Serum concentrations of 25-hydroxyvitamin D and

immunoglobulins in the elderly Swiss cohort:

results of the Senior Labor Study

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Abstract

Background

Vitamin D and the components of humoral immunity play important roles in human health. Elderly people have lower 25-hydroxyvitamin D (25(OH)D) serum levels than younger adults. We aimed to determine the levels of 25(OH)D serum concentrations in healthy senior citizens and to study their relationship to the levels of components of humoral immunity.

Methods

1,470 healthy elderly Swiss men and women, 60 years or more, were recruited for this study. 179 subjects were dropped out of the study because of elevated serum concentrations of C reactive protein. Fasting blood sera were analyzed for 25(OH)D with the high-performance liquid chromatography (HPLC) and for parathyroid hormone (PTH), immunoglobulins and complement C4 & C3 concentrations with immunoassays. The percentage of participants in each of the four different 25(OH)D deficiency groups severely deficient (<10 ng/ml), deficient (10-20), insufficient (21-30 ng/ml) and normal (>31 ng/ml) were statistically compared. A relationship of major components of the humoral system and age to 25(OH)D levels was also assessed.

Results

About 66% of the subjects had insufficient levels of 25(OH)D. Normal levels of 25(OH)D were found in 26.1% of the subjects of which 21% were males and 30.5% were females (total study population). 7.98% of the total study population had severely deficient levels of 25(OH)D. Low levels of 25(OH)D, were positively associated with IgG2 (p<0.01) and with C4 (p<0.02) yet inversely related to levels of IgG1 and IgA (p<0.05) and C3 (p<0.01). Serum levels of total IgA, IgG, IgG2, and IgG4 peaked together with 25(OH)D during late summer.

Conclusions

A majority (~2/3) of the healthy elderly Swiss population presented with Vitamin D insufficiency. The incremental shift in IgA and C3 levels might not necessarily reflect deranged humoral immune defense; however, given the high prevalence of vitamin D deficiency, the importance of this condition in humoral immunity will be worth looking at more closely. This study supports the role of vitamin D in the competent immune system.

Keywords

25(OH)D

immunoglobulins

IgA

Complement

Humoral immunity

Sunlight exposure

Elderly people

Background

The biological role of fat-soluble vitamin D has been shown in recent studies to extend far beyond its role in calcium homeostasis and bone health [1-4]. Currently, the structure of vitamin D receptors and their functions are well understood. Once the receptors bind to their ligand, VDRs dimerize with an isoform of the retinoid X receptor (RXR). Many body tissues express VDR; furthermore, VDR-RXRs are found on a variety of cells, including monocytes/macrophages [3], kidney cells[5], cardiomyocytes [6], skin cells [7] and liver cells [8]. At each corresponding organ site, the vitamin D/VDR system is associated with functional organ performance.

Experimental, epidemiological and clinical studies have shown inverse associations of low vitamin D status with longevity and with retarding immunosenescence [9]. 1,25-dihydroxyvitamin D3 $(1,25(OH)_2D_3)$ blocks the induction of T-helper-1 (T_H1) -cell cytokines (e.g., IFN γ) and promotes T_H2 -cell responses. This activity partly explains the decreased B-cell proliferation, plasma-cell differentiation and IgG secretion [6, 9, 10] which is an activity that conserves efficient immune defense (in particular anti-viral) [3]. The actions of 1,25 $(OH)_2D_3$ on B cell function have been researched for many years [10] and this area has recently received interest regarding the mechanisms of action [6, 11-14]; for example, a recent study described an inverse relation of serum total IgG levels with serum 25-hydroxyvitamin D (25(OH)D) [15]. Although some results remain to be confirmed, low serum 25(OH)D levels have been linked to all-cause cardiovascular, cancer and infection-related mortality and also to stroke. Furthermore, autoimmunity and some infectious agents may also cause disease because of insufficient 25(OH)D [11].

Throughout the life span, a significant proportion of humans have insufficient (<30 ng/ml) or deficient (<20 ng/ml) serum 25(OH)D levels. 25(OH)D severe deficiency may be defined as serum concentrations < 10 ng/ml [16, 17], deficiency as values < 20 and insufficiency as

21-29 ng/ml whereby values >30 ng/ml (75 nmol/L) shall reflect sufficient concentration [2, 18, 19]. Regular exposures to sunlight foster the required hydroxylations to produce the bioactive compound. The first –OH attachment results in 25-hydroxyvitamin D and the second in 1,25-dihydroxyvitamin D (1,25(OH)₂D). Humans acquire much of their vitamin D through a photosynthetic reaction in the skin with a small amount via digestive intake [20]. Those at risk for vitamin D deficiency are those with little sun exposure and/or poor dietary intake. Older people are especially at risk because aging lowers the amount of 7-dehydrocholesterol in the skin reducing the capacity for vitamin D production. People with dark skin have a significantly lower mean serum level of 25(OH)D than those with light skin across the life-span, but dark skin people have a lower level of vitamin D-binding protein and consequently more bioactive Vitamin D [21]. The acquisition of Vitamin D may be hampered by increased melanin production, as is the case with dark skin or with increased age [22]. The European population is known for transient reductions of 10 – 20% of individual 25(OH)D serum levels measured during the winter months compared to the summer months, an observation that is particularly prominent in the elderly [23].

