Exploring Equivalence Testing with the Updated “TOSTER” R Package

Updated TOSTER R Package

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Abstract

Equivalence testing is arguably under utilized by experimental researchers. Due to limited software support for such analyses, and little education on the topic in graduate programs, the utilization of equivalence testings still appares to be low. One option for equivalence testing is the use of two one-sided tests (TOST). The TOSTER R package and jamovi module, originally developed by Daniel Lakens in 2017, was created to make TOST more accessible to the average researcher. In the past two years, I have made significant changes to the TOSTER package in order to increase its accessibility and provide more robust analysis options for researchers. In this paper, I will detail the changes to the package and highlight new analysis options that will make TOST easier for the average quantitative researcher.

*Keywords:* statistics, bootstrap, minimal effects test, NHST, TOST

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# Introduction

Researchers often erroneously declare that no statistical effect exists based on a single “non-significant” p-value (Altman & Bland, 1995). In many of these cases, the data may corroborate the researcher’s claim, but the interpretation of a null hypothesis significance test (NHST), wherein the lack of significance is considered evidence of “no effect”, is nonetheless incorrect. In order to statistically test for whether there is practically no effect, researchers could use equivalence testing. Equivalence testing is used when the goal of a statistical test is to demonstrate that the difference between two conditions is too small to be meaningful. For example, if a researcher wanted to test whether a new drug was no worse than a standard drug, the null hypothesis would be that the new drug is worse than the standard drug by more than a meaningful amount, and the alternative hypothesis would be that the difference between the two drugs is small enough to be meaningless. A very simple equivalence testing approach is the use of “two one-sided tests” (TOST) (Schuirmann, 1987).

The TOST procedure is a statistical test of whether a parameter (e.g., mean difference) is within a specified interval. The TOST procedure can be used to test the equivalence of two means, two proportions, two regression coefficients, and even two variances. An upper () and lower () equivalence bound is specified based on the smallest effect size of interest (SESOI). If the TOST is below a pre-specified alpha level, then the effect can be considered close enough to zero to be practically equivalent (Lakens, 2017).

Both the complaints about erroneous conclusions regarding equivalence (Altman & Bland, 1995) and proposed statistical solutions (Schuirmann, 1987) have existed for decades now. Yet, the problem appears to persist in many applied disciplines. I believe the continued dissonance is due to a general lack of education on equivalence testing and a struggle for many applied researchers to implement equivalence testing. In my experience, most researchers have received some degree of statistical training in their doctoral or master’s studies, but it is rare that any have idea of how to use TOST. It may also be difficult to implement equivalence testing for many researchers. This may be caused by most statistical software defaulting to a null hypothesis of zero, or even completely lacking an ability to change the null hypothesis. Therefore, I feel the continued development of educational content on TOST, and software to help with such analyses, would be beneficial to many quantitative researchers.

The TOSTER R package[[1]](#footnote-20) was originally developed in by Lakens (2017) to introduce experimental psychologists to the concept of equivalence testing and provide an easy-to-use implementation in R. In the years since that publication, I have made a significant update to the package in order to improve the user interface and expand the tools available within the package. An experienced R programmer may have no problem performing equivalence testing within R, but beginners may struggle with both writing the code and interpreting the output. If you fall into that category, I would suggest using jamovi, an open-source statistical software, that has a TOSTER module to perform equivalence/TOST analyses. Not all the features listed in this manuscript are available in the jamovi module, but it is a good starting point for most researchers without statistical programming experience.

In this manuscript, I will detail the updates to the TOSTER package, and give some basic usage examples of some of the new functions. This is meant to just be an introduction to *how* to perform such analyses, and provide a little bit of context for when such analyses are appropriate. For a greater introduction to equivalence testing, I would suggest reading other methodological tutorials (Lakens, 2017; Lakens, McLatchie, Isager, Scheel, & Dienes, 2020; Lakens, Scheel, & Isager, 2018; Mazzolari, Porcelli, Bishop, & Lakens, 2022).

# TOST with t-tests

In an effort to make TOSTER more informative and easier to use, a new function t\_TOST was created. This function operates very similarly to base R’s t.test function, but performs 3 t-tests (one two-tailed and two one-tailed tests). In addition, this function has a generic method where two vectors can be supplied or a formula can be given (e.g.,y ~ group). This function also makes it easier to switch between types of t-tests. All three types (two sample, one sample, and paired samples) can be performed/calculated from the same function. Moreover, the output from this function is verbose, and should make the decisions derived from the function more informative and user-friendly.

Also, t\_TOST is not limited to equivalence tests. Minimal effects testing (MET) is possible. MET is useful for situations where the hypothesis is about a minimal effect and the *null hypothesis is equivalence* (see Figure 1) (Mazzolari et al., 2022).

