main_transcripts

September 2, 2021

1 Enrichment and Overlap of PGC2+CLOZUK

```
[1]: import re
  import os, errno
  import functools
  import numpy as np
  import pandas as pd
  from plotnine import *
  from pandas_plink import read_plink
  from warnings import filterwarnings
  from matplotlib.cbook import mplDeprecation
  from scipy.stats import fisher_exact, binom_test

filterwarnings("ignore", category=mplDeprecation)
  filterwarnings('ignore', category=UserWarning, module='plotnine.*')
  filterwarnings('ignore', category=DeprecationWarning, module='plotnine.*')
```

1.1 Config and Functions

```
feature = "transcripts"
```

```
[3]: Ofunctools.lru_cache()
    def feature_map(feature):
        return {"genes": "Gene", "transcripts": "Transcript",
                 "exons": "Exon", "junctions": "Junction"}[feature]
    @functools.lru_cache()
    def get_de_df():
         HHH
        Load DE analysis
        return pd.read_csv(config_feature['de_file'], sep='\t', index_col=0)
    @functools.lru_cache()
    def get_eqtl_df():
        eqtl_df = pd.read_csv(config feature['fastqtl_output_file'], sep='\t')
        return eqtl_df[(eqtl_df["Type"] == feature_map(feature))]
    @functools.lru cache()
    def get_gwas_snps():
        return pd.read csv(config['gwas snp file'], sep='\t', index col=0,,,
     →low memory=False)
    @functools.lru_cache()
    def get_integration_df():
        return get_gwas_snps().merge(get_eqtl_df(), left_on='our_snp_id',_
     suffixes=['_PGC2', '_eQTL'])\
                               .merge(get_de_df(), left_on='gene_id',__
     →right_index=True)
    @functools.lru_cache()
    def get residual expression df():
        return pd.read_csv(config_feature['residual_expression_file'],
                            sep='\t', index_col=0).transpose()
    @functools.lru_cache()
    def get_pheno_df():
        return pd.read_csv(config['phenotype_file'], index_col=0)
```

```
[4]: def agree_direction(row):
         return [-1, 1][row['pgc2_a1_same_as_our_counted']] * np.sign(row['OR'] - 1)__
      →* np.sign(row['slope']) * np.sign(row['t'])
     def letter_snp(number, a0, a1):
         Example:
         letter_snp(0, 'A', 'G') is 'AA'
         letter_snp(1, 'A', 'G') is 'AG'
         letter_snp(2, 'A', 'G') is 'GG'
         if np.isnan(number):
             return np.nan
         if len(a0) == 1 and len(a1) == 1:
             sep = ''
         else:
             sep = ' '
         return sep.join(sorted([a0]*int(number) + [a1]*(2-int(number))))
     def get_gwas_snp(snp_id):
         gwas = get_gwas_snps()
         r = gwas[gwas['our_snp_id']==snp_id]
         assert len(r) == 1
         return r
[5]: Ofunctools.lru_cache()
     def get_expression_and_pheno_df():
         return pd.merge(get_pheno_df(), get_residual_expression_df(),__
      →left_index=True, right_index=True)
     @functools.lru_cache()
     def get_plink_tuple():
         Usage: (bim, fam, bed) = get_plink_tuple()
         return read_plink(config['plink_file_prefix'])
     @functools.lru_cache()
     def subset_bed():
         11 11 11
         This subsets the bed and bim file and returns the new subsetted
         data with shared brain_ids.
```

```
This is to speed up accessing the bed file.
    (bim, fam, bed) = get_plink_tuple()
    brain_ids = list(set(get_expression_and_pheno_df()['BrNum']).
 →intersection(set(fam['fid'])))
    fam pos = list(fam[(fam["fid"].isin(brain ids))].
→drop_duplicates(subset="fid").loc[:, 'i'])
    unique_snps = get_eqtl_df().variant_id.unique()
    snp_info = bim[(bim["snp"].isin(unique_snps))].copy()
    snp_pos = list(snp_info.loc[:, "i"])
    new bed = bed[snp pos].compute()[:,fam pos]
    new_bim = bim[(bim["i"].isin(snp_pos))].reset_index(drop=True)
    new_bim['ii'] = new_bim.index
    return new_bed, new_bim, brain_ids
@functools.lru_cache()
