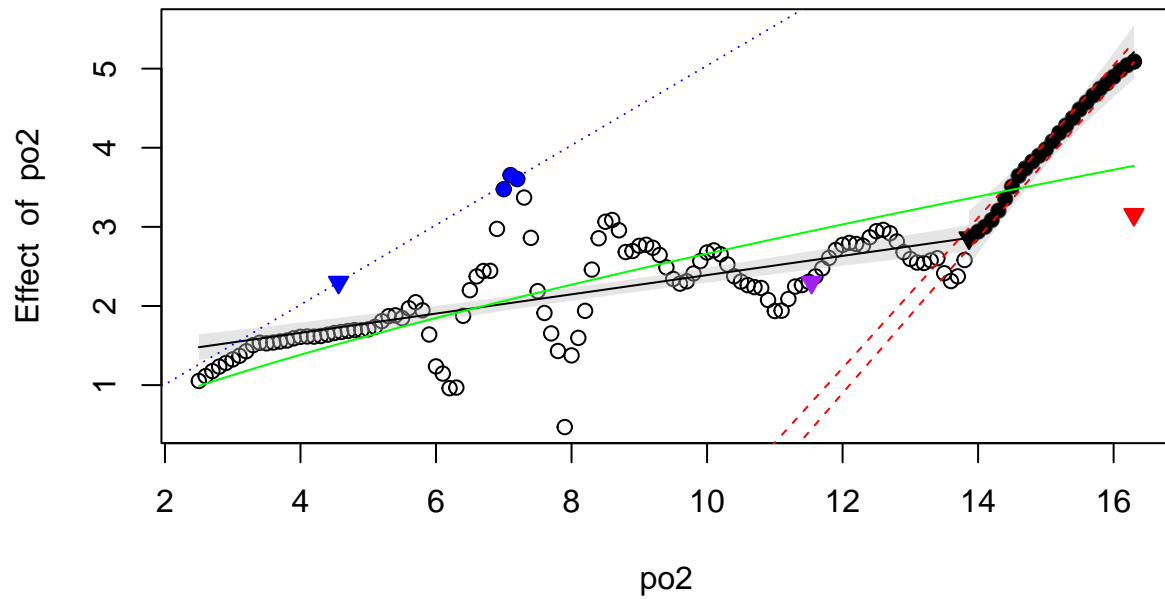
Alpha @ MR of 2.3 = 4.562

Breakpoint = 13.867

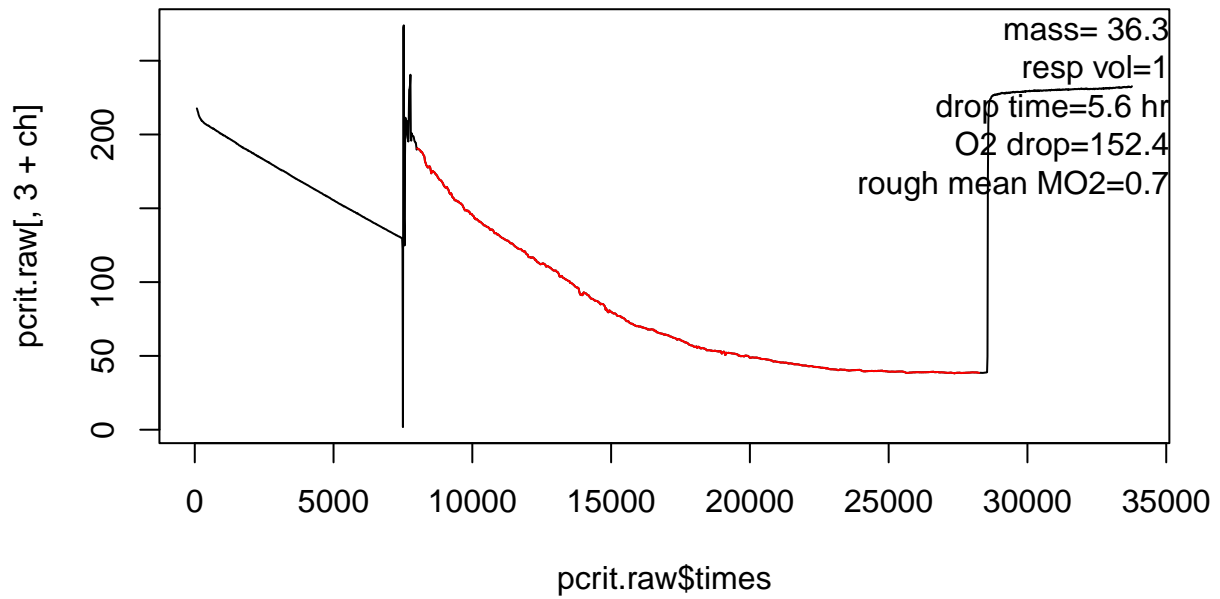
LLO @ MR of 2.3 = 11.542

NLR (Power) = 37.94

Sub-PI = 16.3



tbocto 1000 pcrit tank 1 and 2 day 7 8-19-21.txt



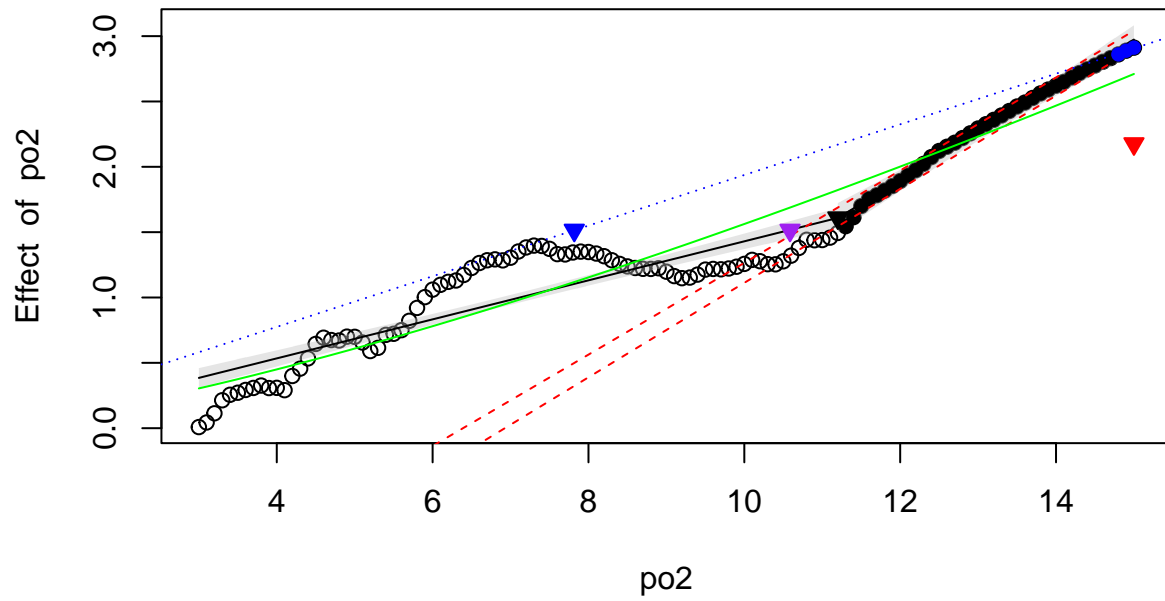
Alpha @ MR of 1.51 = 7.818

Breakpoint = 11.2

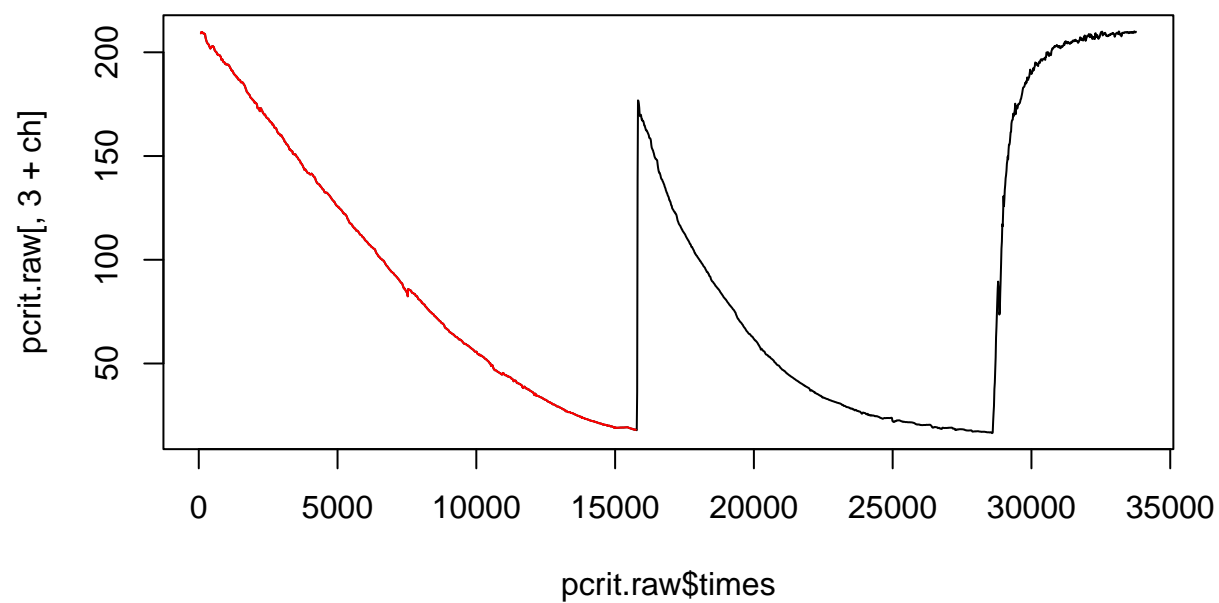
LLO @ MR of 1.51 = 10.587

NLR (Power) = 28.052

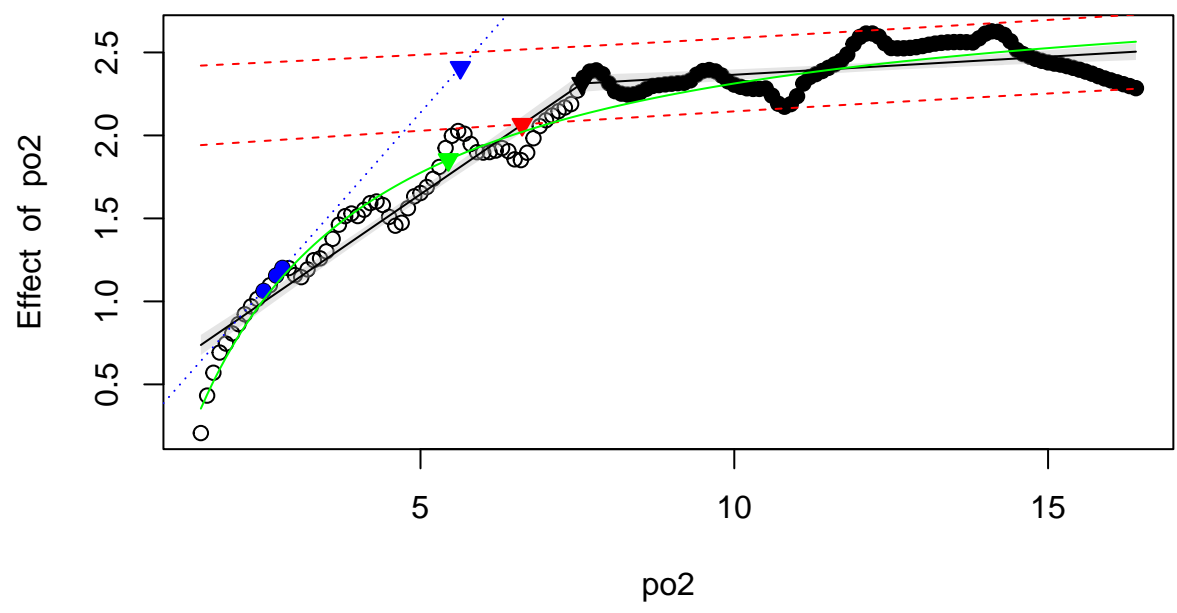
Sub-PI = 15



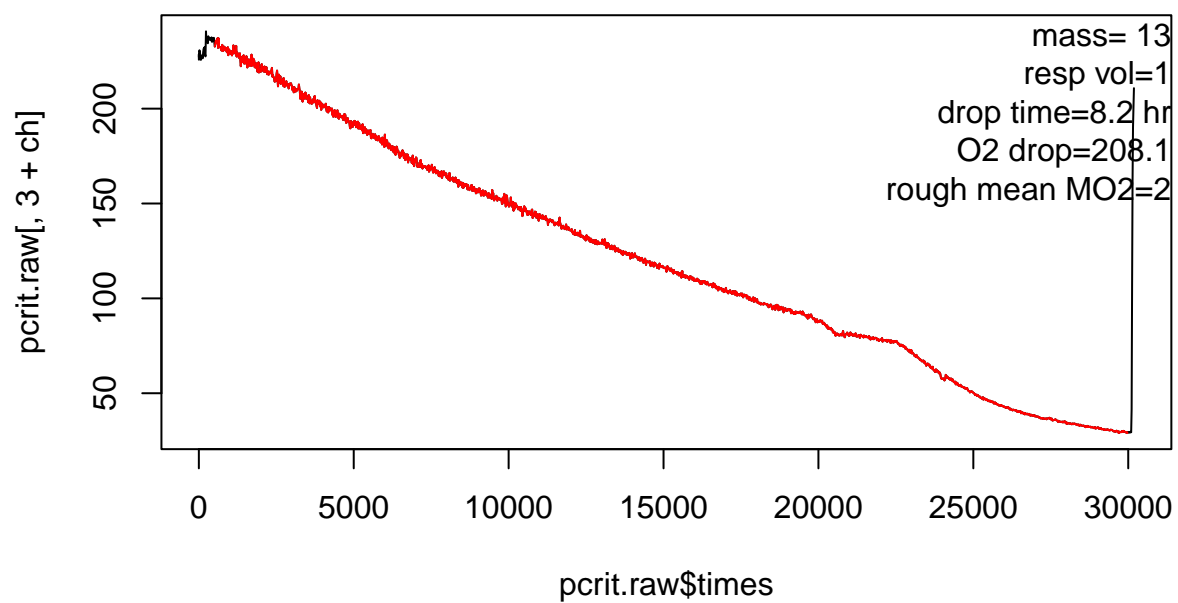
tbocto 1000 pcrit tank 1 and 2 day 7 8–19–21.txt



Alpha @ MR of 2.41 = 5.635
Breakpoint = 7.572
LLO @ MR of NA = NA
NLR (Hyperbola) = 5.444
Sub-PI = 6.62



