

# Estimation of sex ratio heritability in the common snapping turtle

Kyle Hilliard

Master's Defense Seminar

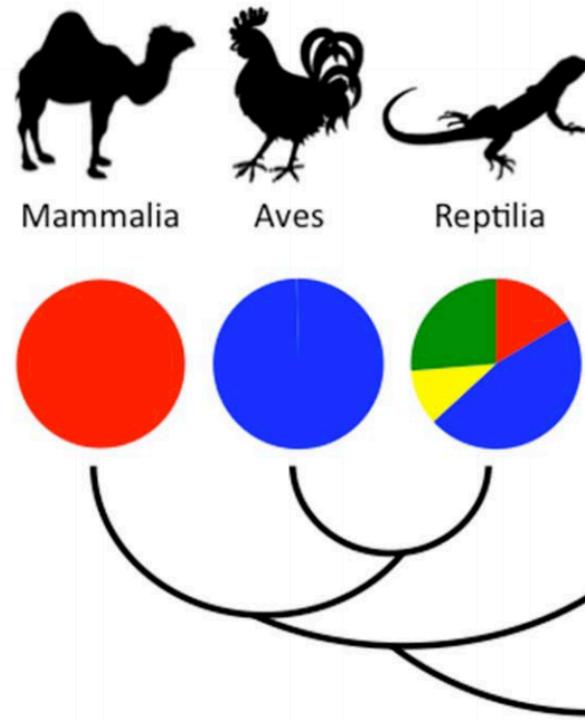
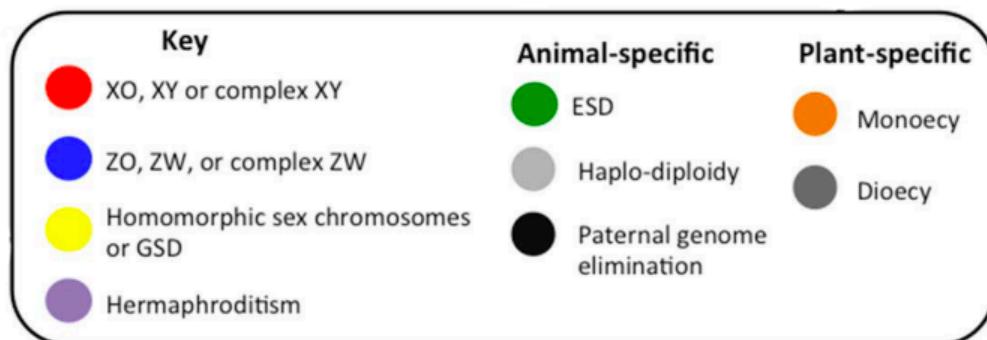
November 26, 2019

Abbott Hall 101, 9 a.m.



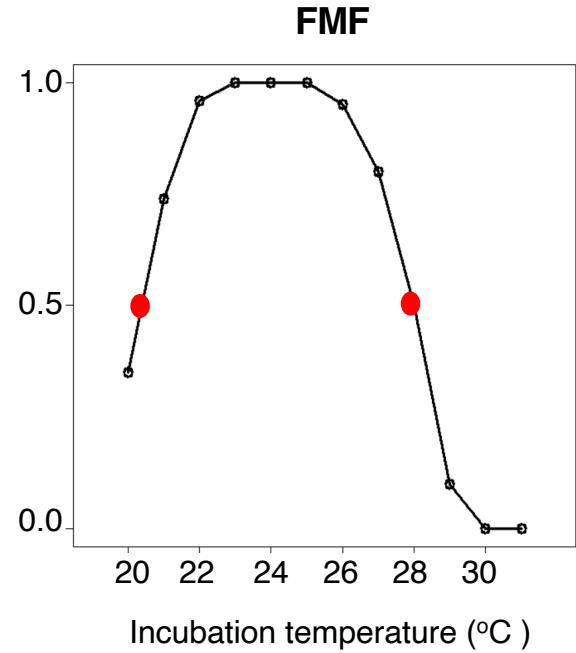
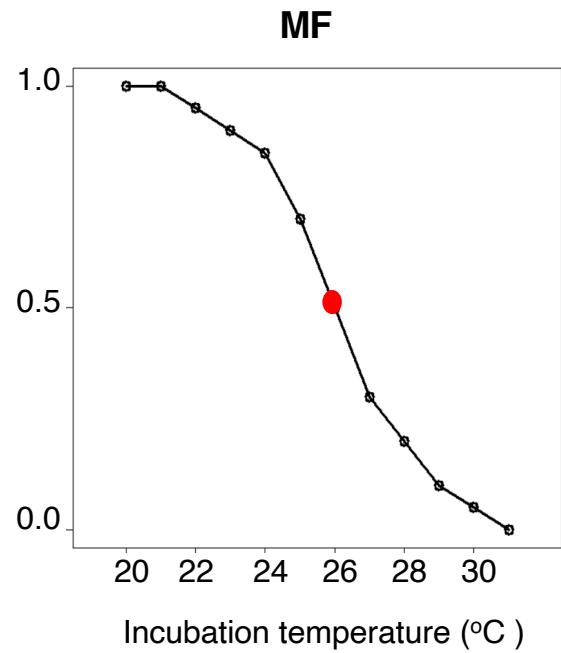
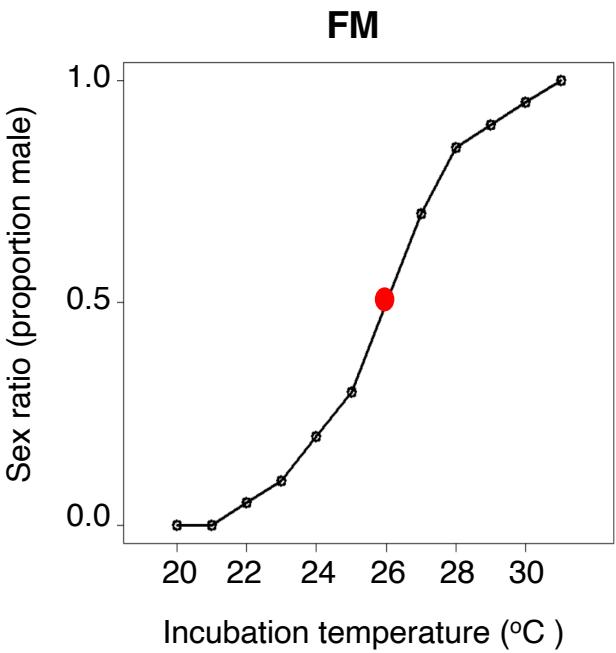
# Sex determination

- Genotypic sex determination (GSD)
- Environmental sex determination (ESD)
  - Temperature-dependent sex determination (TSD)