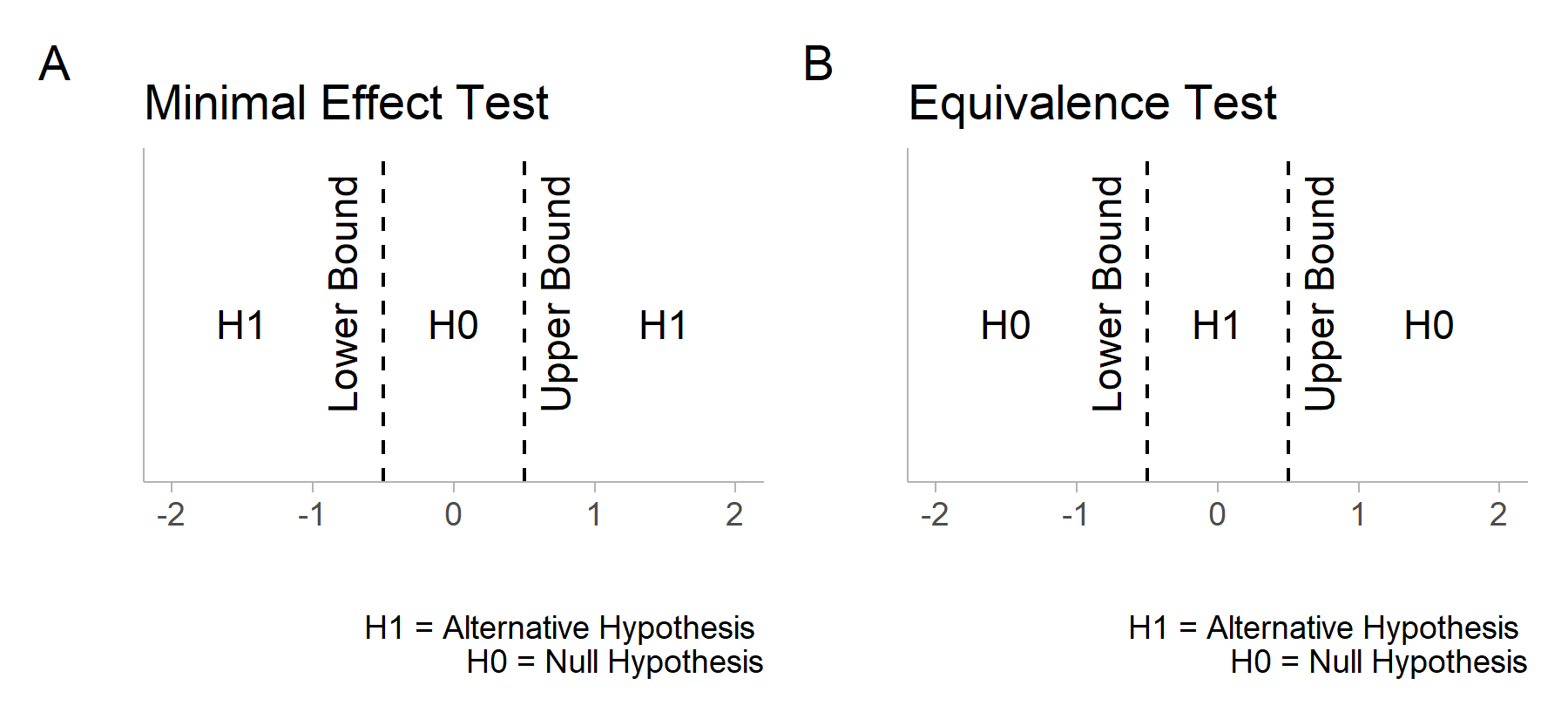


Figure 1: Type of Hypothesis

In these examples of t\_TOST, we will use the bugs data from the jmv R package and the sleep data.

data('sleep')  
library(jmv)  
data('bugs')

## Independent Groups

For this example, we will use the sleep data. In this data, there is a group variable and an outcome extra.

head(sleep,2)

## extra group ID  
## 1 0.7 1 1  
## 2 -1.6 1 2

We will assume the data are independent (in reality this is paired data), and that we have equivalence bounds of +/- 0.5 units of extra. All we need to do is provide the formula, data, and eqb arguments for the function to run appropriately. In addition, we can set the var.equal argument (to assume equal variance), and the paired argument (sets if the data is paired or not). Both are logical indicators that can be set to TRUE or FALSE. The alpha is automatically set to 0.05 but this can also be adjusted by the user depending on the desired alpha-level[[2]](#footnote-27).

Standardize mean differences (SMDs) are provided in the output for any t-test based TOST analysis (e.g., Cohen’s d). The Hedges’s corrected SMD (Hedges, 1981) is automatically calculated, but this can be overridden with the bias\_correction argument[[3]](#footnote-29). In previous versions of this package, the equivalence bounds could be set by the SMD (e.g., equivalence bound of 0.5 SD), but this is an erroneous approach since the bound would be dependent upon the *sample* variance. However, users can opt for such an analysis by setting eqbound\_type to SMD, which will produce a noticeable warning to the R console.

The hypothesis argument is automatically set to “EQU” for equivalence, but if a minimal effect is of interest then “MET” can be supplied.

# Formula Interface  
res1 = t\_TOST(formula = extra ~ group, data = sleep,   
 eqb = .5, smd\_ci = "t")  
# x & y Interface  
res1a = t\_TOST(x = subset(sleep,group==1)$extra,  
 y = subset(sleep,group==2)$extra, eqb =.5)

Once the function has run, we can print the results with the print method. This provides a verbose summary of the results.

print(res1)

##   
## Welch Two Sample t-test  
##   
## The equivalence test was non-significant, t(17.78) = -1.3, p = 0.89  
## The null hypothesis test was non-significant, t(17.78) = -1.86p = 0.08  
## NHST: don't reject null significance hypothesis that the effect is equal to zero   
## TOST: don't reject null equivalence hypothesis  
##   
## TOST Results   
## t df p.value  
## t-test -1.861 17.78 0.079  
## TOST Lower -1.272 17.78 0.890  
## TOST Upper -2.450 17.78 0.012  
##   
## Effect Sizes   
## Estimate SE C.I. Conf. Level  
## Raw -1.5800 0.8491 [-3.0534, -0.1066] 0.9  
## Hedges's g(av) -0.7965 0.5992 [-1.8362, 0.2433] 0.9  
## Note: SMD confidence intervals are an approximation. See vignette("SMD\_calcs").

Another nice feature is the generic plot method that can provide a visual summary of the results. Most of the plots in this package were inspired by the [concurve](https://cran.r-project.org/package=concurve) R package (Rafi & Greenland, 2020). There are two types of plots that can be produced. The first, and default, is the consonance density plot (type = "cd").

plot(res1, type = "cd")

