

# Package ‘LargeMetabo’

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**Type** Package

**Version** 0.1.0

**Title** an out-of-the-box tool for processing and analyzing large-scale metabolomic data

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**Depends** R ( $\geq 3.0$ ), dynamicTreeCut ( $\geq 1.62$ ), fastcluster

**Imports** genefilter, d3heatmap, ggplot2, ggfortify, factoextra, igraph, FSelector, varSelRF, corrplot, mixOmics, ropls, CluMSID, siggenes, e1071, SOMbrero

**Suggests** rmarkdown, knitr, hexbin, shinycssloaders, shinyBS, shinyjs, shinyjqui

**Description** This package for processing and analyzing large-scale metabolomic data, including data combination for metabolomics data, biomarker selection, enhanced metabolite annotation and enrichment analysis.

**License** MIT + file LICENSE

**URL** <https://github.com/LargeMetabo/LargeMetabo>

**RoxygenNote** 7.1.2

**NeedsCompilation** no

**LazyData** true

## R topics documented:

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Integrate_Data	Integrating multiple datasets
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## Description

Integrating multiple datasets from different analytical experiments

## Usage

Integrate\_Data(mutile\_Group, RT\_Tolerance\_1 = 10, mz\_Tolerance\_1 = 0.1, RT\_Tolerance\_2 = 10, mz\_Tolerance\_2 = 0.1)

## Arguments

mutile_Group	Multiple datasets from multiple groups for integrating as a comprehensive dataset.
RT_Tolerance_1	Set the tolerance of retention time for data integration in the primary phase.
mz_Tolerance_1	Set the tolerance of mass-to-charge ratio for data integration in the primary phase.

RT_Tolerance_2	Set the tolerance of retention time for data integration in the secondary phase.
mz_Tolerance_2	Set the tolerance of mass-to-charge ratio for data integration in the secondary phase.

### Value

A matrix of a comprehensive dataset integrated from multiple groups is returned.

### Author(s)

Qingxia Yang <yangqx@cqu.edu.cn>

### Examples

```
Integrate_Data(mutile_Group, RT_Tolerance_1 = 10, mz_Tolerance_1 = 0.1, RT_Tolerance_2 = 10, mz_Tolerance_2 = 0.1)
```

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Removal_Batch	Remove batch effects for the integrated dataset
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### Description

Remove batch effects among different analytical experiments for the integrated dataset

### Usage

```
Removal_Batch(mutile_align, n = 3, algorithm = "BMC/PAMR")
```

### Arguments

mutile_align	The comprehensive dataset integrated from multiple groups.
n	The number of multiple groups.
algorithm	The algorithm for removing batch effects in multiple groups. The algorithm could be BMC/PAMR, ComBat/EB, GlobalNorm, Log2 or None.

### Value

A matrix of a dataset after removing batch effects for the integrated dataset.

### Author(s)

Qingxia Yang <yangqx@cqu.edu.cn>

## Examples

```
Removal_Batch(mutile_align, n = 3, algorithm = "BMC/PAMR")
```

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Sample_Separation	Plot of sample separation
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## Description

Plot of sample separation for visualizing the clustering and separation of different samples

## Usage

```
Sample_Separation(finalData, finalLabel, clusters=2, method = "HCA")
```

## Arguments

finalData	The matrix of dataset for sample separation.
finalLabel	The label of dataset for sample separation.
clusters	The number of cluster for sample separation.
method	The method for sample separation. The method could be HCA, KMC, PCA and SOM.

## Value

The plot of sample separation.

## Author(s)

Qingxia Yang <yangqx@cqu.edu.cn>

## Examples

```
finalData <- MarkerData$finalData
```

```
finalLabel <- MarkerData$finalLabel
```

```
Sample_Separation(finalData, finalLabel, clusters=2, method = "HCA")
```

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Marker_Identify	Marker identification in large-scale metabolomics data
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### **Description**

Marker identification in large-scale metabolomics data

### **Usage**

Marker\_Identify(finalData, finalLabel, method = "FC")

### **Arguments**

finalData            The matrix of dataset for marker identification.

finalLabel           The label of dataset for marker identification.

method               The method for marker identification. The method could be FC, PLS-DA, OPLS-DA, t-test, CHIS, CFS, ENTROPY, LMEB, RELIEF, RF, SAM, SVMRFE or WRST.

### **Value**

The result of marker identification for metabolomics data.