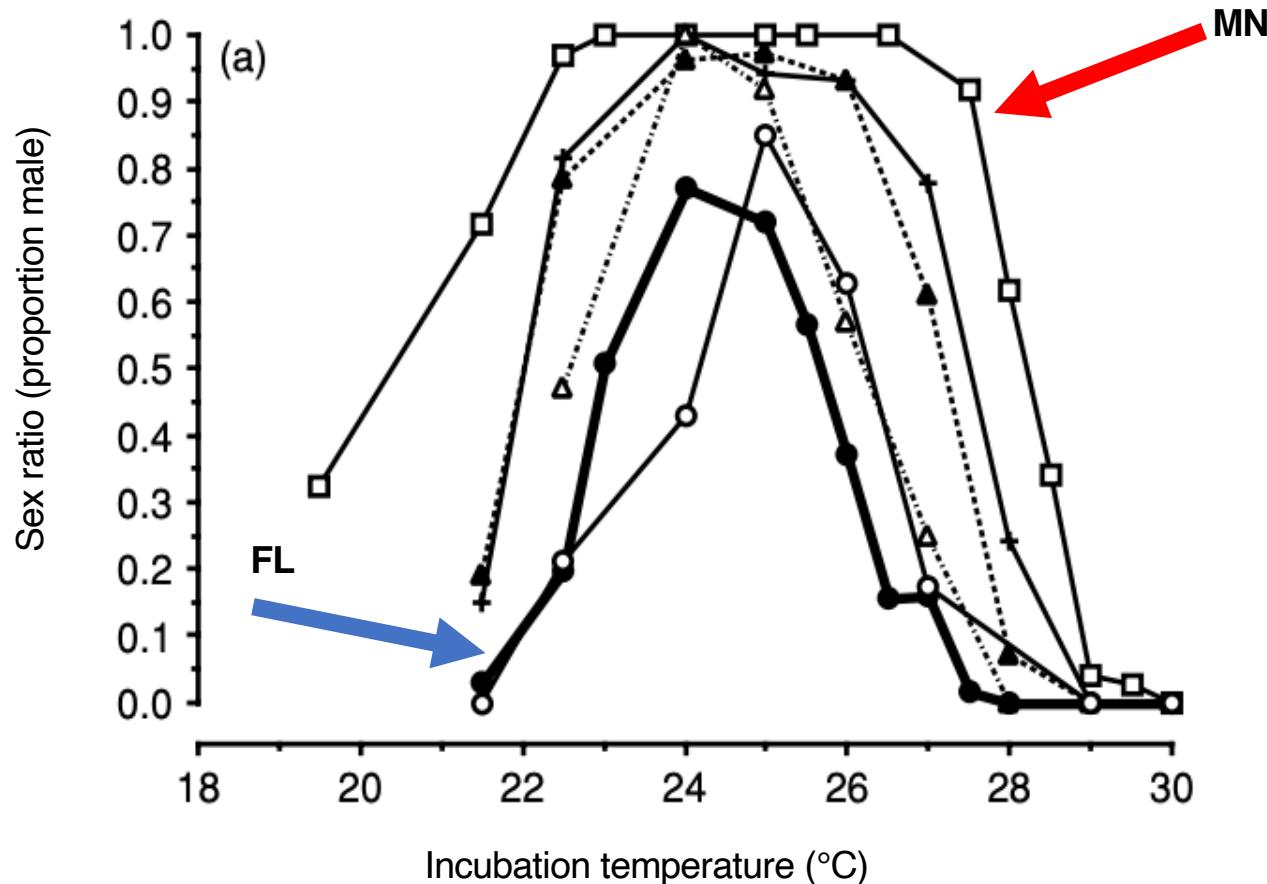


Bachtrog D., Mank J.E., Peichel C.L., Kirkpatrick M., Otto S.P. et al. (2014). Sex determination: why so many ways of doing it? *PLoS Biology* 12: e1001899.

# Variation in TSD among species

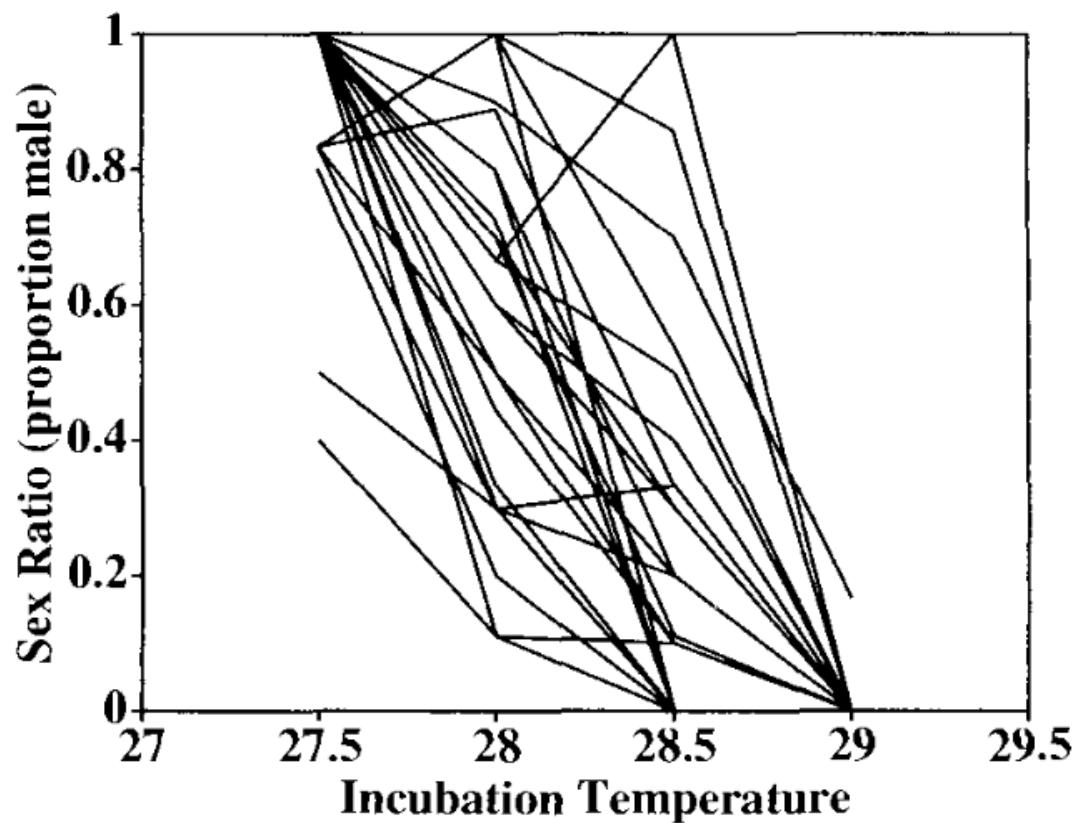


# Variation in TSD among snapping turtle populations



Ewert, M.A., Lang, J.W. and Nelson, C.E. (2005). Geographic variation in the pattern of temperature-dependent sex determination in the American snapping turtle (*Chelydra serpentina*). *Journal of Zoology* 265: 81-95.

# Variation in TSD within snapping turtle populations



Rhen, T. and Lang, J.W. (1998). Among-family variation for environmental sex determination in reptiles. *Evolution* 52: 1514-1520.

# What causes TSD variation in the snapping turtle?

$$V_P = \underline{V_A + V_D + V_I} + V_M + V_E + V_R$$

## 1). Genetic effects

- Embryo genotype influences temperature sensitivity
- $V_G = V_A + V_D + V_I$ 
  - Additive genetic effects ( $V_A$ )
    - $h^2 = V_A/V_P$

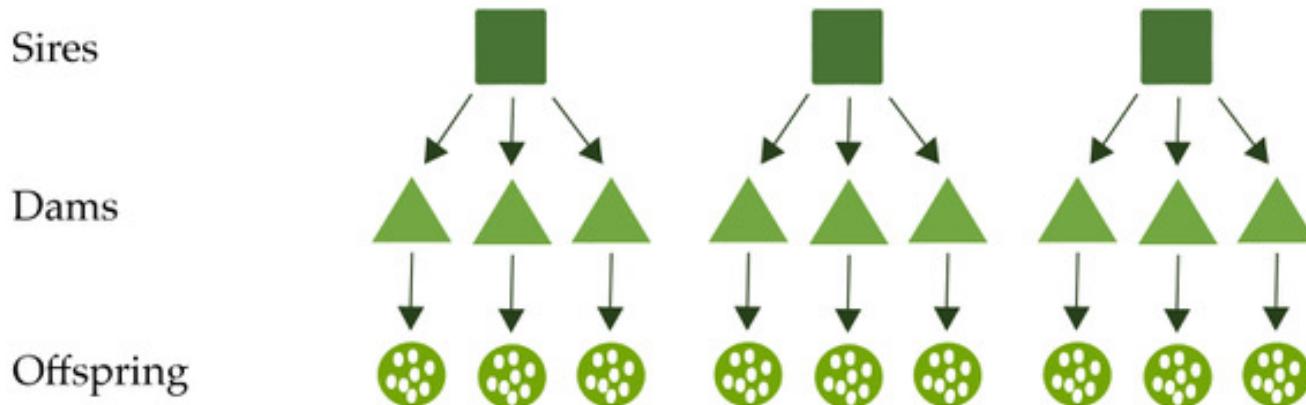
## 2). Non-genetic maternal effects

- Maternally-derived yolk steroids influence temperature sensitivity

# Breeding design



## Full-sib/half-sib breeding design



# Quantitative genetic analysis

- Additive genetic effects TABLE 14.1 vs. non-genetic maternal effects

Interpretation of the observational components of variance  
in a sib analysis

- Egg mass

*Observational component*

- Egg diameter

- Hatching mass

Sires:

$$\sigma_s^2 = cov_{(HS)}$$

Dams:

$$\sigma_d^2 = cov_{(FS)} - cov_{(HS)}$$

- Hatching carapace length

Progenies:

$$\sigma_w^2 = V_p - cov_{(FS)}$$

- Hatching plastron length

Progenies:

$$V_p$$

- Hatching sex

Sires + Dams:

