# Package 'STAARpipeline'

# October 27, 2022

Type Package  Title STAAR Pipeline for Analyzing Whole-Genome/Whole-Exome Sequencing Data  Version 0.9.6  Date 2022-10-27  Author Xihao Li [aut, cre], Zilin Li [aut, cre], Sheila M. Gaynor [aut], Han Chen [aut]  Maintainer Xihao Li <a href="mailto:xihaoli@g.harvard.edu">xihaoli@g.harvard.edu</a> > Zilin Li <a href="mailto:li@hsph.harvard.edu">li@hsph.harvard.edu</a> Description An R package for performing STAAR pipeline in analyzing whole-genome/whole-exome sequencing data.  License GPL-3  Copyright See COPYRIGHTS for details.  Imports Repp, STAAR, SCANG, dplyr, SeqArray, SeqVarTools, GenomicFeatures, TxD-b.Hsapiens.UCSC.hg38.knownGene, GMMAT, GENESIS, Matrix, methods  Encoding UTF-8  LazyData true  Depends R (>= 3.2.0)  LinkingTo Repp, ReppArmadillo  RoxygenNote 7.1.2  Suggests knitr, rmarkdown  VignetteBuilder knitr  R topics documented:  Dynamic_Window_SCANG fit_nullmodel genesis2staar_nullmodel Gene_Centric_Coding_cond Gene_Centric_Coding_cond Gene_Centric_Noncoding In Gene_Centric_Noncoding_cond Individual_Analysis_cond LD_pruning ncRNA ncRNA ncRNA 18 ncRNA_cond 20	
Version 0.9.6  Date 2022-10-27  Author Xihao Li [aut, cre], Zilin Li [aut, cre], Sheila M. Gaynor [aut], Han Chen [aut]  Maintainer Xihao Li <a href="mailto:xihaoli@g.harvard.edu">xilin Li <li@hsph.harvard.edu< a="">  Description An R package for performing STAAR pipeline in analyzing whole-genome/whole-exome sequencing data.  License GPL-3  Copyright See COPYRIGHTS for details.  Imports Repp, STAAR, SCANG, dplyr, SeqArray, SeqVarTools, GenomicFeatures, TxD-b.Hsapiens.UCSC.hg38.knownGene, GMMAT, GENESIS, Matrix, methods  Encoding UTF-8  LazyData true  Depends R (&gt;= 3.2.0)  LinkingTo Rcpp, RcppArmadillo  RoxygenNote 7.1.2  Suggests knitr, rmarkdown  VignetteBuilder knitr  R topics documented:  Dynamic_Window_SCANG fit_nullmodel genesis2staar_nullmodel Gene_Centric_Coding Gene_Centric_Coding_cond Gene_Centric_Noncoding Gene_Centric_Noncoding Individual_Analysis Individual_Analysis_cond LD_pruning neRNA  18</li@hsph.harvard.edu<></a>	Type Package
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Dynamic\_Window\_SCANG Genetic region analysis of dynamic windows using SCANG-STAAR procedure

# **Description**

The Dynamic\_Window\_SCANG function takes in chromosome, starting location, ending location, the object of opened annotated GDS file, and the object from fitting the null model to analyze the association between a quantitative/dichotomous phenotype and variants in a genetic region by using SCANG-STAAR procedure. For each dynamic window, the scan statistic of SCANG-STAAR-O is the set-based p-value of an omnibus test that aggregated p-values across different types of multiple annotation-weighted variant-set tests SKAT(1,1), SKAT(1,25), Burden(1,1) and Burden(1,25) using ACAT method; the scan statistic of SCANG-STAAR-S is the set-based p-value of STAAR-S, which is an omnibus test that aggregated p-values across multiple annotation-weighted variant-set tests SKAT(1,1) and SKAT(1,25) using ACAT method; the scan statistic of SCANG-STAAR-B is the set-based p-value of STAAR-B, which is an omnibus test that aggregated p-values across multiple annotation-weighted variant-set tests Burden(1,1) and Burden(1,25) using ACAT method.

# Usage

```
Dynamic_Window_SCANG(
  chr,
  start_loc,
  end_loc,
  genofile,
  obj_nullmodel,
  Lmin = 40,
  Lmax = 300,
  steplength = 10,
  rare_maf_cutoff = 0.01,
  p_filter = 1e-08,
  f = 0,
  alpha = 0.1,
  QC_label = "annotation/filter",
  variant_type = c("SNV", "Indel", "variant"),
  geno_missing_imputation = c("mean", "minor"),
  Annotation_dir = "annotation/info/FunctionalAnnotation",
  Annotation_name_catalog,
  Use_annotation_weights = c(TRUE, FALSE),
  Annotation_name = NULL,
  silent = FALSE
```

#### **Arguments**

chr chromosome.

start\_loc starting location (position) of the genetic region to be analyzed using SCANG-

STAAR procedure.

end\_loc ending location (position) of the genetic region to be analyzed using SCANG-

STAAR procedure.

genofile an object of opened annotated GDS (aGDS) file.

obj\_nullmodel an object from fitting the null model, which is the output from fit\_nullmodel

function and transformed using the staar2scang\_nullmodel function.

Lmin minimum number of variants in searching windows (default = 40).

Lmax maximum number of variants in searching windows (default = 300).

steplength difference of number of variants in searching windows, that is, the number of

variants in searching windows are Lmin, Lmin+steplength, Lmin+steplength,...,

Lmax (default = 10).

rare\_maf\_cutoff

a cutoff of maximum minor allele frequency in defining rare variants (default =

0.01).

p\_filter a filtering threshold of screening method for SKAT in SCANG-STAAR. SKAT

p-values are calculated for regions whose p-value is possibly smaller than the

filtering threshold (default = 1e-8).

f an overlap fraction, which controls for the overlapping proportion of of detected

regions. For example, when f=0, the detected regions are non-overlapped with each other, and when f=1, we keep every susceptive region as detected regions

(default = 0).

alpha family-wise/genome-wide significance level (default = 0.1).

 $\label{eq:channel name of the QC label in the GDS/aGDS file (default = "annotation/filter").}$ 

variant\_type type of variant included in the analysis. Choices include "SNV", "Indel", or

"variant" (default = "SNV").

geno\_missing\_imputation

method of handling missing genotypes. Either "mean" or "minor" (default =

"mean").

Annotation\_dir channel name of the annotations in the aGDS file

(default = "annotation/info/FunctionalAnnotation").

