

dgeAnalysis v1.1.0

user manual



Leids Universitair
Medisch Centrum

SASC
Sequencing Analysis Support Core

Authors

Tom Kuipers
Hailiang (Leon) Mei
Davy Cats

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1. Preface

With the development of next-generation sequencing techniques, RNA-Seq is becoming a more commonly applied method in various fields, e.g., cancer genomics, cell biology, etc. RNA-Seq gives great insights into a biological species at a specific moment. To find out which genes (or transcripts) are playing an important function at a specific moment or under a specific condition, differential gene expression analysis is used. With the use of this type of analysis, research between different conditions can be conducted.

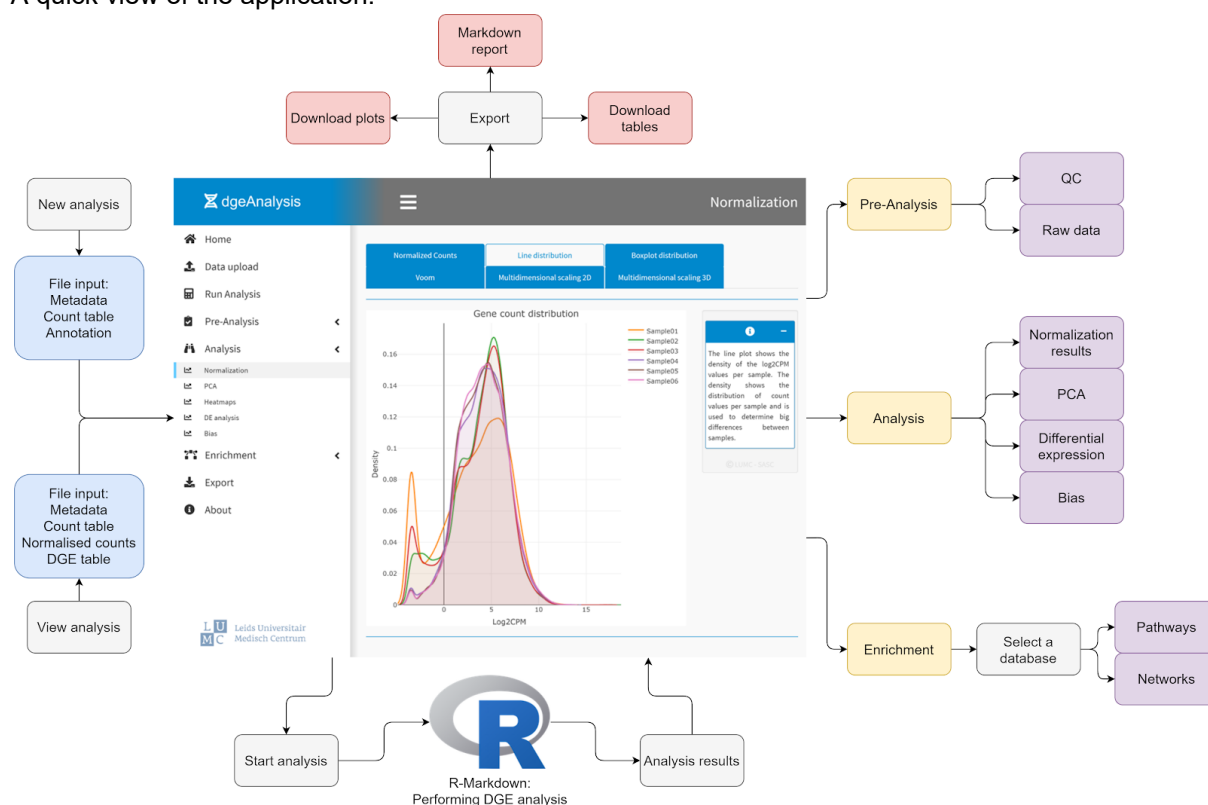
This application is created using primarily R code, together with a variation of R packages including Shiny, edgeR, limma, and DESeq2. The application is built upon a shiny based framework and so can be used in any R environment. Using R-markdown for back-end analysis reproducibility can be ensured. The R-markdown files contain all steps taken to perform an analysis, together with all used input variables and dependencies including version numbers. The front-end images are all created with the use of Plotly to generate interactive plots, with click and hover options.

