

Bioinformatics III

Sixth Assignment

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Exercise 5.1: Boolean Networks

- (a) Listing 1 shows the source code of our propagation matrix class, which behaves like such a matrix. Internally it uses a adjacency matrix to efficiently calculate next states. Therefore it depends on the class AdjacencyMatrix shown in Listing 2. Further, the networks states are encoded in the class State, see 3.

Listing 1: Listing of source code

```
0 from os.path import exists
  from AdjacencyMatrix import AdjacencyMatrix
  from State import State

  class PropagationMatrix:
5      """
      Implements a propagation matrix. The propagated states are not
      preprocessed and are computed on demand to reduce the required
      amount of memory. Thereby increased runtime-changes are negligible
      """
10     def __init__(self, filename):
        """
        Create a class object behaving like a propagation matrix
        for a gene network obtained by textfile
        Textfile specification:
15     A > B indicates that gene A turns gene B activate
        A | B indicates that gene A turns gene B inactive
        (space separated)
        """

        # temporary storage for gene labels (to avoid redundancy)
20     nodes = set()
        # temporary storage for linkages (to avoid double file reading)
        links = list()
        if exists(filename):
            with open(filename) as openfile:
25                 for line in openfile:
                    content = line[0:(len(line)-1)].split(" ")
                    nodes.add(content[0])
                    nodes.add(content[2])
                    if (content[1] == ">"):
30                        links.append((content[0], content[2], 1))
                    elif (content[1] == "|"):
                        links.append((content[0], content[2], -3))

        # sort genes in alphabetiv order by
35     # by making use of list functionalities
        nodes = list(nodes)
        nodes.sort()

        # initialize a adjacency matrix representing the obtained
40     # gene network
```

```
        self.matrix = AdjacencyMatrix(0, nodes)
        for triple in links:
            self.matrix.setByLable(triple[0], triple[1], triple[2])
    else:
        print(filename, "does_not_exist")
45
def propagate(self, state):
    """
    Derive the propagated state form a given state
    Each gene is set inactive if the scalar product
    50 of the given state and its column in the adjacency matrix
    is smaller or equal to zero. Otherwise, the gene is set active
    """
    prop_state = State(state.getLables())
    for lab_y in state.getLables():
        # scalar product
        temp = 0
        for lab_x in state.getLables():
            temp += state.getByLable(lab_x) * self.matrix.getByLable(lab_x, lab_y)
        60 # threshold task specific
        prop_state.setByLable(lab_y, 0 if temp <= 0 else 1)
    return prop_state

def size(self):
    65 """
    Returns the number of states
    """
    return self.matrix.size()

def basinsAndAttractor(self, state):
    70 """
    Track the states obtained by simulating the regulatory
    network until the states rePEAT. Return the results divided in
    orbit states and pure basins
    """
    75 order = list()
    order.append(state.getInt())
    temp = 0
    # simulate network until repetition starts
    80 while True:
        state = self.propagate(state)
        temp = state.getInt()
        if not (temp in order):
            order.append(temp)
        85 else:
            break
    # divide states in upper described distinct subsets
    result = [], []
    switch = 0
    90 for i in range(0, len(order)):
        if order[i] == temp:
            result[0] = order[0:i]
            result[1] = order[i:len(order)]
    return result
95

def simplify(self, basatt):
    """
    Simplify redundand and incomplete basins of attractions and
    orbits
    """
    100 # Since every state can have only one next propagated state,
    # a minimum state-value is unique for every orbit and can be
    # used as key for our simplification
    attractors = dict()
    105 basins = dict()
    for ba in basatt:
        temp = min(ba[1])
```

```

    if not (temp in attractors.keys()):
        attractors[temp] = ba[1]
        basins[temp] = ba[0]
110     else:
        basins[temp].extend(ba[0])
        basins[temp].extend(ba[1])
    simple = list()
115     for i in attractors.keys():
        simple.append([set(basins[i]), attractors[i]])
    return simple

def orbit(self):
120     """
    Find basins of attraction and according orbits
    """
    # Storage for for attractors and basins
    periodes = list()
125     # simulate the network starting from every possible state
    candidates = list(range(0, 2**self.matrix.size()))
    while len(candidates) > 0:
        state = State(self.matrix.getLables())
        state.setInt(candidates[0])
130         # simulate regulatory network
        temp = self.basinsAndAttractor(state)
        # store obtained results in the according categorie,
        # further remove observed states from the candidate list
        # to minimize runtime
135         for basin in temp[0]:
            if basin in candidates:
                candidates.remove(basin)
        for attractor in temp[1]:
            if attractor in candidates:
140                 candidates.remove(attractor)
        periodes.append(temp)
    # results might contain reduncdancies, therefore a last
    # simplification step is possible
    return self.simplify(periodes)

