

10-08-25

THYLAKOID PURIFICATION

We start in LAB. In the cold room (4°C) we prepare B₁ with the leaves ~~to~~. We blend it, then centrifuge it at 4°C.

↳ important to not touch the stems: they contain fibres we don't want

- We blend some leaves with 100 mL of B₁ with the pulse setting.
- With a cheese cloth ~~we~~ and a funnel we filter the mixture into bottles.

(Don't blend too much, this causes heat)

- We centrifuge the mixtures at 2000g in 4°C for 10 minutes

To do this, we first make sure every bottle weighs the same by either adding more buffer or taking out solution.

→ + add buffer B₂

- We discard the supernatant and resuspend the pellet (green stuff stuck to the side) with a soft paintbrush to not disturb it too much.
- We centrifuge again at 4000g in 4°C for 10 minutes
- Discard supernatant + add B₃. Resuspend pellet with brush
- We centrifuge at 8000g in 4°C for 10 minutes
- We resuspend the pellet in B₄.

All of this is done as much in the cold/dark as possible. The blinds in the centrifuge room weren't entirely closed.

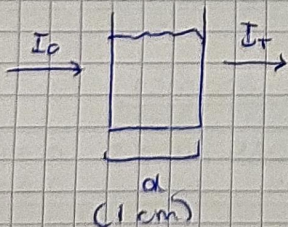
After lunch we took 1 part of our solution and mixed it with 200 parts acetone. After centrifuging it, the proteins were stuck to the bottom.

We pipetted only the not-solid parts into new vials. With these vials we do spectroscopy.

We pipetted around 300 µL into the cuvette of the spectrometer.
 → the mixture should be barely green

We ran the program and got our graphs on USB to further analyse.

spectroscopy



$$T = \frac{I_T}{I_0} \text{ (transmission)}$$

$$\epsilon(d) = \frac{A}{c \cdot d}$$

$$A = -\log_{10} T \text{ (absorbance)}$$

$$A = \epsilon \cdot c \cdot d$$

extinction coefficient

concentration

optical path

We want concentration $C = \frac{A}{\epsilon(d) \cdot d}$

Data analysis

- ① open excel file
- ② filter from low to high
- ③ create extra column $(B_1 - \text{MIN}(B:B))$
and drag it down
- ④ (fitting program) data \rightarrow search \rightarrow solver \rightarrow solver add-in \rightarrow ok
(tick it)
- ⑤ copy column from spectroscopy, paste it in fitting program
under "sample" (values)
- ⑥ click 1, then 2
- ⑦ go to 'chi' \rightarrow put chi f to 0
- ⑧ data \rightarrow solver \rightarrow solve
- ⑨ fitrosso \rightarrow blue and red should be similar
- ⑩ beta car \rightarrow 5
- ⑪ solver \rightarrow solve (good at n fitting)

origin tests:

- chi/car (jasmin 4.190226)
 - chi/a/chi/b (jasmin 2.59)
- ⑫ dilution \rightarrow 200 \rightarrow note down chi content (mg/ml)
jasmin: 1.24
- vol tot \rightarrow 2