Annotation\_name\_catalog

a data frame containing the name and the corresponding channel name in the

aGDS file.

Use\_annotation\_weights

use annotations as weights or not (default = TRUE).

Annotation\_name

a vector of annotation names used in SCANG-STAAR (default = NULL).

silent logical: should the report of error messages be suppressed (default = FALSE).

# Value

The function returns a list with the following members:

SCANG\_O\_res: A matrix that summarizes the significant region detected by SCANG-STAAR-O, including the negative log transformation of SCANG-STAAR-O p-value ("-logp"), chromosome

4 fit\_nullmodel

("chr"), start position ("start\_pos"), end position ("end\_pos"), family-wise/genome-wide error rate (GWER) and the number of variants ("SNV num").

SCANG\_0\_top1: A vector of length 4 which summarizes the top 1 region detected by SCANG-STAAR-O. including the negative log transformation of SCANG-STAAR-O p-value ("-logp"), chromosome ("chr"), start position ("start\_pos"), end position ("end\_pos"), family-wise/genome-wide error rate (GWER) and the number of variants ("SNV\_num").

SCANG\_O\_emthr: A vector of Monte Carlo simulation sample for generating the empirical threshold. The 1-alpha quantile of this vector is the empirical threshold.

SCANG\_S\_res, SCANG\_S\_top1, SCANG\_S\_emthr: Analysis results using SCANG-STAAR-S. Details see SCANG-STAAR-O.

SCANG\_B\_res, SCANG\_B\_top1, SCANG\_B\_emthr: Analysis results using SCANG-STAAR-B. Details see SCANG-STAAR-O.

#### References

Li, Z., Li, X., et al. (2022). A framework for detecting noncoding rare-variant associations of large-scale whole-genome sequencing studies. *Nature Methods*. (pub)

Li, Z., Li, X., et al. (2019). Dynamic scan procedure for detecting rare-variant association regions in whole-genome sequencing studies. *The American Journal of Human Genetics*, 104(5), 802-814. (pub)

Li, X., Li, Z., et al. (2020). Dynamic incorporation of multiple in silico functional annotations empowers rare variant association analysis of large whole-genome sequencing studies at scale. *Nature Genetics*, 52(9), 969-983. (pub)

Liu, Y., et al. (2019). Acat: A fast and powerful p value combination method for rare-variant analysis in sequencing studies. *The American Journal of Human Genetics*, 104(3), 410-421. (pub)

fit\_nullmodel

Fitting generalized linear mixed model with known relationship matrices under the null hypothesis.

# **Description**

The fit\_nullmodel function is a wrapper of the glmmkin function from the GMMAT package that fits a regression model under the null hypothesis for related samples, which provides the preliminary step for subsequent variant-set tests in whole genome sequencing data analysis. See glmmkin for more details.

#### Usage

```
fit_nullmodel(
   fixed,
   data = parent.frame(),
   kins,
   use_sparse = NULL,
   kins_cutoff = 0.022,
   id,
   random.slope = NULL,
   groups = NULL,
   family = binomial(link = "logit"),
```

fit\_nullmodel 5

```
method = "REML",
method.optim = "AI",
maxiter = 500,
tol = 1e-05,
taumin = 1e-05,
taumax = 1e+05,
tauregion = 10,
verbose = FALSE,
...
)
```

#### **Arguments**

fixed an object of class formula (or one that can be coerced to that class): a symbolic

description of the fixed effects model to be fitted.

data a data frame or list (or object coercible by as.data.frame to a data frame)

containing the variables in the model.

kins a known positive semi-definite relationship matrix (e.g. kinship matrix in ge-

netic association studies) or a list of known positive semi-definite relationship matrices. The rownames and colnames of these matrices must at least include all samples as specified in the id column of the data frame data. If kins is

NULL, it will fit a generalized linear model for unrelated samples.

use\_sparse a logical switch of whether the provided dense kins matrix should be trans-

formed to a sparse matrix (default = NULL).

kins\_cutoff the cutoff value for clustering samples to make the output matrix sparse block-

diagonal (default = 0.022).

id a column in the data frame data, indicating the id of samples. When there are

duplicates in id, the data is assumed to be longitudinal with repeated measures.

random. slope an optional column indicating the random slope for time effect used in a mixed

effects model for longitudinal data. It must be included in the names of data. There must be duplicates in id and method.optim must be "AI" (default = NUL-

L).

groups an optional categorical variable indicating the groups used in a heteroscedastic

linear mixed model (allowing residual variances in different groups to be different). This variable must be included in the names of data, and family must be

"gaussian" and method.optim must be "AI" (default = NULL).

family a description of the error distribution and link function to be used in the model.

This can be a character string naming a family function, a family function or the result of a call to a family function. (See family for details of family functions).

method method of fitting the generalized linear mixed model. Either "REML" or "ML"

(default = "REML").

method.optim optimization method of fitting the generalized linear mixed model. Either "AI",

"Brent" or "Nelder-Mead" (default = "AI").

maxiter a positive integer specifying the maximum number of iterations when fitting the

generalized linear mixed model (default = 500).

tol a positive number specifying tolerance, the difference threshold for parameter

estimates below which iterations should be stopped (default = 1e-5).

taumin the lower bound of search space for the variance component parameter  $\tau$  (default

= 1e-5), used when method.optim = "Brent". See Details.

taumax the upper bound of search space for the variance component parameter  $\tau$  (default = 1e5), used when method.optim = "Brent". See Details.

tauregion the number of search intervals for the REML or ML estimate of the variance

component parameter  $\tau$  (default = 10), used when method.optim = "Brent".

See Details.

verbose a logical switch for printing detailed information (parameter estimates in each

iteration) for testing and debugging purpose (default = FALSE).

... additional arguments that could be passed to glm.

#### Value

The function returns an object of the model fit from glmmkin (obj\_nullmodel) and whether the kins matrix is sparse when fitting the null model. See glmmkin for more details.

#### References

Chen, H., et al. (2016). Control for population structure and relatedness for binary traits in genetic association studies via logistic mixed models. *The American Journal of Human Genetics*, 98(4), 653-666. (pub)

Chen, H., et al. (2019). Efficient variant set mixed model association tests for continuous and binary traits in large-scale whole-genome sequencing studies. *The American Journal of Human Genetics*, 104(2), 260-274. (pub)

Chen, H. (2021). GMMAT: Generalized linear Mixed Model Association Tests Version 1.3.2. (web)

genesis2staar\_nullmodel

Transforming the null model object fitted using GENESIS to the null model object to be used for STAAR

#### **Description**

The genesis2staar\_nullmodel function takes in the object from fitting the null model using the GENESIS package and transforms it to the object from fitting the null model to be used for STAAR procedure.