The present study was initiated (i) to study the prevalence of insufficiency and deficiency levels in healthy elderly and (ii) to assess the impact of these deficiencies on immunoglobulin and complement components C4 and C3 production.

Methods

Study population

Consecutive subjectively healthy elderly volunteers aged 60 years and older were recruited between February 2009 and July 2010 as part of the Senior Labor Study, which is an ongoing investigation in the canton of Berne (Switzerland) initially aimed at creating appropriate reference intervals (RIs) for several analytes in the elderly (http://www.seniorlabor.ch). The study participants were contacted through newspaper advertisements and different clubs and associations that had a high probability of having healthy elderly members (e.g., alpine clubs, sports clubs) and through personal contacts of collaborators of the study organization.

A personal history of each subject was taken, anthropometric measurements were performed, and fasting venous blood was drawn into S-Monovette tubes (Sarstedt, Sevelen, Switzerland). Of the 1,470 participants, 179 with high total leukocyte count (>11.5 G/L) and/or high C-reactive protein levels (>5 mg/L) were excluded from further analysis since such laboratory cutoffs are strong indicators of the presence of systemic inflammation, mostly for upper respiratory tract infections. Of the remaining 1291 subjects, 558 were between 60-69 yrs old, 498 between 70-79 yrs old and 235 were \geq 80 yrs old (Table 1).

This study was conducted in accordance with the ethical guidelines of the 1975 Declaration of Helsinki, and informed consent was obtained from all participants. Ethical approval for the present study was obtained from the Berner Kantonal Ethics committee (KEK-Berne-Study Nr 166/08), Bern, Switzerland.

Laboratory testing

All assays were done at our institution. For 25(OH)D concentration measurements, we used the LaChrom Elite apparatus provided by Hitachi (Hitachi Medical Systems Europe, Zug, Switzerland). 25(OH)D₂ and 25(OH)D₃ (derived from cholecalciferol) were distinguishable using this method; for this study, only the latter more stable animal-origin 25(OH)D₃ levels are reported. The US National Institute of Standards and Technology (NIST) calibration standard reference material (www.nist.gov) was consistently followed with each test batch, and only the values complying with the requirements were released; the inter-and intra-assay variations were < 5%. The baseline total 25(OH)D levels were categorized as normal, insufficient, deficient, and severely deficient using levels adapted from the definitions proposed earlier[17, 24]. The IMMULITE 2000 assay was used for quantification of serum 1-84 parathyroid hormone (PTH); this is a solid-phase two-site chemiluminescent immunoassay with a monoclonal mouse capture antibody and a polyclonal goat signal antibody conjugated to alkaline phosphatase.

The levels of immunoglobulins, where applicable, were measured using the BN Prospec $^{\circledR}$ Nephelometry System (Siemens, Zurich, Switzerland), incorporating stabilized human serum as the N Protein Standard SL. Subclass-specific sheep anti-human IgG1 and IgG2 were used in conventional nephelometry and anti-human IgG3 and IgG4 in this system is loaded on polystyrol particles to enhance sensitivity for quantitation of the cognate subclass. For C3 und C4, we used sodium azide preserved anti-C3c antibodies < 6.4 mg/ml and anti-C4 < 5.4 mg/ml. The company recommends application of the following reference intervals established with serum and/or plasma samples from healthy adults: C3/C3c of 0.9 - 1.8 mg/ml and C4/C4c of 01.-0.4 mg/ml.

Statistical procedures

The serum concentration values of 25(OH)D, immunoglobulin classes, IgG subclasses and complements were statistically analyzed using SPSS version 17.0 for Microsoft Windows (SPSS, Inc., Chicago, IL). All statistical analyses were based on two sided hypothesis tests with a significance level of P < 0.05. Kruskal-Wallis test was used to test for statistically significant associations of 25(OH)D and immunoglobulins in more than two groups. Further, the Mann-Whitney rank sum was used to statistically compare the levels of various immunoglobulins and complement components by categories of 25(OH)D levels (normal, insufficient, and deficiency groups). Univariate linear regression analysis was used to examine the relationship between 25(OH)D as the dependent variable and age, immunoglobulin and complement components as the independent variables.

Results

The entire cohort of 1291 people studied revealed a significant correlation between aging and 25(OH)D levels (Figure 1A). To analyze in detail the effect of aging on 25(OH)D, participants were divided into three age groups (60-69 years , 70-79 years and >80 years) (Figure 1B). Only 26.1% of the subjects had normal 25(OH)D levels (>30ng/ml). These consisted of 21% of the males and 30.5% of the female participants of total study population. The detailed distribution of the subjects into the respective 25(OH)D deficiency groups is presented in Table 1. Severe 25(OH)D deficiencies (levels < 10 ng/ml) were observed in 8.0% (M: 8.0%, F: 8.0%) of the subjects. However, it should be noted that subjects became increasingly 25(OH)D deficient with increasing age, with a prevalence of severe deficiency 3.5%, 8.5%, and 16.3% in women and 4.4%, 10.1%, and 13.0% in men in the progressively older age groups (Table 1). About 50% of the healthy elderly Swiss over 70 years have insufficient 25(OH)D levels. Serum PTH levels inversely correlated with 25(OH)D) (p<0.001) (Table 2). 25(OH)D levels were significantly different according to the month the samples were collected (p<0.001) (additional file 1).