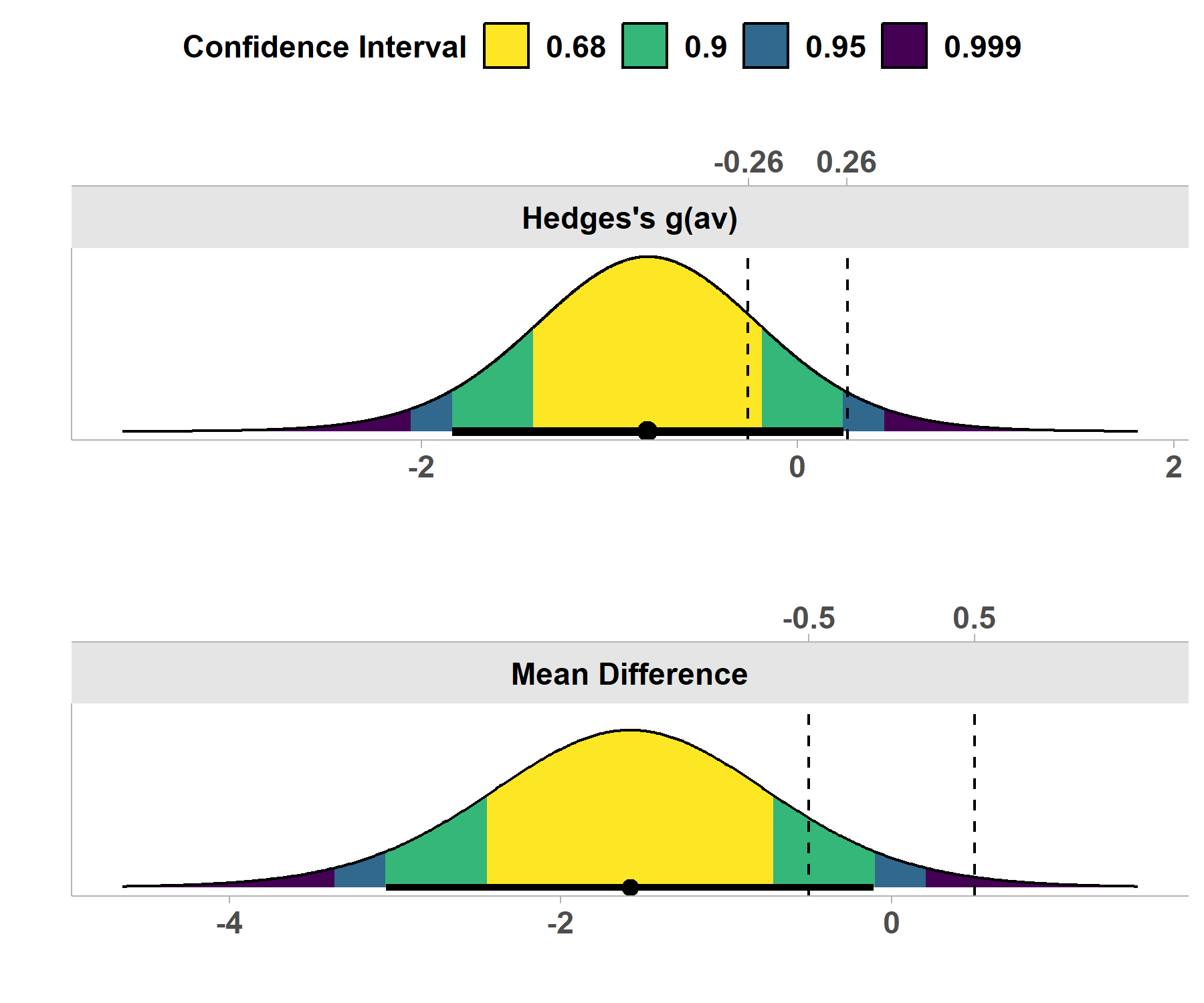


Figure 2: Example of consonance density plot.

The shading pattern can be modified with the ci\_shades.

plot(res1, type = "cd",  
 ci\_shades = c(.9,.95))

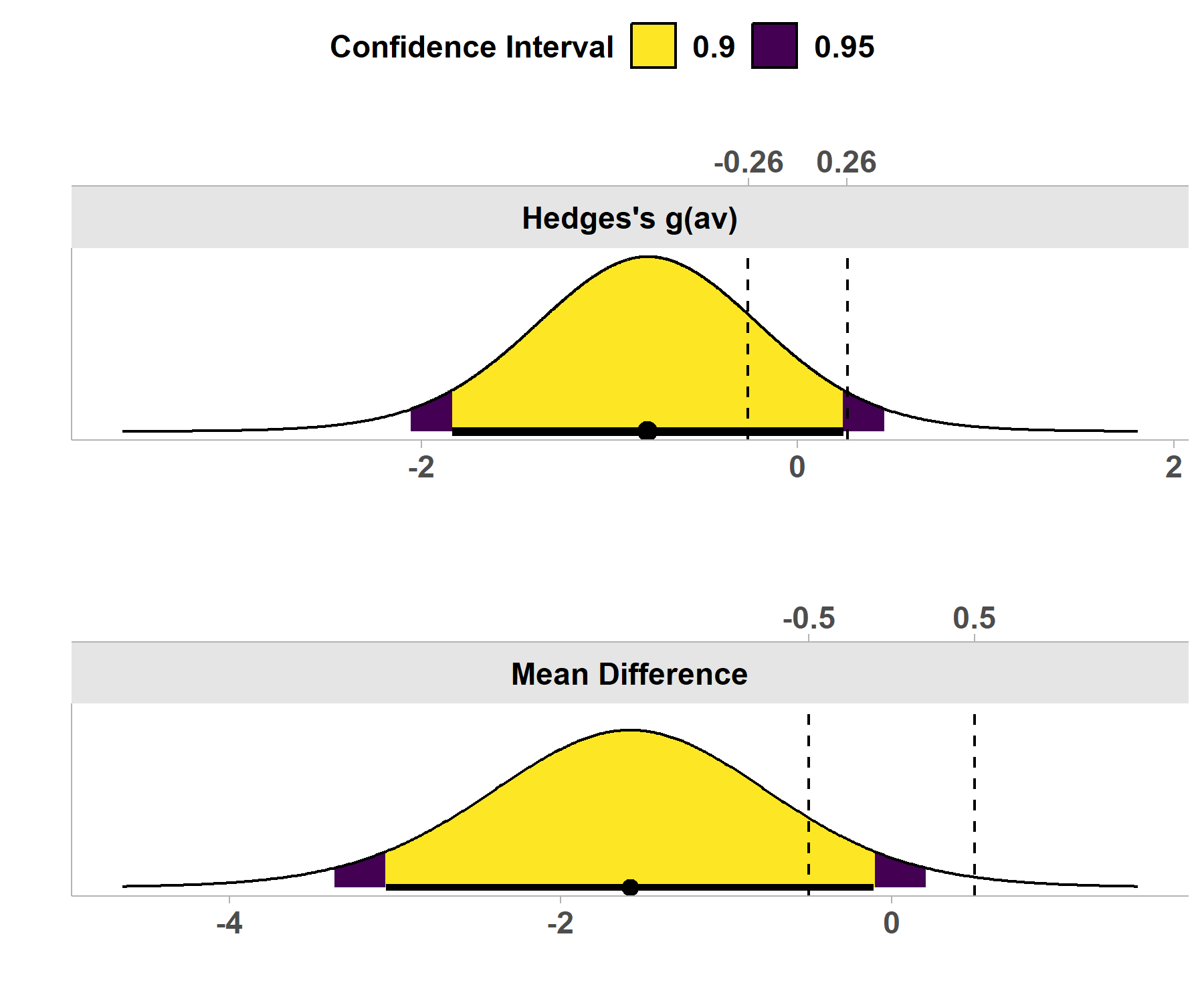


Figure 3: Demonstrating the shading in plot method.

Consonance plots, where all confidence intervals can be simultaneous plotted, can also be produced. The advantage here is multiple confidence interval lines can plotted at once.

plot(res1, type = "c",  
 ci\_lines = c(.9,.95))