### **Author(s)**

Qingxia Yang <yangqx@cqu.edu.cn>

### **Examples**

```
finalData <- MarkerData$finalData
```

```
finalLabel <- MarkerData$finalLabel
```

```
Marker_Identify(finalData, finalLabel, method = "FC")
```

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Marker_Assess	Assessing the markers of different methods by classification
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### **Description**

Classification assessment for the markers identified using different marker identification methods in large-scale metabolomics data

### **Usage**

Marker\_Identify(finalData, finalLabel, method = "PLS-DA")

### Arguments

finalData	The matrix of dataset for marker identification.
finalLabel	The label of dataset for marker identification.
method	The method for marker identification. The method could be FC, PLS-DA, OPLS-DA, t-test, CHIS, CFS, ENTROPY, LMEB, RELIEF, RF, SAM, SVMRFE or WRST.

### Value

The ROC curve and AUC value in the SVM classification using markers.

### Author(s)

Qingxia Yang <yangqx@cqu.edu.cn>

### Examples

```
finalData <- MarkerData$finalData
```

```
finalLabel <- MarkerData$finalLabel
```

```
Marker_Assess(finalData, finalLabel, method = "PLS-DA")
```

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MetaboAnnotation

Metabolite annotation for MS1 spectra

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### Description

Metabolite annotation for MS1 spectra based on a metabolite database

### Usage

```
MetaboAnnotation(AnnotaData, masstole = 10, toleUnit = 1, annotaDB = "metlin", ionMode = "pos")
```

### Arguments

AnnotaData	The value of mass-to-charge ratio for metabolite annotation.
masstole	The tolerance of m/z for metabolite annotation.

toleUnit	The unit of m/z for metabolite annotation. The unit could be 1 for Da, or 2 for ppm.
annotaDB	The database of m/z for metabolite annotation. The database could be metlin or hmdb.
ionMode	The mode of m/z for annotation. The mode could be pos, neg or neu.

### Value

The list of results of metabolite annotation for different charges for primary mass spectrometry.

### Author(s)

Qingxia Yang <yangqx@cqu.edu.cn>

### Examples

```
AnnotaMS <- AnnotaData$AnnotaMS
```

```
MetaboA_res <- MetaboAnnotation(AnnotaMS)
```

```
MetaboA_res$`M+H-2H2O`
```

---

AnnotaTandem	Parameters of metabolite annotation for MS/MS
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### Description

Parameters of metabolite annotation of tandem mass spectrum (MS/MS)

### Usage

```
AnnotaTandem(Parent_mass = 181.04, TandemData, massTandem = 0.1, toleUnitTandem = 1,
massmzTandem = 0.5, toleUnitmzTandem = 1, ModeTandem = "Positive", ionEnergy =
"low(10V)")
```

### Arguments

Parent_mass	The value of parent ion mass for metabolite annotation of tandem mass spectrum.
TandemData	The value of MS/MS peak list (m/z & Intensity) for metabolite annotation of tandem mass spectrum.

massTandem	The tolerance of parent ion mass for metabolite annotation of tandem mass spectrum.
toleUnitTandem	The unit of parent ion mass for metabolite annotation of tandem mass spectrum. The unit could be 1 for Da, or 2 for ppm.
massmzTandem	The tolerance of MS/MS peak mass for metabolite annotation of tandem mass spectrum.
toleUnitmzTandem	The unit of MS/MS peak mass for metabolite annotation of tandem mass spectrum. The unit could be 1 for Da, or 2 for ppm.
ModeTandem	The ionization mode for metabolite annotation of tandem mass spectrum. The mode could be Positive or Negative.
ionEnergy	The CID energy for metabolite annotation of tandem mass spectrum. The mode could be low(10V) Medium(25V) High(40V) or All.

### Value

The parameters of metabolite annotation of tandem mass spectrum (MS/MS).

### Author(s)

Qingxia Yang <yangqx@cqu.edu.cn>

### Examples

```
Parent_mass <- AnnotaData$Parent_mass
```

```
TandemData <- AnnotaData$TandomData
```

```
AnnotaParamTandem <- AnnotaTandem(Parent_mass, TandemData)
```

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annotaDataTandem	Table of metabolite annotation for MS/MS
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### Description

The table of results for metabolite annotation of tandem mass spectrum (MS/MS).

### Usage

```
annotaDataTandem(AnnotaParamTandem)
```



## Arguments

AnnotaParamTandem      The parameters of metabolite annotation of tandem mass spectrum.

## Value

The table of results for metabolite annotation of tandem mass spectrum.

## Author(s)

Qingxia Yang <yangqx@cqu.edu.cn>

## Examples

```
Parent_mass <- AnnotaData$Parent_mass
```

```
TandemData <- AnnotaData$TandomData
```

```
AnnotaParamTandem <- AnnotaTandem(Parent_mass, TandemData)
```

```
annotaDataTandem(AnnotaParamTandem)
```

---

AnnotaTandem_plot	Plot of metabolite annotation of MS/MS
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## Description

The plot of results for metabolite annotation of tandem mass spectrum (MS/MS).