# Usage

```
genesis2staar_nullmodel(obj_nullmodel_genesis)
```

# **Arguments**

obj\_nullmodel\_genesis

an object from fitting the null model, which is the output from fitNullModel function in the GENESIS package.

# Value

an object from fitting the null model for related samples to be used for STAAR procedure, which is the output from fit\_nullmodel function.

Gene\_Centric\_Coding

#### References

Gogarten, S.M., Sofer, T., Chen, H., et al. (2019). Genetic association testing using the GENESIS R/Bioconductor package. *Bioinformatics*, *35*(24), 5346-5348. (pub)

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Li, X., Li, Z., et al. (2020). Dynamic incorporation of multiple in silico functional annotations empowers rare variant association analysis of large whole-genome sequencing studies at scale. *Nature Genetics*, 52(9), 969-983. (pub)

Li, Z., Li, X., et al. (2022). A framework for detecting noncoding rare-variant associations of large-scale whole-genome sequencing studies. *Nature Methods*. (pub)

Gene\_Centric\_Coding Gene-centric analysis of coding functional categories using STAAR procedure

# **Description**

The Gene\_Centric\_Coding function takes in chromosome, gene name, functional category, the object of opened annotated GDS file, and the object from fitting the null model to analyze the association between a quantitative/dichotomous phenotype and coding functional categories of a gene by using STAAR procedure. For each coding functional category, the STAAR-O p-value is a p-value from an omnibus test that aggregated SKAT(1,25), SKAT(1,1), Burden(1,25), Burden(1,1), ACAT-V(1,25), and ACAT-V(1,1) together with p-values of each test weighted by each annotation using Cauchy method.

# Usage

```
Gene_Centric_Coding(
  chr,
  gene_name,
  category = c("all_categories", "plof", "plof_ds", "missense", "disruptive_missense",
    "synonymous"),
  genofile,
  obj_nullmodel,
  rare_maf_cutoff = 0.01,
  rv_num_cutoff = 2,
  QC_label = "annotation/filter"
  variant_type = c("SNV", "Indel", "variant"),
  geno_missing_imputation = c("mean", "minor"),
  Annotation_dir = "annotation/info/FunctionalAnnotation",
  Annotation_name_catalog,
 Use_annotation_weights = c(TRUE, FALSE),
 Annotation_name = NULL,
  silent = FALSE
)
```

# Arguments

chr chromosome.

gene\_name name of the gene to be analyzed using STAAR procedure.

category the coding functional category to be analyzed using STAAR procedure. Choices

 $include \ all\_categories, \ plof, \ plof\_ds, \ missense, \ disruptive\_missense,$ 

synonymous (default = all\_categories).

genofile an object of opened annotated GDS (aGDS) file.

obj\_nullmodel an object from fitting the null model, which is either the output from fit\_nullmodel

function, or the output from fitNullModel function in the GENESIS package  $\,$ 

and transformed using the genesis2staar\_nullmodel function.

rare\_maf\_cutoff

the cutoff of maximum minor allele frequency in defining rare variants (default

= 0.01).

rv\_num\_cutoff the cutoff of minimum number of variants of analyzing a given variant-set (de-

fault = 2).

QC\_label channel name of the QC label in the GDS/aGDS file (default = "annotation/filter").

variant\_type type of variant included in the analysis. Choices include "SNV", "Indel", or

"variant" (default = "SNV").

geno\_missing\_imputation

method of handling missing genotypes. Either "mean" or "minor" (default =

"mean").

Annotation\_dir channel name of the annotations in the aGDS file

(default = "annotation/info/FunctionalAnnotation").

Annotation\_name\_catalog

a data frame containing the name and the corresponding channel name in the

aGDS file.

Use\_annotation\_weights

use annotations as weights or not (default = TRUE).

Annotation\_name

a vector of annotation names used in STAAR (default = NULL).

silent logical: should the report of error messages be suppressed (default = FALSE).

# Value

a list of data frames containing the STAAR p-values (including STAAR-O) corresponding to the coding functional category of the given gene.

#### References

Li, Z., Li, X., et al. (2022). A framework for detecting noncoding rare-variant associations of large-scale whole-genome sequencing studies. *Nature Methods*. (pub)

Li, X., Li, Z., et al. (2020). Dynamic incorporation of multiple in silico functional annotations empowers rare variant association analysis of large whole-genome sequencing studies at scale. *Nature Genetics*, 52(9), 969-983. (pub)

```
Gene_Centric_Coding_cond
```

Gene-centric conditional analysis of coding functional categories using STAAR procedure

#### **Description**

The Gene\_Centric\_Coding\_cond function takes in chromosome, gene name, functional category, the object of opened annotated GDS file, the object from fitting the null model, and the set of known variants to be adjusted for in conditional analysis to analyze the conditional association between a quantitative/dichotomous phenotype and coding functional categories of a gene by using STAAR procedure. For each coding functional category, the conditional STAAR-O p-value is a p-value from an omnibus test that aggregated conditional SKAT(1,25), SKAT(1,1), Burden(1,25), Burden(1,1), ACAT-V(1,25), and ACAT-V(1,1) together with conditional p-values of each test weighted by each annotation using Cauchy method.

#### Usage

```
Gene_Centric_Coding_cond(
  chr,
  gene_name,
  category = c("plof", "plof_ds", "missense", "disruptive_missense", "synonymous"),
  genofile,
  obj_nullmodel,
  known_loci = NULL,
  rare_maf_cutoff = 0.01,
  rv_num_cutoff = 2,
 method_cond = c("optimal", "naive"),
 QC_label = "annotation/filter",
  variant_type = c("SNV", "Indel", "variant"),
  geno_missing_imputation = c("mean", "minor"),
 Annotation_dir = "annotation/info/FunctionalAnnotation",
 Annotation_name_catalog,
 Use_annotation_weights = c(TRUE, FALSE),
 Annotation_name = NULL
)
```

# **Arguments**

chr	chromosome.
gene_name	name of the gene to be analyzed using STAAR procedure.
category	the coding functional category to be analyzed using STAAR procedure. Choices include plof, plof_ds, missense, disruptive_missense, synonymous (default = plof).
genofile	an object of opened annotated GDS (aGDS) file.
obj_nullmodel	an object from fitting the null model, which is either the output from fit_nullmodel function, or the output from fitNullModel function in the GENESIS package and transformed using the genesis2staar_nullmodel function.

known\_loci the data frame of variants to be adjusted for in conditional analysis and should

contain 4 columns in the following order: chromosome (CHR), position (POS), reference allele (REF), and alternative allele (ALT) (default = NULL).

rare\_maf\_cutoff

the cutoff of maximum minor allele frequency in defining rare variants (default

= 0.01).

rv\_num\_cutoff the cutoff of minimum number of variants of analyzing a given variant-set (de-

fault = 2).

method\_cond a character value indicating the method for conditional analysis. optimal refers

to regressing residuals from the null model on known\_loci as well as all covariates used in fitting the null model (fully adjusted) and taking the residuals; naive refers to regressing residuals from the null model on known\_loci and

taking the residuals (default = optimal).