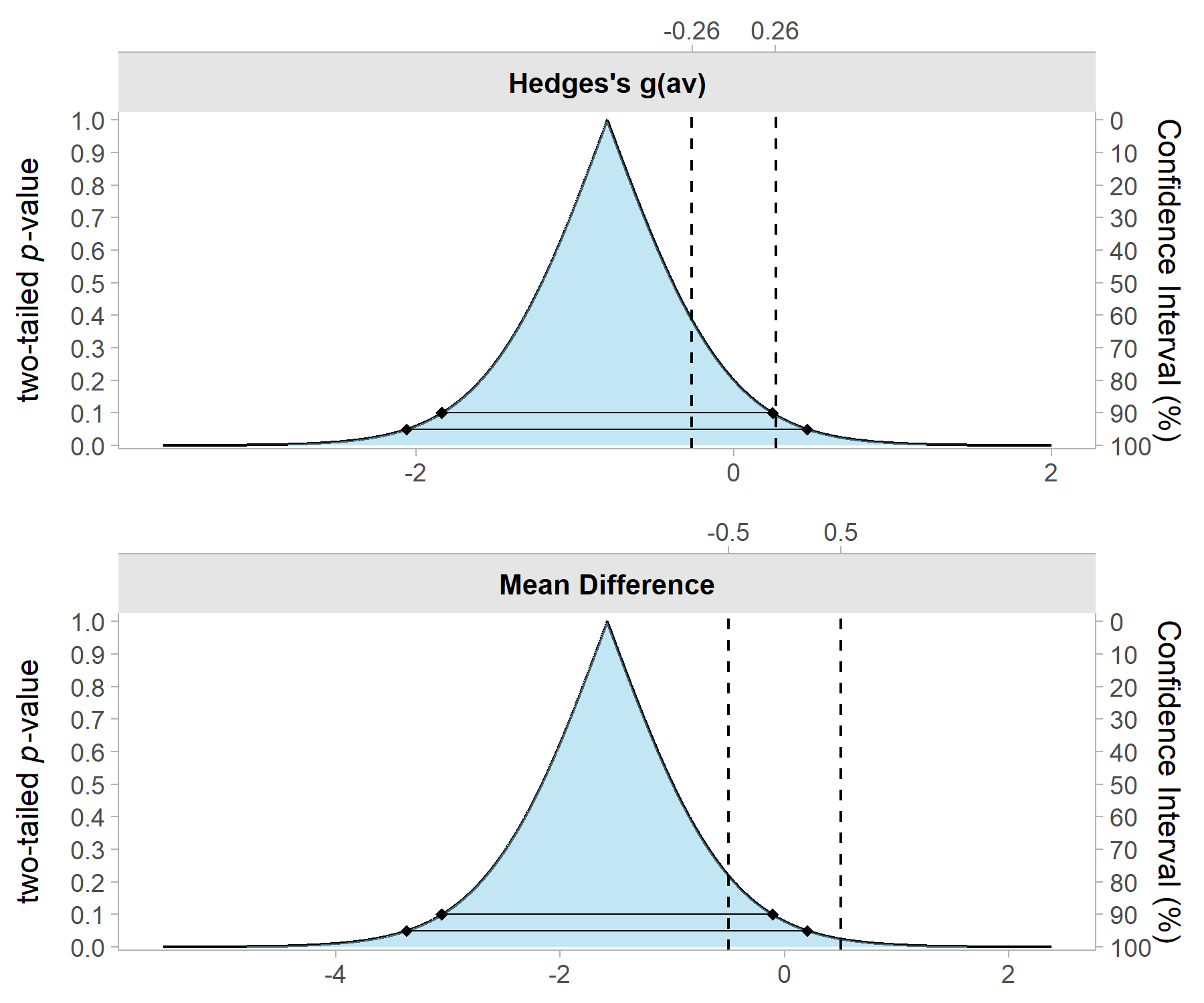


Figure 4: Example of consonance plot.

## Paired Sample

To perform TOST on paired samples, the process does not change much. We could process the test the same way by providing a formula. All we would need to then is change paired to TRUE.

res2 = t\_TOST(formula = extra ~ group,  
 data = sleep,  
 paired = TRUE,  
 eqb = .5)  
res2

##   
## Paired t-test  
##   
## The equivalence test was non-significant, t(9) = -2.8, p = 0.99  
## The null hypothesis test was significant, t(9) = -4.06p < 0.01  
## NHST: reject null significance hypothesis that the effect is equal to zero   
## TOST: don't reject null equivalence hypothesis  
##   
## TOST Results   
## t df p.value  
## t-test -4.062 9 0.003  
## TOST Lower -2.777 9 0.989  
## TOST Upper -5.348 9 < 0.001  
##   
## Effect Sizes   
## Estimate SE C.I. Conf. Level  
## Raw -1.580 0.389 [-2.293, -0.867] 0.9  
## Hedges's g(z) -1.174 0.411 [-1.8046, -0.4977] 0.9  
## Note: SMD confidence intervals are an approximation. See vignette("SMD\_calcs").

However, we may have two vectors of data that are paired. So instead we may want to just provide those separately rather than using a data set and setting the formula. This can be demonstrated with the “bugs” data.

res3 = t\_TOST(x = bugs$LDHF,  
 y = bugs$LDLF,  
 paired = TRUE,  
 eqb = 1)  
res3

##   
## Paired t-test  
##   
## The equivalence test was non-significant, t(90) = 2.66, p = 1  
## The null hypothesis test was significant, t(90) = 6.649p < 0.01  
## NHST: reject null significance hypothesis that the effect is equal to zero   
## TOST: don't reject null equivalence hypothesis  
##   
## TOST Results   
## t df p.value  
## t-test 6.649 90 < 0.001  
## TOST Lower 10.642 90 < 0.001  
## TOST Upper 2.655 90 0.995  
##   
## Effect Sizes   
## Estimate SE C.I. Conf. Level  
## Raw 1.6648 0.2504 [1.2487, 2.081] 0.9  
## Hedges's g(z) 0.6911 0.1167 [0.4987, 0.8802] 0.9  
## Note: SMD confidence intervals are an approximation. See vignette("SMD\_calcs").

Additionally, a MET, instead of equivalence testing, can be performed with the hypothesis argument set to “MET”. With this setting, the hypothesis being tested is whether the effect is *greater* than the equivalence bound.

res3a = t\_TOST(x = bugs$LDHF,  
 y = bugs$LDLF,  
 paired = TRUE,  
 hypothesis = "MET",  
 eqb = 1)  
res3a

##   
## Paired t-test  
##   
## The minimal effect test was significant, t(90) = 10.64, p < 0.01  
## The null hypothesis test was significant, t(90) = 6.649p < 0.01  
## NHST: reject null significance hypothesis that the effect is equal to zero   
## TOST: reject null MET hypothesis  
##   
## TOST Results   
## t df p.value  
## t-test 6.649 90 < 0.001  
## TOST Lower 10.642 90 1  
## TOST Upper 2.655 90 0.005  
##   
## Effect Sizes   
## Estimate SE C.I. Conf. Level  
## Raw 1.6648 0.2504 [1.2487, 2.081] 0.9  
## Hedges's g(z) 0.6911 0.1167 [0.4987, 0.8802] 0.9  
## Note: SMD confidence intervals are an approximation. See vignette("SMD\_calcs").

The data would indicate that we should accept the MET hypothesis.