$$\sigma_s^2 + \sigma_d^2 = cov_{(FS)}$$

*Covariance and causal components*

*estimated*

$$= \frac{1}{4} V_A$$

$$= \frac{1}{4} V_A + \frac{1}{4} V_D + V_{Ec}$$

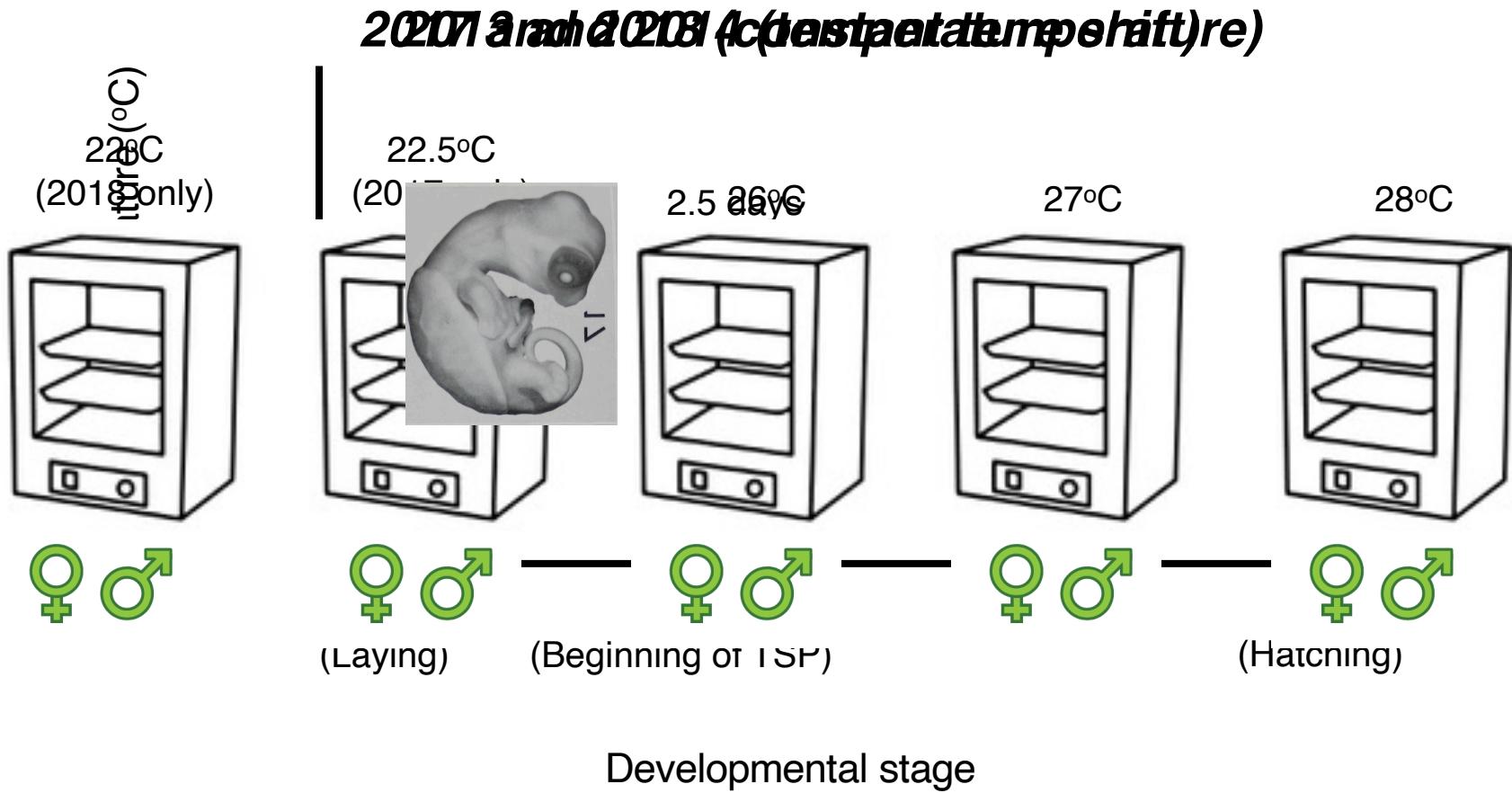
$$= \frac{1}{2} V_A + \frac{3}{4} V_D + V_{Ew}$$

$$= V_A + V_D + V_{Ec} + V_{Ew}$$

$$= \frac{1}{2} V_A + \frac{1}{4} V_D + V_{Ec}$$

Falconer, D.S. and Mackay, T.F.C. (1996). *Introduction to Quantitative Genetics*. 4<sup>th</sup> edition Longman House: Essex, England.

# Egg collection and incubation



# Measurement of phenotypic traits and tissue collection

- In 2013, 2014, 2017 and 2018 we sexed hatchling turtles and collected tissue for genotyping
- In 2017 and 2018, we also collected egg and hatchling size data

## Sample sizes

2013: 1062  
2014: 1035  
2017: 1358  
2018: 1422

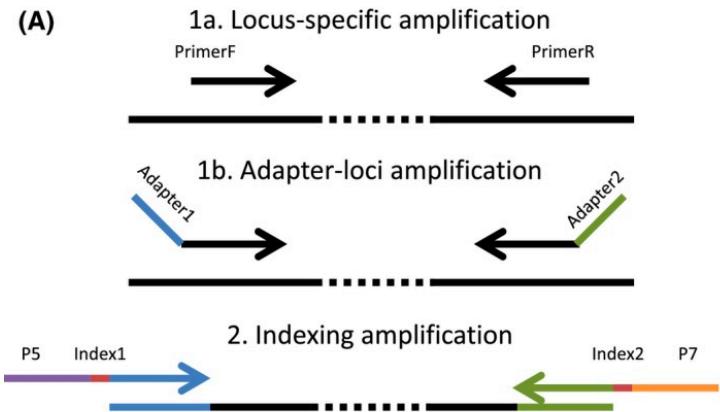
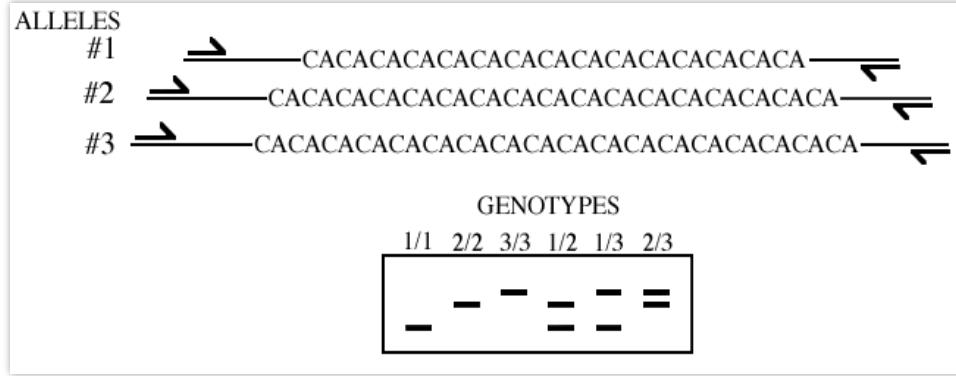
---