QC\_label channel name of the QC label in the GDS/aGDS file (default = "annotation/filter").

variant\_type type of variant included in the analysis. Choices include "SNV", "Indel", or

"variant" (default = "SNV").

geno\_missing\_imputation

method of handling missing genotypes. Either "mean" or "minor" (default = "mean").

Annotation\_dir channel name of the annotations in the aGDS file

(default = "annotation/info/FunctionalAnnotation").

Annotation\_name\_catalog

a data frame containing the name and the corresponding channel name in the aGDS file.

Use\_annotation\_weights

use annotations as weights or not (default = TRUE).

Annotation\_name

a vector of annotation names used in STAAR (default = NULL).

# Value

a data frame containing the conditional STAAR p-values (including STAAR-O) corresponding to each coding functional category of the given gene.

#### References

Li, Z., Li, X., et al. (2022). A framework for detecting noncoding rare-variant associations of large-scale whole-genome sequencing studies. *Nature Methods*. (pub)

Li, X., Li, Z., et al. (2020). Dynamic incorporation of multiple in silico functional annotations empowers rare variant association analysis of large whole-genome sequencing studies at scale. *Nature Genetics*, 52(9), 969-983. (pub)

Sofer, T., et al. (2019). A fully adjusted two-stage procedure for rank-normalization in genetic association studies. *Genetic Epidemiology*, 43(3), 263-275. (pub)

```
Gene_Centric_Noncoding
```

Gene-centric analysis of noncoding functional categories using S-TAAR procedure

# **Description**

The Gene\_Centric\_Noncoding function takes in chromosome, gene name, functional category, the object of opened annotated GDS file, and the object from fitting the null model to analyze the association between a quantitative/dichotomous phenotype and noncoding functional categories of a gene by using STAAR procedure. For each noncoding functional category, the STAAR-O p-value is a p-value from an omnibus test that aggregated SKAT(1,25), SKAT(1,1), Burden(1,25), Burden(1,1), ACAT-V(1,25), and ACAT-V(1,1) together with p-values of each test weighted by each annotation using Cauchy method.

# Usage

```
Gene_Centric_Noncoding(
 chr,
 gene_name,
  category = c("all_categories", "downstream", "upstream", "UTR", "promoter_CAGE",
    "promoter_DHS", "enhancer_CAGE", "enhancer_DHS"),
  genofile.
 obj_nullmodel,
  rare_maf_cutoff = 0.01,
  rv_num_cutoff = 2,
  QC_label = "annotation/filter",
  variant_type = c("SNV", "Indel", "variant"),
  geno_missing_imputation = c("mean", "minor"),
  Annotation_dir = "annotation/info/FunctionalAnnotation",
  Annotation_name_catalog,
 Use_annotation_weights = c(TRUE, FALSE),
  Annotation_name = NULL,
  silent = FALSE
)
```

# **Arguments**

= 0.01).

chr chromosome. gene\_name name of the gene to be analyzed using STAAR procedure. the noncoding functional category to be analyzed using STAAR procedure. Choiccategory es include all\_categories, downstream, upstream, UTR, promoter\_CAGE, promoter\_DHS, enhancer\_CAGE, enhancer\_DHS (default = all\_categories). an object of opened annotated GDS (aGDS) file. genofile obj\_nullmodel an object from fitting the null model, which is either the output from fit\_nullmodel function, or the output from fitNullModel function in the GENESIS package and transformed using the genesis2staar\_nullmodel function. rare\_maf\_cutoff the cutoff of maximum minor allele frequency in defining rare variants (default

rv\_num\_cutoff the cutoff of minimum number of variants of analyzing a given variant-set (de-

fault = 2).

QC\_label channel name of the QC label in the GDS/aGDS file (default = "annotation/filter").

variant\_type type of variant included in the analysis. Choices include "SNV", "Indel", or

"variant" (default = "SNV").

 ${\tt geno\_missing\_imputation}$ 

method of handling missing genotypes. Either "mean" or "minor" (default = "mean").

Annotation\_dir channel name of the annotations in the aGDS file

(default = "annotation/info/FunctionalAnnotation").

Annotation\_name\_catalog

a data frame containing the name and the corresponding channel name in the aGDS file.

Use\_annotation\_weights

use annotations as weights or not (default = TRUE).

Annotation\_name

a vector of annotation names used in STAAR (default = NULL).

silent logical: should the report of error messages be suppressed (default = FALSE).

#### Value

a list of data frames containing the STAAR p-values (including STAAR-O) corresponding to each noncoding functional category of the given gene.

# References

Li, Z., Li, X., et al. (2022). A framework for detecting noncoding rare-variant associations of large-scale whole-genome sequencing studies. *Nature Methods*. (pub)

Li, X., Li, Z., et al. (2020). Dynamic incorporation of multiple in silico functional annotations empowers rare variant association analysis of large whole-genome sequencing studies at scale. *Nature Genetics*, 52(9), 969-983. (pub)

Gene\_Centric\_Noncoding\_cond

Gene-centric conditional analysis of noncoding functional categories using STAAR procedure

# **Description**

The Gene\_Centric\_Noncoding\_cond function takes in chromosome, gene name, functional category, the object of opened annotated GDS file, the object from fitting the null model, and the set of known variants to be adjusted for in conditional analysis to analyze the conditional association between a quantitative/dichotomous phenotype and noncoding functional categories of a gene by using STAAR procedure. For each noncoding functional category, the conditional STAAR-O p-value is a p-value from an omnibus test that aggregated conditional SKAT(1,25), SKAT(1,1), Burden(1,25), Burden(1,1), ACAT-V(1,25), and ACAT-V(1,1) together with conditional p-values of each test weighted by each annotation using Cauchy method.