## One Sample t-test

In other cases we may have a one sample test. If that is the case, only x argument for the data is needed. This is useful in situations where you may have hypotheses to test about a single samples mean. In order for the two-sample test to be correct, we also need to supply the mu argument. In the example below, we hypothesize that the mean of LDHF is not more than 1.5 points greater or less than 7. With the way the mu and eqb arguments are set, we are testing whether the mean of LDHF is significantly different from 7.5 (two-tailed tests) and () than 1.5 points 7.5 as well (equivalence bounds at 5.5 and 8.5).

res4 = t\_TOST(x = bugs$LDHF,  
 hypothesis = "EQU",  
 mu = 7.5,  
 eqb = c(5.5,8.5))  
res4

##   
## One Sample t-test  
##   
## The equivalence test was significant, t(90) = -4.2, p < 0.01  
## The null hypothesis test was non-significant, t(90) = -0.458p = 0.65  
## NHST: don't reject null significance hypothesis that the effect is equal to 7.5   
## TOST: reject null equivalence hypothesis  
##   
## TOST Results   
## t df p.value  
## t-test -0.4577 90 0.648  
## TOST Lower 7.1156 90 < 0.001  
## TOST Upper -4.2444 90 < 0.001  
##   
## Effect Sizes   
## Estimate SE C.I. Conf. Level  
## Raw -0.12088 0.2641 [6.9402, 7.818] 0.9  
## Hedges's g -0.04758 0.1049 [-0.2185, 0.1236] 0.9  
## Note: SMD confidence intervals are an approximation. See vignette("SMD\_calcs").

We would conclude that LDHF is practically equivalent to the hypothesized mean (7.5).

## Using Summary Statistics

In some cases you may only have access to the summary statistics (e.g., when reviewing an article or attempting to perform a meta-analysis). Therefore, I created a function, tsum\_TOST, to perform the same tests just based on the summary statistics. This involves providing the function with a number of different arguments.

* n1 & n2 the sample sizes (only n1 needs to be provided for one sample case)
* m1 & m2 the sample means
* sd1 & sd2 the sample standard deviation
* r12 the correlation between each if paired is set to TRUE[[4]](#footnote-46)

The results from the bugs example can be replicated with the tsum\_TOST:

res\_tsum = tsum\_TOST(  
 m1 = mean(bugs$LDHF, na.rm=TRUE), sd1 = sd(bugs$LDHF, na.rm=TRUE),  
 n1 = length(na.omit(bugs$LDHF)),  
 hypothesis = "EQU", smd\_ci = "t", eqb = c(5.5, 8.5)  
)  
  
res\_tsum

##   
## One-sample t-Test  
##   
## The equivalence test was significant, t(90) = -4.244, p = 2.66e-05  
## The null hypothesis test was significant, t(90) = 27.942, p = 3.91e-46  
## NHST: reject null significance hypothesis that the effect is equal to zero   
## TOST: reject null equivalence hypothesis  
##   
## TOST Results   
## t df p.value  
## t-test 27.942 90 < 0.001  
## TOST Lower 7.116 90 < 0.001  
## TOST Upper -4.244 90 < 0.001  
##   
## Effect Sizes   
## Estimate SE C.I. Conf. Level  
## Raw 7.379 0.2641 [6.9402, 7.818] 0.9  
## Hedges's g 2.905 0.2395 [2.4289, 3.3804] 0.9  
## Note: SMD confidence intervals are an approximation. See vignette("SMD\_calcs").

# Robust Methods for Equivalence Testing

In some cases, the use of t-test may be less than ideal. Any serious violation to the assumptions of a t-test (e.g., normality or homoscedasticity) could greatly inflate the type 1 error rate of TOST. Therefore, it may be useful to explore alternatives to the t-test for TOST that either do not have those assumptions or are robust to violating those assumptions.

The TOSTER package currently provides 4 robust alternatives to the t-test for TOST. First, there is the wilcox\_TOST function which uses the Wilcoxon-Mann-Whitney (WMW) type tests (i.e., wilcox.test) to perform TOST as a test of symmetry. Second, there is the boot\_t\_TOST function which uses the bootstrap method outlined by Efron and Tibshirani (1993). Third, there is the log\_TOST function which performs log-transformed t-tests, which is a parametric approach commonly used in pharmaceutical bioequivalence studies on ratio data (He, Deng, You, & Zhou, 2022). Fourth, there is the boot\_log\_TOST function which uses the same bootstrap method outlined by Efron and Tibshirani (1993) but on the log-transformed data, which is more robust than parametric log t-test (He et al., 2022).

In the following sections, I will briefly outline the available robust TOST functions within the TOSTER package.

## Tests of Symmetry (rank based tests)

The WMW group of tests (e.g., Mann-Whitney U-test) provide a non-parametric test of differences between groups, or within samples, based on *ranks*. This provides a test of location shift, which is a fancy way of saying differences in the center of the distribution (i.e., in parametric tests the location is mean). Within the TOST framework, there are two separate tests of directional location shift to determine if the location shift is within (equivalence) or outside (minimal effect) the equivalence bounds. Many researchers mistakenly think these are tests of medians, but this is not the case (See Divine, Norton, Barón, and Juarez-Colunga (2018) for details). Using a WMW-based TOST is useful for testing whether the differences between groups/conditions is symmetric around the equivalence bounds[[5]](#footnote-49). For equivalence testing, the TOST would be testing whether there is asymmetry towards no effect with a null hypothesis of symmetry at the equivalence bound.

In the TOSTER package, we accomplish this “test of symmetry” with the wilcox\_TOST function. This function operates in an extremely similar implementation to the t\_TOST function. The exact calculations utilized in this function can be explored via the documentation of the wilcox.test function. A standardized mean difference (SMD) is *not* calculated in this function since this would be an inappropriate measure of effect size alongside the non-parametric test statistics. Instead, a standardized effect size (SES) is calculated for *all* types of comparisons (e.g., two sample, one sample, and paired samples). The function can produce a rank-biserial correlation (Kerby, 2014), a WMW Odds (O’Brien & Castelloe, 2006), or a “common language effect size” (Kerby, 2014) (Also known as the non-parametric probability of superiority, or concordance probability).[[6]](#footnote-50)

As an example, we can use the sleep data to make a non-parametric comparison of equivalence.

test1 = wilcox\_TOST(formula = extra ~ group,  
 data = sleep,  
 paired = FALSE,  
 eqb = .5)  
print(test1)

##   
## Wilcoxon rank sum test with continuity correction  
##   
## The equivalence test was non-significant W = 20.000, p = 8.94e-01  
## The null hypothesis test was non-significant W = 25.500, p = 6.93e-02  
## NHST: don't reject null significance hypothesis that the effect is equal to zero   
## TOST: don't reject null equivalence hypothesis  
##   
## TOST Results   
## Test Statistic p.value  
## NHST 25.5 0.069  
## TOST Lower 34.0 0.894  
## TOST Upper 20.0 0.013  
##   
## Effect Sizes   
## Estimate C.I. Conf. Level  
## Median of Differences -1.346 [-3.4, -0.1] 0.9  
## Rank-Biserial Correlation -0.490 [-0.7493, -0.1005] 0.9

Based on these results, we would have conclude there is no significant difference but not equivalent differences either (i.e., inconclusive result).

