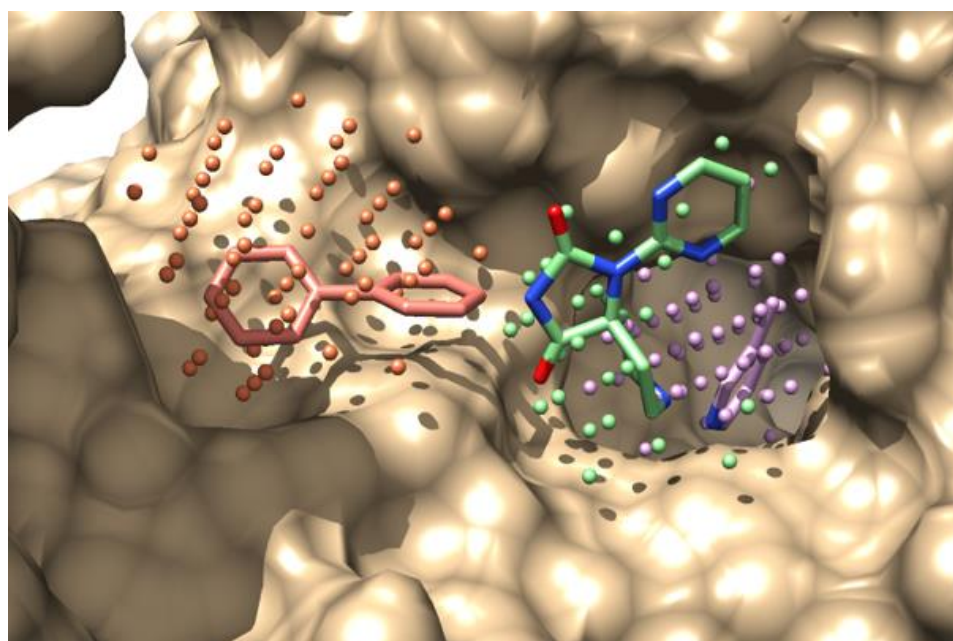


ProCare & utils

A Point Cloud Registration Approach to Compare and Align
Protein Cavities

Version 1.0.0



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1. Description and theory

ProCare¹ is a Python package for protein cavities comparison and alignment, based on a point cloud registration algorithm.²⁻⁴ A protein cavity by IChem VolSite module is represented as an ensemble of points that imprint the cavity shape and pharmacophoric properties (**Figure 1**).⁵

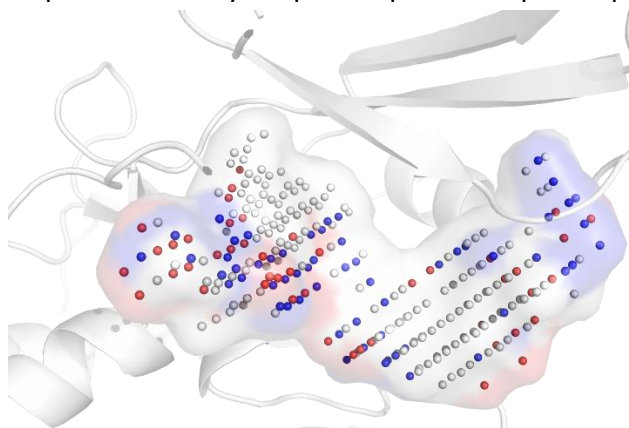


Figure 1. Example of CDK8 (PDB code: 5HBH) ATP cavity representation by IChem VolSite. Each point is associated to one of the eight possible pharmacophoric features according to the nearest protein atom. Blue: h-bond donor, positive ionizable; red: h-bond acceptor and donceptor, negative ionizable; white: hydrophobic, aromatic, dummy. The transparent molecular surface of the cavity was illustrated par Pymol 2.1 (Schrödinger, New York, USA).

A comparison is carried in 4 steps (**Figure 2**):

1. Computation of the points descriptors. A point descriptor is encoding both shape and position of the eight pharmacologic features (h-bond donor, acceptor and donceptor, positive ionizable, negative ionizable, hydrophobic, aromatic, dummy).
2. Initial RANSAC search by sampling randomly a few points at a time in the mobile cavity to match the most similar points in the reference cavity. Point similarity is estimated by the Euclidian distance of their respective descriptors. Given that the most similar points are searched, a point is always associated to a presumed equivalent in the target cavity, regardless of the distance. Then, false positive matches will be filtered out by checking the sample topology (pairwise distances should match withing a certain distance tolerance). Then an initial alignment is estimated and scored by a property-agnostic scoring function (fitness = proportion of aligned points in source cavity). This search is repeated until convergence.
3. Refinement of the initial alignment by the iterative closest point algorithm. A point is associated to its nearest neighbor in the Euclidian space.

4. Proposed alignment is scored by different schemes and metrics, accounting for the number of associated source/target pairs of points of the same or compatible pharmacophoric feature.