Total: 4877

# DNA isolation, quantification, normalization

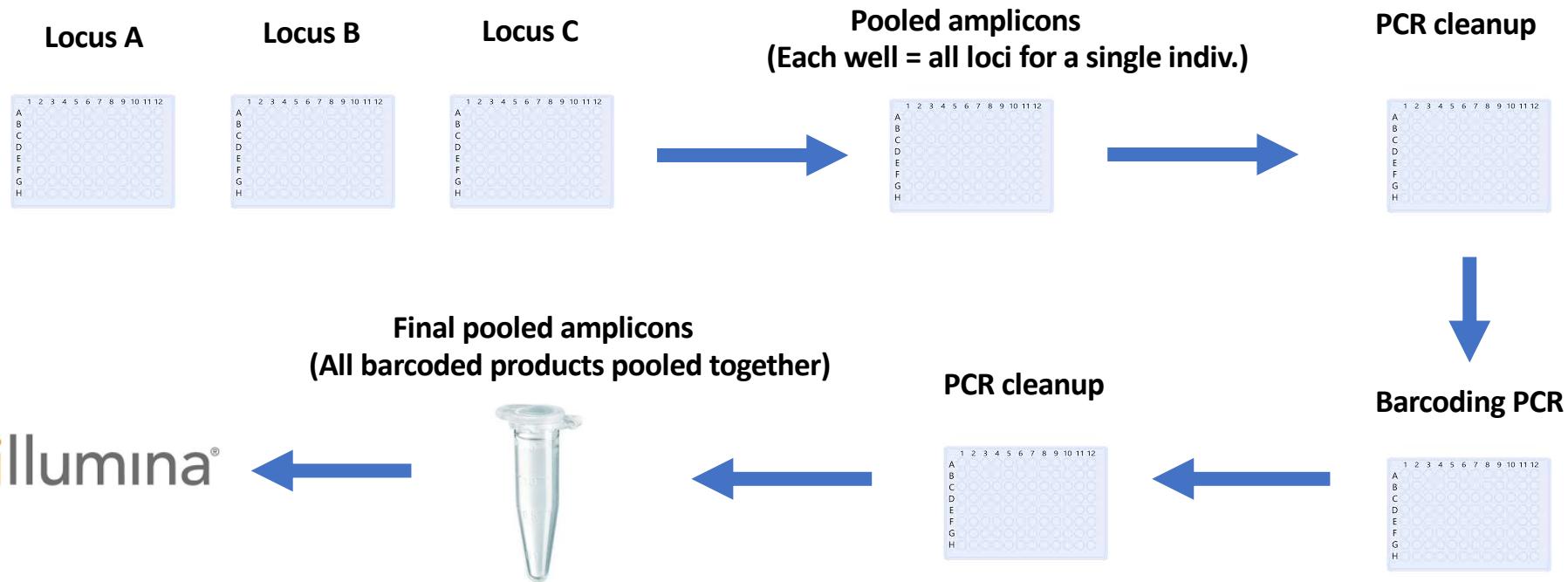


# Amplicon sequencing



Darby B.J., Erickson S.F., Hervey S.D., Ellis-Felege S.N. (2016). Digital fragment analysis of short tandem repeats by high-throughput amplicon sequencing. *Ecology and Evolution* 6: 4502–4512.

# Amplicon sequencing



# Bioinformatics and data processing

```
>CStetra11.2;size=992;
GCCAGAATTAAAGCTGTCTATGCAGCCTGAATT CAGAGGATGGATAGATAGATAGATAGATAGATAGATAGA  

TAGATAGATAGATAGATAGATAGATAGATAGATGCATTATGATACTCTTCACAGTATTATATGGAAAATCTTCAGCAGAGCCT  

GGAAAAGAACATGGAACAGAAAAGCCAGTTCTTCTATGTCCAAGTTCAAGTTCTAAACAATAAGCCCCCTTTCTTCTT  

GCAATCTTCTGCTTATTGGTGCAACCTGTGTACTGAA
>CStetra12_davy.3;size=837;
GGCATTCTAGCCAACAGGAAAACAGCAGCTCAGGTAGCTCAGATAACCATGGTATGGCATGGATAGATAGATAGAT  

AGATAGATAGATAGATAGAGATCTGTTAACCTACATGCATATATTTTAGAAGTCATTAACCCTCATATACAGG  

AAATAAAACACTGAGAAAAGCACACCGCT
>CStetra12_davy.4;size=598;
GGCATTCTAGCCAACAGGAAAACAGCAGCTCAGGTAGCTCAGATAACCATGGTATGGCATGGATAGATAGATAGAT  

AGATAGATAGATAGATAGATAGATAGAGATCTGTTAACCCACATGCATATATTTAGAAGTCATTAACCCTCATATACAGG  

ACCCTCATATACAGGAAATAAACACTGAGAAAAGCACACCGCT
>CStetra9.5;size=373;
GGCTCATTAGCATAGAACTCAGAACCAAAATGCAATGAAATGCAAAACTCATTGATCTTCATCAAATAGATAAATAA  

AACCCCTCAGAGAGAAGGGGTTTCATGATGATCATGGTCTCAAAATGATTAGATAGATAGATAGATAGATAGATA  

GATAGATAGATAGATAGATAGATAGAGTCAAATTATTGTCTATATTGAAGGGTGCCTGGCACTCTAATTACAAG  

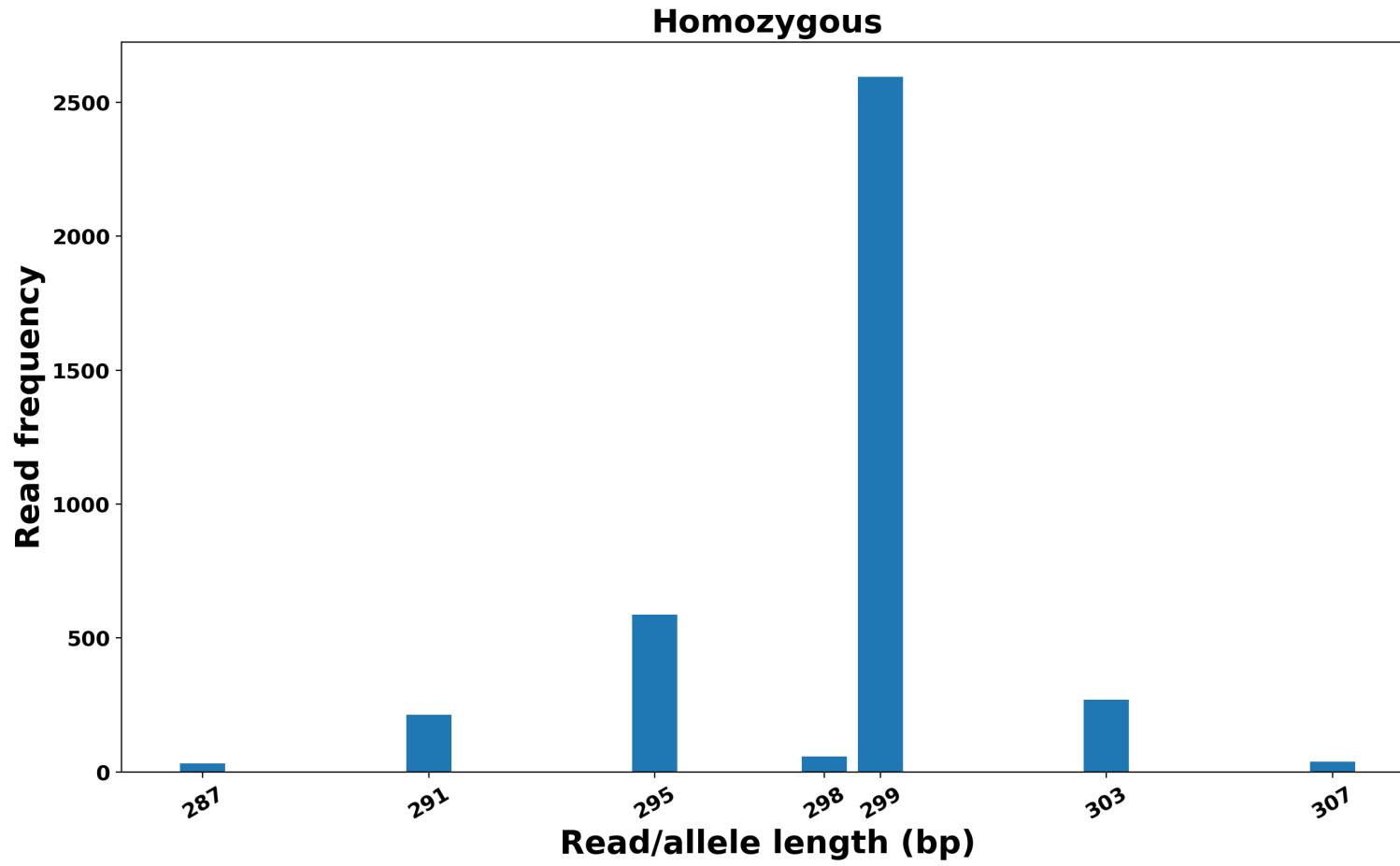
ACCTTATTCAGTTGTTATAACTGCCAACTGTAGC
>CStetra16_Davy.6;size=350;
TCCAATGCAGCTCTTCACTGTACAGTAATAATAAATAAAAAAAAGCTCACCAATTATTGCGATCTCTCGCTT  

GATTCTGCACGAGAAGATTAGTGTAAAGATCTGCAGGCCAGTTGCTCCACCAGCAAAGACTAGCCCATAATTGTC  

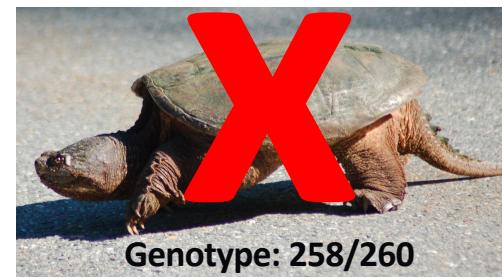
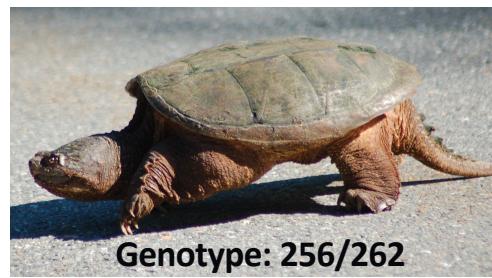
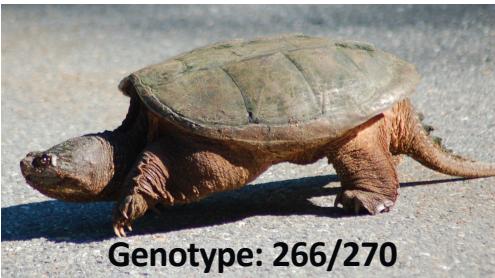
GAGATGGCAAGG
CCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCC  

GGGGGGGGGGGGGGGGGGGGGGGGGGGGFDGGGGFFGFFFFF
```