Using the application for differential gene analysis has proven to be very helpful in many projects. Next to checking data quality, it also provides a platform to carry out a DGE analysis quickly. This makes it an easy to use platform capable of performing a variety of analysis possibilities.

The package is available from GitHub:

<https://github.com/LUMC/dgeAnalysis>

A quick view of the application:



1. Preface	2
2. Start dgeAnalysis	4
2.1. Rstudio	4
2.2. Docker image	5
2.3. Application successfully started	5
2.4. Example data	5
3. Procedures	6
3.1. Perform a new analysis	6
3.1.1. Upload data	6
3.1.2. Check Pre-Analysis	7
3.1.3. Run the analysis	7
3.1.4. Check the analysis results	8
3.1.5. Export results	8
3.1.6. Enrichment	8
3.2. View an analysis	8
4. Install packages manually	9

2. Start dgeAnalysis

2.1. Rstudio

To use the *dgeAnalysis* package has a number of requirements before installing the package in R.

1. R v3.6+ <https://www.r-project.org/>
2. Rstudio v1.0.153+ <https://rstudio.com/>
3. A web browser (pref. Google Chrome or Mozilla Firefox)

If installing on Windows, make sure RTools is installed:

<https://cran.r-project.org/bin/windows/Rtools/Rtools35.exe>

If installing on Linux, make sure these libraries are installed. This can be done by running this in the linux terminal:

```
sudo apt-get update && apt-get install \  
  build-essential \  
  gdebi-core \  
  pandoc \  
  pandoc-citeproc \  
  libcairo2-dev \  
  libjpeg-dev \  
  libxt-dev \  
  libssl-dev \  
  libcurl4-gnutls-dev \  
  libxml2-dev \  
  libcurl4-openssl-dev \  
  -y
```

With R and Rstudio installed the *dgeAnalysis* package can be installed. First, open Rstudio and type the following steps in the **Console**:

1. Install the “devtools” package (if not already installed)

```
install.packages("devtools")
```

2. Install the *dgeAnalysis* package. Sometimes installing the package can cause problems. These are often related to other packages failing to install. Check

```
library("devtools")  
devtools::install_github("LUMC/dgeAnalysis")
```

3. Launch the application

```
library("dgeAnalysis")  
dgeAnalysis::startApp()
```

Running the command `dgeAnalysis::startApp()` first initializes the application:

```
> dgeAnalysis::startApp()  
Initializing dgeAnalysis...  
  
Listening on http://0.0.0.0:1402
```

2.2. Docker image

This method is specifically for Linux or Mac OS users (A browser (pref. Google Chrome or Mozilla Firefox is required)

Make sure Docker is installed: <https://docs.docker.com/get-docker/>

To start the docker image follow these steps:

1. Open a terminal
2. Load docker image

```
docker load < dgeAnalysis.tar.gz
```

3. Run docker image

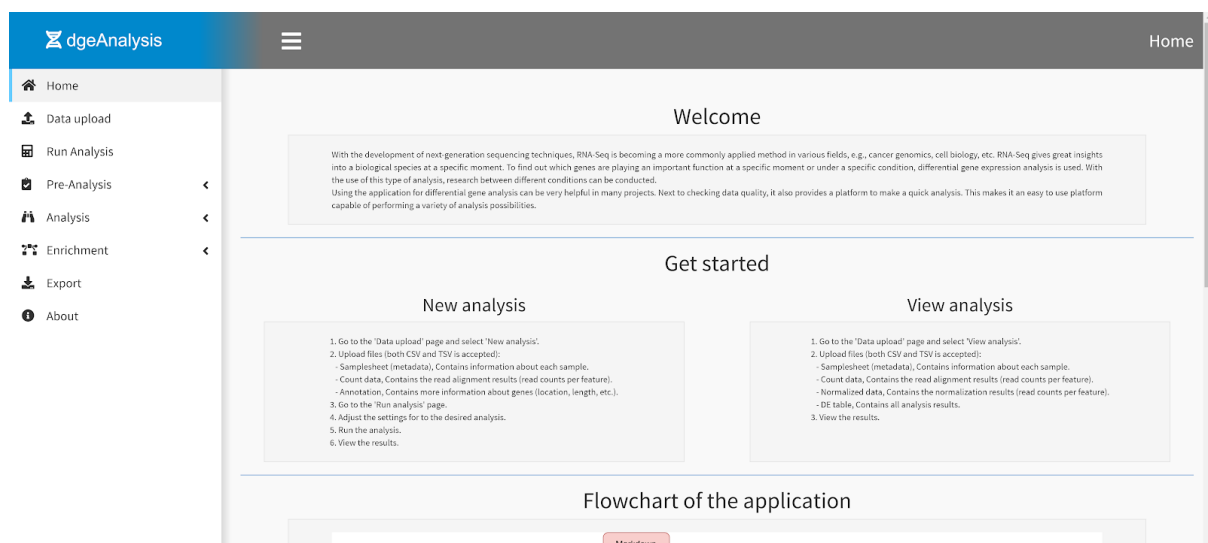
```
docker run -p 1402:1402 dgeAnalysis
```

4. If a browser isn't opened automatically, click or copy the link in the generated output.

```
Listening on http://0.0.0.0:1402
```

2.3. Application successfully started

Once all packages are loaded the application will look like this:



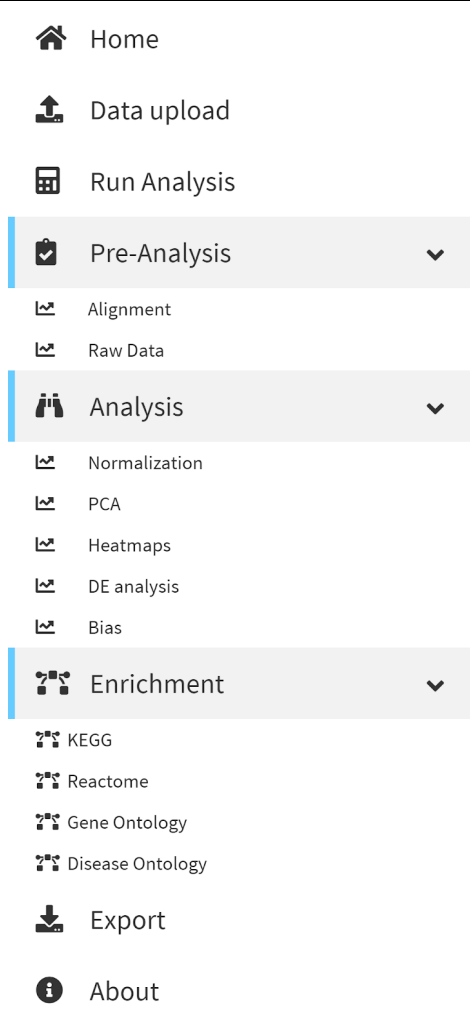
The application is now ready to use!

2.4. Example data

On the GitHub page of *dgeAnalysis* there is a folder called “exampleData”. This folder includes a zip file with a sample sheet, count table, and an annotation file. These files can be viewed to check the data format and perform a test analysis. Note: The test data is of no use, so the results should be treated as such.