We also evaluated the relationship between 25(OH)D levels and major humoral components of innate and acquired immunity. While levels of IgA inversely correlated to 25(OH)D, IgM nor of total IgG or IgE differed from changing 25(OH)D levels (Figure 2 and Table 2). The p values were obtained either using Kruskal Wallis analysis across the board for trend or by comparing the 25(OH)D severely deficient and the normal group.

The mean serum level of IgG2 significantly differed among 25(OH)D level groups (Table 2). Further, complement component C4 increased with increasing 25(OH)D (Figure 3 A & B), whereas complement component C3 decreased with increasing 25(OH)D (Figure 3 C & D). Seasonal variation in immunoglobulin levels and complement components revealed differences according to the month samples were taken (additional file 2). Significant

differences were observed for total IgG (p=0.001), IgG2 (p=0.003), IgG4 (p=0.04), complement component C4 (p=0.01), and complement component C3 (p=0.006). Total IgG, IgG2, and IgG4 peaked together with 25(OH)D during August.

Discussion

This is the first time, that the 25(OH)D status is confronted to humoral immunity features in healthy elderly subjects not receiving any real-time medical care; in fact, most studies reporting insufficient/deficient 25(OH)D levels have been performed in the context of disease, such as patients assigned to primary care [25], with (risk for) bone fractures [26], prostate cancer [27], cardiovascular disease [28], and evaluations in response to influenza or recombinant hepatitis B vaccine [29, 30]. Some authors have questioned the quality of such studies, mainly for technical reasons [31]. Our study is in line with recent epidemiological investigations demonstrating a high prevalence of mild and severe 25(OH)D deficiency; in one of such studies, this deficiency did not significantly increase with aging [32], whereas it did in another [33].

Here, a majority (over 50%) of the elderly subjectively healthy Swiss population presented with 25(OH)D insufficiency/deficiency. We observed a progressive decline in 25(OH)D levels with aging both in women and men, most likely due to lower sunlight exposure and decline in balanced food composition for those not (yet) part of homes for retired elderly, as those under study here. In the Osteoporotic Fractures in Men Study (MrOS) from North America, 31% of men aged 80-84 yrs had levels < 20 mg/ml [33], and the proportion increased to 40% in those >85 yrs [33]. It can be assumed that Vitamin D insufficiency causes a substantial burden of disease (e.g., falls, osteoporosis, arterial stiffness [34],

cardiovascular diseases [21], autoimmune disease, infectious disease, and neoplasia that would classify insufficiency as a risk factor in our subjects rather than a past event. In conjunction with these new insights, 25(OH)D testing requests received by medical analytical laboratories are surging worldwide [35]. The present study was performed in compliance with current good laboratory practice, GLP. The cutoffs for normal, insufficient, or (severely) deficient levels of vitamin D are defined at different levels based on an original proposal [36]. Currently, the deficiency decision limit of vitamin D levels is set by an international agreement < 20 ng/ml, although this cutoff is not recognized by all [18, 24]. Here we considered values < 10 ng/ml as indicative of severe deficiency. Since the separation into 4 different effectual groups cannot not relate to clinical features in the healthy elderly studied, the cutoffs for 25(OH)D levels do not target risk viz. no risk considerations [37].

Depending on latitude and climate, seasonal/monthly variations serum levels of 25(OH) do occur [33] or remain similar[38] and in the cohort under study here did vary, reflected by incremental levels towards end of summer/fall. To the best of our knowledge, it is the first time that we here report a seasonal variation of humoral immunity components more or less consistently with 25(OH)D levels. 25(OH)D insufficiency might also occur in deliberately sunny environment [38]. The variations of levels in humoral immunity components along the year observed in our study might depend on changes in the microbiological mosaic offending us with Gram positive bacteria [39] or else influenza H1N1 [40], examples of the most recently studied pathogens eliciting differential humoral immune responses. In the light of the here found interdependency 25/OH)D/humoral immunity, Vitamin D supplementation therefore must be considered the whole year round on an individual risk/efficiency basis. A recent update of circannual fluctuation of 25(OH)D in 50 kidney transplant recipients found that (i) summer/winter peak-to-peak variations occur in our latitudes with ensuing (ii) an

increased frequency of 25(OH)D insufficiency during winter time [41]; this finding contrasts somewhat to recent reports from Israel [38] and South Africa [42].

To establish reference intervals (RIs), different statistical procedures are commonly applied at each step. The sample size in the >80-yr group of the present study exceeds the 100-observation barrier. Indeed, the significant p-values produced by the SSPS analysis under non-parametric conditions highlight the findings that incremental insufficiency progresses to deficiency with increasing age, a finding not taken into account by the Endocrine Society's Clinical Guidelines [24].

The ability of vitamin D to influence normal human immunity is highly dependent on the 25(OH)D status of individuals, and may lead to aberrant response to infection or autoimmunity in those with insufficient vitamin D. This article describes some of the recent developments with respect to vitamin D and the immune system, and possible clinical implications. The influence that 1,25(OH) vitamin D exerts on the immune system sparks interest for its potential for reducing infections and preventing allograft rejection after transplantation [43].