## Usage

```
AnnotaTandem_plot(AnnotaParamTandem, TandemData)
```

## Arguments

AnnotaParamTandem      The parameters of metabolite annotation of tandem mass spectrum.

TandemData              The value of MS/MS peak list (m/z & Intensity) for metabolite annotation of tandem mass spectrum.

## Value

The plot of results for metabolite annotation of tandem mass spectrum.

## Author(s)

Qingxia Yang <yangqx@cqu.edu.cn>

## Examples

```
Parent_mass <- AnnotaData$Parent_mass
```

```
TandemData <- AnnotaData$TandomData
```

```
AnnotaParamTandem <- AnnotaTandem(Parent_mass, TandemData)
```

```
AnnotaTandem_plot(AnnotaParamTandem, TandemData)
```

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## KEGGENrichPlotPanel

Parameters for metabolite enrichment

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## Description

The parameters for metabolite enrichment based on the metabolite database.

## Usage

```
KEGGENrichPlotPanel(sampleData, enrichDB = "kegg", pvalcutoff = 0.05, IDtype = 1, cateIdx = 1)
```

## Arguments

sampleData	The charactor of input for metabolite enrichment.
enrichDB	The database of the input data. The database could be kegg, smpdb, cfam, foodb, biofunc, tcm, spectax or toxin.
pvalcutoff	The cutoff of p value for metabolite enrichment.
IDtype	The number for name type for metabolite enrichment, such as KEGG ID, CAS ID, PubChem ID, Name or HMDB ID.
cateIdx	The number of category for metabolite enrichment.

## Value

The parameters for metabolite enrichment.

## Author(s)

Qingxia Yang <yangqx@cqu.edu.cn>

## Examples

```
sampleDatakegg <- EnrichData$sampleDatakegg
```

```
KEGGENrichPlotPanel(sampleDatakegg, enrichDB = "kegg", pvalcutoff = 0.05, IDtype = 1,  
cateIdx = 1)
```

---

Enrichment

Table for metabolite enrichment

---

### Description

The table for metabolite enrichment.

### Usage

```
Enrichment(param)
```

### Arguments

param                      The parameters for metabolite enrichment.

### Value

The table of results for metabolite enrichment.

### Author(s)

Qingxia Yang <yangqx@cqu.edu.cn>

### Examples

```
sampleDatakegg <- EnrichData$sampleDatakegg
```

```
EnrichParam <- KEGGENrichPlotPanel(sampleDatakegg, enrichDB = "kegg", pvalcutoff = 0.05,  
IDtype = 1, cateIdx = 1)
```

```
EnrichResultList <- Enrichment(EnrichParam); EnrichResultList$Table.Result
```

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KEGGENrichPlot

Plot of metabolite enrichment using KEGG database

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### Description

The plot of metabolite enrichment using KEGG database.

### Usage

```
KEGGENrichPlot(EnrichResultList, cpdID, cpdFC)
```

## Arguments

**EnrichResultList**      The table of results for metabolite enrichment using KEGG database.

**cpdID**                      The character of input for metabolite enrichment.

**cpdFC**                      The distance in the metabolite enrichment plot using KEGG database.

## Value

The plot of results for metabolite enrichment using KEGG database.

## Author(s)

Qingxia Yang <yangqx@cqu.edu.cn>

## Examples

```
sampleDatakegg <- EnrichData$sampleDatakegg

EnrichParam <- KEGGEnrichPlotPanel(sampleDatakegg, enrichDB = "kegg", pvalcutoff = 0.05,
IDtype = 1, cateIdx = 1)

EnrichResultList <- Enrichment(EnrichParam)

EnrichFC <- seq(from = -2, to = 2, length.out = 24)

KEGGEnrichPlot(EnrichResultList = EnrichResultList, cpdID = sampleDatakegg, cpdFC =
EnrichFC)
```

---

EnrichPlot	Plot of results for metabolite enrichment
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## Description

The plot of results for metabolite enrichment.

## Usage

```
EnrichPlot(db, EnrichResultList)
```

## Arguments

**db**                              The database of input for metabolite enrichment.

**EnrichResultList**      The table of results for metabolite enrichment.

**Value**

The plot of results for metabolite enrichment.

**Author(s)**

Qingxia Yang <yangqx@cqu.edu.cn>

**Examples**

```
sampleDataacas <- EnrichData$sampleDataacas
```

```
enrichDB <- EnrichData$enrichDB
```

```
EnrichParam <- KEGGEnrichPlotPanel(sampleDataacas, enrichDB = enrichDB, pvalcutoff =  
0.05, IDtype = 2, cateIdx = 1)
```

```
EnrichResultList <- Enrichment(EnrichParam)
```

```
dbChoice <- enrichDB
```

```
EnrichPlot(dbChoice, EnrichResultList)
```