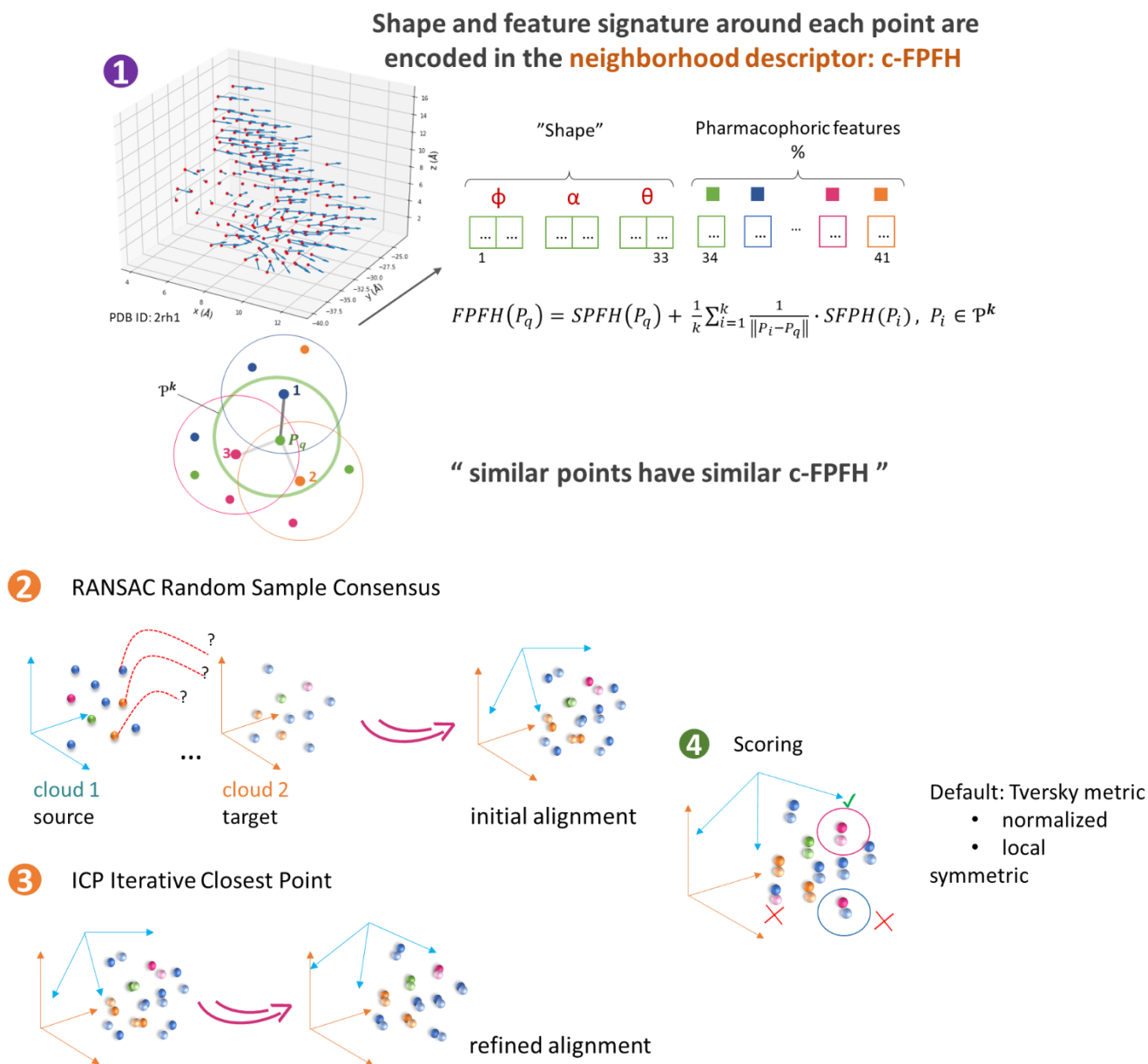


Figure 2. ProCare steps for comparing protein cavities.

2. Install

The installation procedure is summarized in:

#easy install

<https://github.com/kimeguida/ProCare/blob/master/install.sh>

- Install conda if not already there

See: <https://docs.conda.io/en/latest/miniconda.html>

- Create a conda environment

```
$ conda env create -n procare -f procare_environment.yml
```

- Activate procare conda environment

```
$ conda activate procare
```

If successful, your prompt will turn into: (procare) \$

- Download procare

```
$ git clone https://github.com/kimeguida/ProCare.git
$ cd ProCare/
```

- Install procare in the conda environment

```
(procare)$ pip install procare_python_package/
```

3. Help and issues

Signal issues at <https://github.com/kimeguida/ProCare/issues>

4. Usage

4.1. Compare cavities with **procare_launcher.py**

```
(procare)$ python procare_launcher.py -s 2rh1_cavity.mol2 -t  
5d6l_cavity.mol2 --transform --ligandtransform 2rh1_ligand.mol2
```