Beyond the risk for rickets and osteoporosis, other diseases are increasingly being noticed for their association with Vitamin D status [44]. Therefore, we were interested in the recently suggested role of vitamin D and its active metabolite 1,25 (OH)₂D₃ in modulating immune responses [11, 45, 46]. Specific cell types involved in the immune response express the enzyme CYP27B1, which produces 1,25(OH)₂D₃ in situ [47]. Recent studies suggest a role of vitamin D in the reduction in the proliferation of immune cells, their maturation to plasma cells and their production of immunoglobulin [6]. Furthermore, 25(OH)D and human IgG levels have been directly linked in recent reports [15]. A potential explanation for the

significant relationship between IgA, IgG2 and vitamin D discovered in the present study could be explained by the recently suggested interplay between innate and adaptive immunity and the role of T-cell cytokines and cytokine signaling [48, 49], which also might explain the dependency of 25(OH)D levels and C4 and C3. In fact, interferon-γ potentiation of Toll-like receptor–induced antimicrobial peptides is associated with CYP27B1 activity that leads to enhanced bioconversion of 25D₃ to 1,25D₃ in the vitamin D-dependent antimicrobial pathway [50].

In vitro, the effects of 1,25(OH)2D3 on lymphocyte proliferation and cytokine production by PBMCs are pleiotropic [51]. Total serum IgG levels were previously reported to have a negative association with serum 25(OH)D [15], although such association was not present in cerebrospinal fluid [52] nor in our study. Low levels of 25(OH)D are thus a finding occurring in healthy elderly and might have an impact on immune defense.

Conclusions

The majority (~2/3) of healthy elderly Swiss subjects presents with vitamin D insufficiency. With impacts on serum concentrations of IgG1, IgG2, IgA and complement, more so in severe cases of vitamin D deficiency, but also with insufficiency of 25(OH)D, this study supports a role of vitamin D in the immune system under immunocompetent conditions.

Competing interests

The authors declare no competing interests. None of the authors has had in the past 12 months any financial relationship with a biotechnology manufacturer, a pharmaceutical company, or other commercial entity that has any interest in the subject matter, materials, or processes discussed in the manuscript.

Authors' contributions

UN, LR and MR participated in the design of the study; BS, CN, ZS, UN and LR drafted the manuscript and analyzed the data. BS, PM and CN performed the statistical analysis. All authors read and approved the final manuscript.

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References

- 1. Holick MF: The D-lemma: to screen or not to screen for 25-hydroxyvitamin D concentrations. Clin Chem 2010, 56:729-731.
- 2. Holick MF, Binkley NC, Bischoff-Ferrari HA, Gordon CM, Hanley DA, Heaney RP, Murad MH, Weaver CM: Guidelines for preventing and treating vitamin d deficiency and insufficiency revisited. *J Clin Endocrinol Metab* 2012, 97:1153-1158.
- 3. Beard JA, Bearden A, Striker R: **Vitamin D and the anti-viral state.** *J Clin Virol* 2011, **50**:194-200.
- 4. Hajas A, Sandor J, Csathy L, Csipo I, Barath S, Paragh G, Seres I, Szegedi G, Shoenfeld Y, Bodolay E: **Vitamin D insufficiency in a large MCTD population.** *Autoimmunity Rev* 2011, **10:**317-324.
- 5. Messa P, Alfieri C, Rastaldi MP: **Recent insights into vitamin D and its receptor.** *J Nephrol* 2011, **24 Suppl 18:**S30-37.
- 6. Chen S, Sims GP, Chen XX, Gu YY, Lipsky PE: **Modulatory effects of 1,25-dihydroxyvitamin D3 on human B cell differentiation.** *J Immunol* 2007, **179:**1634-1647.
- 7. Luderer HF, Demay MB: **The vitamin D receptor, the skin and stem cells.** *J Steroid Biochem Mol Biol* 2010, **121:**314-316.
- 8. Barchetta I, Carotti S, Labbadia G, Vespasiani GU, Onetti MA, Angelico F, Silecchia G, Leonetti F, Fraioli A, Picardi A, et al: Liver VDR, CYP2R1 and CYP27A1 expression: Relationship with liver histology and vitamin D3 levels in patients with NASH or HCV hepatitis. *Hepatology* 2012.
- 9. Mora JR, Iwata M, von Andrian UH: Vitamin effects on the immune system: vitamins A and D take centre stage. *Nat Rev Immunol* 2008, **8:**685-698.
- 10. Lemire JM, Adams JS, Sakai R, Jordan SC: 1 alpha,25-dihydroxyvitamin D3 suppresses proliferation and immunoglobulin production by normal human peripheral blood mononuclear cells. *J Clin Invest* 1984, 74:657-661.
- 11. Bikle DD: **Vitamin D and immune function: understanding common pathways.** *Curr Osteoporos Rep* 2009, **7:**58-63.
- 12. Enioutina EY, Bareyan D, Daynes RA: **TLR-induced local metabolism of vitamin D3 plays an important role in the diversification of adaptive immune responses.** *J Immunol* 2009, **182:**4296-4305.
- 13. Hewison M: Vitamin D and the intracrinology of innate immunity. *Mol Cell Endocrinol* 2010, **321:**103-111.
- 14. Wimalawansa SJ: **Vitamin D in the new millennium.** *Curr Osteoporos Rep* 2012, **10:**4-15.
- 15. Pincikova T, Nilsson K, Moen IE, Karpati F, Fluge G, Hollsing A, Knudsen PK, Lindblad A, Mared L, Pressler T, Hjelte L: **Inverse relation between vitamin D and serum total immunoglobulin G in the Scandinavian Cystic Fibrosis Nutritional Study.** *Eur J Clin Nutr* 2011, **65:**102-109.
- 16. Kennel KA, Drake MT, Hurley DL: Vitamin D deficiency in adults: when to test and how to treat. *Mayo Clin Proc* 2010, **85:**752-758.
- 17. Ginde AA, Liu MC, Camargo CA: **Demographic differences and trends of vitamin D insufficiency in the US population, 1988-2004.** Arch Intern Med 2009, **169:**626-632.
- 18. Rosen CJ: Clinical practice. Vitamin D insufficiency. N Engl J Med 2011, **364:**248-254.

