

# Class09: Structural Bioinformatics Part 1

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## The PDB Database

The main database for structural biology is called the PDB (Protein Data Bank). Let's have a look at what it contains:

Download a CSV file from the PDB site (accessible from “Analyze” > “PDB Statistics” > “by Experimental Method and Molecular Type”. Move this CSV file into your RStudio project and use it to answer the following questions:

```
pdbtable <- read.csv("DataExportSummary.csv")  
pdbtable
```

	Molecular.Type	X.ray	EM	NMR	Integrative	Multiple.methods
1	Protein (only)	176,204	20,299	12,708	342	218
2	Protein/Oligosaccharide	10,279	3,385	34	8	11
3	Protein/NA	9,007	5,897	287	24	7
4	Nucleic acid (only)	3,066	200	1,553	2	15
5	Other	173	13	33	3	0
6	Oligosaccharide (only)	11	0	6	0	1
	Neutron	Other	Total			
1	83	32	209,886			
2	1	0	13,718			
3	0	0	15,222			
4	3	1	4,840			
5	0	0	222			
6	0	4	22			

```
 pdbtable$Total
```

```
[1] "209,886" "13,718" "15,222" "4,840" "222" "22"
```

```
# sum(pdbtable[, "X.ray"]) not working because they are characters, not numeric.  
pdbtable$Total
```

```
[1] "209,886" "13,718" "15,222" "4,840" "222" "22"
```

```
sub(", ", "---", pdbtable$Total)
```

```
[1] "209---886" "13---718" "15---222" "4---840" "222" "22"
```

```
library(readr)  
stats <- read_csv("DataExportSummary.csv")  
stats
```

```
# A tibble: 6 x 9  
#> #> `Molecular Type` `X-ray`   EM    NMR Integrative `Multiple methods` Neutron  
#> #> <chr>       <dbl> <dbl> <dbl>      <dbl>           <dbl>     <dbl>  
#> 1 Protein (only) 176204 20299 12708      342          218      83  
#> 2 Protein/Oligosacch~ 10279  3385   34          8          11       1  
#> 3 Protein/NA        9007   5897   287         24          7       0  
#> 4 Nucleic acid (only) 3066   200    1553        2          15      3  
#> 5 Other              173    13     33          3          0       0  
#> 6 Oligosaccharide (o~ 11     0      6          0          1       0  
#> # i 2 more variables: Other <dbl>, Total <dbl>
```

```
stats$Total
```

```
[1] 209886 13718 15222 4840 222 22
```

Q1: What percentage of structures in the PDB are solved by X-Ray and Electron Microscopy. Give to 2 sig figs.

```
n.xray <- sum(stats$`X-ray`) #Add `` in X-ray  
n.total <- sum(stats$Total)  
round(n.xray/n.total*100, 2) # Use round(number,2)
```

```
[1] 81.48
```

```
n.em <- sum(stats$EM)
round(n.em/n.total*100, 2)
```

```
[1] 12.22
```

Around 81.48% of structures in the PDB are solved by X-Ray and 12.22% are solved by Electron Microscopy.

Q2: What proportion of structures in the PDB are protein?

```
round(stats[1, "Total"]/ n.total* 100, 2)
```

```
Total
1 86.05
```

```
# or stats$Total[1]
```

86.05% are protein (only).

Q3: Type HIV in the PDB website search box on the home page and determine how many HIV-1 protease structures are in the current PDB?

In PDB website, there are 4865 structures for HIV and 1150 for HIV-1 protease.

## Exploring PDB structures

Package for structural bioinformatics.

```
library(bio3d)
hiv <- read.pdb("1hsg")
```

Note: Accessing on-line PDB file

```
hiv
```

```
Call: read.pdb(file = "1hsg")

Total Models#: 1
Total Atoms#: 1686, XYZs#: 5058 Chains#: 2 (values: A B)

Protein Atoms#: 1514 (residues/Calpha atoms#: 198)
Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)

Non-protein/nucleic Atoms#: 172 (residues: 128)
Non-protein/nucleic resid values: [ HOH (127), MK1 (1) ]

Protein sequence:
PQITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWPKMIGGIGGF1KVRQYD
QILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNFPQITLWQRPLVTIKIGGQLKE
ALLDTGADDTVLEEMSLPGRWPKMIGGIGGF1KVRQYDQILIEICGHKAIGTVLVGPTP
VNIIGRNLLTQIGCTLNF

+ attr: atom, xyz, seqres, helix, sheet,
calpha, remark, call
```