```

Listing 2: Listing of source code

```

0 class AdjacencyMatrix:
    """
    Adjancency matrix, encoding a squared matrix with edge weight
    information between nodes.
    Initilialization and data access is only possible over the
5     row and column lables, not by positions
    """

    def __init__(self, initial, lables):
        """
10         Initialization of the adjacency matrix, required are an
        initial default weight and row lables, which are also used as
        column lables -> leading to a squared matrix
        """
        self.lables = lables
15         # default setup
        self.matrix = [0] * len(lables)
        for i in range(0, len(self.matrix)):
            self.matrix[i] = [initial] * len(lables)

20         # Similar to the state class, data access is only possible
        # with row and column lables, dicts enable an acces in linear
        # time
        self.access = dict()
        counter = 0
25         for i in self.lables:
            self.access[i] = counter
            counter += 1

```

```
def setByLable(self, a, b, value):
    """
    Set the weight for the edge from a to b
    """
    self.matrix[self.access[a]][self.access[b]] = value

def getByLable(self, a, b):
    """
    Get the weight for the edge from a to b
    """
    return self.matrix[self.access[a]][self.access[b]]

def size(self):
    """
    Returns the size of the matrix
    expressed by the number of lables
    """
    return len(self.lables)

def getLables(self):
    return self.lables

def show(self):
    """
    Output the matrix as collection of lists
    """
    print("Square_Matrix:")
    for i in range(0, len(self.lables)):
        print(self.matrix[i])
```

Listing 3: Listing of source code

```
class State:
    """
    Encode if genes are active (1) or inactive (0) or inactive
    Covers a simple list and extends it with functionality required
    by the PropagationMatrix and SquareMatrix class
    Data acces can only be done by using the gene lables.
    """

    def __init__(self, lables):
        """
        Initialize a state with the set of all gene lables
        """
        self.lables = lables
        # dict allows to access the data by lable in linear time
        self.access = dict()
        counter = 0
        for l in lables:
            self.access[l] = counter
            counter += 1
        # encondes if genes are active or inactive
        self.state = [0] * len(lables)

    def setByLable(self, lable, value):
        """
        Set the gene with a given lable avtive or inactive
        Every input will be translated to active (1) or inactive (0)
        """
        self.state[self.access[lable]] = 0 if value <= 0 else 1

    def getByLable(self, lable):
        """
        Returns 1 if the gene with a certain lable is active,
        0 otherwise
        """
        return self.state[self.access[lable]]
```

```
35     def getLables(self):
        """
        Returns the lables of genes
        """
40     return self.lables

    def size(self):
        """
        Returns the lenght of the state,
        i.g. the number of encoded genes
45     """
        return len(self.lables)

    def getInt(self):
        """
50     Returns the unique integer obtained by the binary encoded
        genes (active or inactive)
        """
        value = 0
55     for n in range(0, len(self.lables)):
        value += self.state[n] * (2**n)
        return value

    def setInt(self, value):
60     """
        Initializes a state whose binary representation equals the
        provided integer value
        """
        if value == 0:
65         return
        binaries = 2**(len(self.lables)-1)
        pos = len(self.lables) - 1
        while pos >= 0:
            if value >= binaries:
70                 self.state[pos] = 1
                value -= binaries
                binaries /= 2
                pos -= 1
        return

75     def show(self):
        """
        Output the state as binary list
        """
80     print("State:")
        print(self.state)
```

- (b) Listing 4 shows source code applying the the functionality of the code shown in Listing 1, which includes the network simulation.

1) It makes sense to stop the propagation when a state is observed a second time, from then on we will only observe orbiting behavior of the network states. The results of 2) are shown in this way. e.g. the first repeting state is the last shown.

2) Programs output for the required initial states:

Initial state 1:

1 -> 3 -> 7 -> 23 -> 55 -> 63 -> 13 -> 1

Initial state 4:

4 -> 18 -> 36 -> 26 -> 4

Initial state 21:

21 -> 51 -> 47 -> 13 -> 1 -> 3 -> 7 -> 23 -> 55 -> 63 -> 13

Initial state 33:

33 -> 11 -> 5 -> 19 -> 39 -> 31 -> 5

Listing 4: Listing of source code

```
0 from PropagationMatrix import PropagationMatrix
  from State import State

  def trackPropagation(state, repeats):
      """
      5   Visualize the propagations by a sequence of integers
          """
          track = str(state.getInt())
          for i in range(1, repeats):
              state = prop.propagate(state)
              track += " -> "
              track += str(state.getInt())
          print(track)

      # Initialize propagation network with text file
      15 # File contains structural informations of the given
          # gene regulatory network
          prop = PropagationMatrix("net.txt")

          # initialize with the state integer 13
      20 state_a = State(['A', 'B', 'C', 'D', 'E', 'F'])
          print("-----Exercise 6.1 b)-----")
          print("-----")
          print("\nInitial state 1:")
          state_a.setInt(1)
      25 trackPropagation(state_a, 8)

          # initialize with the state integer 13
          print("\nInitial state 4:")
          state_b = State(['A', 'B', 'C', 'D', 'E', 'F'])
      30 state_b.setInt(4)
          trackPropagation(state_b, 5)

          # initialize with the state integer 13
          print("\nInitial state 21:")
      35 state_c = State(['A', 'B', 'C', 'D', 'E', 'F'])
          state_c.setInt(21)
          trackPropagation(state_c, 11)

          # initialize with the state integer 13
```

```

40 print("\nInitial_state_33:")
    state_d = State(['A', 'B', 'C', 'D', 'E', 'F'])
    state_d.setInt(33)
    trackPropagation(state_d, 7)

45 print()
    print(".....Exercise_6.1_c).....")
    print(".....")
    orbits = prop.orbit()
    for i in range(0, len(orbits)):
50     print()
        length = len(orbits[i][1])
        print("Orbit_" + str(i + 1) + "_with_length_" + str(length) + ":")
        print(orbits[i][1])
        print("Set_of_basins:")
55     print(orbits[i][0])
        coverage = float(len(orbits[i][0]))
        coverage /= float(2**prop.size())
        coverage *= 100.0
        print("Relative_coverage:" + str(coverage) + "%")

```

(c) Output of the program listing the orbits:

Orbit 1 with length 1:

0

Set of basins:

0, 6, 8, 12, 16, 20, 22, 24, 28, 32, 34, 40, 42, 44, 46, 48, 50, 52, 54, 56, 58,
 60, 62

Relative coverage: 35.9375%

Orbit 2 with length 7:

1, 3, 7, 23, 55, 63, 13

Set of basins:

1, 3, 7, 9, 13, 21, 23, 25, 29, 41, 43, 45, 47, 49, 51, 53, 55, 57, 59, 61, 63

Relative coverage: 32.8125%

Orbit 3 with length 4:

4, 18, 36, 26

Set of basins:

2, 4, 36, 38, 10, 14, 18, 26, 30

Relative coverage: 14.0625%

Orbit 4 with length 4:

5, 19, 39, 31

Set of basins:

33, 35, 5, 37, 39, 11, 15, 17, 19, 27, 31

Relative coverage: 17.1875%

(d) Interpretation:

1) ???

2) Two special are gene A and D. If A is active, the network becomes permanent impulses; keeping the network in motion. Further, D hinders the network to remain fully active, i.g. if gene D is activated, it throws the network back into an earlier state, initiating the orbiting behavior. This is further ensured since D inactivates its own activating gene.

Table 1 shows the two shorter orbits including the genes activation status at each state of the network. The upper described principles, are demonstrated for Orbit 4, where A activates the network step by step along the regulatory linkages until D gets activated. Afterwards, D inactivates most genes to reset the network to an earlier state.

For Orbit 3, the driving mechanisms are different. Here, B and C activate each other so

Table 1: Comparison of the two orbits with length 4

Orbit 3							Orbit 4						
Decimal	A	B	C	D	E	F	Decimal	A	B	C	D	E	F
4			X				5	X		X			
18		X			X		19	X	X			X	
36			X			X	39	X	X	X			X
26		X		X	X		31	X	X	X	X	X	

that only one of both is active at the same time. When C is active, it further activates E what initiates D to inhibit B, E and F after 3 propagations. Since this inhibition occurs when C is active, the orbit closes and starts again. If D would have even distance (number of activating forward linkages), D would inhibit the active B what would turn the complete network inactive. In this case we would not observe orbiting behavior.

Exercise 5.2: Differential Expression Analysis

The task specific code is shown in Listing 5

Listing 5: Listing of source code

```

0 library(samr)
  library(preprocessCore)

play <- function(rna_sample){
  # store number of up and down regulated genes
5  # per fdr - nperm combination
  low_count = matrix(0, ncol = 10, nrow = 10)
  up_count = matrix(0, ncol = 10, nrow = 10)
  for(fdr in seq(from = 0.1, to = 1.0, by = 0.1)){
    for(nperms in seq(from = 10, to = 100, by = 10)){
10      sink("/dev/null")
      sam <- SAM(x = rna1,
                  y = c(1,1,1,2,2,2),
                  resp.type="Two-class-unpaired",
                  nperms=nperms,
15      logged2 = TRUE,
                  fdr.output = fdr)

      sink()
      low_count[fdr*10,nperms*0.1] = nrow(sam$siggenes.table$genes.lo)
      up_count[fdr*10,nperms*0.1] = nrow(sam$siggenes.table$genes.up)
20    }
  }
  print(low_count)
  print(up_count)
}

25 # Read in data table
DFrame <- read.table(file = "ms_data.txt", sep = "\t", header = TRUE)
DTable <- as.matrix(as.data.frame(lapply(DFrame[,1:9], as.numeric)))
rownames(DTable) <- DFrame[,11]

30 # Log2 transform the data (apply log2 on every cell)
DTable <- log2(DTable)

# Apply quantil normalization of library preprocessCore
35 # split the two siRNA-induced samples
rna1 <- normalize.quantiles(DTable[,1:6], FALSE)
rna2 <- normalize.quantiles(DTable[,c(1:3,7:9)], FALSE)

# Apply SAM function of samr library on sample 1
40 sink("/dev/null")
sam <- SAM(x = rna1,