- 19. Alvarez-Rodriguez L, Lopez-Hoyos M, Garcia-Unzueta M, Amado JA, Cacho PM, Martinez-Taboada VM: **Age and low levels of circulating vitamin D are associated with impaired innate immune function.** *J Leukoc Biol* 2012.
- 20. Heaney RP, Davies KM, Chen TC, Holick MF, Barger-Lux MJ: **Human serum 25-hydroxycholecalciferol response to extended oral dosing with cholecalciferol.** *Am J Clin Nutr* 2003, **77:**204-210.
- 21. Martins D, Wolf M, Pan D, Zadshir A, Tareen N, Thadhani R, Felsenfeld A, Levine B, Mehrotra R, Norris K: **Prevalence of cardiovascular risk factors and the serum levels of 25-hydroxyvitamin D in the United States: data from the Third National Health and Nutrition Examination Survey.** *Arch Intern Med* 2007, **167:**1159-1165.
- 22. Perez-Lopez FR, Fernandez-Alonso AM, Mannella P, Chedraui P: **Vitamin D, sunlight and longevity.** *Minerva Endocrinol* 2011, **36:**257-266.
- 23. Schilling S: Epidemischer Vitamin-D-Mangel bei Patienten einer geriatrischen Rehabilitationsklinik. *Dtsch Arztebl Int* 2012, **109:**33-38.
- 24. Holick MF, Binkley NC, Bischoff-Ferrari HA, Gordon CM, Hanley DA, Heaney RP, Murad MH, Weaver CM: Evaluation, treatment, and prevention of vitamin D deficiency: an Endocrine Society clinical practice guideline. *J Clin Endocrinol Metab* 2011, 96:1911-1930.
- 25. Lapid MI, Cha SS, Takahashi PY: **Vitamin D and depression in geriatric primary care patients.** *Clinical Intervention in Aging* 2013, **8:**509-514.
- 26. Bischoff-Ferrari HA, Willett WC, Orav EJ, Lips P, Meunier PJ, Lyons RA, Flicker L, Wark J, Jackson RD, Cauley JA, et al: **A pooled analysis of vitamin D dose requirements for fracture prevention.** *N Engl J Med* 2012, **367:**40-49.
- 27. Donkena KV, Young CY: **Vitamin d, sunlight and prostate cancer risk.** *Adv Prev Med* 2011, **2011:**281863.
- 28. McGreevy C, Williams D: **New insights about vitamin D and cardiovascular disease: a narrative review.** *Ann Intern Med* 2011, **155:**820-826.
- 29. Chadha MK, Fakih M, Muindi J, Tian L, Mashtare T, Johnson CS, Trump D: **Effect** of 25-hydroxyvitamin D status on serological response to influenza vaccine in prostate cancer patients. *Prostate* 2011, 71:368-372.
- 30. Zitt E, Sprenger-Maehr H, Knoll F, Neyer U, Lhotta K: Vitamin D deficiency is associated with poor response to active hepatitis B immunistion in patients with chronic kidney disease. *Vaccine* 2012, **30:**931-935.
- 31. Elamin MB, Abu Elnour NO, Elamin KB, Fatourechi MM, Alkatib AA, Almandoz JP, Liu H, Lane MA, Mullan RJ, Hazem A, et al: Vitamin D and cardiovascular outcomes: a systematic review and meta-analysis. *J Clin Endocrinol Metab* 2011, 96:1931-1942.
- 32. Lippi G, Montagnana M, Meschi T, Borghi L: Vitamin D concentration and deficiency across different ages and genders. *Aging Clin Exp Res* 2012.
- 33. Orwoll E: **Vitamin D deficiency in older men.** *J Clin Endocrinol Metab* 2009, **94:**1244.
- 34. Giallauria F, Milaneschi Y, Tanaka T, Maggio M, Canepa M, Elango P, Vigorito C, Lakatta EG, Ferrucci L, Strait J: **Arterial Stiffness and Vitamin D Levels: the Baltimore Longitudinal Study of Aging.** *J Clin Endocrinol Metab* 2012.
- 35. Farrell CJ, Martin S, McWhinney B, Straub I, Williams P, Herrmann M: **State-of-the-art vitamin D assays: a comparison of automated immunoassays with liquid chromatography-tandem mass spectrometry methods.** Clin Chem 2012, **58:**531-542.
- 36. Holick MF: Vitamin D deficiency. New Engl J Med 2007, 357:266-281.
- 37. Rosen CJ, Abrams SA, Aloia JF, Brannon PM, Clinton SK, Durazo-Arvizu RA, Gallagher JC, Gallo RL, Jones G, Kovacs CS, et al: **IOM committee members**