3. Procedures

Once the application is loaded and started in a web browser it's ready to use. There are multiple pages in the application, which will be explained here. Navigation through the application is easy using the navigation bar on the left of the screen:

<ul style="list-style-type: none"> • Home The home page contains a short explanation of the application and a quick startup guide. • Data upload Here all raw data can be uploaded and viewed to check that data is loaded correctly. • Run Analysis Change settings and run an analysis with the uploaded data. • Pre-Analysis View distribution of raw data and perform quality checks based on raw data. • Analysis View the distribution of normalized data, PCA, and the differential gene expression analysis results. • Enrichment Perform enrichment analysis on the analyzed data, with the use of multiple databases. • Export Download generated tables or a markdown report containing all used analysis steps that have been running on the background. • About Check application information and information about the current session (like package versions, etc.). 	 <ul style="list-style-type: none"> Home Data upload Run Analysis Pre-Analysis Alignment Raw Data Analysis Normalization PCA Heatmaps DE analysis Bias Enrichment KEGG Reactome Gene Ontology Disease Ontology Export About
---	---

3.1. Perform a new analysis

3.1.1. Upload data

Before starting an analysis load all data in the “Data upload” page. Here there is an option to start a new analysis. To start a new analysis, there are three file upload options:

1. Metadata/sample sheet
 - This contains the sample information.
2. Count table
 - This contains the read alignment results (read counts per feature).
3. Annotation (optional)
 - This contains more information about genes (location, length, etc.).

Where the metadata and count table are mandatory, the annotation is optional. This is because they are not needed to perform the analysis itself. It is recommended to do upload an annotation as well

because this can add more insight into the data. To perform gene set enrichment an annotation is required. To perform the enrichment, EntrezIDs are required, which can be set in the annotation file. All uploaded data is shown in table format. This can be used as a control check to see if all data is loaded correctly.

3.1.2. Check Pre-Analysis

Before starting the actual analysis it's a good idea to first check the distribution of raw data. This can be done in the tabs below "Pre-Analysis". Check how the alignment results to see how many reads actually are aligned and how the reads are distributed across the biggest genes. Next, check how the raw data is distributed, possible outliers can already be detected here.

3.1.3. Run the analysis

Navigate to the "Run analysis" page to start a new analysis.

The screenshot shows the 'Run Analysis' page of the dgeAnalysis web application. The interface includes a sidebar with navigation options: Home, Data upload, Run Analysis (selected), Pre-Analysis, Analysis, Enrichment, Export, and About. The main content area is titled 'Settings' and contains various configuration options for the analysis. Numbered callouts (1-9) highlight specific features: (1) 'Choose analysis method' with buttons for Limma/Voom, EdgeR (selected), and DESeq2; (2) 'Set FDR cutoff (adjusted P-Value)' with a slider; (3) 'Set Log2CPM cutoff' with a slider; (4) 'Select base column for comparison' with a dropdown menu; (5) 'Select values for comparison' with input fields for 'AA' and 'BB'; (6) 'Find genes that respond to' with a text input; (7) 'Exclude samples' with a list of sample IDs (Sample01 to Sample06); (8) 'Use gene ID or gene symbols' with radio buttons for 'Gene ID' and 'Gene Symbol'; and (9) the 'Run Analysis' button at the bottom.

- (1) Choose between Limma/Voom, EdgeR, or DESeq2 as method to run the analysis with.
- (2) Change the p-value cutoff (default="0.05") to define differentially expressed genes.
- (3) Change the Log2CPM cutoff (default="1") to set a CPM filter for low expressed genes.
- (4) Set the base column to start the comparison and select a design type (Basic or advanced).
- (5) Select values to set a comparison in the inputs (based on selected columns).
- (6) The text shows the design matrix (left) and the comparison (right).
- (7) By selecting samples they will be excluded from the analysis.
- (8) Set if genes should be shown in plots/tables with ID or symbol.
- (9) With the button "Run Analysis" the analysis is started.

There are three analysis methods: Limma, EdgeR, and DESeq2. The method selected, will be the method used to perform the differential gene expression analysis. There are also two cutoff values to be used in the analysis: p-value and Log2CPM. The Log2CPM is used during the filtering of low expressed genes. The p-value is used to define the differential expressed genes.

To set a comparison, a column is selected from the sample sheet. Based on this column it's possible to create an analysis with a basic- or advanced design. The design of the analysis categorizes the samples based on the metadata input. The exact values to compare can be chosen based on the selected design columns and determine which samples are compared with each other.