def get_snp_df(snp_id):
    Returns a dataframe containing the genotype on snp snp_id.
    The allele count is the same as in the plink files.
    Example:
    qet_snp_df('rs653953').head(5)
            rs653953 num rs653953 letter rs653953
    Br5168
                        0
                                        GG
                                              0 \setminus nGG
    Br2582
                        1
                                       AG
                                             1 \setminus nAG
    Br2378
                        1
                                        AG
                                             1 \setminus nAG
    Br5155
                        2
                                        AA
                                             2 \backslash nAA
    Br5182
                        2
                                              2 \backslash nAA
                                       AA
    bed, bim, brain ids = subset bed()
    snp_info = bim[bim['snp']==snp_id]
    snp_pos = snp_info.iloc[0]['ii']
    dfsnp = pd.DataFrame(bed[[snp_pos]], columns=brain_ids, index=[snp_id + u
→'_num']).transpose().dropna()
    my_letter_snp = functools.partial(letter_snp, a0=snp_info.iloc[0]['a0'],_
 \rightarrowa1=snp_info.iloc[0]['a1'])
    # the 2 - in next line is to workaround a possible bug in pandas plink? a1_{\sqcup}
\rightarrow and a0 inverted
    dfsnp[[snp_id + '_num']] = 2 - dfsnp[[snp_id + '_num']].astype('int')
    dfsnp[snp_id + '_letter'] = dfsnp[snp_id + '_num'].apply(my_letter_snp)
    dfsnp[snp_id] = (dfsnp[snp_id + '_num'].astype('str') + '\n' +
                      dfsnp[snp_id + '_letter'].astype('str')).astype('category')
    return dfsnp
```

```
@functools.lru_cache()
     def get_gwas_ordered_snp_df(snp_id):
         Returns a dataframe containing the genotype on snp snp_id.
         The allele count is the number of risk alleles according to GWAS.
         Example:
         get_gwas_ordered_snp_df('rs653953').head(5)
                 rs653953_num rs653953_letter rs653953
         Br5168
                             2
                                             GG
                                                   2 \backslash nGG
         Br2582
                             1
                                             AG
                                                   1 \setminus nAG
         Br2378
                             1
                                                  1 \setminus nAG
                                            AG
                             0
         Br5155
                                             AA \quad O \setminus nAA
                             0
                                                   O \setminus nAA
         Br5182
                                            AA
         111
         pgc = get_gwas_snps()
         dfsnp = get_snp_df(snp_id).copy()
         gwas_snp = get_gwas_snp(snp_id)
         if gwas_snp['pgc2_a1_same_as_our_counted'].iloc[0]:
             if gwas snp['OR'].iloc[0] > 1:
                 pass
             else:
                 dfsnp[[snp_id + '_num']] = 2 - dfsnp[[snp_id + '_num']]
         else:
             if gwas_snp['OR'].iloc[0] > 1:
                 dfsnp[[snp_id + '_num']] = 2 - dfsnp[[snp_id + '_num']]
             else:
                 pass
         dfsnp[snp_id] = (dfsnp[snp_id + '_num'].astype('str') + '\n' +
                           dfsnp[snp_id + '_letter'].astype('str')).astype('category')
         return dfsnp
[6]: Ofunctools.lru_cache()
     def get_biomart_df():
         biomart = pd.read_csv(config['biomart_file'])
         biomart['description'] = biomart['description'].str.replace('\[Source.
      →*$','', regex=True)
         return biomart
     def get_gene_symbol(gene_id, biomart=get_biomart_df()):
```