#### Usage

```
Gene_Centric_Noncoding_cond(
 chr,
 gene_name,
  category = c("downstream", "upstream", "UTR", "promoter_CAGE", "promoter_DHS",
    "enhancer_CAGE", "enhancer_DHS"),
  genofile,
  obj_nullmodel,
  known_loci = NULL,
  rare_maf_cutoff = 0.01,
  rv_num_cutoff = 2,
 method_cond = c("optimal", "naive"),
 QC_label = "annotation/filter",
  variant_type = c("SNV", "Indel", "variant"),
  geno_missing_imputation = c("mean", "minor"),
  Annotation_dir = "annotation/info/FunctionalAnnotation",
  Annotation_name_catalog,
 Use_annotation_weights = c(TRUE, FALSE),
  Annotation_name = NULL
)
```

# **Arguments**

chr chromosome.

gene\_name name of the gene to be analyzed using STAAR procedure.

category the noncoding functional category to be analyzed using STAAR procedure. Choic-

es include downstream, upstream, UTR, promoter\_CAGE, promoter\_DHS, enhancer\_CAGE,

enhancer\_DHS (default = downstream).

genofile an object of opened annotated GDS (aGDS) file.

obj\_nullmodel an object from fitting the null model, which is either the output from fit\_nullmodel

function, or the output from fitNullModel function in the GENESIS package

and transformed using the genesis2staar\_nullmodel function.

known\_loci the data frame of variants to be adjusted for in conditional analysis and should

contain 4 columns in the following order: chromosome (CHR), position (POS),

reference allele (REF), and alternative allele (ALT) (default = NULL).

rare\_maf\_cutoff

the cutoff of maximum minor allele frequency in defining rare variants (default

= 0.01).

rv\_num\_cutoff the cutoff of minimum number of variants of analyzing a given variant-set (de-

fault = 2).

method\_cond a character value indicating the method for conditional analysis. optimal refers

to regressing residuals from the null model on known\_loci as well as all covariates used in fitting the null model (fully adjusted) and taking the residuals; naive refers to regressing residuals from the null model on known\_loci and

taking the residuals (default = optimal).

QC\_label channel name of the QC label in the GDS/aGDS file (default = "annotation/filter").

variant\_type type of variant included in the analysis. Choices include "SNV", "Indel", or

"variant" (default = "SNV").

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#### Value

a data frame containing the conditional STAAR p-values (including STAAR-O) corresponding to the noncoding functional category of the given gene.

#### References

Li, Z., Li, X., et al. (2022). A framework for detecting noncoding rare-variant associations of large-scale whole-genome sequencing studies. *Nature Methods*. (pub)

Li, X., Li, Z., et al. (2020). Dynamic incorporation of multiple in silico functional annotations empowers rare variant association analysis of large whole-genome sequencing studies at scale. *Nature Genetics*, 52(9), 969-983. (pub)

Sofer, T., et al. (2019). A fully adjusted two-stage procedure for rank-normalization in genetic association studies. *Genetic Epidemiology*, 43(3), 263-275. (pub)

Individual\_Analysis Individual-variant

Individual-variant analysis using score test

#### **Description**

The Individual\_Analysis function takes in chromosome, starting location, ending location, the object of opened annotated GDS file, and the object from fitting the null model to analyze the association between a quantitative/dichotomous phenotype and each individual variant in a genetic region by using score test.

# Usage

```
Individual_Analysis(
   chr,
   start_loc,
   end_loc,
   genofile,
   obj_nullmodel,
   mac_cutoff = 20,
   subset_variants_num = 5000,
   QC_label = "annotation/filter",
   variant_type = c("variant", "SNV", "Indel"),
   geno_missing_imputation = c("mean", "minor")
)
```

#### **Arguments**

chr	chromosome.	
start_loc	starting location (position) of the genetic region for each individual variant to be analyzed using score test.	
end_loc	ending location (position) of the genetic region for each individual variant to be analyzed using score test.	
genofile	an object of opened annotated GDS (aGDS) file.	
obj_nullmodel	an object from fitting the null model, which is either the output from fit_nullmodel function, or the output from fitNullModel function in the GENESIS package and transformed using the genesis2staar_nullmodel function.	
mac_cutoff	the cutoff of minimum minor allele count in defining individual variants (default = 20).	
subset_variants_num		
	the number of variants to run per subset for each time (default = $5e3$ ).	
QC_label	channel name of the QC label in the GDS/aGDS file (default = "annotation/filter").	
variant_type	type of variant included in the analysis. Choices include "variant", "SNV", or "Indel" (default = "variant").	
<pre>geno_missing_imputation</pre>		

#### Value

a data frame containing the score test p-value and the estimated effect size for each individual variant in the given genetic region. The first 4 columns correspond to chromosome (CHR), position (POS), reference allele (REF), and alternative allele (ALT).

# References

Chen, H., et al. (2016). Control for population structure and relatedness for binary traits in genetic association studies via logistic mixed models. *The American Journal of Human Genetics*, 98(4), 653-666. (pub)

Individual\_Analysis\_cond

Individual-variant conditional analysis using score test

# **Description**

The Individual\_Analysis\_cond function takes in the data frame of individual variants, the object of opened annotated GDS file, the object from fitting the null model, and the set of known variants to be adjusted for in conditional analysis to analyze the conditional association between a quantitative/dichotomous phenotype and each (significant) individual variant by using score test.

#### Usage

```
Individual_Analysis_cond(
   chr,
   individual_results,
   genofile,
   obj_nullmodel,
   known_loci = NULL,
   method_cond = c("optimal", "naive"),
   QC_label = "annotation/filter",
   variant_type = c("variant", "SNV", "Indel"),
   geno_missing_imputation = c("mean", "minor"),
   geno_position_ascending = TRUE
)
```

# **Arguments**

chr chromosome.

individual\_results

the data frame of (significant) individual variants for conditional analysis using score test. The first 4 columns should correspond to chromosome (CHR), position (POS), reference allele (REF), and alternative allele (ALT).

genofile an object of opened annotated GDS (aGDS) file.

obj\_nullmodel an object from fitting the null model, which is either the output from fit\_nullmodel

function, or the output from fitNullModel function in the GENESIS package

and transformed using the genesis2staar\_nullmodel function.

known\_loci the data frame of variants to be adjusted for in conditional analysis and should

contain 4 columns in the following order: chromosome (CHR), position (POS),

reference allele (REF), and alternative allele (ALT) (default = NULL).

method\_cond a character value indicating the method for conditional analysis. optimal refers

to regressing residuals from the null model on known\_loci as well as all covariates used in fitting the null model (fully adjusted) and taking the residuals; naive refers to regressing residuals from the null model on known\_loci and

taking the residuals (default = optimal).