tbocto 1000 pcrit tank 3 and 4 8-11-21-ch1.txt



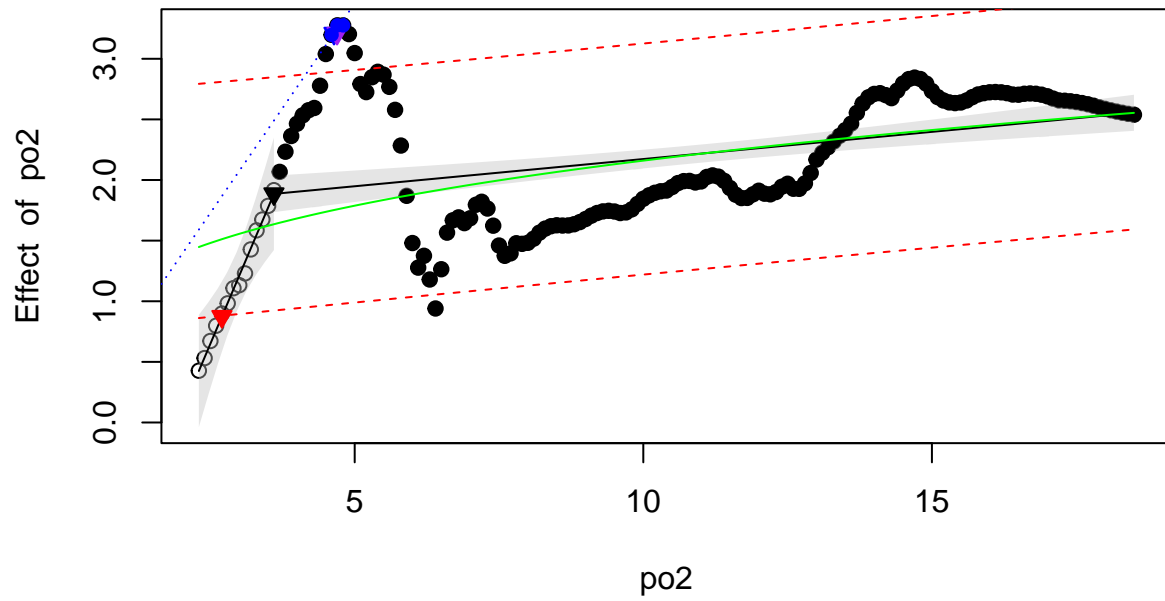
Alpha @ MR of 3.21 = 4.642

Breakpoint = 3.601

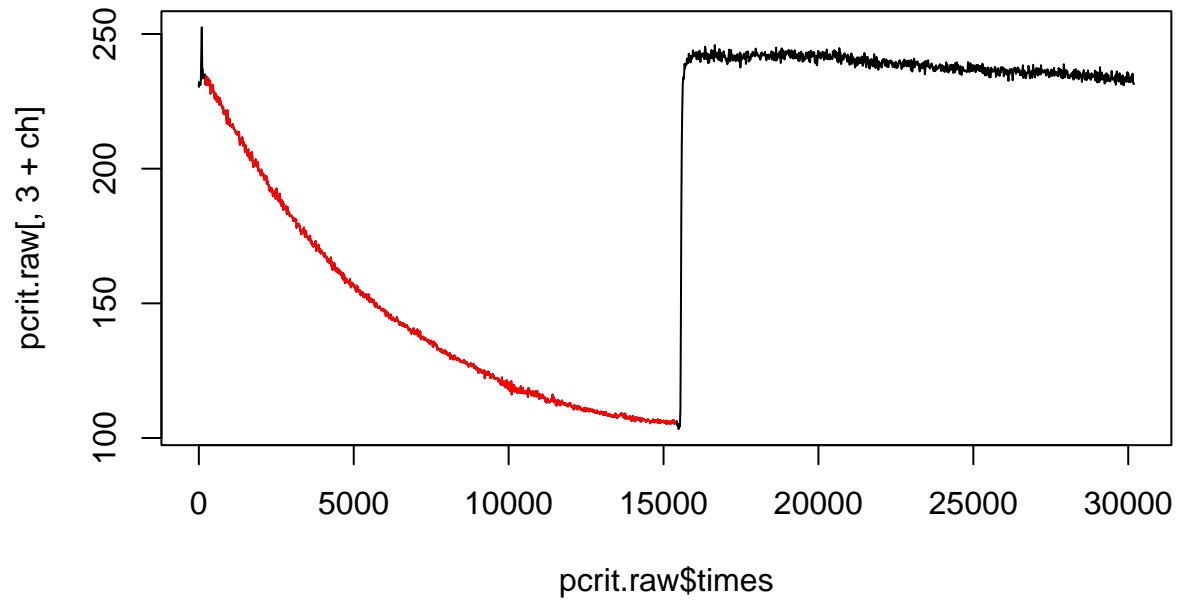
LLO @ MR of 3.21 = 4.695

NLR (Power) = 77.005

Sub-PI = 2.7



tbocto 1000 pcrit tank 3 and 4 8-11-21-ch1.txt



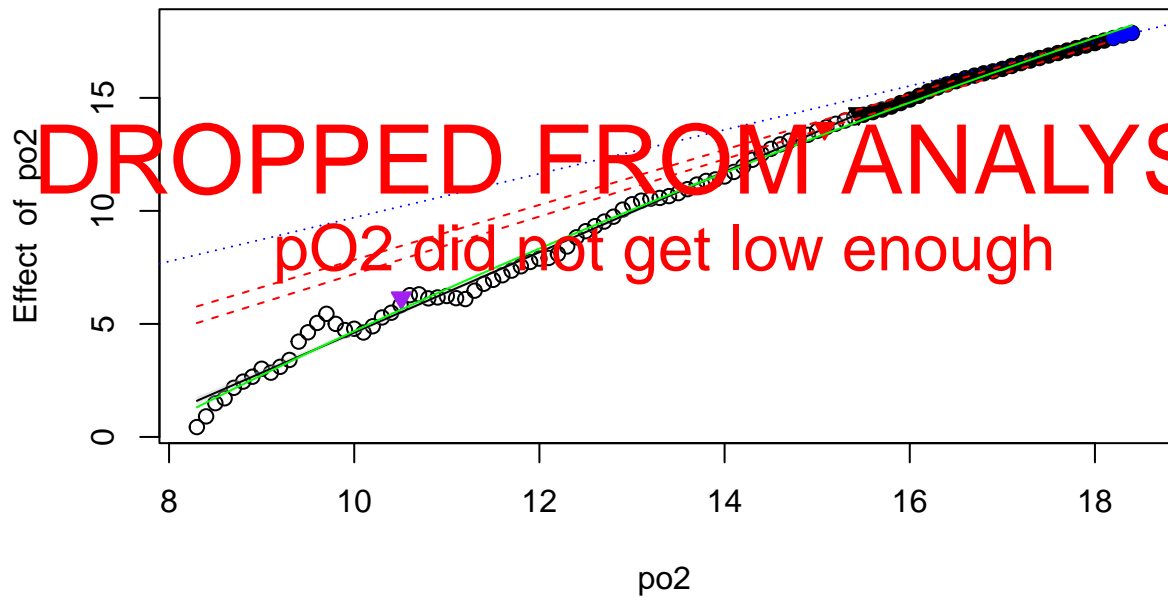
Alpha @ MR of 6.15 = 6.337

Breakpoint = 15.447

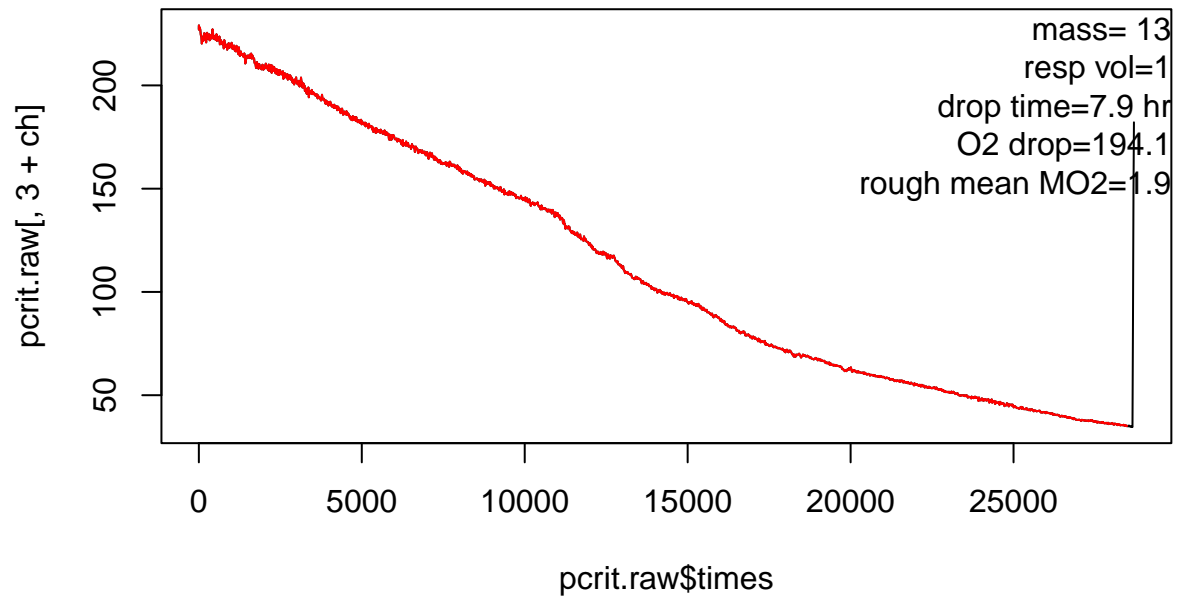
LLO @ MR of 6.15 = 10.509

NLR (Hyperbola) = 24.757

Sub-PI = 15.08



tbocto 1000 pcrit tank 3 and 4 day 7 8-19-21-ch1.txt



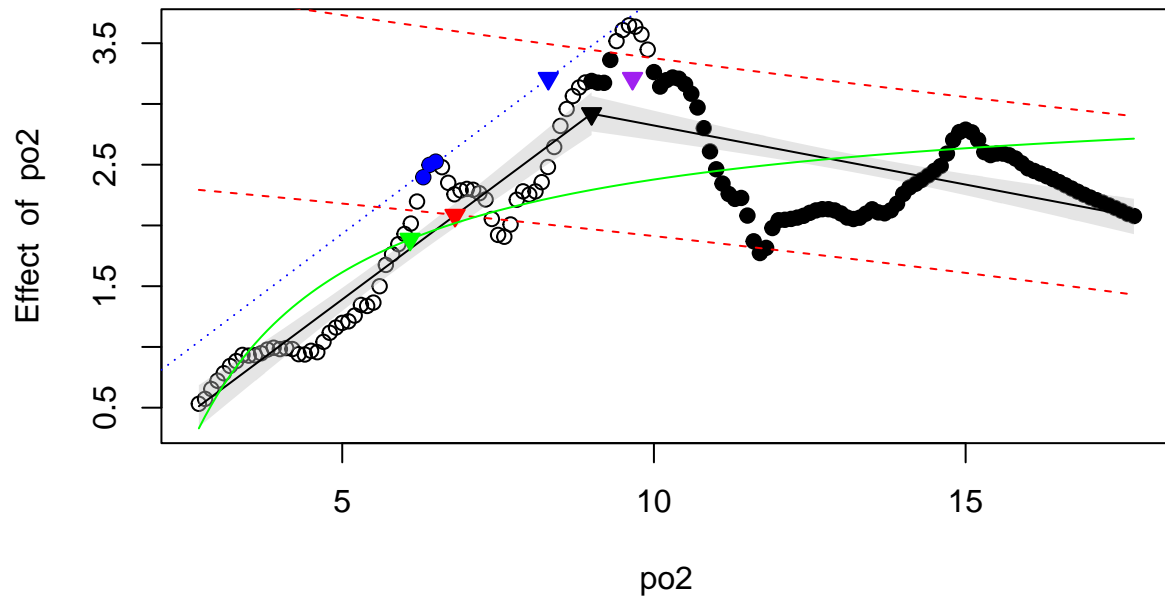
Alpha @ MR of 3.21 = 8.306

Breakpoint = 9

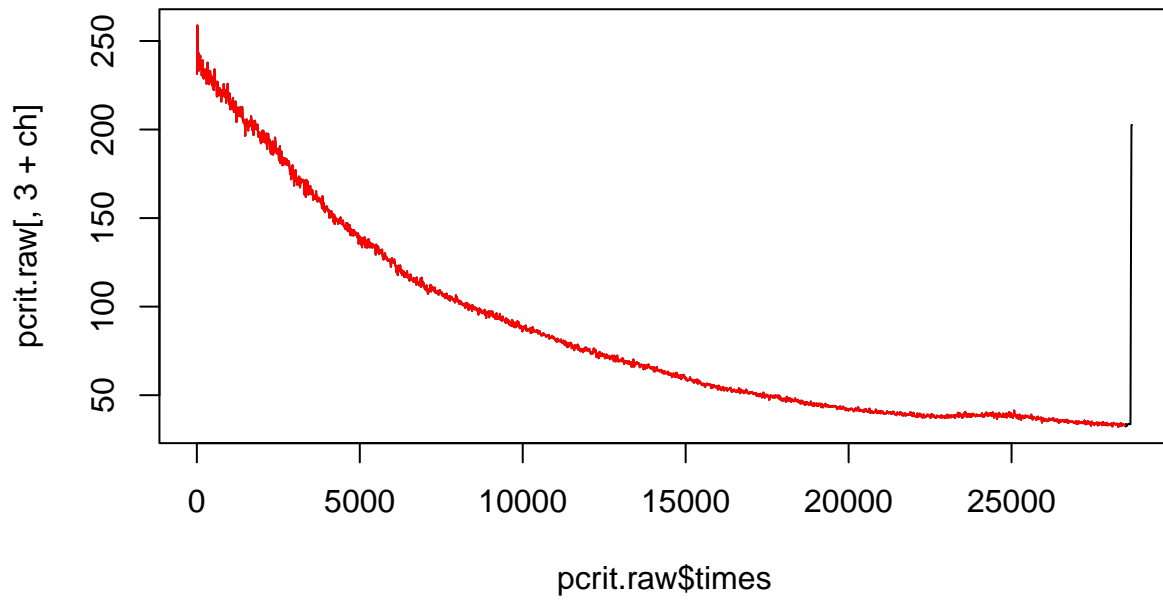
LLO @ MR of 3.21 = 9.657

NLR (Pareto) = 6.088

Sub-PI = 6.81



tbocto 1000 pcrit tank 3 and 4 day 7 8-19-21-ch1.txt



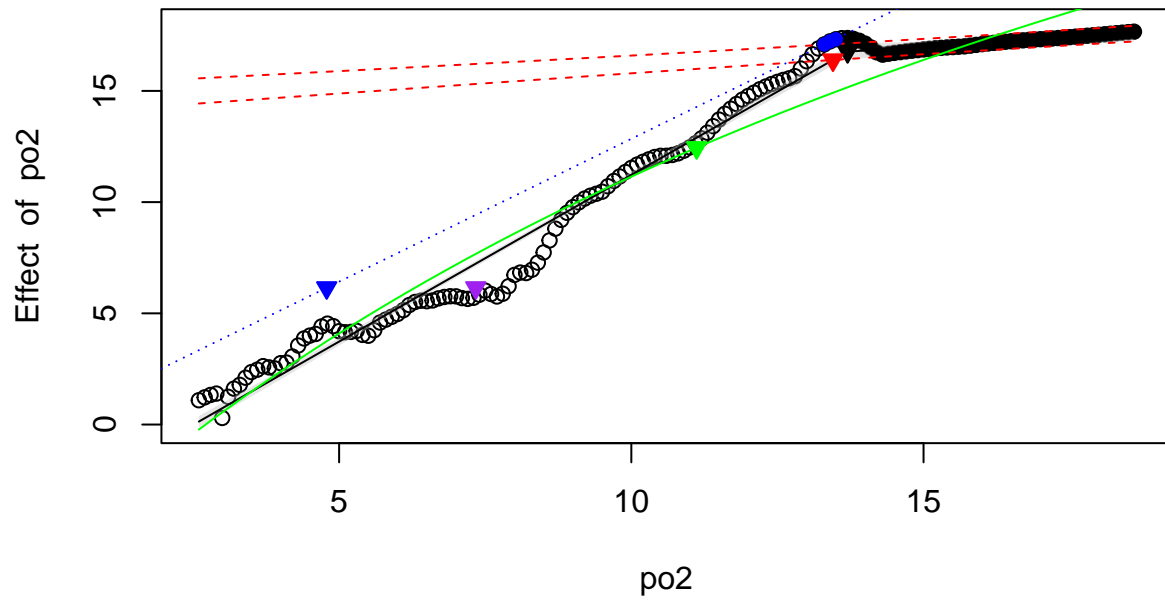
Alpha @ MR of 6.15 = 4.788

Breakpoint = 13.7