ensge = re.sub('\..+\$','', gene_id)

```
ggg = biomart[biomart['ensembl_gene_id']==ensge]
    if ggg.shape[0]==0:
        return '', ''
    gs = ggg['external_gene_name'].values[0]
    de = ggg['description'].values[0]
    if type(de)!=str:
        de = ''
    de = re.sub('\[Source:.*$','',de)
    return gs, de
@functools.lru cache()
def get_risk_allele(snp_id):
    gwas_snp = get_gwas_snp(snp_id)
    if gwas_snp['OR'].iloc[0] > 1:
        ra = gwas_snp['A1'].iloc[0]
        ra = gwas_snp['A2'].iloc[0]
    return ra
```

```
[7]: def get_snp_gene_pheno_df(snp_id, gene_id, snp_df_func):
        pheno_columns = list(get_pheno_df().columns)
        expr_df = get_expression_and_pheno_df()[pheno_columns + [gene_id]]
        snp df = snp df func(snp id)
        return expr_df.merge(snp_df, left_on='BrNum', right_index=True)
    def simple_snp_expression_pheno_plot_impl(snp_id, gene_id, snp_df_func,_u
     →pheno_var):
        df = get_snp_gene_pheno_df(snp_id, gene_id, snp_df_func)
        df['Dx'] = df.Dx.astype('category').cat.rename_categories({'Control':
     y0 = df[gene_id].quantile(.01) - 0.26
        y1 = df[gene_id].quantile(.99) + 0.26
        pjd = position_jitterdodge(jitter_width=0.27)
        p = ggplot(df, aes(x=snp_id, y=gene_id, fill=pheno_var)) \
        + geom_boxplot(alpha=0.4, outlier_alpha=0) \
        + geom_jitter(position=pjd, stroke=0, alpha=0.6) + ylim(y0, y1) \
        + labs(y='Residualized expression', fill='Diagnosis') \
        + theme_bw(base_size=20)\
        + theme(legend title=element text(face='bold'),
                panel_grid_major=element_blank(),
                panel grid minor=element blank())
        return p
```

```
[8]: def save_plot(p, fn):
         for ext in ['png', 'pdf', 'svg']:
             p.save(fn + '.' + ext)
     def gwas_annotation(snp_id):
         return 'SZ GWAS pvalue: %.1e' % get_gwas_snp(snp_id).iloc[0]['P']
     def eqtl_annotation(snp_id, gene_id):
         r = get_eqtl_df()[(get_eqtl_df()['variant_id']==snp_id) &
                           (get_eqtl_df()['gene_id']==gene_id)]
         assert len(r)==1
         return 'eQTL nominal p-value: %.1e' % r.iloc[0]['pval_nominal']
     def de_annotation(gene_id):
         g = get_de_df()[(get_de_df()['transcript_id'] == gene_id)]
         return 'DE adj.P.Val: %.3f' % g.iloc[0]['adj.P.Val']
     def risk_allele_annotation(snp_id):
         return 'SZ risk allele: %s' % get_risk_allele(snp_id)
     def gwas_annotated_eqtl_pheno_plot(snp_id, gene_id, pheno_var):
         p = simple_gwas_ordered_snp_expression_pheno_plot(snp_id, gene_id,_
     →pheno_var)
         de_df = get_de_df()[(get_de_df()['transcript_id'] == gene_id)]
         gene_symbol, gene_description = get_gene_symbol(de_df.iloc[0]['gene_id'])
         title ="\n".join([gene_symbol,
                           gene_id,
                           gwas_annotation(snp_id),
                           risk_allele_annotation(snp_id),
                           eqtl_annotation(snp_id, gene_id),
                           de_annotation(gene_id)])
         p += ggtitle(title)
         return p
```