QC\_label channel name of the QC label in the GDS/aGDS file (default = "annotation/filter").

variant\_type type of variant included in the analysis. Choices include "variant", "SNV", or

"Indel" (default = "variant").

geno\_missing\_imputation

method of handling missing genotypes. Either "mean" or "minor" (default = "mean").

geno\_position\_ascending

logical: are the variant positions in ascending order in the GDS/aGDS file (default = TRUE).

#### Value

a data frame containing the conditional score test p-value and the estimated effect size of the minor allele for each (significant) individual variant in individual\_results.

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#### References

Chen, H., et al. (2016). Control for population structure and relatedness for binary traits in genetic association studies via logistic mixed models. *The American Journal of Human Genetics*, 98(4), 653-666. (pub)

Sofer, T., et al. (2019). A fully adjusted two-stage procedure for rank-normalization in genetic association studies. *Genetic Epidemiology*, 43(3), 263-275. (pub)

LD\_pruning

Linkage disequilibrium (LD) pruning procedure

# **Description**

The LD\_pruning function takes in chromosome, the object of opened annotated GDS file, the object from fitting the null model, and a given list of variants to perform LD pruning among these variants in sequential conditional analysis by using score test.

# Usage

```
LD_pruning(
   chr,
   genofile,
   obj_nullmodel,
   variants_list,
   maf_cutoff = 0.01,
   cond_p_thresh = 1e-04,
   method_cond = c("optimal", "naive"),
   QC_label = "annotation/filter",
   variant_type = c("variant", "SNV", "Indel"),
   geno_missing_imputation = c("mean", "minor"),
   geno_position_ascending = TRUE
)
```

#### Arguments

chr	chromosome.
genofile	an object of opened annotated GDS (aGDS) file.
obj_nullmodel	an object from fitting the null model, which is either the output from fit_nullmodel function, or the output from fitNullModel function in the GENESIS package and transformed using the genesis2staar_nullmodel function.
variants_list	the data frame of variants to be LD-pruned in sequential conditional analysis and should contain 4 columns in the following order: chromosome (CHR), position (POS), reference allele (REF), and alternative allele (ALT).
maf_cutoff	the cutoff of minimum minor allele frequency in defining individual variants to be LD-pruned (default = $0.01$ ).
cond_p_thresh	the cutoff of maximum conditional p-value allowed for variants to be kept in the LD-pruned list of variants (default = 1e-04).

ncRNA

method_cond	a character value indicating the method for conditional analysis. optimal refers to regressing residuals from the null model on known_loci as well as all covariates used in fitting the null model (fully adjusted) and taking the residuals; naive refers to regressing residuals from the null model on known_loci and taking the residuals (default = optimal).	
QC_label	$channel\ name\ of\ the\ QC\ label\ in\ the\ GDS/aGDS\ file\ (default="annotation/filter").$	
variant_type	type of variant included in the analysis. Choices include "variant", "SNV", or "Indel" (default = "variant").	
<pre>geno_missing_imputation</pre>		
	method of handling missing genotypes. Either "mean" or "minor" (default = "mean").	
geno_position_ascending		
	logical: are the variant positions in ascending order in the GDS/aGDS file (default = TRUE).	

#### Value

a data frame containing the list of LD-pruned variants in the given chromosome.

ncRNA

Gene-centric analysis of long noncoding RNA (ncRNA) category using STAAR procedure

# **Description**

The ncRNA function takes in chromosome, gene name, the object of opened annotated GDS file, and the object from fitting the null model to analyze the association between a quantitative/dichotomous phenotype and the exonic and splicing category of an ncRNA gene by using STAAR procedure. For each ncRNA category, the STAAR-O p-value is a p-value from an omnibus test that aggregated SKAT(1,25), SKAT(1,1), Burden(1,25), Burden(1,1), ACAT-V(1,25), and ACAT-V(1,1) together with p-values of each test weighted by each annotation using Cauchy method.

# Usage

```
ncRNA(
   chr,
   gene_name,
   genofile,
   obj_nullmodel,
   rare_maf_cutoff = 0.01,
   rv_num_cutoff = 2,
   QC_label = "annotation/filter",
   variant_type = c("SNV", "Indel", "variant"),
   geno_missing_imputation = c("mean", "minor"),
   Annotation_dir = "annotation/info/FunctionalAnnotation",
   Annotation_name_catalog,
   Use_annotation_weights = c(TRUE, FALSE),
   Annotation_name = NULL,
   silent = FALSE
)
```

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# **Arguments**

chr chromosome.

gene\_name name of the ncRNA gene to be analyzed using STAAR procedure.

genofile an object of opened annotated GDS (aGDS) file.

obj\_nullmodel an object from fitting the null model, which is either the output from fit\_nullmodel

function, or the output from fitNullModel function in the GENESIS package

and transformed using the genesis2staar\_nullmodel function.

rare\_maf\_cutoff

the cutoff of maximum minor allele frequency in defining rare variants (default

= 0.01).

rv\_num\_cutoff the cutoff of minimum number of variants of analyzing a given variant-set (de-

fault = 2).

QC\_label channel name of the QC label in the GDS/aGDS file (default = "annotation/filter").

variant\_type type of variant included in the analysis. Choices include "SNV", "Indel", or

"variant" (default = "SNV").

geno\_missing\_imputation

method of handling missing genotypes. Either "mean" or "minor" (default =

"mean").

Annotation\_dir channel name of the annotations in the aGDS file

(default = "annotation/info/FunctionalAnnotation").

Annotation\_name\_catalog

a data frame containing the name and the corresponding channel name in the

aGDS file.

Use\_annotation\_weights

use annotations as weights or not (default = TRUE).

Annotation\_name

a vector of annotation names used in STAAR (default = NULL).

silent logical: should the report of error messages be suppressed (default = FALSE).

#### Value

a data frame containing the STAAR p-values (including STAAR-O) corresponding to the exonic and splicing category of the given ncRNA gene.