## Bootstrap TOST

The bootstrap refers to resampling with replacement and can be used for statistical estimation and inference. Bootstrapping techniques are very useful because they are considered somewhat robust to the violations of assumptions for a simple t-test and provide better estimations of SMDs (Kirby & Gerlanc, 2013). Therefore, I added a bootstrapping function, boot\_t\_TOST, to the package to provide another robust alternative to the t\_TOST function.

In this function we provide a percentile bootstrap solution outlined by Efron and Tibshirani (1993) (see chapter 16, page 220). The bootstrapped p-values are derived from the “studentized” version of a test of mean differences (Efron & Tibshirani, 1993). Overall, the results should be similar to the results of t\_TOST. **However**, for paired samples, the Cohen’s d(rm) effect size *cannot* be calculated by this function.

### Two Sample Algorithm.

The steps by which the bootstrapping occurs are fairly simple.

1. Form B bootstrap data sets from x\* and y\* wherein x\* is sampled with replacement from and y\* is sampled with replacement from
2. t is then evaluated on each sample, but the mean of each sample (y or x) and the overall average (z) are subtracted from each (i.e., null distribution is formed)
3. An approximate p-value can then be calculated as the number of bootstrapped results greater than the observed t-statistic from the sample.

The same process is completed for the one sample case but with the one sample solution for the equation outlined by . The paired sample case in this bootstrap procedure is equivalent to the one sample solution because the test is based on the difference scores.

### Example of Bootsrapping.

We can use the sleep data to see an example of the bootstrapped results. If you plot the bootstrap samples, it will show how the resampling via bootstrapping indicates the instability of Hedges’ d(z). Just looking at the printed results you will notice some differences between confidence intervals from the bootstrapped result and the t-test.

set.seed(891111)  
test1 = boot\_t\_TOST(formula = extra ~ group,  
 data = sleep,  
 paired = TRUE,  
 eqb = .5,  
 R = 999)  
  
  
print(test1)

##   
## Bootstrapped Paired t-test  
##   
## The equivalence test was non-significant, t(9) = -2.777, p = 1e+00  
## The null hypothesis test was significant, t(9) = -4.062, p = 0e+00  
## NHST: reject null significance hypothesis that the effect is equal to zero   
## TOST: don't reject null equivalence hypothesis  
##   
## TOST Results   
## t df p.value  
## t-test -4.062 9 < 0.001  
## TOST Lower -2.777 9 1  
## TOST Upper -5.348 9 < 0.001  
##   
## Effect Sizes   
## Estimate SE C.I. Conf. Level  
## Raw -1.580 0.3699 [-2.26, -1.038] 0.9  
## Hedges's g(z) -1.174 0.6491 [-2.7507, -0.9285] 0.9  
## Note: percentile bootstrap method utilized.

## Log TOST

The natural logarithmic (log) transformation is often utilized to stabilize the variance of a measure, and it often provides the best approximation of the normal distribution (Bland & Altman, 1996). However, another, less often reported, advantage of the log transformation is that the back transformation of the differences of the log-transformed data is a *ratio* (Bland & Altman, 1996). For example, if we had a two samples (x & y) with an geometric mean[[7]](#footnote-55) or 7 and 10.5, x and y respectively in the code below, we could represent the differences as ratio of y:x where y is 1.5 times greater than x.

x = 7; y = 10.5  
log(y) - log(x)

## [1] 0.4054651

log(y/x)

## [1] 0.4054651

exp(log(y) - log(x))

## [1] 1.5

y/x

## [1] 1.5

The log transformation thereby acts as a useful tool help tame data into conforming to the normality assumption, and makes the interpretation fairly simple. In addition, some regulatory agencies, such as the United States Food and Drug Administration (FDA) (Food and Drug Administration, 2014), specifically require bioequivalence studies to report the geometric means and make statistical comparisons on the log transformed data (He et al., 2022). In pharmaceutical reserach, bioequivalence testing involves determining whether two drugs, a test drug and a reference drug, have the same rate and extent of absorption in the body. This is typically accomplished by testing whether the blood concentrations of the drug after administration of the test drug are sufficiently close to the blood concentrations after administration of the reference drug. If the two drugs are bioequivalent, they can be used interchangeably. The area under the curve (AUC) is the measure of the extent of absorption, and the peak concentration is the measure of the rate of absorption. In order to determine bioequivalence, the AUC and peak concentration of the test drug must be within a certain percentage of the AUC and peak concentration of the reference drug.

In my personal experience as a physiologist, it is not uncommon that biological/physiological phenomenon present have longer right-tailed distributions, and are often adequately normalized with a natural log transformation. The additional advantage is the how equivalence bounds can, almost, be universally applied when making comparisons on the log scale. The FDA considers to drugs to be bioequivalent when the maximal concentration and AUC differences between drugs are less than 1.25. To put it another way, ratio between two means must be between 1.25 and 0.8 (i.e., 1/1.25) (Food and Drug Administration, 2014).

Therefore, I have implemented two functions to allow for the comparison of data that is believed to be left skewed (long right tail), and is on a ratio scale[[8]](#footnote-56). The first function is a parametric t-test on the log transformed scale while the second function is a bootstrapping test which is more robust than parametric version (He et al., 2022).

### Example of Log TOST.

The log\_TOST function is almost exactly the same as the t\_TOST function. First, the primary differences is that it only accepts paired and two sample comparisons. One sample tests are not support (i.e., there is no ratio to calculate). Second, standardized mean differences are not calculated, but a ratio of means is instead reported (Lajeunesse, 2015)[[9]](#footnote-57). Third, the default equivalence bounds are by default set to the FDA standards (i.e., eqb = 1.25), but can be changed by the user[[10]](#footnote-58).

As an example we can use the mtcars data to compare the type of transmission (am) effects on the gas mileage (mpg). We can see from the data below there are significant, non-equivalent, differences in mpg between transmission types.

log\_TOST(mpg ~ am, data = mtcars)

##   
## Log-transformed Welch Two Sample t-test  
##   
## The equivalence test was non-significant, t(23.96) = -1.363, p = 9.07e-01  
## The null hypothesis test was significant, t(23.96) = -3.826, p = 8.19e-04  
## NHST: reject null significance hypothesis that the effect is equal to one   
## TOST: don't reject null equivalence hypothesis  
##   
## TOST Results   
## t df p.value  
## t-test -3.826 23.96 < 0.001  
## TOST Lower -1.363 23.96 0.907  
## TOST Upper -6.288 23.96 < 0.001  
##   
## Effect Sizes   
## Estimate SE C.I. Conf. Level  
## log(Means Ratio) -0.3466 0.09061 [-0.5017, -0.1916] 0.9  
## Means Ratio 0.7071 NA [0.6055, 0.8256] 0.9

### Example of Bootstrap Log TOST.

The bootstrap version of log\_TOST, boot\_log\_TOST, uses the same bootstrapping method detailed above (boot\_t\_TOST), but it uses the log-transformed values and produces the ratio of means as the effect size.

boot\_log\_TOST(mpg ~ am, data = mtcars, R=999)

##   
## Bootstrapped Log Welch Two Sample t-test  
##   
## The equivalence test was non-significant, t(23.96) = -1.363, p = 9.57e-01  
## The null hypothesis test was significant, t(23.96) = -3.826, p = 0e+00  
## NHST: reject null significance hypothesis that the effect is equal to 1   
## TOST: don't reject null equivalence hypothesis  
##   
## TOST Results   
## t df p.value  
## t-test -3.826 23.96 < 0.001  
## TOST Lower -1.363 23.96 0.957  
## TOST Upper -6.288 23.96 < 0.001  
##   
## Effect Sizes   
## Estimate SE C.I. Conf. Level  
## log(Means Ratio) -0.3466 0.08634 [-0.4871, -0.2039] 0.9  
## Means Ratio 0.7071 0.06119 [0.6144, 0.8156] 0.9  
## Note: percentile bootstrap method utilized.