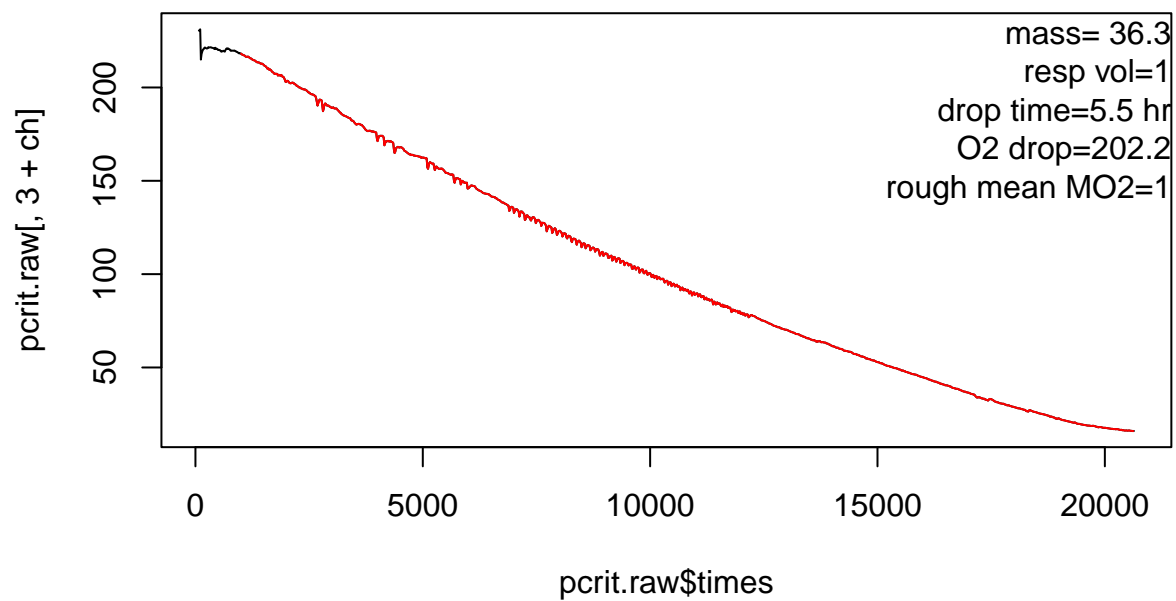
LLO @ MR of 6.15 = 7.338

NLR (Hyperbola) = 11.118

Sub-PI = 13.45



Tbocto 1000 pcrti tank 1 and 2 8-11-21.txt



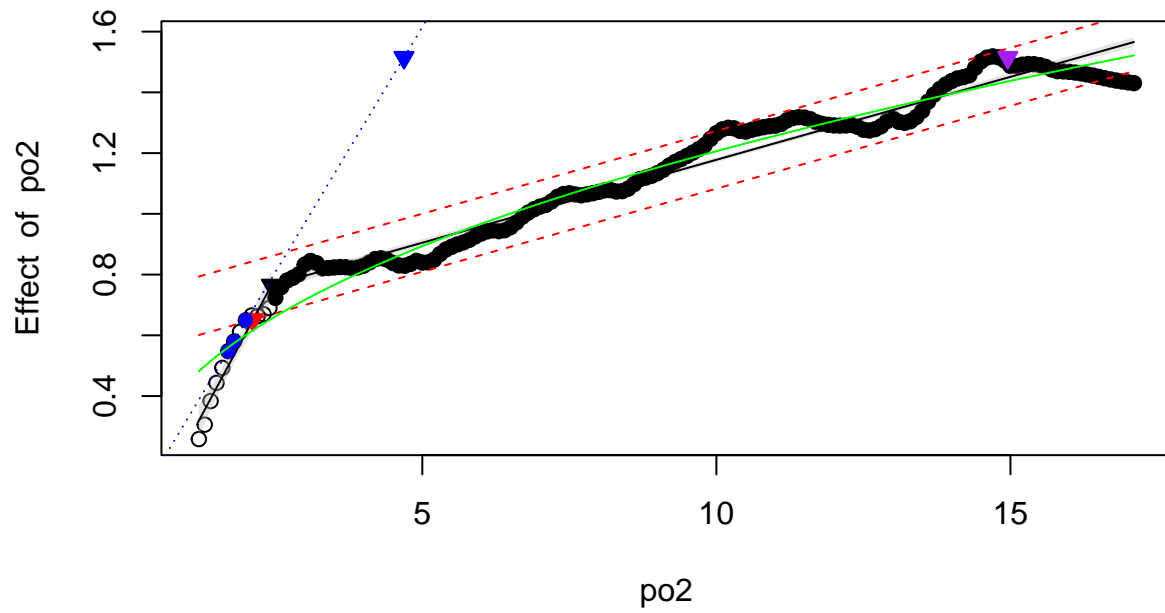
Alpha @ MR of 1.51 = 4.689

Breakpoint = 2.435

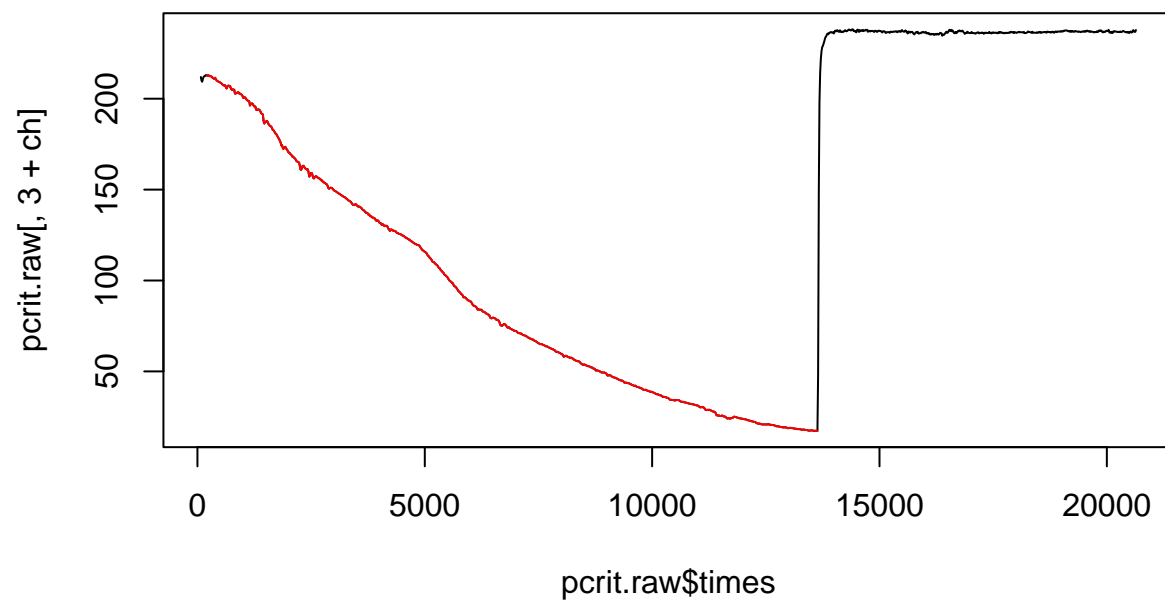
LLO @ MR of 1.51 = 14.954

NLR (Power) = 66.218

Sub-PI = 2.12



Tbocto 1000 pcrti tank 1 and 2 8-11-21.txt



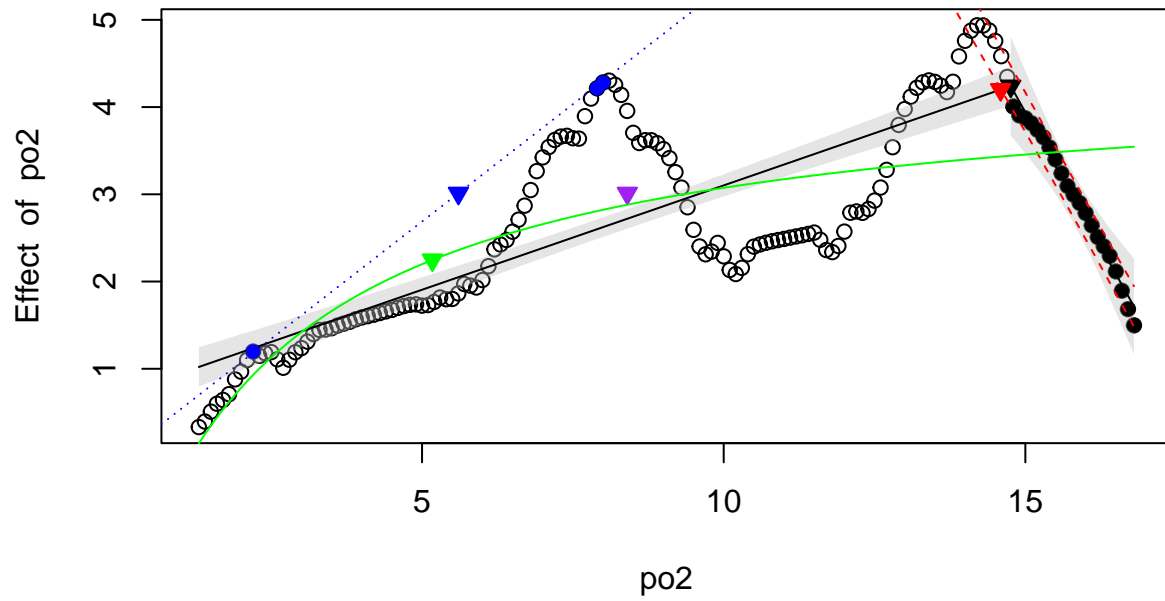
Alpha @ MR of 3.01 = 5.603

Breakpoint = 14.758

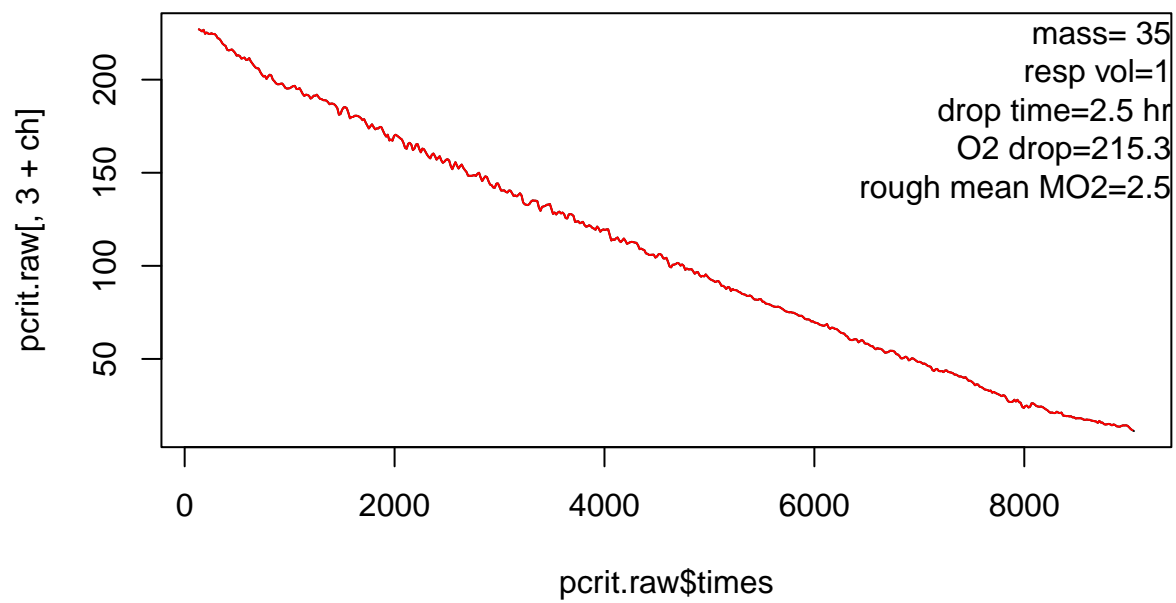
LLO @ MR of 3.01 = 8.403

NLR (Hyperbola) = 5.171

Sub-PI = 14.59



gr2muus1000 pcrit 7-21-21.txt



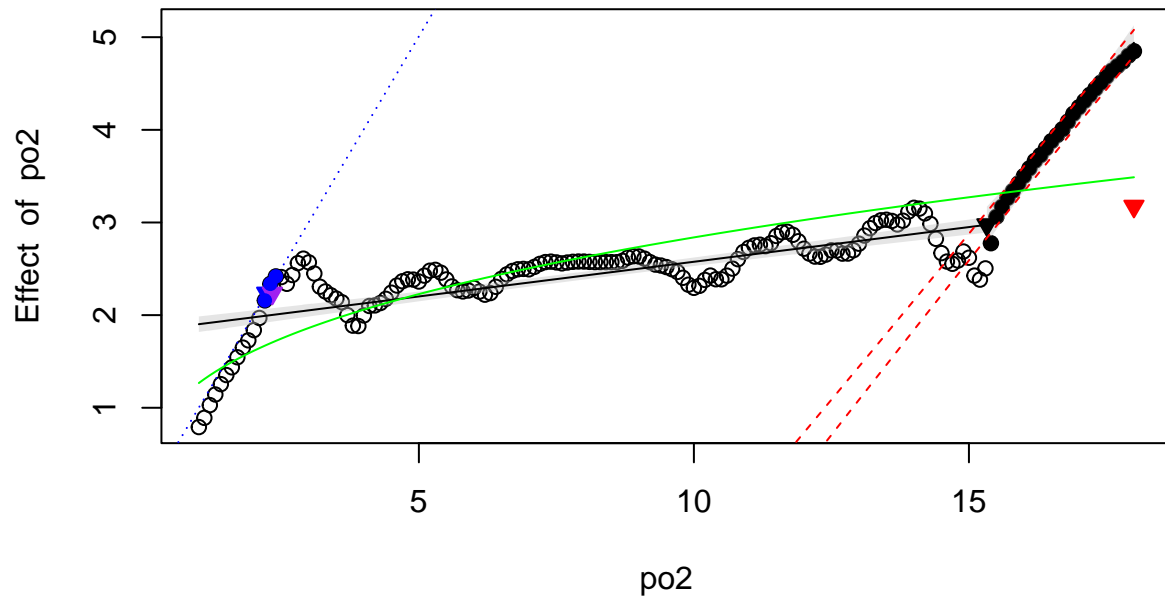
Alpha @ MR of 2.24 = 2.233

Breakpoint = 15.327

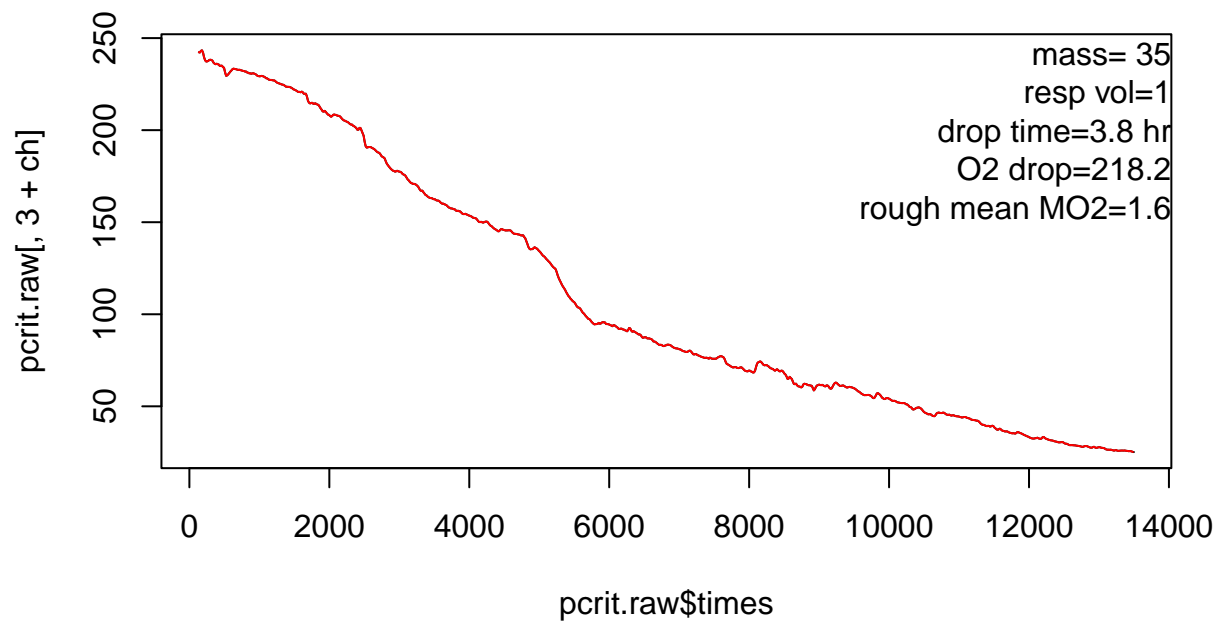
LLO @ MR of 2.24 = 2.337

NLR (Power) = 64.181

Sub-PI = 18



gr2muus1000-2 pcrit 7-26-21.txt



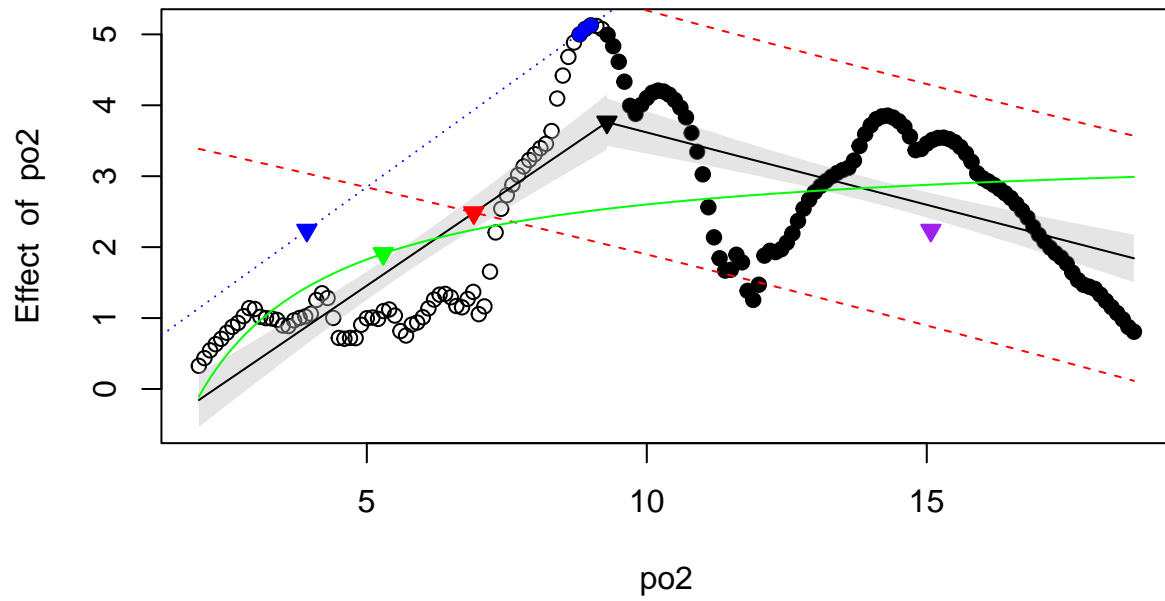