Let's first use the Mol\* viewer to explore this structure. molstar.org

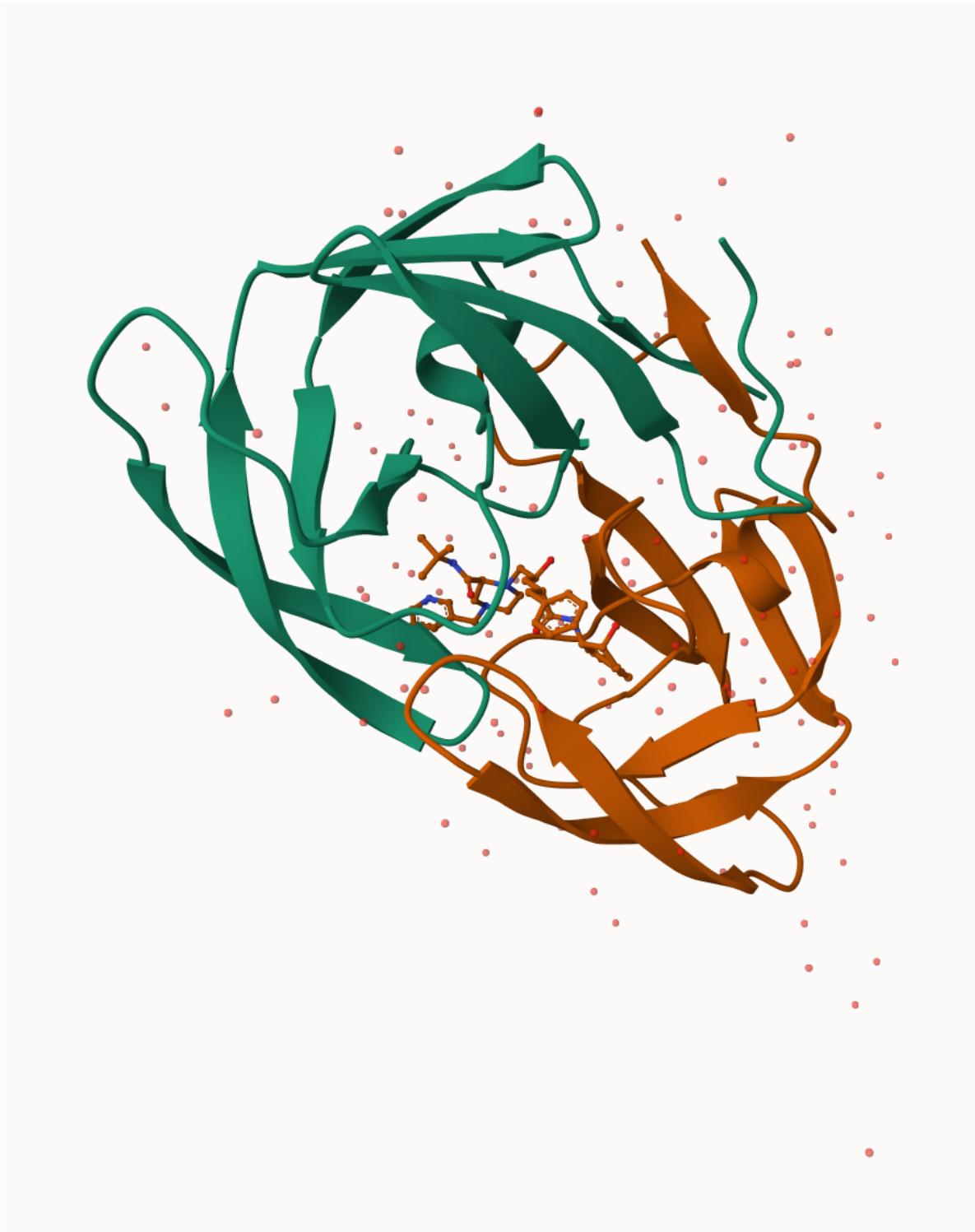


Figure 1: My first view of HIV-Protease

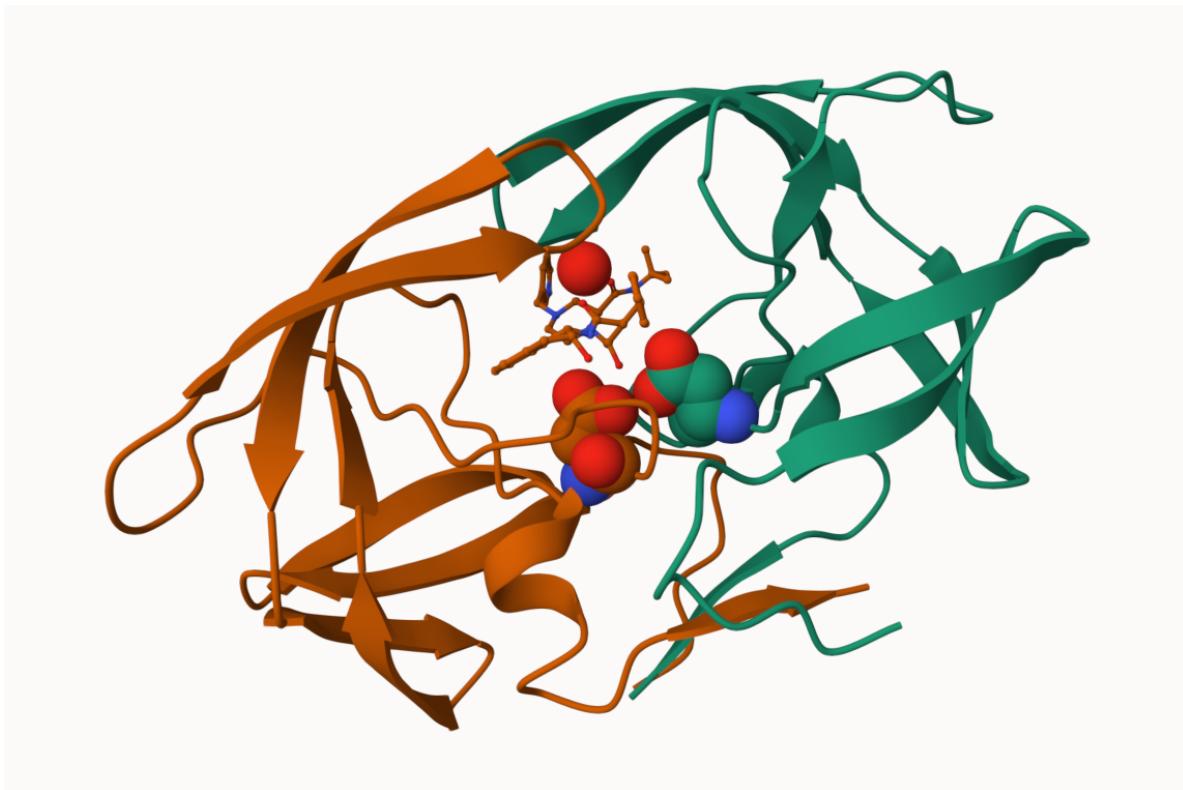
Q4: Water molecules normally have 3 atoms. Why do we see just one atom per water molecule in this structure?

It's for simplification. Now it's easier to view.

Q5: There is a critical "conserved" water molecule in the binding site. Can you identify this water molecule? What residue number does this water molecule have?

This water molecule is important for binding with ligand. The residue number is HOH 308.

And a view of the ligand (ball and stick) with catalytic ASP25 amino-acids (spacefill) and the all important active site water molecules (spacefill):



## PDB objects in R

```
head(hiv$atom)
```

	type	eleno	elety	alt	resid	chain	resno	insert	x	y	z	o	b		
1	ATOM		1		N	<NA>	PRO	A	1	<NA>	29.361	39.686	5.862	1	38.10

```

2 ATOM      2      CA <NA>    PRO      A      1      <NA> 30.307 38.663 5.319 1 40.62
3 ATOM      3      C <NA>     PRO      A      1      <NA> 29.760 38.071 4.022 1 42.64
4 ATOM      4      O <NA>     PRO      A      1      <NA> 28.600 38.302 3.676 1 43.40
5 ATOM      5      CB <NA>    PRO      A      1      <NA> 30.508 37.541 6.342 1 37.87
6 ATOM      6      CG <NA>    PRO      A      1      <NA> 29.296 37.591 7.162 1 38.40
segid elesy charge
1 <NA>      N      <NA>
2 <NA>      C      <NA>
3 <NA>      C      <NA>
4 <NA>      O      <NA>
5 <NA>      C      <NA>
6 <NA>      C      <NA>