```



```

    y = c(1,1,1,2,2,2),
    resp.type="Two-class-unpaired",
    nperms=100,
45     genenames = rownames(DTable),
        logged2 = TRUE,
        fdr.output = 0.05)

sink()
cat("10-up-regulated genes for sample 1:\n")
50 print(sam$siggenes.table$genes.up[1:10,c(1,2,6)])
cat("10-down-regulated genes for sample 1:\n")
print(sam$siggenes.table$genes.lo[1:10,c(1,2,6)])

# Apply SAM function of samr library on sample 2
55 sink("/dev/null")
sam <- SAM(x = rna2,
    y = c(1,1,1,2,2,2),
    resp.type="Two-class-unpaired",
    nperms=100,
60     genenames = rownames(DTable),
        logged2 = TRUE,
        fdr.output = 0.05)

sink()
cat("10-up-regulated genes for sample 2:\n")
65 print(sam$siggenes.table$genes.up[1:10,c(1,2,6)])
cat("10-down-regulated genes for sample 2:\n")
print(sam$siggenes.table$genes.lo[1:10,c(1,2,6)])

#play(rna1)
70 #play(rna2)
```

We played with fdr parameter and found, that an increasing value for fdr increases the number of significant genes. Reasonable since a higher fdr builds a less harsh threshold. This leads to more genes identified as significant. More permutations on the other hand do not affect the number of genes identified as significant. The numbers are fluctuating within a small range. But an increasing number of permutations refines the method so that the found results are more precise. So that the false positive and negative rate reduces.

10 up-regulated genes for sample 1:

Gene Name	Fold Change
Polypyrimidine tract-binding protein 2	21.679
Cellular retinoic acid-binding protein 2	6.432
Creatine kinase U-type, mitochondrial	14.359
Disabled homolog 1	6.345
Calcium-binding and coiled-coil domain-containing protein 2	4.017
Multidrug resistance protein 1	5.337
Neurobeachin	4.166
Protein Tob2;Protein Tob1	3.483
Lanosterol 14-alpha demethylase	2.898
Interferon-related developmental regulator 1	2.357

10 down-regulated genes for sample 1:

Gene Name	Fold Change
Cathepsin B	0.091
Integrin beta-5	0.092
Integrin alpha-V	0.097
Fibrillin-1	0.103
Polypeptide N-acetylgalactosaminyltransferase 1	0.083
Glia-derived nexin	0.25
Solute carrier family 2, facilitated glucose transporter member 1	0.121
DnaJ homolog subfamily C member 3	0.173
Erlin-1	0.229
UPF0501 protein KIAA1430	0.164

10 up-regulated genes for sample 2:

Gene Name	Fold Change
Disabled homolog 2	3.036
Heme oxygenase 1	3.765
Interferon-related developmental regulator 1	3.005
EPM2A-interacting protein 1	8.654
Disabled homolog 1	3.21
Cellular retinoic acid-binding protein 2	2.605
Asparagine synthetase [glutamine-hydrolyzing]	2.355
EKC/KEOPS complex subunit LAGE3	3.012
Dehydrogenase/reductase SDR family member 7	2.278
Exonuclease 3-5 domain-containing protein 2	1.928

10 down-regulated genes for sample 2:

Gene Name	Fold Change
Receptor-type tyrosine-protein phosphatase eta	0.303
Alpha-galactosidase A	0.245
Protein disulfide-isomerase A5	0.149
Integrin alpha-V;Integrin alpha-V heavy chain;Integrin alpha-V light chain	0.144
EGF-like repeat and discoidin I-like domain-containing protein 3	0.209
Spectrin beta chain, non-erythrocytic 1	0.261
DnaJ homolog subfamily C member 10	0.187
Spectrin alpha chain, non-erythrocytic 1	0.368
Cathepsin B;Cathepsin B light chain;Cathepsin B heavy chain	0.282
Endoplasmic reticulum-Golgi intermediate compartment protein 1	0.243