# Microsatellite genotyping



# Parentage analysis



# Quantitative genetic analysis

## Traits

- Egg mass
- Egg diameter
- Hatchling mass
- Hatchling carapace length
- Hatchling plastron length
- Hatchling sex

## Model parameters

- **Incubation temperature** → factor, fixed effect
- **Year** → factor, random effect
- **Sire** → factor, random effect
- **Dam** → (nested within sire) → factor, random effect
- **Sire x temperature** → random effect
- **Dam x temperature** (nested within sire) → random effect



Fixed effects tests = Type III ANOVA, Tukey HSD

Random effects tests = Likelihood ratio test

# Analysis of egg mass

**Table 3.** Likelihood ratio tests of random effects on egg mass in snapping turtles.

<i>Effect</i>	$LR_{\chi^2}$	<i>Df</i>	<i>p</i>
Sire	0	1	1.0
Dam (nested within sire)	1719.06	1	<0.0001
Year	91.31	1	<0.0001

**Table 5.** Estimates of narrow-sense heritability ( $h^2$ ) and maximum maternal effects ( $m^2_{Max}$ ) of egg mass  $\pm$  standard error.

<i>Parameter</i>	<i>Estimate</i>
$h^2$	$0.00 \pm 0.48$
$m^2_{Max}$	$0.59 \pm 0.24$

# Analysis of egg diameter

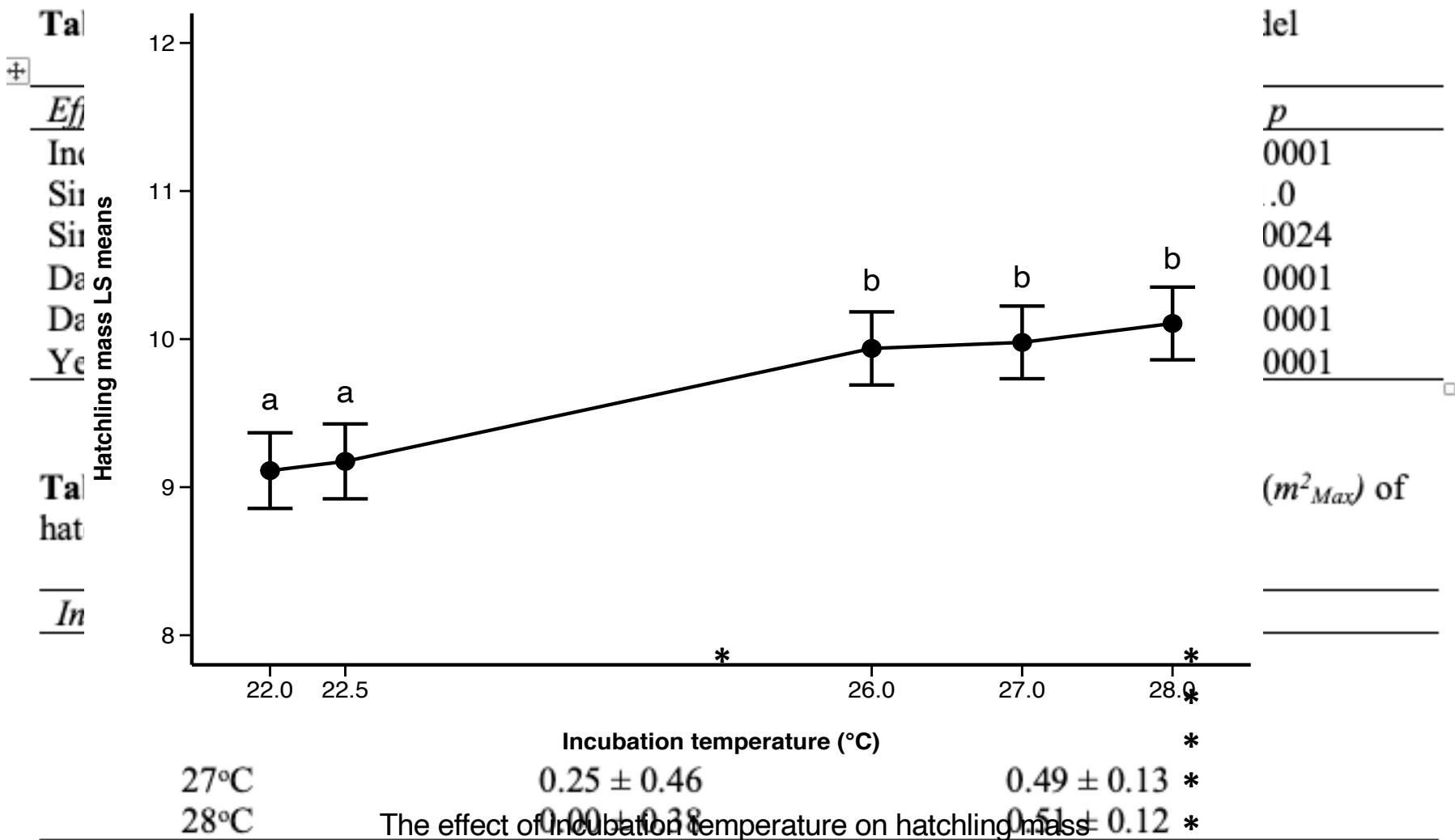
**Table 4.** Likelihood ratio tests of random effects on egg diameter in snapping turtles.

Effect	$LR_{\chi^2}$	Df	p
Sire	0	1	1.0
Dam (nested within sire)	1211.20	1	<0.0001
Year	202.00	1	<0.0001

**Table 6.** Estimates of narrow-sense heritability ( $h^2$ ) and maximum maternal effects ( $m^2_{Max}$ ) of egg diameter  $\pm$  standard error.