From this analysis, we would conclude there is a significant effect that is not practically equivalent.

# Equivalence Testing with ANOVAs

Many researchers utilize ANOVA as an omnibus test for the absence/presence of effects before inspecting multiple pairwise comparisons. This is very useful when implementing factorial designs wherein multiple experimental factors are tested and/or manipulated. As Campbell and Lakens (2021) suggest, the lack of a significant result at the ANOVA-level does not necessarily indicate that a factor or interaction of factors have no effect. However, Campbell and Lakens (2021) only suggest an equivalence test for one-way ANOVAs and therefore exclude multi-factor or factorial ANOVAs. Therefore, I have extended the work of Campbell and Lakens (2021) to include functions that allow for equivalence testing of the partial (eta-squared) effect size from ANOVAs.

## F-test Calculations

Statistical equivalence testing[[11]](#footnote-63) for *F*-tests are special use case of the cumulative distribution function of the non-central *F* distribution. As Campbell and Lakens (2021) states, this type of statistical test answers the question: “Can we reject the hypothesis that the total proportion of variance in outcome Y attributable to X is greater than or equal to the equivalence bound ?”

### Hypothesis Tests.

In TOSTER, I have gone a tad farther than Campbell and Lakens (2021), and have included a calculation for a generalization of the non-centrality parameter that allows the equivalence test for *F*-tests to be applied to variety of designs.

Campbell and Lakens (2021) calculate the *p*-value as:

The non-centrality parameter (ncp = ) can be calculated with the equivalence bound and the degrees of freedom:

The *p*-value for the equivalence test () could then be calculated from traditional ANOVA results and the distribution function:

## Example of Equivalence ANOVA Testing

Using the InsectSprays data set in R and the base R aov function, I can demonstrate how this omnibus equivalence testing can be applied with TOSTER. From the initial analysis we an see a clear “significant” effect (very small p-value) of the inspect spray. However, we *may* be interested in testing if the effect is practically equivalent. I will arbitrarily set the equivalence bound to a partial eta-squared of 0.35 ().

data("InsectSprays")  
aovtest = aov(count ~ spray, data = InsectSprays)  
anova(aovtest)

## Analysis of Variance Table  
##   
## Response: count  
## Df Sum Sq Mean Sq F value Pr(>F)   
## spray 5 2668.8 533.77 34.702 < 2.2e-16 \*\*\*  
## Residuals 66 1015.2 15.38   
## ---  
## Signif. codes: 0 '\*\*\*' 0.001 '\*\*' 0.01 '\*' 0.05 '.' 0.1 ' ' 1

We can then use the information in the table above to perform an equivalence test using the equ\_ftest function. This function returns an object of the S3 class htest and the output will look very familiar to that of the t-test. The main difference is the estimates, and confidence interval, are for partial .

equ\_ftest(Fstat = 34.70228, df1 = 5, df2 = 66, eqb = 0.35)

##   
## Equivalence Test from F-test  
##   
## data: Summary Statistics  
## F = 34.702, df1 = 5, df2 = 66, p-value = 1  
## 95 percent confidence interval:  
## 0.5806263 0.7804439  
## sample estimates:  
## [1] 0.724439

Based on the results above we would conclude there is a significant effect of “spray” and the differences due to spray are *not* statistically equivalent. In essence, we reject the traditional null hypothesis of “no effect” but accept the null hypothesis of the equivalence test.

The equ\_ftest function is very useful because all you need is very basic summary statistics. However, if you are doing all your analyses in R then you can use the equ\_anova function. This function accepts objects produced from stats::aov, car::Anova and afex::aov\_car (or any ANOVA from derived from afex).

As a second example, we can use the afex package’s data and ANOVA (Singmann, Bolker, Westfall, Aust, & Ben-Shachar, 2022). Again, we will use the equivalence bound of 0.35, which is a completely arbitrary (and baseless) equivalence bound. Notice that the output contains 2 p-values: one for the significance (p.null) and another for the equivalence test (p.equ).

# Example using a purely within-subjects design   
# (Maxwell & Delaney, 2004, Chapter 12, Table 12.5, p. 578):  
library(afex)  
data(md\_12.1)  
aovtest2 = aov\_ez("id", "rt", md\_12.1, within = c("angle", "noise"),   
 anova\_table=list(correction = "none", es = "none"))  
equ\_anova(aovtest2,  
 eqb = 0.35)

## effect df1 df2 F.value p.null pes eqbound p.equ  
## 1 (Intercept) 1 9 598.44917 1.526600e-09 0.9851839 0.35 0.9999997  
## 2 angle 2 18 40.71910 2.086763e-07 0.8189831 0.35 0.9992557  
## 3 noise 1 9 33.76596 2.559737e-04 0.7895522 0.35 0.9763228  
## 4 angle:noise 2 18 45.31034 9.424093e-08 0.8342857 0.35 0.9996103

# Equivalence Between Replication Studies

During the development of this TOSTER update, I was helping advise a team of researchers on a massive replication project for sport and exercise science (Murphy, Mesquida, Caldwell, Earp, & Warne, 2022). How to determine whether a direct[[12]](#footnote-68) replication was a successful replication of the original study was contentious topic of conversation among the team. Inspired by these discussions, I created 2 functions that would utilize the basic principles of SMDs[[13]](#footnote-69) to test for differences between two studies.

Overall, the concept is simple: if we have estimates of SMDs from two very similar studies we can use the large-sample approximation to compute the sampling variances[[14]](#footnote-70) to estimate the degree to which the two studies differ from one another (i.e., calculate p-values). The users of TOSTER then have the option to test whether the two SMDs significantly differ, or use TOST to estimate if they are practically equivalent. Additionally, there are two options for comparing SMDs: using the summary statistics or using bootstrapping (assuming original data is available).

## Example using Summary Statistics

In this example, let us imagine an “original” study that reports an effect of Cohen’s dz = 0.95 in a paired samples design with 25 subjects. However, a replication doubled the sample size, found a non-significant effect at an SMD of 0.2. Are these two studies compatible (the lower the p-value the lower the compatibility)? Or, to put it another way, should the replication be considered a “failure” to replicate the original study?