When it's necessary to exclude certain samples from the analysis, then these samples can be selected. All selected samples will be excluded during the analysis, so they don't have any effect on the results. This is very useful in case a sample is a clear outlier.

If an annotation file is uploaded, there is an extra option to show results with the gene IDs (Ensembl IDs) or with gene symbols (gene names). To start the analysis with all uploaded files and settings, there is a button with "Run Analysis" to perform the analysis.

3.1.4. Check the analysis results

After the analysis has finished view all results from top to bottom in the tabs below "Analysis". The normalization tabs are the same as the raw data tabs. Now the data can be compared to see the difference before and after normalization. Check the PCA on possible outliers as well. If these plots show a clear outlier, it might be worth removing them from the analysis. This means rerunning the analysis in "Run Analysis" and select the samples that need to be excluded. The DE analysis tab contains plots that visualize the actual analysis results based on the set comparison.

3.1.5. Export results

Downloading a plot can be done from the plot itself. Hover with your mouse somewhere over a plot and see this taskbar opening in the top-right corner:



Click on the camera icon on the left to download a plot.

To export tables navigate to the page "Export". Here choose a table in the select menu and download the results as CSV or TSV by clicking on one of the two buttons.

!! IMPORTANT !!

Always download the Rmarkdown report after ANY analysis. The Rmarkdown report contains all information about the current analysis, including all used settings, intermediate steps, and important results. This HTML file is important to ensure reproducibility!

3.1.6. Enrichment

There are four databases (KEGG, Reactome, GO, and DO) and two types of analysis available for the enrichment analysis: "Over-represented enrichment" and "Gene set enrichment". Over-represented enrichment is performed with the entrezID's of only the differentially expressed genes. Gene set enrichment is performed with the entrezID's of all analyzed genes and the corresponding Log2FC.

3.2. View an analysis

To view an analysis, these files should be uploaded for the full use of the application:

1. Metadata/sample sheet
 - This contains the sample information.
2. Count table
 - This contains the read alignment results (read counts per feature).
3. Normalized counts
 - This contains the normalization results of the counts per feature.
4. DE table
 - This contains all the analysis results.

The metadata is required, but the other three files are only needed to view specific parts of the application: A count table to view raw data and QC, Normalized counts to view normalization results, and PCA and DE table to view differential expression results.

4. Install packages manually

If there is trouble installing some packages, copy the line of a specific packages into the R/Rstudio console to install the package manually:

```
## Install or import required packages
if (!require("shiny")) install.packages("shiny")
if (!require("shinydashboard")) install.packages("shinydashboard")
if (!require("shinyWidgets")) install.packages("shinyWidgets")
if (!require("shinycssloaders")) install.packages("shinycssloaders")
if (!require("shinyjs")) install.packages("shinyjs")

## Differential expression analysis
if (!require("BiocManager")) install.packages("BiocManager")
if (!require("knitr")) install.packages("knitr")
if (!require("SummarizedExperiment")) BiocManager::install("SummarizedExperiment")
if (!require("edgeR")) BiocManager::install("edgeR")
if (!require("limma")) BiocManager::install("limma")
if (!require("DESeq2")) BiocManager::install("DESeq2")
if (!require("tidyr")) install.packages("tidyr")
if (!require("scales")) install.packages("scales")
if (!require("broom")) install.packages("broom")
if (!require("plotly")) install.packages("plotly")

## Pathway analysis
if (!require("clusterProfiler")) BiocManager::install("clusterProfiler")
if (!require("DOSE")) BiocManager::install("DOSE")
if (!require("graphite")) BiocManager::install("graphite")
if (!require("ReactomePA")) BiocManager::install("ReactomePA")
if (!require("igraph")) install.packages("igraph")
if (!require("org.Hs.eg.db")) BiocManager::install("org.Hs.eg.db")
if (!require("org.Mm.eg.db")) BiocManager::install("org.Mm.eg.db")
if (!require("reshape2")) install.packages("reshape2")
```