Parameter	Estimate
$h^2$	$0.00 \pm 0.38$
$m^2_{Max}$	$0.46 \pm 0.20$

# Analysis of hatchling mass



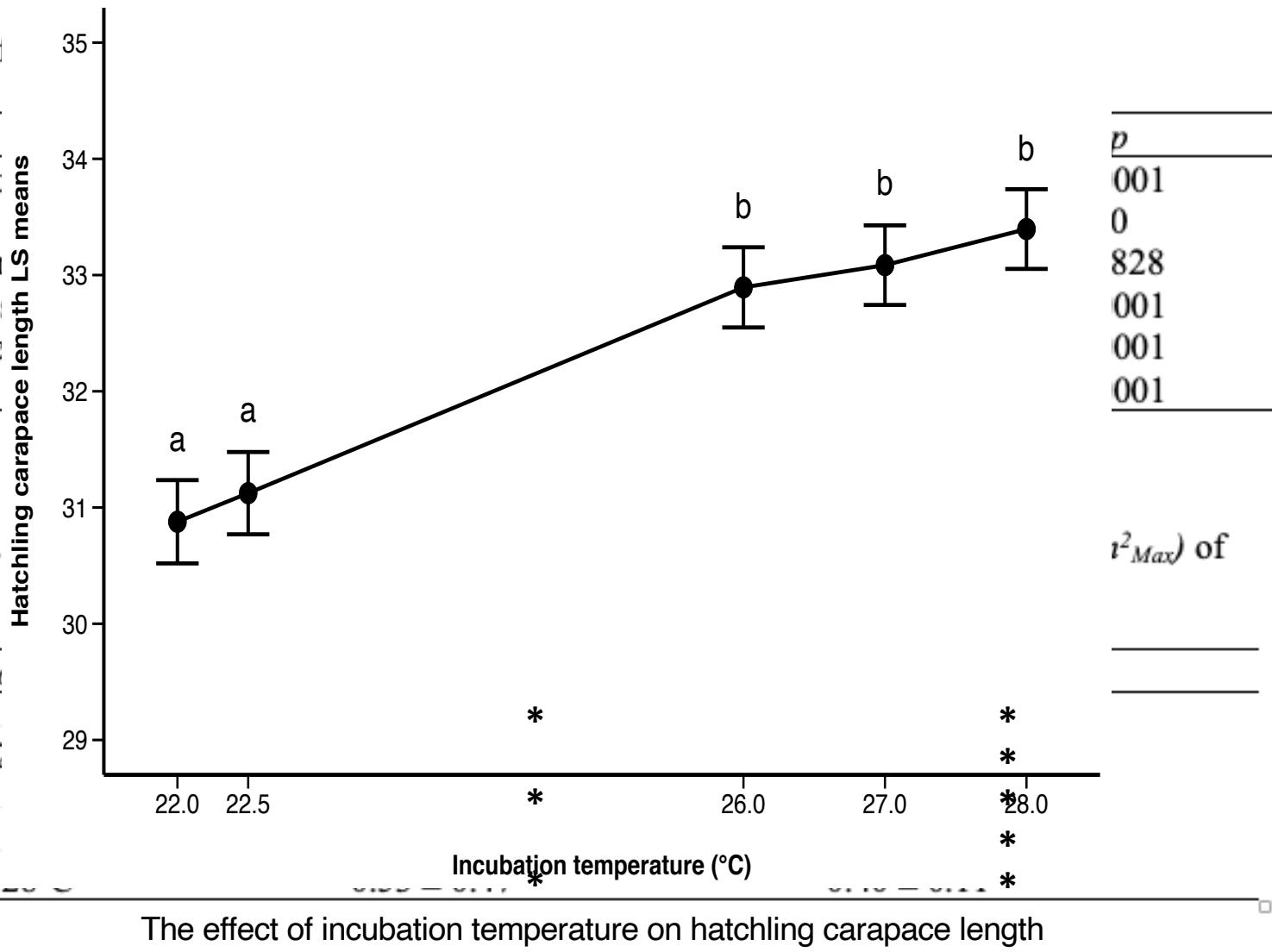
# Analysis of hatchling carapace length

**Table 18.**  
using the i

Effect	p
Incubation	0.001
Sire	0
Sire x te	828
Dam (ne	0.001
Dam x te	0.001
Year	0.001

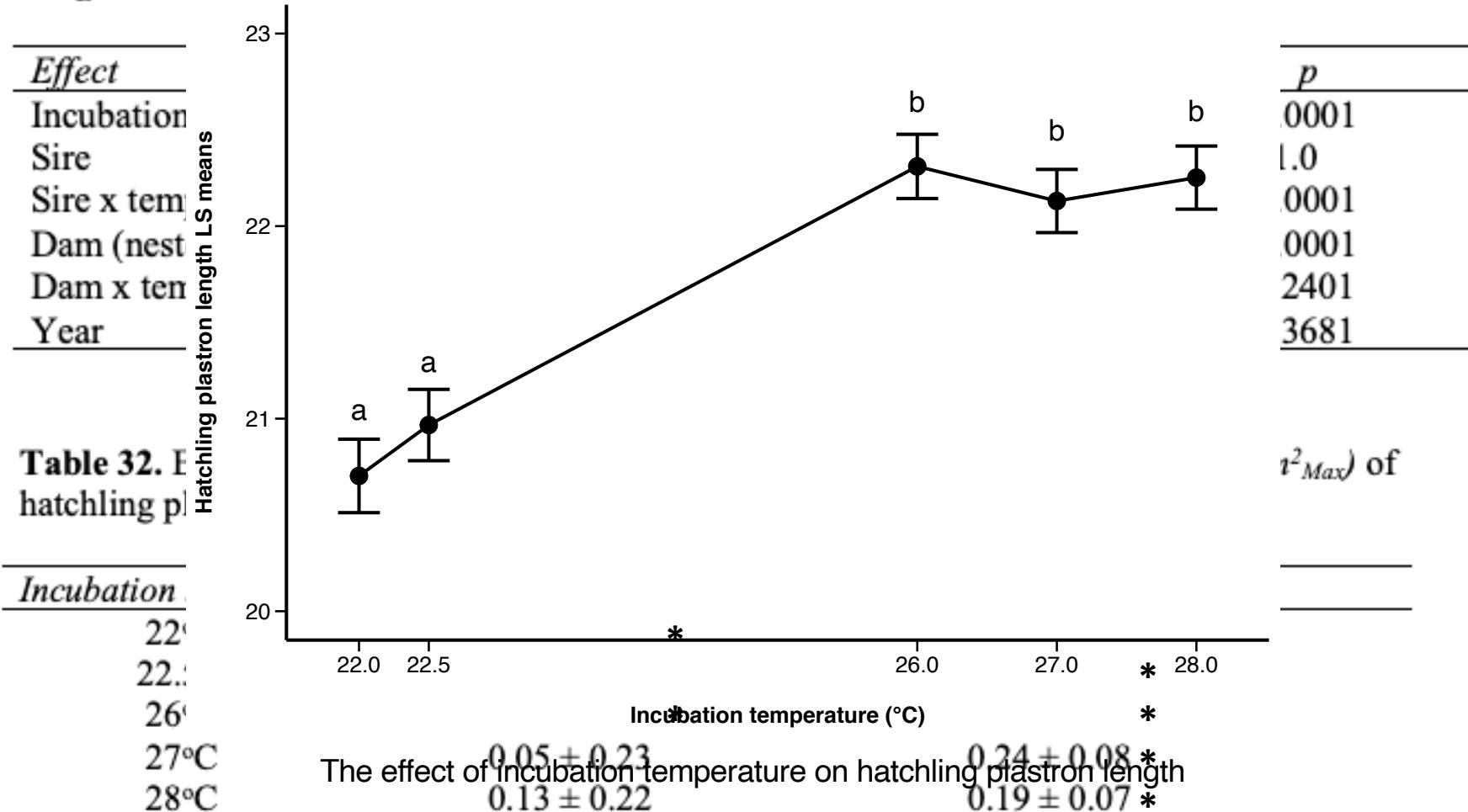
**Table 24.**  
hatching

Incubation	$r^2_{Max}$
22.0	0.001



# Analysis of hatchling plastron length

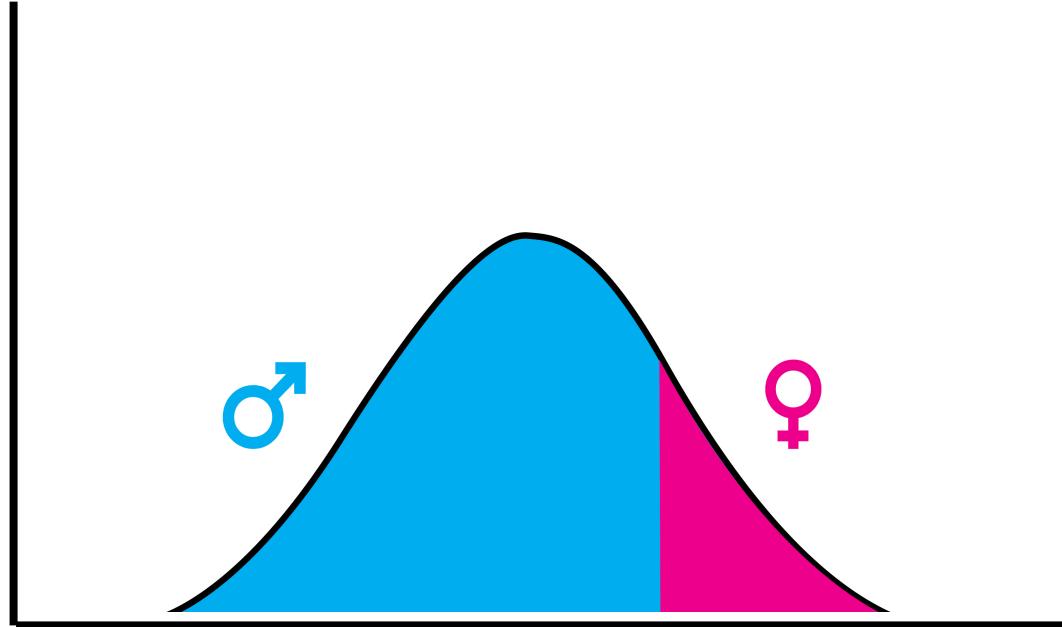
**Table 26.** Analysis of fixed and random effects on hatchling plastron length using the full model