We can use the compare\_smd function to at least measure how often we would expect a discrepancy between the original and replication study if the same underlying effect was being measured (also assuming no publication bias).

We can see from the results below that, if the null hypothesis were true, we would only expect to see a discrepancy in SMDs between studies at least this large ~1% of the time.

compare\_smd(smd1 = 0.95,  
 n1 = 25,  
 smd2 = 0.23,  
 n2 = 50,  
 paired = TRUE)

##   
## Difference in Cohen's dz (paired)  
##   
## data: Summary Statistics  
## z = 2.5685, p-value = 0.01021  
## alternative hypothesis: true difference in SMDs is not equal to 0  
## sample estimates:  
## difference in SMDs   
## 0.72

Let us also imagine a scenario where a replication team considers a replication successful if the SMDs are within 0.25 units of each other. We can set the TOST argument to TRUE, and then set the equivalence bound using null argument.

compare\_smd(smd1 = 0.95, n1 = 25, smd2 = 0.23,n2 = 50,  
 paired = TRUE, TOST = TRUE, null = .25)

##   
## Difference in Cohen's dz (paired)  
##   
## data: Summary Statistics  
## z = 1.6767, p-value = 0.9532  
## alternative hypothesis: equivalence  
## null values:  
## difference in SMDs difference in SMDs   
## 0.25 -0.25   
## sample estimates:  
## difference in SMDs   
## 0.72

Based on the imaginary studies we outlined above, we would not reject the null equivalence hypothesis, but reject the null significance hypothesis. Therefore, we would could conclude that there are significant differences between the studies that are not practically equivalent.

## Example using Bootstrapping

The above results are only based on an approximating the differences between the SMDs. If the raw data is available, then the optimal solution is the bootstrap. This can be accomplished with the boot\_compare\_smd function. The only drawback to this function is that TOST is currently not avaiable, and users would instead have to run 2 one-sided tests manually using the null and alternative arguments.

For this example, we will simulate some data. As an alternative approach to TOST, we can just set the alpha to 0.1, and then check to see if the confidence interval is within the preset equivalence bounds.

set.seed(4522)  
boot\_test = boot\_compare\_smd(x1 = rnorm(25,.95), x2 = rnorm(50),   
 paired = TRUE, alpha = .1)  
boot\_test

##   
## Bootstrapped Differences in SMDs (paired)  
##   
## data: Bootstrapped  
## z (observed) = 2.887, p-value = 0.006003  
## alternative hypothesis: true difference in SMDs is not equal to 0  
## 90 percent confidence interval:  
## 0.4070761 1.3508435  
## sample estimates:  
## difference in SMDs   
## 0.8058872

# Conclusions

In this manuscript I have demonstrated most of the new functions and features within the TOSTER R package. This constitutes a major update to the package over the past 2 years. I hope that updates to the package builds upon the original impact of the TOSTER package[[15]](#footnote-74), and has been made TOST more accessible to the average researcher. In addition, I have added a number of other functions that offer robust alternatives to the t-test for performing TOST analyses. I would strongly recommend users of TOSTER to explore these functions, and, at the very least, compare the robust results to the t-test results to ensure that the conclusions do not change due to the chosen analysis[[16]](#footnote-75). Lastly, to my knowledge, this is the first package to offer equivalence testing options for ANOVAs or for comparing SMDs between studies. Overall, this package and its functions offer an easily accessible option for researchers to explore equivalence testing, and hopefully improve their statistical analyses.

# Additional Information

All analyses/code in this manuscript are from TOSTER v0.6.0:

# Install the exact release with this code  
devtools::install\_github("Lakens/TOSTER@v0.6.0")

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## Notes on contributor(s)

Daniel Lakens provided a review of many of the materials that have been incorporated into the update of TOSTER, and was the original author of this package. Without his help and encouragment, the TOSTER package and this update would not exist.

## Nomenclature/Notation

* ANOVA: Analysis of Variance
* Bootstrapping: the use of random sampling with replacement to estimate statistics
* FDA: Food and Drug Administration (United States of America)
* MET: Minimal Effects Test
* ncp: non-centrality parameter
* SESOI: Smallest Effect Size of Interest
* SMD: Standardized Mean Difference (e.g., Cohen’s d)
* TOST: Two-One Sided Tests
* WMW: Wilcoxon-Mann-Whitney

## Notes

The R package is also (partially) implemented in jamovi as the TOSTER module.

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1. All updates to the package can be found on the package’s website <https://aaroncaldwell.us/TOSTERpkg> [↑](#footnote-ref-20)
2. I strongly recommend users “justify their alpha” (Lakens, D., et al, 2018; Maier & Lakens, 2022), and the justification process can be aided by my other R package [Superpower](https://aaroncaldwell.us/Superpower) [↑](#footnote-ref-27)
3. Glass’s delta can also be produced in the output by using the glass argument [↑](#footnote-ref-29)
4. The extract\_r\_paired function can be used if the correlation between paired observations is not readily available. [↑](#footnote-ref-46)
5. Care should be taken when considering paired samples; a test on the rank transformed data (Kornbrot, 1990) or another robust test may be more prudent. [↑](#footnote-ref-49)
6. There is no plotting capability at this time for the output of this function. [↑](#footnote-ref-50)
7. The mean of log-transformed data is the *geometric* not *arithmetic* mean. I highly recommend reading Bland and Altman (1996) and Caldwell and Cheuvront (2019) for more details [↑](#footnote-ref-55)
8. Ratio scale means the outcome is measured on a numerical scale that has equal distances between adjacent values and true zero. [↑](#footnote-ref-56)
9. Also, referred to as a “response ratio” in ecology. Like an SMD, the response ratio can be utilized in meta-analysis. [↑](#footnote-ref-57)
10. Only one value needs to be supplied to eqb; the reciprocal value of eqb is taken as the other equivalence bound. For example, if eqb = 0.85 then the upper equivalence bound is 1/0.85 (~1.333) [↑](#footnote-ref-58)
11. Also called “omnibus non-inferiority testing” by Campbell and Lakens (2021) [↑](#footnote-ref-63)
12. Defined as being a as-close-as possible replication to the original study, in contrast to “conceptual” replications. [↑](#footnote-ref-68)
13. The textbook by Borenstein, Hedges, Higgins, and Rothstein (2021) and the some of the works of Wolfgang Vietchbauer, metafor R package author, were a large source of information for developing these functions. [↑](#footnote-ref-69)
14. Users can also supply their own sampling variances using the se1 and se2 arguments. [↑](#footnote-ref-70)
15. In my opinion, the impact of the Lakens (2017) cannot be overstated considering it is cited by over 1000 other papers! [↑](#footnote-ref-74)
16. If they do change, then it would be prudent to explore what features in the data might explain this discrepancy. [↑](#footnote-ref-75)