- respond to Endocrine Society vitamin D guideline. *J Clin Endocrinol Metab* 2012, **97:**1146-1152.
- 38. Oren Y, Shapira Y, Agmon-Levin N, Kivity S, Zafrir Y, Altman A, Lerner A, Shoenfeld Y: **Vitamin D insufficiency in a sunny environment: a demographic and seasonal analysis.** *The Israel Medical Association journal: IMAJ* 2010, **12:**751-756.
- 39. Ademokun A, Wu YC, Martin V, Mitra R, Saöck U, Baxendale H, Kipling D, Dunn-Walters DK: Vaccination-induced changes in human B-cell repertoire and pneumococcal IgM and IgA antibody at different ages. *Aging Cell* 2011, **10:**922-930.
- 40. Frasca D, Diaz A, Romero M, Mendez NV, Landin AM, Blomberg BB: Effects of age on H1N1-specific serum IgG1 and IgG3 levels evaluated during the 2011-2012 influenza vaccine season. *Immn Ageing* 2013, 10:14.
- 41. Burkhalter F, Schaub S, Dickenmann M: **Preserved circannual rhythm of vitamin D** in kidney transplant patients. *Swiss Medical Weekly* 2012, **10.4414/smw** 2012.13672.
- 42. Martineau AR, Nhamoyebonde S, Oni T, Rangaka MX, Marais S, Bangani N, Tsekela R, Bashe L, de Azevedo V, Caldwell J, et al: **Reciprocal seasonal variation in vitamin D status and tuberculosis notifications in Cape Town, South Africa.** *Proc Natl Acad Sci U S A* 2011, **108:**19013-19017.
- 43. Stein EM, Shane E: **Vitamin D in organ transplantation.** *Osteoporos Int* 2011, **22:**2107-2118.
- 44. Christakos S, Hewison M, Gardner DG, Wagner CL, Sergeev IN, Rutten E, Pittas AG, Boland R, Ferrucci L: **Vitamin D beyond bone.** *Ann N Y Acad Sci* 2013, **1287:**45-58.
- 45. Lalor MK, Floyd S, Goark-Stlinska P, Weir RE, Blitz R, Bransion K, Fine PE, Dockrell HM: **BCG Vaccination: a role for vitamin D?** *PLoS ONE* 2011, **6:**1-11.
- 46. Ovsyannikova IG, Dhiman N, Haralabmieva IH, Vierkant RA, OByrne MM, Jacobson RM, Poland GA: Rubella vaccine-induced cellular immunity: evidence of associations with polymorphisms in the toll-like vitamin A and D receptors, and innate immune response genes. *Hum Genet* 2009, 127:207-221.
- 47. Sigmundsdottir H, Pan J, Debes GF, Alt C, Habtezion A, Soler D, Butcher EC: **DCs** metabolize sunlight-induced vitamin **D3 to 'program' T cell attraction to the** epidermal chemokine CCL27. *Nat Immunol* 2007, 8:285-293.
- 48. Edfeldt K, Liu PT, Chun R, Fabri M, Schenk M, Wheelwright M, Keegan C, Krutzik SR, Adams JS, Hewison M, Modlin RL: **T-cell cytokines differentially control human monocyte antimicrobial responses by regulating vitamin D metabolism.**Proc Natl Acad Sci U S A 2010, **107**:22593-22598.
- 49. Chen Y, Liu W, Sun T, Huang Y, Wang Y, Deb DK, Yoon D, Kong J, Thadhani R, Li YC: 1,25-Dihydroxyvitamin D promotes negative feedback regultation of TLR signaling via targeting microRNA-155-SOCS1 in macrophages. *J Immunol* 2013, 190:3687-3695.
- 50. Liu PT, Wheelwright M, Teles R, Komisopoulou E, Edfeldt K, Ferguson B, Mehta MD, Vazirnia A, Rea TH, Sarno EN, et al: MicroRNA-21 targets the vitamin D-dependent antimicrobial pathway in leprosy. *Nat Med* 2012, **18:**267-273.
- 51. Hustmyer FG, Girasole G, Manolagas SC: **Signal-dependent pleiotropic regulation of lymphocyte proliferation and cytokine production by 1,25-dihydroxyvitamin D3: potent modulation of the hormonal effects by phorbol esters.** *Immunology* 1992, **77:**520-526.
- 52. Holmoy T, Lossius A, Gundersen TE, Moen SM, Castellazzi M, Fainardi E, Casetta I: **Intrathecal levels of vitamin D and IgG in multiple sclerosis.** *Acta Neurol Scand* 2012, **125:**e28-31.

Table 1. Percentage of healthy elderly subjects in the different groups of vitamin D deficiency according to gender

	25(OH)D (ng/ml)											
	Men (600)				Women (691)				Total (1291)			
Age groups	< 10	10 - 20	21 - 29	≥ 30	< 10	10 - 20	21 - 29	≥ 30	< 10	10 - 20	21 - 29	≥ 30
60-69 (558)	4.4	29.3	39.9	26.4	3.5	26.7	32.3	37.5	3.9	28.5	36.0	32.1
70-79 (498)	10.1	38.3	29.1	22.5	8.5	26.6	30.3	34.7	9.2	31.9	29.7	29.1
≥ 80 (235)	13.0	36.0	28.0	23.0	16.3	38.5	20.0	25.2	14.9	37.5	23.4	24.6
Total (1291)	8.0	33.8	33.8	24.3	8.0	28.9	29.1	34.0	8.0	31.2	31.3	29.5

Numbers are percentages of participants classified according to 25(OH)D level group: <10 ng/ml (severe deficiency), 10-20 ng/ ml (deficiency), 21-29 (insufficiency) ng/ml and ≥30 ng/ml (normal)

Table 2. PTH, immunoglobulins and complement C4 and C3 levels compared among 4 concentration ranges of 25(OH)D in 1291 healthy elderly subjects. Differences between groups were assessed by Kruskal-Wallis test.