# Analysis of sex ratio

Approaches to analyze sex:

- Continuous model
- Logistic model
- Liability threshold model

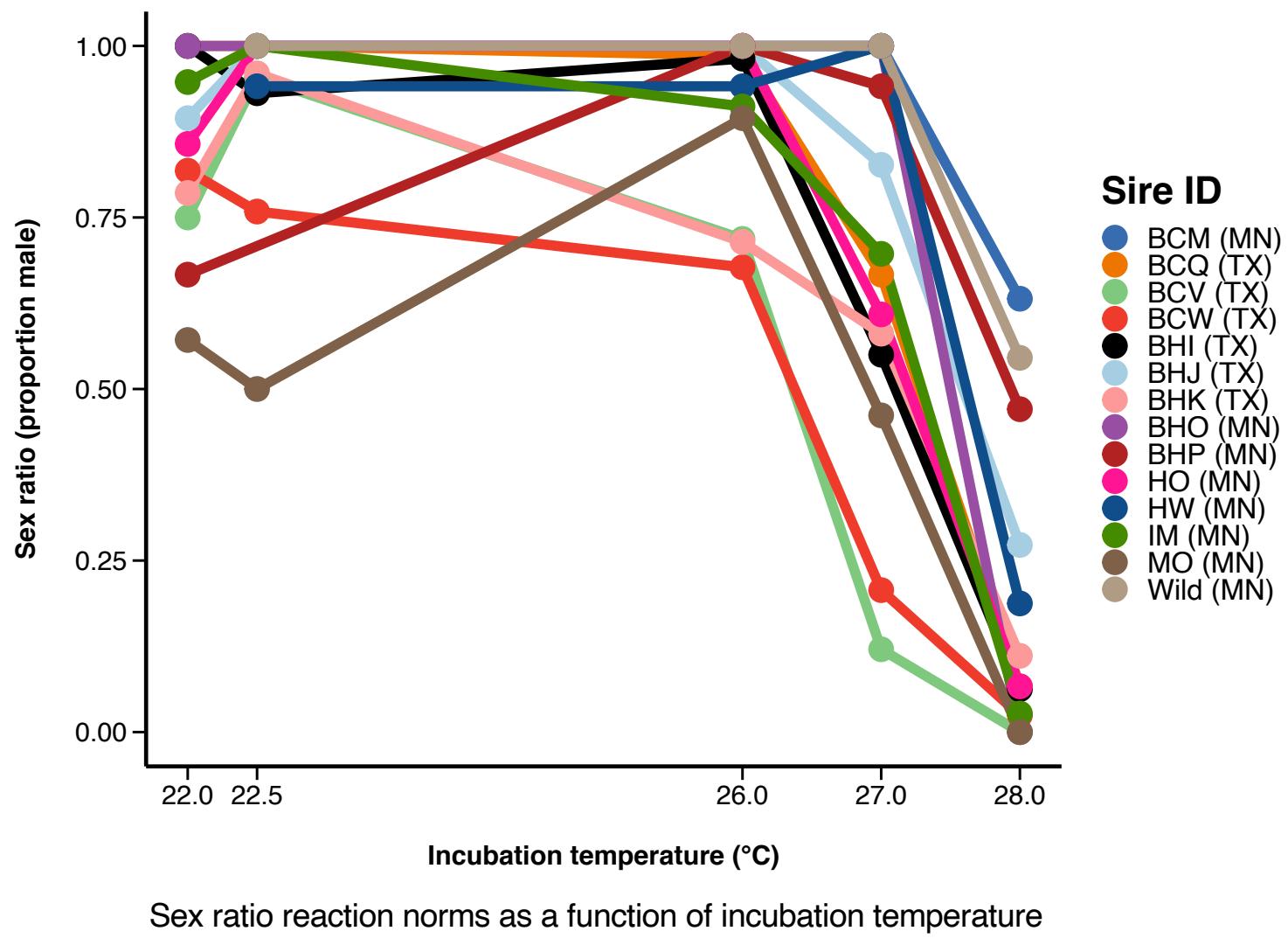


# Analysis of sex ratio

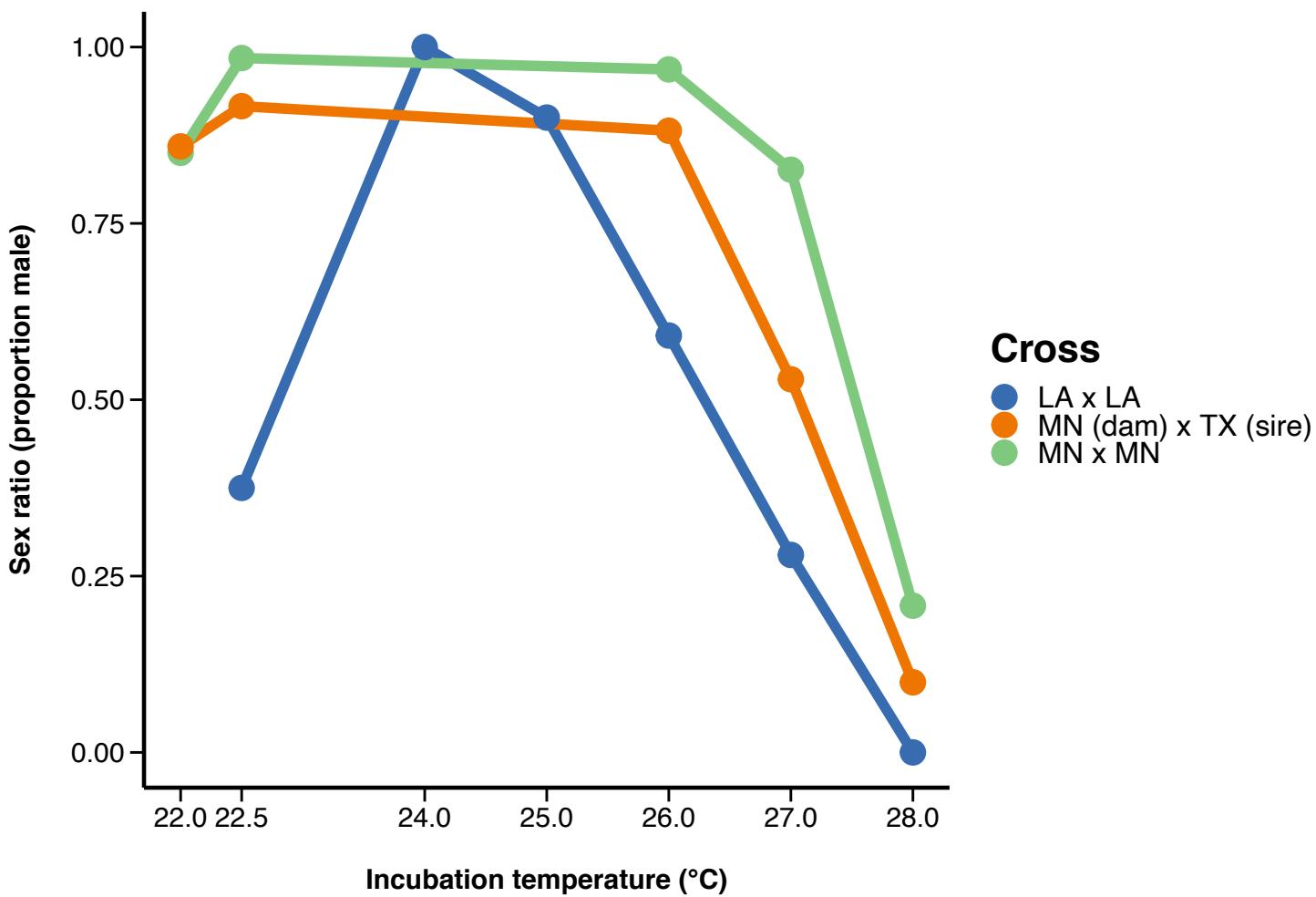
**Table 41.** Estimates of narrow-sense heritability ( $h^2$ ) of hatchling sex at individual incubation temperatures with their 95% confidence intervals. Estimates are shown using the observed scale, and two liability methods as described in the text.