Alpha @ MR of 2.24 = 3.929

Breakpoint = 9.288

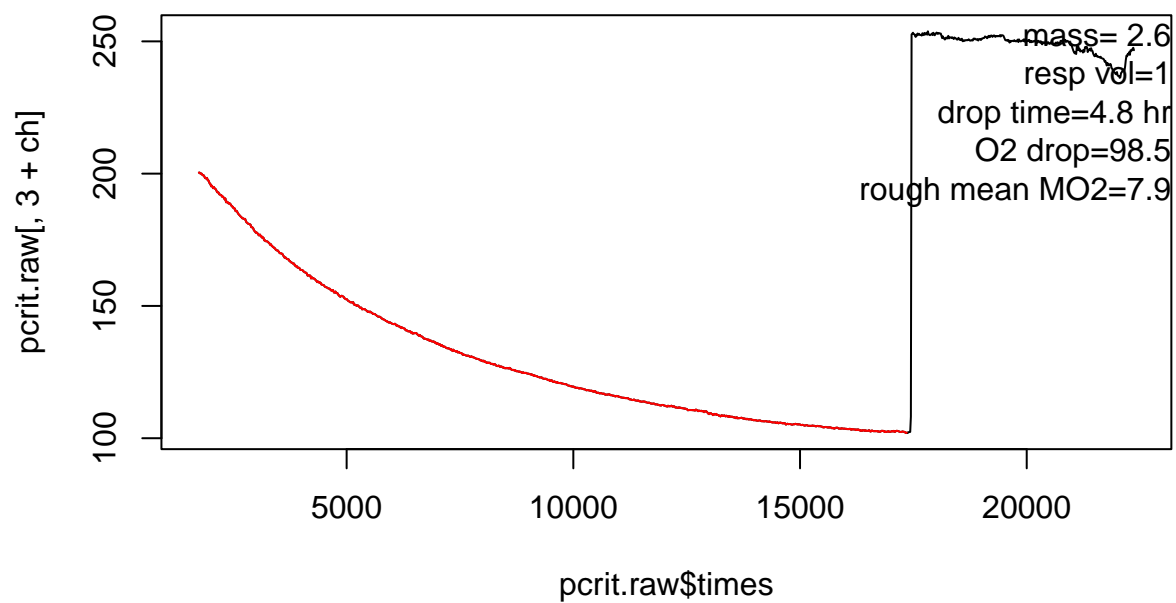
LLO @ MR of 2.24 = 15.073

NLR (Hyperbola) = 5.293

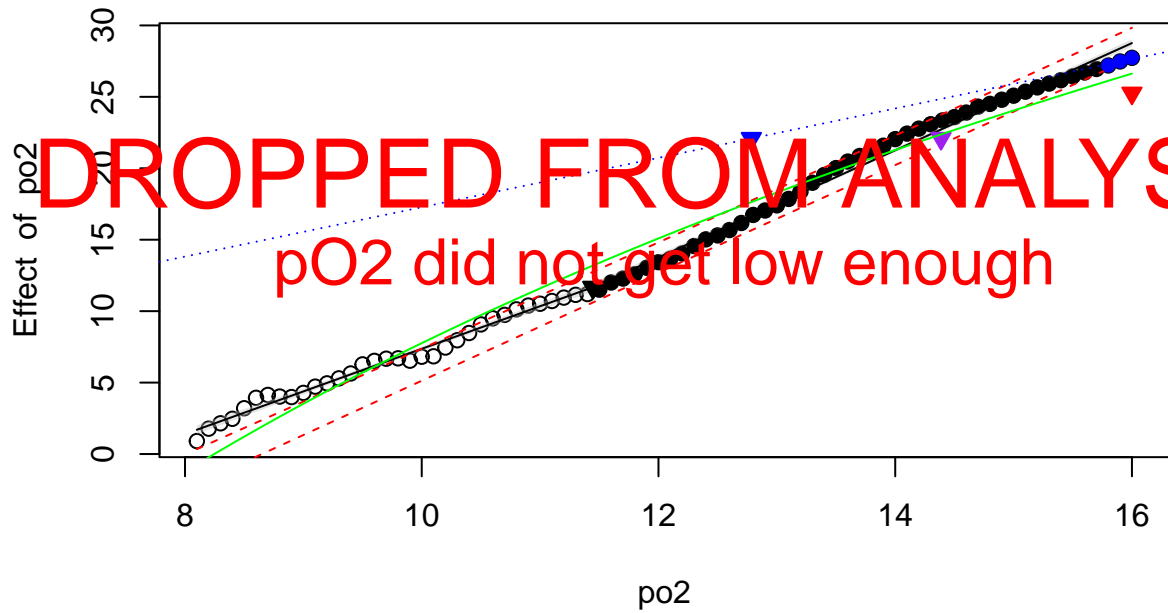
Sub-PI = 6.91



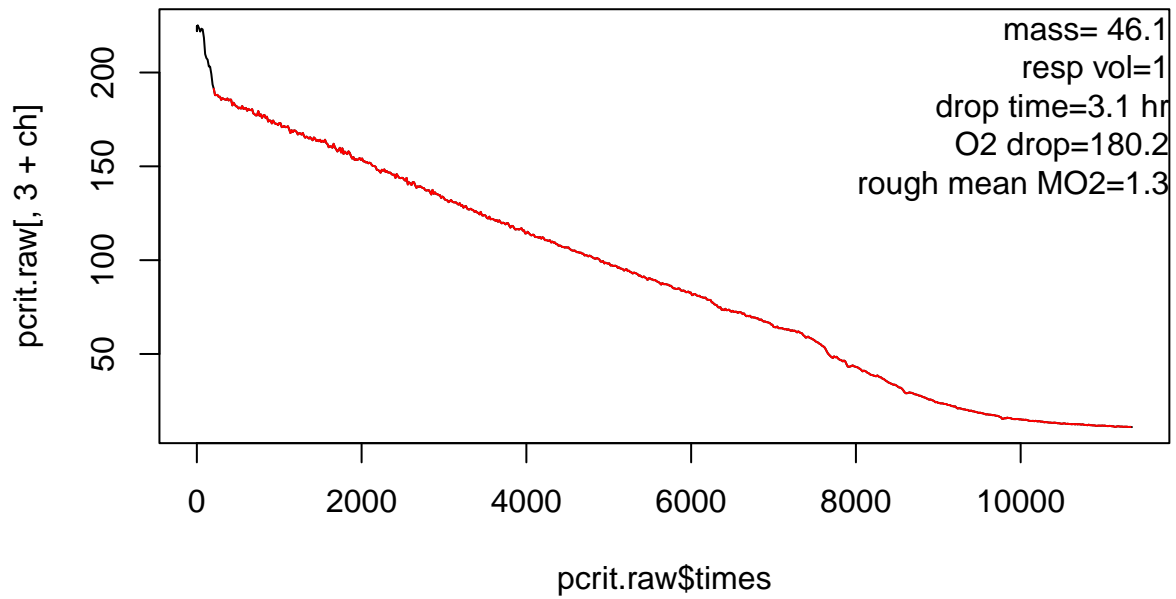
gr2MUUS1800-2pcritday7.8-3-21.txt



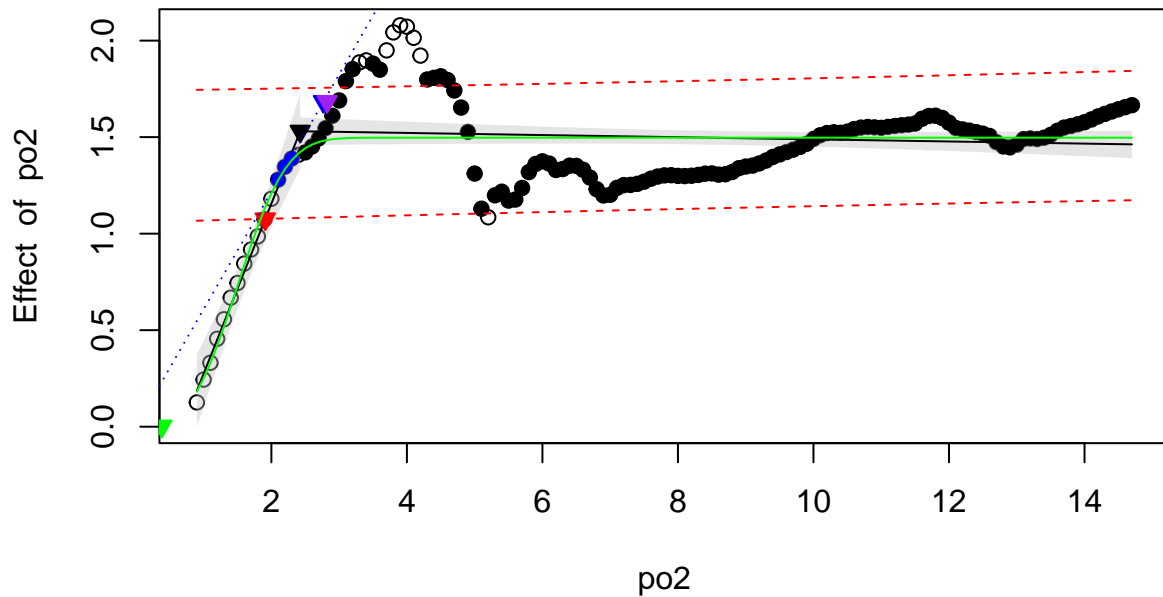
Alpha @ MR of 22.09 = 12.785
Breakpoint = 11.444
LLO @ MR of 22.09 = 14.387
NLR (Weibull with intercept) = 21
Sub-PI = 16



tbocto 1800 pcrit day 7 tank 10 blank ch 3 4 8-20-21-ch1.txt



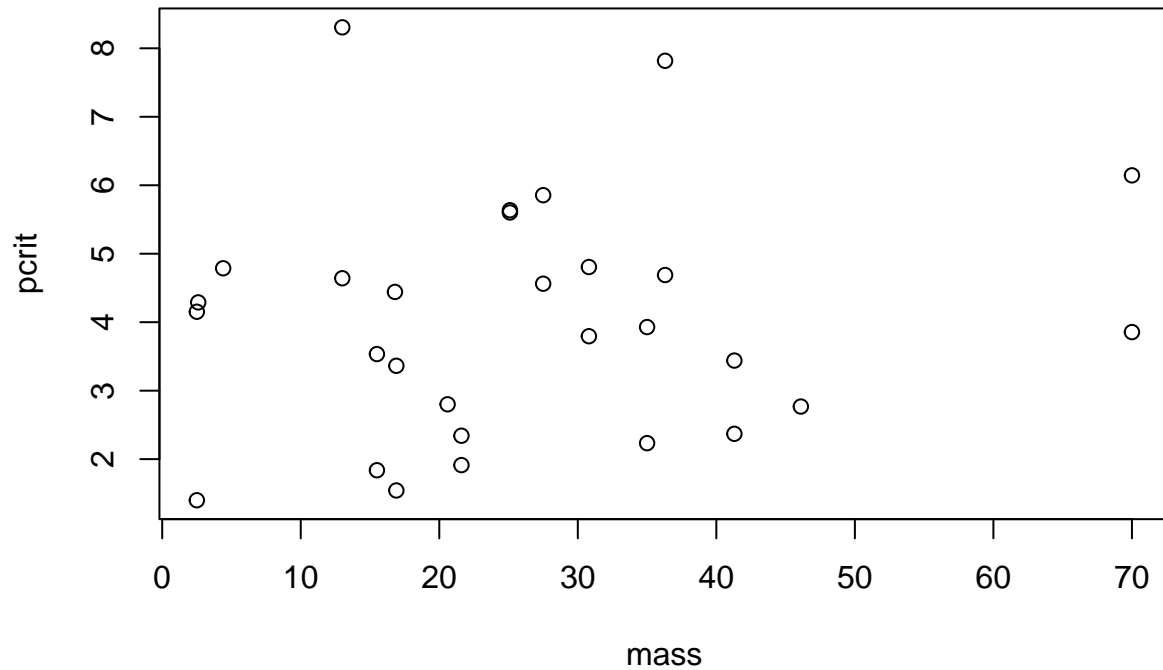
Alpha @ MR of 1.68 = 2.768
 Breakpoint = 2.426
 LLO @ MR of 1.68 = 2.824
 NLR (Weibull with intercept) = 0.384
 Sub-PI = 1.91



Next, I remove the measurements for which the data was excluded (runs did not reach oxygen level of 50 mmHgO₂). Then I remove the file for which there is no initial data.

```