# References

Li, Z., Li, X., et al. (2022). A framework for detecting noncoding rare-variant associations of large-scale whole-genome sequencing studies. *Nature Methods*. (pub)

Li, X., Li, Z., et al. (2020). Dynamic incorporation of multiple in silico functional annotations empowers rare variant association analysis of large whole-genome sequencing studies at scale. *Nature Genetics*, 52(9), 969-983. (pub)

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or a DNIA and and	C
ncRNA_cond	Gene-centric conditional analysis of long noncoding RNA (ncRNA)
	category using STAAR procedure

# **Description**

The ncRNA\_cond function takes in chromosome, gene name, the object of opened annotated GDS file, the object from fitting the null model, and the set of known variants to be adjusted for in conditional analysis to analyze the conditional association between a quantitative/dichotomous phenotype and the noncoding RNA (ncRNA) category of an ncRNA gene by using STAAR procedure. For each ncRNA category, the conditional STAAR-O p-value is a p-value from an omnibus test that aggregated conditional SKAT(1,25), SKAT(1,1), Burden(1,25), Burden(1,1), ACAT-V(1,25), and ACAT-V(1,1) together with conditional p-values of each test weighted by each annotation using Cauchy method.

# Usage

```
ncRNA_cond(
  chr,
  gene_name,
  genofile,
  obj_nullmodel,
  known_loci = NULL,
  rare_maf_cutoff = 0.01,
  rv_num_cutoff = 2,
  method_cond = c("optimal", "naive"),
  QC_label = "annotation/filter",
  variant_type = c("SNV", "Indel", "variant"),
  geno_missing_imputation = c("mean", "minor"),
  Annotation_dir = "annotation/info/FunctionalAnnotation",
  Annotation_name_catalog,
  Use_annotation_weights = c(TRUE, FALSE),
  Annotation_name = NULL
)
```

# **Arguments**

chr	chromosome.	
gene_name	name of the ncRNA gene to be analyzed using STAAR procedure.	
genofile	an object of opened annotated GDS (aGDS) file.	
obj_nullmodel	an object from fitting the null model, which is either the output from fit_nullmodel function, or the output from fitNullModel function in the GENESIS package and transformed using the genesis2staar_nullmodel function.	
known_loci	the data frame of variants to be adjusted for in conditional analysis and should contain 4 columns in the following order: chromosome (chr), position (pos), reference allele (ref), and alternative allele (alt) (default = NULL).	
rare_maf_cutoff		
	the cutoff of maximum minor allele frequency in defining rare variants (default $= 0.01$ ).	

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rv\_num\_cutoff the cutoff of minimum number of variants of analyzing a given variant-set (de-

fault = 2).

method\_cond a character value indicating the method for conditional analysis. optimal refers

to regressing residuals from the null model on known\_loci as well as all covariates used in fitting the null model (fully adjusted) and taking the residuals; naive refers to regressing residuals from the null model on known\_loci and

taking the residuals (default = optimal).

QC\_label channel name of the QC label in the GDS/aGDS file (default = "annotation/filter").

variant\_type type of variant included in the analysis. Choices include "SNV", "Indel", or

"variant" (default = "SNV").

geno\_missing\_imputation

method of handling missing genotypes. Either "mean" or "minor" (default =

"mean").

Annotation\_dir channel name of the annotations in the aGDS file

(default = "annotation/info/FunctionalAnnotation").

Annotation\_name\_catalog

a data frame containing the name and the corresponding channel name in the aGDS file.

Use\_annotation\_weights

use annotations as weights or not (default = TRUE).

Annotation\_name

a vector of annotation names used in STAAR (default = NULL).

#### Value

a data frame containing the conditional STAAR p-values (including STAAR-O) corresponding to the ncRNA category of the given ncRNA gene.

# References

Li, Z., Li, X., et al. (2022). A framework for detecting noncoding rare-variant associations of large-scale whole-genome sequencing studies. *Nature Methods*. (pub)

Li, X., Li, Z., et al. (2020). Dynamic incorporation of multiple in silico functional annotations empowers rare variant association analysis of large whole-genome sequencing studies at scale. *Nature Genetics*, 52(9), 969-983. (pub)

Sofer, T., et al. (2019). A fully adjusted two-stage procedure for rank-normalization in genetic association studies. *Genetic Epidemiology*, 43(3), 263-275. (pub)

Sliding\_Window

Genetic region analysis of sliding windows using STAAR procedure

# Description

The Sliding\_Window function takes in chromosome, starting location, ending location, sliding window length, the object of opened annotated GDS file, and the object from fitting the null model to analyze the association between a quantitative/dichotomous phenotype and variants in a genetic region by using STAAR procedure. For each sliding window, the STAAR-O p-value is a p-value from an omnibus test that aggregated SKAT(1,25), SKAT(1,1), Burden(1,25), Burden(1,1), ACAT-V(1,25), and ACAT-V(1,1) together with p-values of each test weighted by each annotation using Cauchy method.

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#### Usage

```
Sliding_Window(
  chr,
  start_loc,
  end_loc,
  sliding_window_length = 2000,
  type = c("single", "multiple"),
  genofile,
 obj_nullmodel,
  rare_maf_cutoff = 0.01,
  rv_num_cutoff = 2,
  QC_label = "annotation/filter",
  variant_type = c("SNV", "Indel", "variant"),
  geno_missing_imputation = c("mean", "minor"),
  Annotation_dir = "annotation/info/FunctionalAnnotation",
 Annotation_name_catalog,
 Use_annotation_weights = c(TRUE, FALSE),
 Annotation_name = NULL,
  silent = FALSE
)
```

#### **Arguments**

chr chromosome. start\_loc starting location (position) of the genetic region to be analyzed using STAAR procedure. ending location (position) of the genetic region to be analyzed using STAAR end\_loc procedure. sliding\_window\_length the (fixed) length of the sliding window to be analyzed using STAAR procedure. type the type of sliding window to be analyzed using STAAR procedure. Choices include single,multiple (default = single). genofile an object of opened annotated GDS (aGDS) file. obj\_nullmodel an object from fitting the null model, which is either the output from fit\_nullmodel function, or the output from fitNullModel function in the GENESIS package and transformed using the genesis2staar\_nullmodel function. rare\_maf\_cutoff the cutoff of maximum minor allele frequency in defining rare variants (default = 0.01). the cutoff of minimum number of variants of analyzing a given variant-set (derv\_num\_cutoff fault = 2). QC\_label channel name of the QC label in the GDS/aGDS file (default = "annotation/filter"). type of variant included in the analysis. Choices include "SNV", "Indel", or variant\_type "variant" (default = "SNV"). geno\_missing\_imputation method of handling missing genotypes. Either "mean" or "minor" (default = "mean"). Annotation\_dir channel name of the annotations in the aGDS file

(default = "annotation/info/FunctionalAnnotation").

```
Annotation_name_catalog

a data frame containing the name and the corresponding channel name in the aGDS file.

Use_annotation_weights

use annotations as weights or not (default = TRUE).

Annotation_name

a vector of annotation names used in STAAR (default = NULL).

silent logical: should the report of error messages be suppressed (default = FALSE).
```

## Value

a data frame containing the STAAR p-values (including STAAR-O) corresponding to each sliding window in the given genetic region.