1.2 Transcripts

```
[9]: try:
    os.makedirs(feature)
except OSError as e:
    if e.errno != errno.EEXIST:
        raise
```

1.2.1 Enrichment

Integrate DEG with PGC2+CLOZUK SNPs

```
[10]: dft = get_integration_df()
dft.shape
```

/home/jbenja13/.local/lib/python3.9/site-packages/numpy/lib/arraysetops.py:583: FutureWarning: elementwise comparison failed; returning scalar instead, but in the future will perform elementwise comparison

```
[10]: (2280801, 65)
```

[[989, 54364], [48243, 2177205]]

[11]: (0.8210127548449392, 4.079815690149912e-10)

```
[12]: dft1 = dft[(dft['P']<5e-8) & ((dft['adj.P.Val']<.05))]
    df = dft1.groupby('agree_direction').size().reset_index()
    df</pre>
```

```
[12]: agree_direction 0
0 No 231
1 Yes 758
```

```
[13]: binom_test(df[0].iloc[1], df[0].sum())
```

[13]: 4.805232318453914e-66

```
[14]: dft2 = dft[(dft['P']<=5e-8) & (dft['adj.P.Val'] < 0.05)].copy() dft2['risk_allele'] = dft2['our_snp_id'].apply(get_risk_allele)
```

```
direction = {-1: 'Down', 1: 'Up'}
      boolean_conv = {True: 1, False: -1}
      dft2.pgc2_a1_same_as_our_counted = [boolean_conv[item] for item in_
      →dft2['pgc2_a1_same_as_our_counted']]
      dft2['eqtl gwas dir'] = [direction[item] for item in np.
      ⇒sign(dft2['pgc2_a1_same_as_our_counted']) * np.sign(dft2['slope']) * np.

sign(dft2['OR'] - 1)]
      dft2['de_dir'] = [direction[item] for item in np.sign(dft2['t'])]
      dft2['eqtl_slope'] = np.sign(dft2['pgc2_a1_same_as_our_counted']) * np.

→sign(dft2['OR'] - 1) * dft2['slope']
      dft2 = dft2[['gene_id', 'gene_name', 'variant_id', 'A1', 'A2', 'risk_allele',
      \hookrightarrow 'OR',
                   'P', 'pval_nominal', 'adj.P.Val', 'logFC', 't', 'eqtl_slope',
                   'de_dir', 'eqtl_gwas_dir', 'agree_direction']]
      dft2.to_csv('%s/integration_by_symbol.txt' % feature, sep='\t', index=False)
[15]: df2 = dft2.groupby(['gene_id']).first().reset_index().sort_values('P')
      df2.groupby(['agree_direction']).size()
[15]: agree_direction
      No
             3
      Yes
             6
      dtype: int64
[16]: df2.set_index('gene_name')
[16]:
                           gene_id
                                            variant_id A1 A2 risk_allele
                                                                                OR \
      gene_name
     HCG11
                ENST00000411553.2
                                      chr6:26466161:G:A G A
                                                                       A 0.91432
      ZNF391
                ENST00000244576.8
                                      chr6:27280994:A:C A
                                                           C
                                                                       A 1.07340
                                                                       G 0.92873
     PLCH2
                ENST00000378486.7
                                       chr1:2440958:A:G A
                                                           G
      ZSCAN26
                ENST00000617168.4
                                      chr6:27910960:T:G T
                                                                       T 1.07250
                                                                       C 1.06040
      CNNM2
                ENST00000369878.8 chr10:102825368:C:A C
                                                           Α
      IP6K3
                ENST00000293756.4
                                      chr6:33741539:G:A G
                                                           Α
                                                                       G 1.07670
      SREBF2
                ENST00000361204.8
                                    chr22:41885425:A:G A
                                                                       A 1.05670
     HCG4
                ENST00000418983.1
                                     chr6:29388857:G:C G
                                                           C
                                                                       C 0.93808
      ZNF14
                ENST00000344099.3
                                    chr19:19623068:T:C T
                                                                       T 1.06130
                           P pval_nominal adj.P.Val
                                                           logFC
                                                                        t \
      gene_name
                 1.020000e-14 1.890010e-05
                                             0.002565 0.102920 4.413076
      HCG11
      ZNF391
                 1.610000e-13 1.931590e-04
                                              0.036305 0.086460 3.470055
      PLCH2
                4.630000e-11 1.180570e-08
                                              0.010192 -0.193953 -3.957680
      ZSCAN26
                6.280000e-11 1.656890e-04
                                             0.016166 -0.419030 -3.793507
                                              0.000852 0.084962 4.753266
      CNNM2
                1.120000e-09 4.928250e-11
      IP6K3
                4.780000e-09 6.262500e-08
                                              0.005209 -0.262250 -4.180640
```

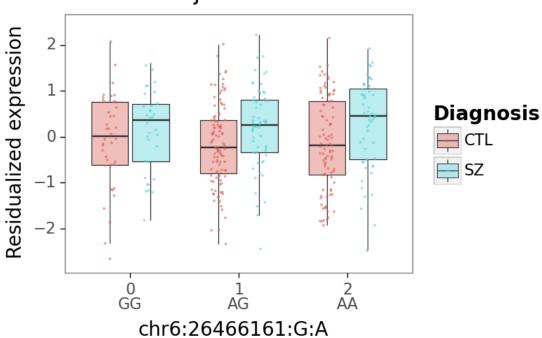
```
8.110000e-09 6.456540e-05
SREBF2
                                       0.018145 -0.073932 -3.747975
HCG4
           2.870000e-08 7.612650e-05
                                       0.028722
                                                 0.251395 3.568189
ZNF14
           4.300000e-08 2.146240e-05
                                       0.047456
                                                 0.077718 3.357046
           eqtl_slope de_dir eqtl_gwas_dir agree_direction
gene_name
HCG11
            0.201059
                         Uр
                                       Uр
                                                      Yes
ZNF391
           -0.170013
                                     Down
                                                       No
                         Uр
PLCH2
           -0.266826
                                     Down
                                                      Yes
                       Down
ZSCAN26
           -0.238187
                       Down
                                     Down
                                                      Yes
CNNM2
            -0.179745
                                     Down
                                                       No
                         Uр
IP6K3
            0.221272
                       Down
                                       Uр
                                                       No
SREBF2
           -0.182903
                       Down
                                     Down
                                                      Yes
HCG4
            0.220492
                         Uр
                                       Uр
                                                      Yes
ZNF14
            0.156479
                                                      Yes
                         Uр
                                       Uр
```