pcrits=pcrits[complete.cases(pcrits),]
pcrits=pcrits[pcrits$filename!="GR1 Muus1000 pcrit 7-21-21.txt",]
```

```
plot(pcrit~mass,data=pcrits)
```

Complete table of final pcrit and alpha values

```
pcrits.kable=pcrits[,c(3:6,8:9)]
#pcrits.kable$rmr=round(pcrits.kable$rmr,2)
pcrits.kable$pcrit=round(pcrits.kable$pcrit,2)
pcrits.kable$alpha=round(pcrits.kable$alpha,2)
colnames(pcrits.kable)=c("Octopus ID",
                          "mass (g)",
                          "pCO2 (μatm)",
                          "day",
                          "Routine Metabolic Rate (μmol O2 g-1 hr-1)",
                          "P-CRIT~ (kPa)",
                          "Oxygen Supply Capacity (α)")
pcrits.kable=pcrits.kable[order(pcrits.kable$day),]
pcrits.kable=pcrits.kable[order(pcrits.kable$`Octopus ID`),]
kable(pcrits.kable,align=c("l",rep("c",4),"r"),row.names = F)
```

Octopus ID	mass (g)	pCO ₂ (μatm)	day	P _{CRIT} (kPa)	Oxygen Supply Capacity (α)
1-1	30.8	1800	1	3.80	0.66
1-1	30.8	1800	7	4.81	0.52
1-2	20.6	1000	1	2.80	0.79
1-3	2.5	1800	1	1.40	1.64
1-3	2.5	1800	7	4.15	0.55
2-1	16.8	1800	7	4.44	0.33

Octopus ID	mass (g)	pCO ₂ (μ atm)	day	P _{CRIT} (kPa)	Oxygen Supply Capacity (α)
2-2	35.0	1000	1	2.23	1.00
2-2	35.0	1000	7	3.93	0.57
2-3	2.6	1800	1	4.29	0.85
3-1	70.0	1800	1	6.14	0.24
3-1	70.0	1800	7	3.86	0.48
3-2	21.6	1000	1	1.91	1.14
3-2	21.6	1000	7	2.34	0.93
3-3	16.9	1800	1	1.54	1.40
3-3	16.9	1800	7	3.36	0.64
4-1	27.5	1800	1	4.56	0.50
4-1	27.5	1800	7	5.85	0.39
4-2	15.5	1000	1	1.84	1.37
4-2	15.5	1000	7	3.53	0.71
4-3	41.3	1800	1	2.37	0.89
4-3	41.3	1800	7	3.44	0.62
5-1	36.3	1000	1	4.69	0.32
5-1	36.3	1000	7	7.82	0.19
5-2	25.1	1000	1	5.60	0.54
5-2	25.1	1000	7	5.63	0.43
5-3	13.0	1000	1	4.64	0.69
5-3	13.0	1000	7	8.31	0.39
5-4	4.4	1000	7	4.79	1.28
5-5	46.1	1800	1	2.77	0.61