	25(OH)D										
Analyte	< 10	10-20	21-29	≥ 30	P-Value						
	N = 103	N = 403	N = 404	N = 381	trend	Sd/Nor ⁺					
PTH pg/ml	7.87 ± 0.51 *	4.78 ± 0.13	4.25 ± 0.10	3.73 ± 0.10	< 0.01	< 0.01					
IgA mg/ml	2.32 ± 0.10	2.12 ± 0.05	2.22 ± 0.05	2.12 ± 0.05	0.12	0.036					
IgE kU/l	145.38 ± 50.1	75.45 ± 10.08	85.01 ± 13.17	70.37 ± 7.89	0.28	> 0.05					
IgM mg/ml	0.88 ± 0.05	0.94 ± 0.04	0.96 ± 0.03	0.98 ± 0.03	0.09	> 0.05					
IgG mg/ml	10.63 ± 0.25	10.05 ± 0.11	10.28 ± 0.10	10.28 ± 0.12	0.24	> 0.05					
IgG1 mg/ml	6.74 ± 0.21	6.16 ± 0.08	6.17 ± 0.07	6.41 ± 0.11	0.14	0.033					
IgG2 mg/ml	3.13 ± 0.15	3.23 ± 0.07	3.45 ± 0.07	3.24 ± 0.06	0.01	>0.05					
IgG3 mg/ml	0.38 ± 0.02	0.37 ± 0.01	0.38 ± 0.01	0.41 ± 0.03	0.37	>0.05					
IgG4 mg/ml	0.66 ± 0.06	0.60 ± 0.03	0.66 ± 0.03	0.59 ± 0.02	0.12	>0.05					
C4 mg/ml	0.22 ± 0.01	0.23 ± 0.00	0.23 ± 0.00	0.23 ± 0.00	0.02	0.01					
C3 mg/ml	1.11 ± 0.02	1.09 ± 0.01	1.7 ± 0.007	1.07 ± 0.01	0.03	0.01					

^{*} Mean <u>+</u> 1 SD

⁺Sd/Nor: between the groups of Severely deficient 25 and Normal serum concentrations of 25(OH)D

Figure legends

Figure 1 - 25(OH)D levels in healthy elderly Swiss citizens

Figure 1 A The regression line illustrates reduction of these levels in 1291 men and women (p<0.01).

Figure 1 B The same data as in A were sorted into 3 age groups and are here represented in box plots. They further illustrate the decrease of 25(OH)D levels by decrements of median and interquartile ranges in relation to aging.

Figure 2 - Box and Whiskers plot of the relationship between immunoglobulins levels and 25(OH)D.

Figure 2A: IgA levels, Figure 2B: IgE levels, Figure 2C: IgM levels and Figure 2D: IgG levels, compared among the different deficiency groups of 25(OH)D in the healthy elderly Swiss population. P values are indicated only when statistically significant.

Figure 3 - 25(OH)D levels and their relationship to C4 and C3

Comparison of concentrations of complement components C4 and C3 in 4 different 25(OH)D serum level groups and analysis of statistical significance. A & C: set of data points with

linear regression line. B and D: box-and-whisker plot illustrating trend and statistical significance with variability outside the upper and lower quartiles.