1.2.2 Plot with PGC2 risk allele

```
[17]: for xx in range(df2.shape[0]):
    gg = gwas_annotated_eqtl_pheno_plot(df2.iloc[xx, :].variant_id, df2.
    →iloc[xx, :].gene_id, 'Dx')
    print(gg)
    label = '%s/eqtl_gwas_%s' % (feature, df2.iloc[xx, :].gene_name)
    save_plot(gg, label)
```

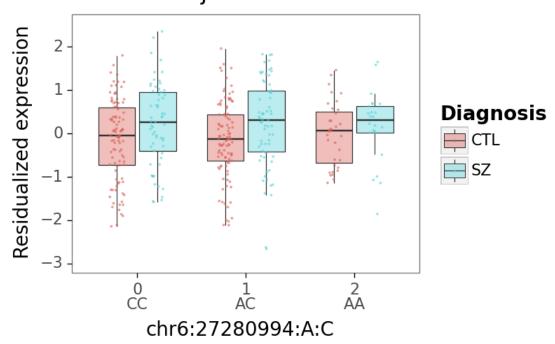
Mapping files: 100% | 3/3 [00:25<00:00, 8.64s/it]

HCG11 ENST00000411553.2 SZ GWAS pvalue: 1.0e-14 SZ risk allele: A eQTL nominal p-value: 1.9e-05 DE adj.P.Val: 0.003



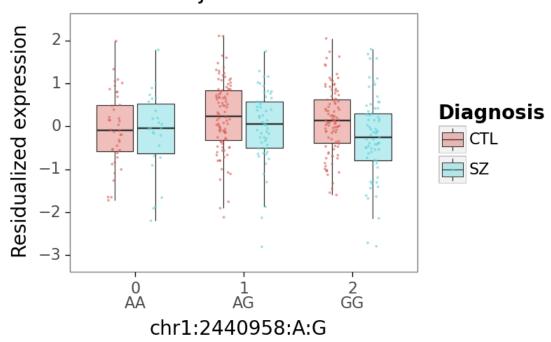
<ggplot: (8729546831080)>

ZNF391 ENST00000244576.8 SZ GWAS pvalue: 1.6e-13 SZ risk allele: A eQTL nominal p-value: 1.9e-04 DE adj.P.Val: 0.036



<ggplot: (8729548902684)>

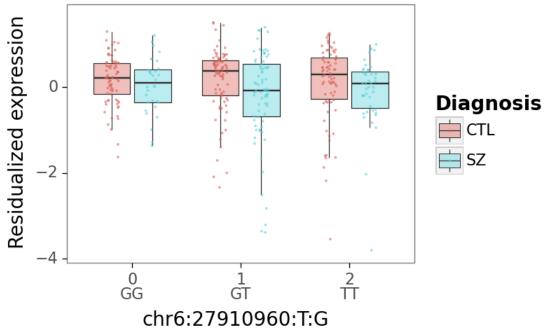
PLCH2 ENST00000378486.7 SZ GWAS pvalue: 4.6e-11 SZ risk allele: G eQTL nominal p-value: 1.2e-08 DE adj.P.Val: 0.010



<ggplot: (8729548537971)>

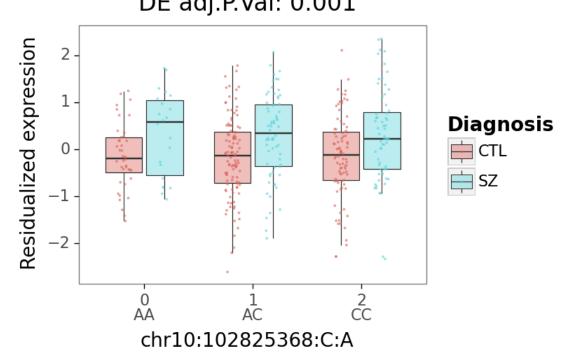
ZSCAN26 ENST00000617168.4 SZ GWAS pvalue: 6.3e-11 SZ risk allele: T eQTL nominal p-value: 1.7e-04 DE adj.P.Val: 0.016