5 P_{CRIT} Linear mixed effect model

5.1 setting pCO₂ to factor class:

```
pcrits$pco2=as.factor(pcrits$pco2)
```

5.2 Next I set orthogonal contrasts:

```
contrasts(pcrits$pco2)=contr.poly(2)
```

5.3 Running the linear mixed effects model and ANOVA using type III sum of squares:

```
pcrits.lme=lme(pcrit~mass+pco2+day,random=~1|octo,
               correlation=corAR1(form=~day|octo),
               data=pcrits,na.action=na.omit)

Anova(pcrits.lme,type="III")
```

```
## Analysis of Deviance Table (Type III tests)
##
## Response: pcrit
##           Chisq Df Pr(>Chisq)
## (Intercept) 14.8709  1  0.0001151 ***
## mass         0.4563  1  0.4993495
## pco2         0.4646  1  0.4954673
## day         10.5386  1  0.0011691 **
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

5.4 Summary of LME

```
pcrit.em=data.frame(emmeans(pcrits.lme,~pco2+day+mass))
```

```
## Warning: contrasts dropped from factor pco2
```

```
pcrit.em
```

```
##   pco2 day    mass  emmean      SE df lower.CL upper.CL
## 1 1000   1 26.25862 3.635840 0.5676006 11 2.386560 4.885121
## 2 1800   1 26.25862 3.135719 0.5341027 11 1.960167 4.311271
## 3 1000   7 26.25862 4.998403 0.5699236 11 3.744010 6.252797
## 4 1800   7 26.25862 4.498282 0.5479037 11 3.292354 5.704210
```

```
pcrit.df=
data.frame(cbind(
  as.numeric(as.character(pcrit.em$pco2)),
  pcrit.em$day,
  sprintf("%.2f",signif(pcrit.em$emmean,3)),
  paste(sprintf("%.2f",signif(data.frame(pcrit.em)$lower.CL,3)),
    "_",
    sprintf("%.2f",signif(data.frame(pcrit.em)$upper.CL,3)))
))
pcrit.df=pcrit.df[order(pcrit.df[,1]),]
pcrit.df
```

```
##      X1 X2  X3      X4
## 1 1000  1 3.64 2.39 - 4.89
## 3 1000  7 5.00 3.74 - 6.25
## 2 1800  1 3.14 1.96 - 4.31
## 4 1800  7 4.50 3.29 - 5.70
```

```
colnames(pcrit.df)=c("pCO~2~ ($\\mu$atm)",
  "day",
  "P~CRIT~ (kPa)",
  "P~CRIT~ 95% CI")
kable(pcrit.df,align="c",row.names = F)
```

pCO ₂ (μ atm)	day	P _{CRIT} (kPa)	P _{CRIT} 95% CI
1000	1	3.64	2.39 - 4.89
1000	7	5.00	3.74 - 6.25
1800	1	3.14	1.96 - 4.31
1800	7	4.50	3.29 - 5.70

6 Plotting the Critical Oxygen Pressure results

Assigning the colors for the treatments.

```
hi.co2.col="#790000ff"
lo.co2.col="#838fd5ff"
```

Next, making the figures in SVG format.

```
svg(filename = "Figure_4.svg",width=3.5,height=3.5,pointsize=6)
par(fig=c(0.04,1,0,1))
boxplot(pcrit~pco2+day,data=pcrits,range=0,
        axes=F,col=c(lo.co2.col,hi.co2.col,lo.co2.col,hi.co2.col),ylab="",xlab="")
box(lwd=2)
abline(v=2.5)
axis(1, at=c(1.5,3.5),labels = c("Day 1","Day 7"),tick=F,cex.axis=1.5)
axis(2,at=c(2,4,6,8),cex.axis=1.5)
mtext(expression("Critical Oxygen Pressure (kPa)"),side=2,cex=1.8,line=3)
legend(1,8,pt.bg=c(lo.co2.col,hi.co2.col),legend=c("1000","1800"),
      pch=22,cex=1.3,pt.cex=3.2,bty="n",adj=c(0,0.5),title=expression("pCO"[2]*" ("*mu*"atm)"))
dev.off()
```

```
## pdf
## 2
```

Converting the image to a png and jpg to be displayed in the RMarkdown.

```
cairosvg Figure_4.svg -o Figure_4.png -d 300
convert Figure_4.png Figure_4.jpg
```

Converting to eps for submission.

```
inkscape Figure_4.svg -o Figure_4.eps --export-ignore-filters --export-ps-level=3
```

7 Oxygen supply capacity (α) linear mixed effect model

7.1 Running the linear mixed effects model and ANOVA using type III sum of squares:

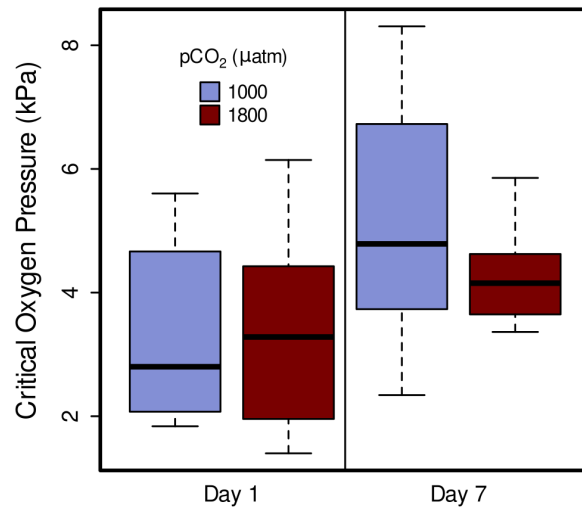


Figure 1: Critical oxygen pressure of *Muusoctopus leioderma* at differing pCO₂ treatments

```
alpha.lme=lme(alpha~mass*pco2*day,random=~1|octo,
              correlation=corAR1(form=~day|octo),
              data=pcrits,na.action=na.omit)
```

```
alpha.anova=Anova(alpha.lme,type="III")
```

```
options(scipen=10)
alpha.lme.table=cbind(
  c("Mass","pCO~2~","Day"),
  round(alpha.anova$Chisq[2:4],2),
  alpha.anova$Df[2:4],
  round(alpha.anova$`Pr(>Chisq)`[2:4],6)
)
```

```
colnames(alpha.lme.table)=c("Factor","Chi-square", "DF", "p-value")
```

```
kable(alpha.lme.table)
```

Factor	Chi-square	DF	p-value
Mass	9.84	1	0.00171
pCO ₂	0.06	1	0.808008
Day	17.35	1	0.000031

7.2 Summary of LME

```
alpha.em=data.frame(emmeans(alpha.lme,~pco2+day+mass))

## Warning: contrasts dropped from factor pco2

alpha.em

##   pco2 day    mass    emmean      SE df lower.CL upper.CL
## 1 1000   1 26.25862 0.8040022 0.1151260  8 0.5385212 1.0694832
## 2 1800   1 26.25862 0.8925850 0.1019162  8 0.6575658 1.1276041
## 3 1000   7 26.25862 0.5266816 0.1181234  8 0.2542887 0.7990746
## 4 1800   7 26.25862 0.4591574 0.1058851  8 0.2149860 0.7033289

alpha.df=
data.frame(cbind(
  as.numeric(as.character(alpha.em$pco2)),
  alpha.em$day,
  sprintf("%.2f",signif(alpha.em$emmean,3)),
  paste(sprintf("%.2f",signif(data.frame(alpha.em)$lower.CL,3)),
        "_",
        sprintf("%.2f",signif(data.frame(alpha.em)$upper.CL,3)))
))
alpha.df=alpha.df[order(alpha.df[,1]),]
alpha.df

##      X1 X2  X3      X4
## 1 1000  1 0.80 0.54 - 1.07
## 3 1000  7 0.53 0.25 - 0.80
## 2 1800  1 0.89 0.66 - 1.13
## 4 1800  7 0.46 0.21 - 0.70

colnames(alpha.df)=c("pCO~2~ ($\\mu$atm)",
  "day",
  "Oxygen Supply Capacity ($\\alpha$)",
  "Oxygen Supply Capacity 95% CI")
kable(alpha.df,align="c",row.names = F)
```

pCO ₂ (μ atm)	day	Oxygen Supply Capacity (α)	Oxygen Supply Capacity 95% CI
1000	1	0.80	0.54 - 1.07
1000	7	0.53	0.25 - 0.80
1800	1	0.89	0.66 - 1.13
1800	7	0.46	0.21 - 0.70

8 Oxygen Supply Capacity figure

8.1 Predicted values

First I am getting the model predicted values for each treatment between the max an minimum mass values.

```

seq1.1800=seq(from=min(pcrits$mass[pcrits$pco2==1800]),
              to=max(pcrits$mass[pcrits$pco2==1800]),
              length.out=100)

df1.1800=data.frame(
  day=rep(1,100),
  mass=seq1.1800,
  pco2=as.factor(rep(1800,100))
)
pred1.1800= predict(alpha.lme,newdata = df1.1800,level=0)

seq1.1000=seq(from=min(pcrits$mass[pcrits$pco2==1000]),
              to=max(pcrits$mass[pcrits$pco2==1000]),
              length.out=100)

df1.1000=data.frame(
  day=rep(1,100),
  mass=seq1.1000,
  pco2=as.factor(rep(1000,100))
)

pred1.1000=predict(alpha.lme,newdata = df1.1000,level=0)

seq7.1800=seq(from=min(pcrits$mass[pcrits$pco2==1800]),
              to=max(pcrits$mass[pcrits$pco2==1800]),
              length.out=100)

df7.1800=data.frame(
  day=rep(7,100),
  mass=seq7.1800,
  pco2=as.factor(rep(1800,100))
)

pred7.1800=predict(alpha.lme,newdata = df7.1800,level=0)

seq7.1000=seq(from=min(pcrits$mass[pcrits$pco2==1000]),
              to=max(pcrits$mass[pcrits$pco2==1000]),
              length.out=100)

df7.1000=data.frame(
  day=rep(7,100),
  mass=seq7.1000,
  pco2=as.factor(rep(1000,100))
)

pred7.1000=predict(alpha.lme,newdata = df7.1000,level=0)

```

Assigning the colors for the treatments.

```

hi.co2.col="#790000ff"
lo.co2.col="#838fd5ff"

```

Next, I am actually plotting it.