```

Extract the sequence:

```
pdbseq(hiv)
```

```

 1   2   3   4   5   6   7   8   9   10  11  12  13  14  15  16  17  18  19  20
"P" "Q" "I" "T" "L" "W" "Q" "R" "P" "L" "V" "T" "I" "K" "I" "G" "G" "Q" "L" "K"
21  22  23  24  25  26  27  28  29  30  31  32  33  34  35  36  37  38  39  40
"E" "A" "L" "L" "D" "T" "G" "A" "D" "D" "T" "V" "L" "E" "E" "M" "S" "L" "P" "G"
41  42  43  44  45  46  47  48  49  50  51  52  53  54  55  56  57  58  59  60
"R" "W" "K" "P" "K" "M" "I" "G" "G" "I" "G" "G" "F" "I" "K" "V" "R" "Q" "Y" "D"
61  62  63  64  65  66  67  68  69  70  71  72  73  74  75  76  77  78  79  80
"Q" "I" "L" "I" "E" "I" "C" "G" "H" "K" "A" "I" "G" "T" "V" "L" "V" "G" "P" "T"
81  82  83  84  85  86  87  88  89  90  91  92  93  94  95  96  97  98  99  1
"P" "V" "N" "I" "I" "G" "R" "N" "L" "L" "T" "Q" "I" "G" "C" "T" "L" "N" "F" "P"
2   3   4   5   6   7   8   9   10  11  12  13  14  15  16  17  18  19  20  21
"Q" "I" "T" "L" "W" "Q" "R" "P" "L" "V" "T" "I" "K" "I" "G" "G" "Q" "L" "K" "E"
22  23  24  25  26  27  28  29  30  31  32  33  34  35  36  37  38  39  40  41
"A" "L" "L" "D" "T" "G" "A" "D" "D" "T" "V" "L" "E" "E" "M" "S" "L" "P" "G" "R"
42  43  44  45  46  47  48  49  50  51  52  53  54  55  56  57  58  59  60  61
"W" "K" "P" "K" "M" "I" "G" "G" "I" "G" "G" "F" "I" "K" "V" "R" "Q" "Y" "D" "Q"
62  63  64  65  66  67  68  69  70  71  72  73  74  75  76  77  78  79  80  81
"I" "L" "I" "E" "I" "C" "G" "H" "K" "A" "I" "G" "T" "V" "L" "V" "G" "P" "T" "P"
82  83  84  85  86  87  88  89  90  91  92  93  94  95  96  97  98  99
"V" "N" "I" "I" "G" "R" "N" "L" "L" "T" "Q" "I" "G" "C" "T" "L" "N" "F"

```

## Predict protein flexibility

We can run a bioinformatics calculation to predict protein dynamics - i.e. functional motions.

We will use the `nma()` function:

```
adk <- read.pdb("6s36")
```

Note: Accessing on-line PDB file  
PDB has ALT records, taking A only, rm.alt=TRUE

```
adk
```

Call: `read.pdb(file = "6s36")`

Total Models#: 1

Total Atoms#: 1898, XYZs#: 5694 Chains#: 1 (values: A)

Protein Atoms#: 1654 (residues/Calpha atoms#: 214)

Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)

Non-protein/nucleic Atoms#: 244 (residues: 244)

Non-protein/nucleic resid values: [ CL (3), HOH (238), MG (2), NA (1) ]

Protein sequence:

```
MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGDMRLAAVKSGSELGKQAKDIMDAGKLVT  
DELVIALVKERIAQEDCRNGFLLDGFPRTIPQADAMKEAGINVVDYVLEFDVPDELVDKI  
VGRRVHAPSGRVYHVKNPPKVEGKDDVTGEELTTRKDDQEETVRKRLVEYHQMTAPLIG  
YYSKAEAGNTKYAKVDGTPVAEVRADLEKILG
```

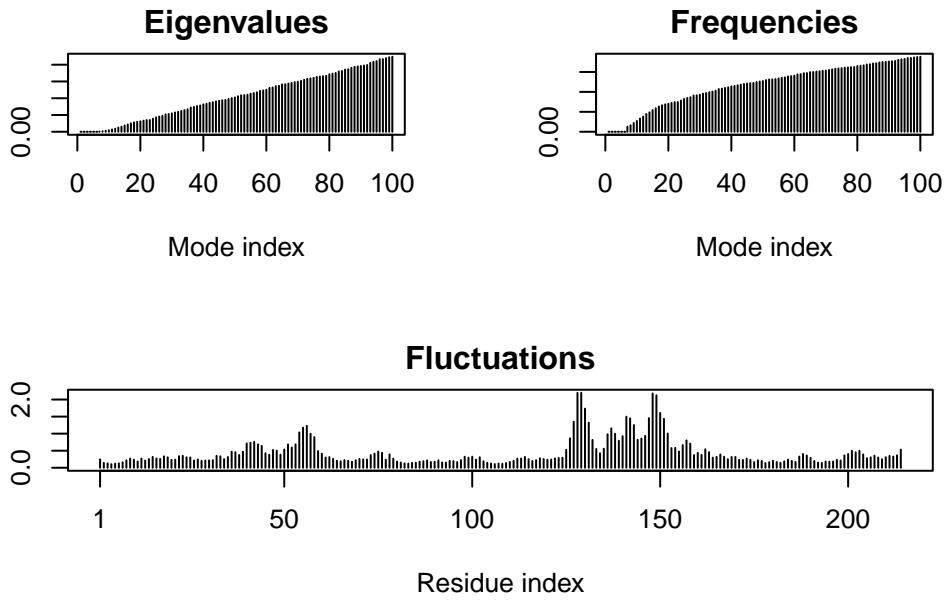
+ attr: atom, xyz, seqres, helix, sheet,  
calpha, remark, call

```
m <- nma(adk)
```

Building Hessian... Done in 0.045 seconds.

Diagonalizing Hessian... Done in 0.269 seconds.

```
plot(m)
```



Generate a “trajectory” of predicted motion

```
mktrj(m, file="ADK_nma.pdb")
```