#### References

Li, Z., Li, X., et al. (2022). A framework for detecting noncoding rare-variant associations of large-scale whole-genome sequencing studies. *Nature Methods*. (pub)

Li, X., Li, Z., et al. (2020). Dynamic incorporation of multiple in silico functional annotations empowers rare variant association analysis of large whole-genome sequencing studies at scale. *Nature Genetics*, 52(9), 969-983. (pub)

 $Sliding\_Window\_cond$ 

Genetic region conditional analysis of sliding windows using STAAR procedure

# **Description**

The Sliding\_Window\_cond function takes in chromosome, starting location, ending location, the object of opened annotated GDS file, the object from fitting the null model, and the set of known variants to be adjusted for in conditional analysis to analyze the conditional association between a quantitative/dichotomous phenotype and variants in a genetic region by using STAAR procedure. For each sliding window, the conditional STAAR-O p-value is a p-value from an omnibus test that aggregated conditional SKAT(1,25), SKAT(1,1), Burden(1,25), Burden(1,1), ACAT-V(1,25), and ACAT-V(1,1) together with conditional p-values of each test weighted by each annotation using Cauchy method.

#### Usage

```
Sliding_Window_cond(
   chr,
   start_loc,
   end_loc,
   genofile,
   obj_nullmodel,
   known_loci = NULL,
   rare_maf_cutoff = 0.01,
   rv_num_cutoff = 2,
   method_cond = c("optimal", "naive"),
   QC_label = "annotation/filter",
   variant_type = c("SNV", "Indel", "variant"),
```

```
geno_missing_imputation = c("mean", "minor"),
Annotation_dir = "annotation/info/FunctionalAnnotation",
Annotation_name_catalog,
Use_annotation_weights = c(TRUE, FALSE),
Annotation_name = NULL
)
```

# **Arguments**

chr chromosome. start\_loc starting location (position) of the sliding window to be analyzed using STAAR procedure. end\_loc ending location (position) of the sliding window to be analyzed using STAAR procedure. genofile an object of opened annotated GDS (aGDS) file. obi\_nullmodel an object from fitting the null model, which is either the output from fit\_nullmodel function, or the output from fitNullModel function in the GENESIS package and transformed using the genesis2staar\_nullmodel function. the data frame of variants to be adjusted for in conditional analysis and should known\_loci contain 4 columns in the following order: chromosome (CHR), position (POS), reference allele (REF), and alternative allele (ALT) (default = NULL). rare\_maf\_cutoff the cutoff of maximum minor allele frequency in defining rare variants (default = 0.01). the cutoff of minimum number of variants of analyzing a given variant-set (derv\_num\_cutoff fault = 2). a character value indicating the method for conditional analysis. optimal refers method\_cond to regressing residuals from the null model on known\_loci as well as all covariates used in fitting the null model (fully adjusted) and taking the residuals; naive refers to regressing residuals from the null model on known\_loci and taking the residuals (default = optimal). QC\_label channel name of the QC label in the GDS/aGDS file (default = "annotation/filter"). type of variant included in the analysis. Choices include "SNV", "Indel", or variant\_type "variant" (default = "SNV").  ${\tt geno\_missing\_imputation}$ method of handling missing genotypes. Either "mean" or "minor" (default = "mean"). Annotation\_dir channel name of the annotations in the aGDS file (default = "annotation/info/FunctionalAnnotation"). Annotation\_name\_catalog a data frame containing the name and the corresponding channel name in the aGDS file. Use\_annotation\_weights use annotations as weights or not (default = TRUE).

# Value

Annotation\_name

a data frame containing the conditional STAAR p-values (including STAAR-O) corresponding to the sliding window in the given genetic region.

a vector of annotation names used in STAAR (default = NULL).

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#### References

Li, Z., Li, X., et al. (2022). A framework for detecting noncoding rare-variant associations of large-scale whole-genome sequencing studies. *Nature Methods*. (pub)

Li, X., Li, Z., et al. (2020). Dynamic incorporation of multiple in silico functional annotations empowers rare variant association analysis of large whole-genome sequencing studies at scale. *Nature Genetics*, 52(9), 969-983. (pub)

Sofer, T., et al. (2019). A fully adjusted two-stage procedure for rank-normalization in genetic association studies. *Genetic Epidemiology*, 43(3), 263-275. (pub)

staar2scang\_nullmodel Transforming the null model object fitted using STAAR to the null model object to be used for SCANG-STAAR

# **Description**

The staar2scang\_nullmodel function takes in the object from fitting the null model and transforms it to the object from fitting the null model to be used for SCANG-STAAR procedure.

#### Usage

staar2scang\_nullmodel(obj\_nullmodel)

#### **Arguments**

obj\_nullmodel an object from fitting the null model, which is either the output from fit\_nullmodel function, or the output from fitNullModel function in the GENESIS package and transformed using the genesis2staar\_nullmodel function.

# Value

an object from fitting the null model for related samples to be used for SCANG-STAAR procedure, which is the output from fit\_null\_glmmkin\_SCANG function for related samples in the SCANG package.

# References

- Li, Z., Li, X., et al. (2022). A framework for detecting noncoding rare-variant associations of large-scale whole-genome sequencing studies. *Nature Methods*. (pub)
- Li, X., Li, Z., et al. (2020). Dynamic incorporation of multiple in silico functional annotations empowers rare variant association analysis of large whole-genome sequencing studies at scale. *Nature Genetics*, 52(9), 969-983. (pub)
- Li, Z., Li, X., et al. (2019). Dynamic scan procedure for detecting rare-variant association regions in whole-genome sequencing studies. *The American Journal of Human Genetics*, 104(5), 802-814. (pub)

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