```

svg(filename="Figure_5.svg",height=3.5,width=3.5,pointsize=6)
par(fig=c(0.04,1,0,1))
plot(alpha~mass,data=pcrits,axes=F,ylab="",xlab="",type="n")
box(lwd=2)
axis(1,lwd=2,cex.axis=2)
axis(2,lwd=2,cex.axis=1.5)
mtext(expression("Oxygen Supply Capacity ("*mu*"molO"[2]*" g"^-1*"hr"^-1*"kPa"^-1*")"),
      side=2,cex=1.8,line=2.5)
mtext("Mass (g)",side=1,cex=1.8,line=2.5)

points(alpha~mass,data=pcrits[pcrits$pcO2==1000&pcrits$day==1,],
      pch=22,bg="white",col=lo.co2.col,cex=1.5)
points(alpha~mass,data=pcrits[pcrits$pcO2==1000&pcrits$day==7,],
      pch=22,bg=lo.co2.col,cex=1.5)
points(alpha~mass,data=pcrits[pcrits$pcO2==1800&pcrits$day==7,],
      pch=21,bg=hi.co2.col,cex=1.5)
points(alpha~mass,data=pcrits[pcrits$pcO2==1800&pcrits$day==1,],
      pch=21,bg="white",col=hi.co2.col,cex=1.5)
lines(seq1.1800,pred1.1800,col=hi.co2.col,lwd=2,lty=2)
lines(seq1.1000,pred1.1000,col=lo.co2.col,lwd=2,lty=2)
lines(seq7.1800,pred7.1800,col=hi.co2.col,lwd=2,lty=1)
lines(seq7.1000,pred7.1000,col=lo.co2.col,lwd=2,lty=1)
legend("topright", c(expression("1000 "*mu*"atm pCO"["2"]*", day 1"),
                      expression("1000 "*mu*"atm pCO"["2"]*", day 7"),
                      expression("1800 "*mu*"atm pCO"["2"]*", day 1"),
                      expression("1800 "*mu*"atm pCO"["2"]*", day 7")),
      pch = c(22,22,21,21),bty="n",title = expression("Treatment pCO"["2"]"),
      pt.bg=c("white",lo.co2.col,"white",hi.co2.col),col=c(lo.co2.col,"black",hi.co2.col,"black"),
      inset = .02,cex=1.3,box.lwd=2,pt.lwd=1,pt.cex=2)

dev.off()

## pdf
## 2

```

Converting the image to a png and jpg to be displayed in the RMarkdown.

```

cairosvg Figure_5.svg -o Figure_5.png -d 300
convert Figure_5.png Figure_5.jpg

```

Converting to eps for submission.

```

inkscape Figure_5.svg -o Figure_5.eps --export-ignore-filters --export-ps-level=3

```

9 Finding the slopes of the treatments

Here I am producing a table of the slope and intercepts of the linear mixed effects model of routine metabolic rates. These values correspond to the logged mass and logged RMR, and do not translate to the untransformed data. The relationship between the untransformed data is not linear, and therefore has no slope. Also, because the log of 0 is infinite, these intercepts correspond instead to a mass of 1g ($\exp(0)=1$).

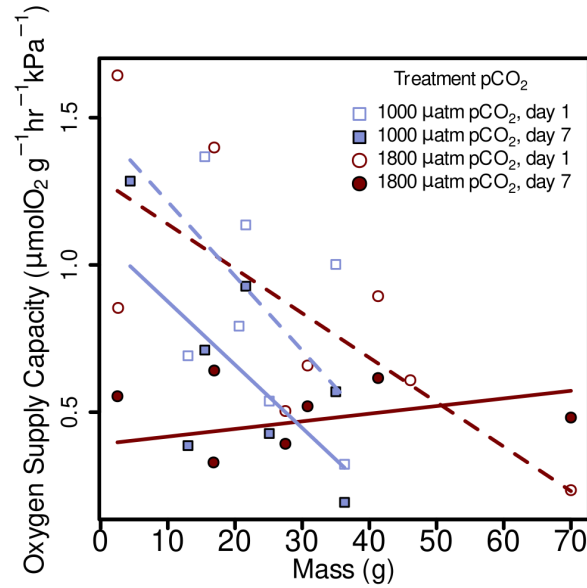


Figure 2: Oxygen supply capacity from *M.leioderma* in Burrows Bay, Anacortes Washington

```

mass=0
alpha.slope=data.frame(pCO2=c(1000,1000,1800,1800),
  Day=c(1,7,1,7),
  Intercept=round(predict(alpha.lme,
    newdata=data.frame(
      day=c(1,7,1,7),
      mass=rep(mass,4),
      pco2=as.factor(c(1000,1000,1800,1800))),
    level=0)
    [1:4],2),
  Slope=c(
    round(-1*(pred1.1000[1]-pred1.1000[length(pred1.1000)])/diff(range(seq1.1000)),3),
    round(-1*(pred7.1000[1]-pred7.1000[length(pred7.1000)])/diff(range(seq7.1000)),3),
    round(-1*(pred1.1800[1]-pred1.1800[length(pred1.1800)])/diff(range(seq1.1800)),3),
    round(-1*(pred7.1800[1]-pred7.1800[length(pred7.1800)])/diff(range(seq7.1800)),3)
  )
)

colnames(alpha.slope)[1]="pCO~2~ ($\\mu$atm)"
kable(alpha.slope,align="c",row.names=F)

```

pCO ₂ (μatm)	Day	Intercept	Slope
1000	1	1.47	-0.025
1000	7	1.09	-0.021
1800	1	1.29	-0.015
1800	7	0.39	0.003

10 Is there a relationship between RMR and alpha?

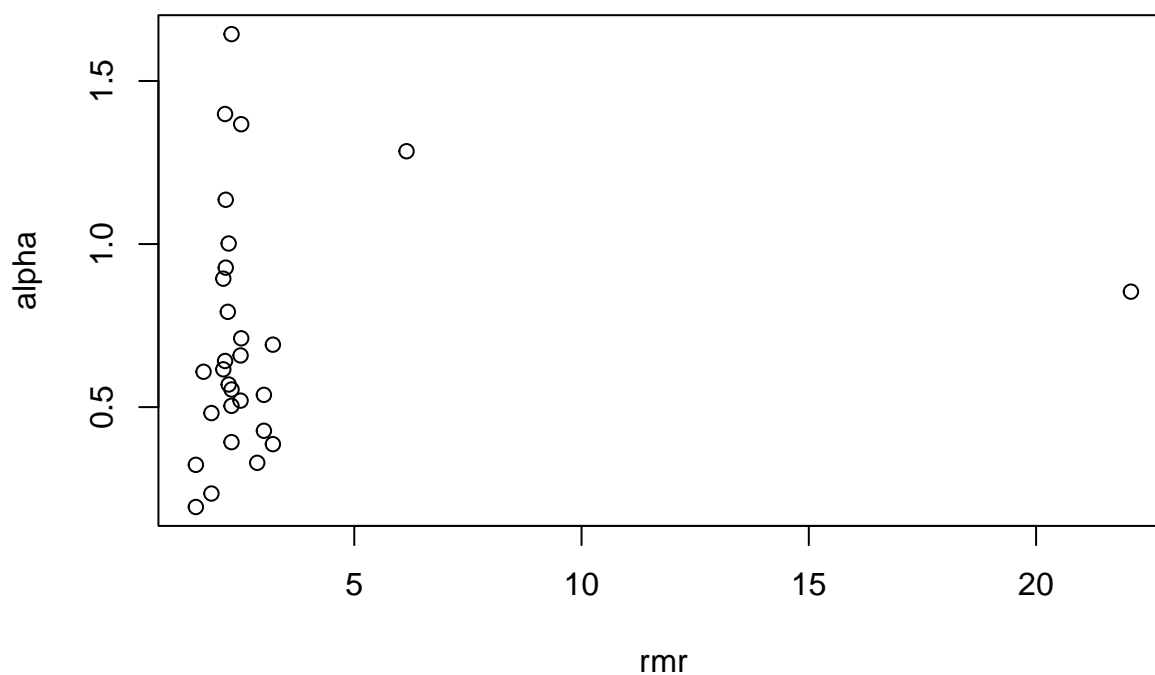
One reviewer asked if alpha was correlated with mass or with RMR. The previous analysis answers the first part of that question. Here I attempt to answer the second. I did not include RMR in the linear mixed effects model because there is a well-documented relationship between RMR and mass, as can be seen in Figure 3 of this analysis, and I did not want to include a

```
summary(aov(lm(alpha~rmr,data=pcrits)))
```

```
##           Df Sum Sq Mean Sq F value Pr(>F)
## rmr        1  0.067  0.06668    0.484  0.493
## Residuals  27  3.720  0.13778
```

With a p-value of 0.493, there appears to be no significant relationship between alpha and rmr. Here is the graph.

```
plot(alpha~rmr,data=pcrits)
```



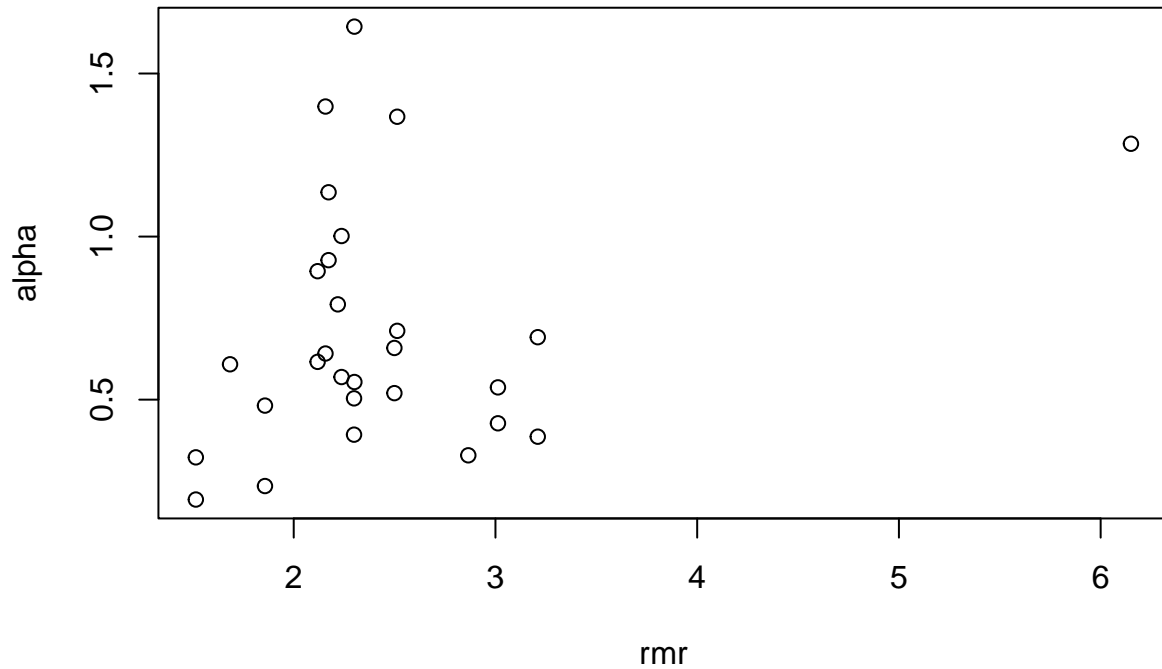
It seems the analysis could be significantly influenced by the one very high rmr of over 20. Just to cover all my bases, we should see if the linear regression is still not significant when that datum is excluded.

```
summary(aov(lm(alpha~rmr,data=pcrits[pcrits$rmr<20,])))
```

```
##           Df Sum Sq Mean Sq F value Pr(>F)
## rmr        1  0.282  0.2819    2.103  0.159
## Residuals  26  3.484  0.1340
```

It is still not significantly related.

```
plot(alpha~rmr,data=pcrits[pcrits$rmr<20,])
```



11 N values for each measurement for flowchart

```
sum(pcrits$pco2==1000&pcrits$day==1)
```

```
## [1] 7
```

```
sum(pcrits$pco2==1000&pcrits$day==7)
```

```
## [1] 7
```

```
sum(pcrits$pco2==1800&pcrits$day==1)
```

```
## [1] 8
```

```
sum(pcrits$pco2==1800&pcrits$day==7)
```

```
## [1] 7
```