DE adj.P.Val: 0.016



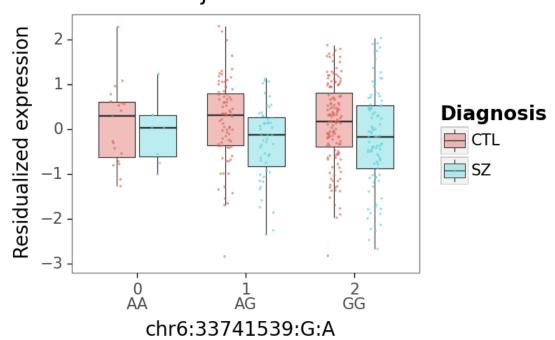
<ggplot: (8729549316563)>

CNNM2 ENST00000369878.8 SZ GWAS pvalue: 1.1e-09 SZ risk allele: C eQTL nominal p-value: 4.9e-11 DE adj.P.Val: 0.001



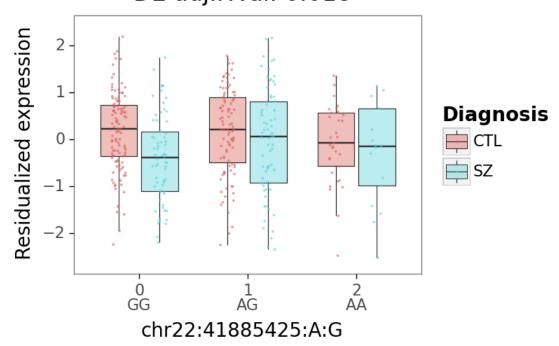
<ggplot: (8729515558193)>

IP6K3 ENST00000293756.4 SZ GWAS pvalue: 4.8e-09 SZ risk allele: G eQTL nominal p-value: 6.3e-08 DE adj.P.Val: 0.005



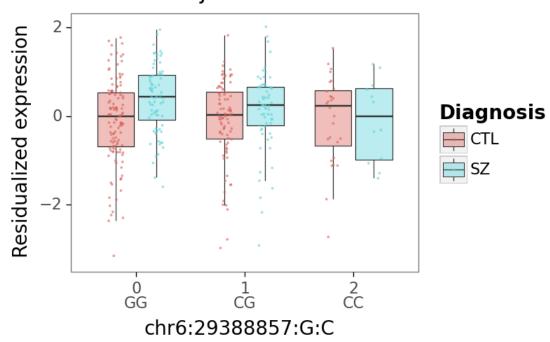
<ggplot: (8729515224149)>

SREBF2 ENST00000361204.8 SZ GWAS pvalue: 8.1e-09 SZ risk allele: A eQTL nominal p-value: 6.5e-05 DE adj.P.Val: 0.018



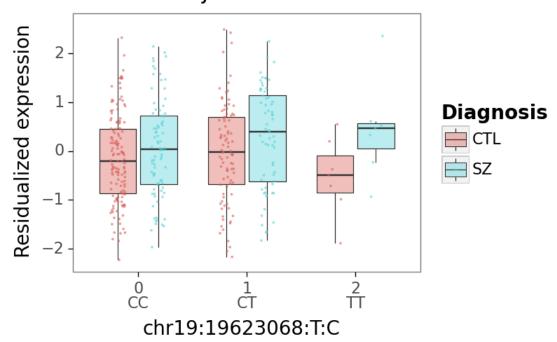
<ggplot: (8729514559657)>

HCG4
ENST00000418983.1
SZ GWAS pvalue: 2.9e-08
SZ risk allele: C
eQTL nominal p-value: 7.6e-05
DE adj.P.Val: 0.029



<ggplot: (8729513879065)>

ZNF14 ENST00000344099.3 SZ GWAS pvalue: 4.3e-08 SZ risk allele: T eQTL nominal p-value: 2.1e-05 DE adj.P.Val: 0.047



<ggplot: (8729515214743)>