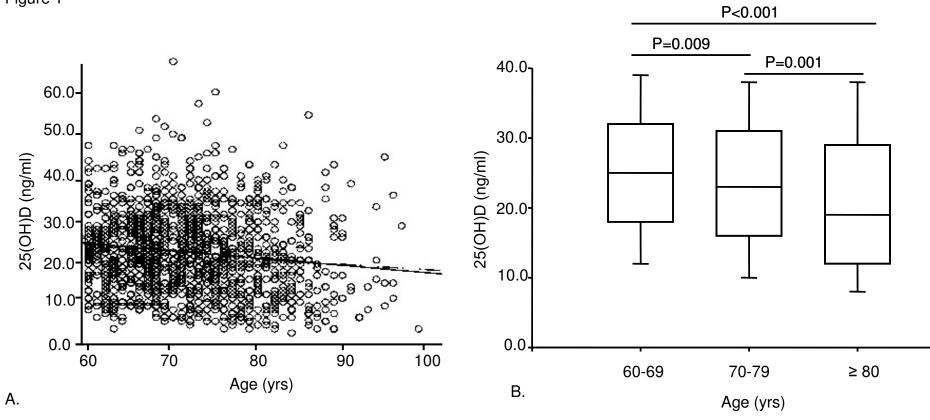
Additional file 1

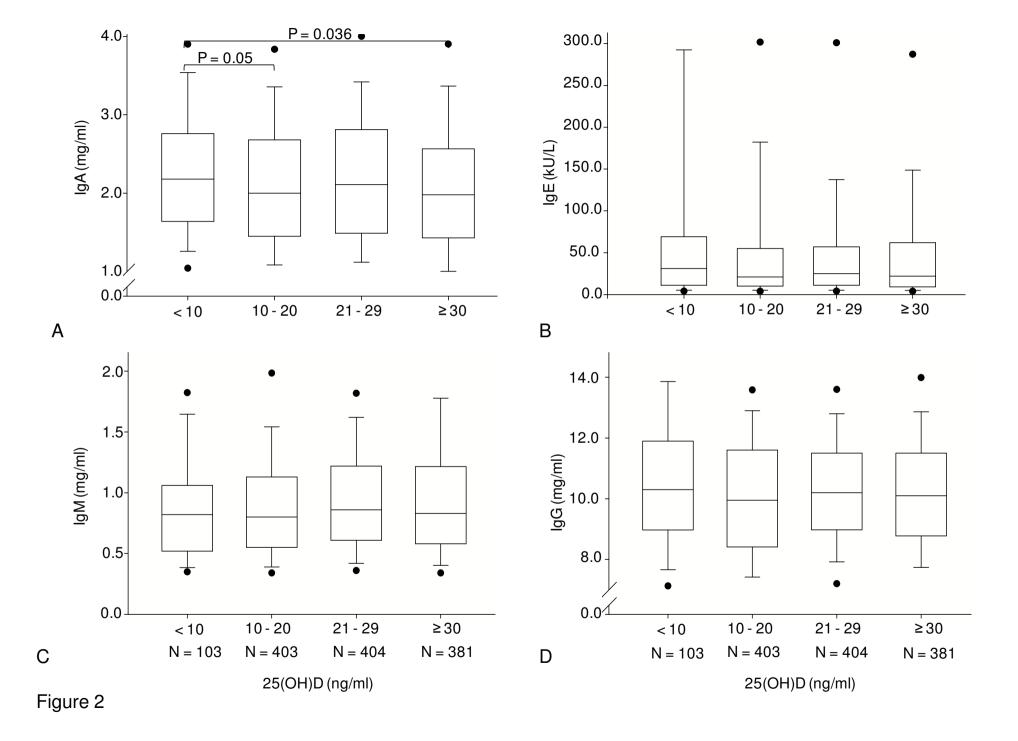
Monthly follow-up of mean \pm 1 SD serum levels of 25(OH)D and humoral immune components. The IgE increment in summer likely reflects pollen exposure of the subjects. Number of samples collected per month are listed in brackets on the abscissa.

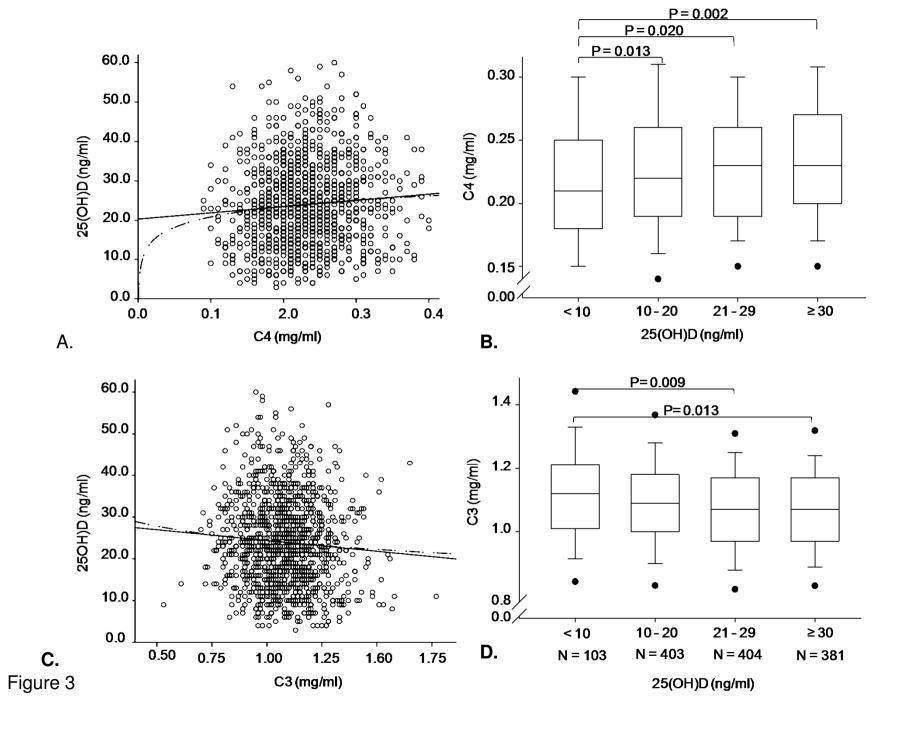
Additional file 2

The percentage increase or decrease in seasonal variation of the humoral immune components showing significant correlation in seasonal variation with 25(OH)D, compared to the month of January (100%) is further illustrated here.

Figure 1







Additional files provided with this submission:

Additional file 1: follow_up monthly_12.pptx, 224K http://www.biomedcentral.com/imedia/5363009541026450/supp1.pptx
Additional file 2: From100PercentOnwards.pptx, 51K http://www.biomedcentral.com/imedia/4969013410264510/supp2.pptx