<i>Incubation temperature</i>	<i>Continuous</i>	<i>Liability (Bull)</i>	<i>Liability (F &amp; M)</i>
22°C	0.34, CI = [0.11, 1.00]	0.78, CI = [0.29, 1.00]	0.80, CI = [0.27, 1.00]
22.5°C	0.61, CI = [0.20, 1.00]	1.00, CI = [0.71, 1.00]	1.00, CI = [0.76, 1.00]
26°C	0.29, CI = [0.08, 1.00]	0.80, CI = [0.34, 1.00]	0.86, CI = [0.22, 1.00]
26-31-26°C	0.50, CI = [0.21, 1.00]	1.00, CI = [0.50, 1.00]	1.00, CI = [0.50, 1.00]
27°C	1.00, CI = [0.52, 1.00]	1.00, CI = [0.80, 1.00]	1.00, CI = [0.83, 1.00]
28°C	0.90, CI = [0.36, 1.00]	1.00, CI = [0.86, 1.00]	1.00, CI = [0.92, 1.00]

# Significant sire x temperature interaction

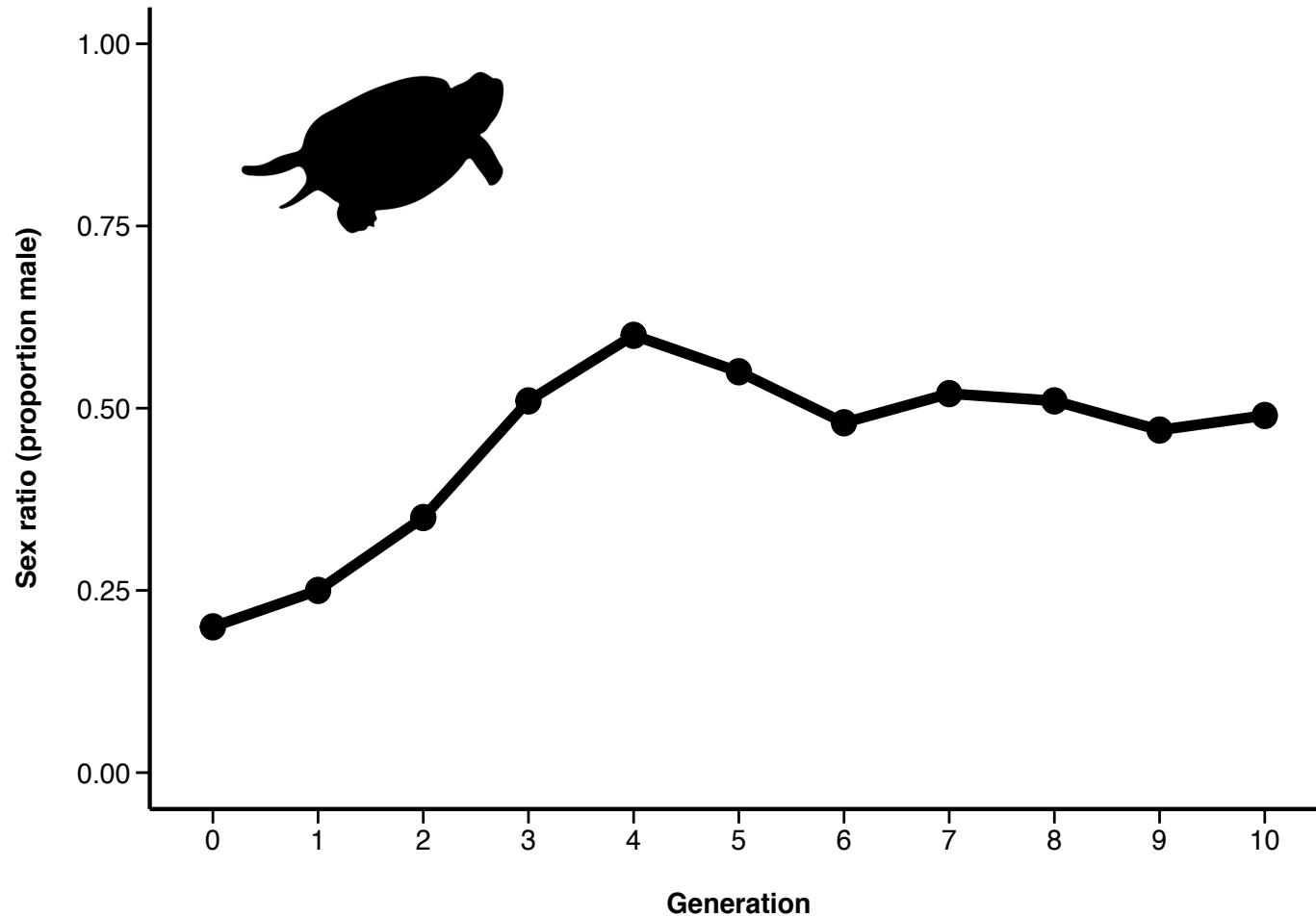


# Hybrid status effect



Sex ratio reaction norms of three snapping turtle population crosses

# Sex ratio evolution through frequency-dependent selection



# Conclusion

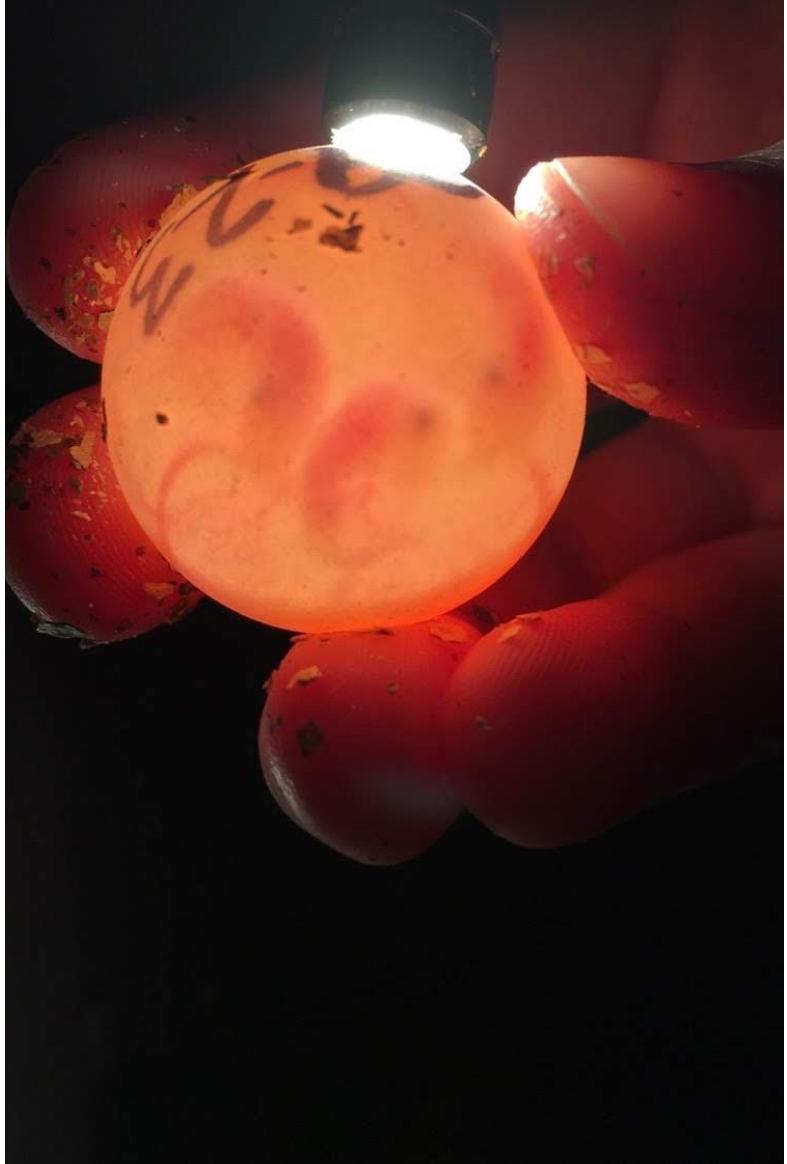
1. Variation in egg size and hatchling size is caused by non-genetic maternal effects.
2. In the same offspring, sexual phenotype is highly heritable. We also detected a significant sire-by-temperature interaction.
3. Genetic variation in sex determination would allow the evolution of TSD pattern in response to climate change.

# Acknowledgments

- Committee
  - Turk Rhen
  - Brian Darby
  - Robert Newman
- Rhen lab
- Sam Hervey
- Matt Flom
- UND Biology Department
- NSF Grants IBN IOS 0923300 and IBN IOS-1558034
- IACUC Protocol 1509-6C



# Questions?



# Sex ratio reaction norms: Texas v.s. Minnesota