Outputs:

- **procare_scores.tsv** (tab-separated): simplified scores output
- **procare.tsv**: complete output containing transformation matrices elements, pharmacophore contributions, scores
- using the **--transform** option will output rotated cavity mol2 (**cfpfh_2rh1_cavity.mol2**)
- using the **--ligandtransform** option with a ligand/protein/anything mol2 file as argument will output aligned ligand/protein/anything in the rotated cavity reference frame in mol2 format (**cfpfh_2rh1_ligand.mol2**). Can handle multiple files:

```
[...] --ligandtransform 2rh1_ligand.mol2 <anything>.mol2
```

Help and options:

```
(procare)$ python procare_launcher.py -help
```

```
(procare)$ python procare_launcher.py [options] <values>
```

Input options

-s, --source <cavity>	source cavity (mobile) mol2
-t, --target <cavity>	target cavity (stationary) mol2

Alignment options

-rv, --ransacvalid default : 500	maximum RANSAC validation
-ri, --ransaciter default: 4000000	maximum RANSAC iteration
-rn, --ransacn default: 4	RANSAC clique size

-gt, --globaltranstype	transformation estimation method for global registration
default:	TransformationEstimationPointToPoint
-gd, --globaldist	distance (Å) threshold to define aligned points in global registration
default:	1.5
-cs, --checkersim	relative distance tolerance of RANSAC clique: between 0 and 1
default:	0.9
-it, --icptranstype	transformation estimation method for ICP registration
default:	TransformationEstimationPointToPoint
-id', '--icpdist'	distance (Å) threshold to define aligned points in ICP registration
default:	3
-ir, --icprmse	relative RMSE threshold for ICP terminaison
default:	10e-6
-if, --icpfitness	relative fitness score threshold for ICP terminaison
default:	10e-6
-ii, --icpiter	maximum ICP iteration
default:	100
-nr, --normalrad	radius (Å) for local surface normal estimation
default:	3.1
-nm, --normalmaxn	maximum number of neighbors to consider for local surface normal estimation
default:	471 (based on 4.5 Å radius)
-fr, --featurerad	radius (Å) for local surface feature calculation
default:	3.1

-fm, --featuremaxn maximum number of neighbors to
 consider for local surface
 feature calculation
 default: 135 (based on 3 Å radius)

Output options

-o, --output complete output containing
 transformation matrices elements,
 pharmacophore contributions,
 scores
 default: procare.tsv

-so, --scoreoutput simplified scores output
 default: procare_scores.tsv

-p, --paramid user-defined ID for parameters
 default: default

-c, --classification class for retrospective
 screening: 0 or 1
 default: NAN

--transform if set, output roto/translated
 cavity. Does not require <value>

--ligandtransform <mol2> apply transformation to other
 object(s) mol2

Utils

4.2. Inspect superposed points with: **procare_aligned_points.py**

Superposed points are source/target points of the same pharmacophoric feature and within 1.5 Å distance of each other.

```
(procare)$ python procare_aligned_points.py  
-c1 cfpfh_2rh1_cavity.mol2 -c2 5d6l_cavity.mol2  
-o1 aligned_2rh1_cavity.mol2 -o2 aligned_5d6l_cavity.mol2
```

Outputs:

- superposed points in cavities (**aligned_2rh1_cavity.mol2** and **aligned_5d6l_cavity.mol2**)

Help and options:

```
(procare)$ python procare_aligned_points.py -help
```

```
(procare)$ python procare_aligned_points.py [options] <values>
```

Input options

-c1, --cav1 <cavity> source cavity mol2

-c2, --cav2 <cavity> target cavity mol2

Output options

-o1, --ocav1 <cavity> output mol2 of aligned
points in source

-o2, --ocav2 <cavity> output mol2 of aligned
points in target

4.3. Reapply stored transformation matrices to objects with `procare_apply_transformation.py`

```
(procare)$ python utils/procare_apply_transformation.py -f
procare.tsv -a 2rh1_ligand.mol2 2rh1_cavity.mol2 -l 1
```

Outputs:

- `rot_2rh1_ligand.mol2` and `rot_2rh1_cavity.mol2` as rotated mol2 objects
`2rh1_ligand.mol2` and `2rh1_cavity.mol2`

Help and options:

```
(procare)$ python procare_apply_transformation.py -help
```

```
(procare)$ python procare_apply_transformation.py [options]
<values>
```

Input options

<code>-f, --fileprocare</code>	ProCare output file, e.g. <code>procare.tsv</code>
<code>-l, --line</code>	line index where results are stored in file, e.g. 2. File indexes start with 0 (header)
<code>-a, --appliedtomol2</code>	mol2 file(s) to apply transformation

Output options

<code>--prefix</code>	prefix to roto/translated output(s)
default: <code>rot (rot_<name>)</code>	

4.4. Rescoring of previously superposed cavities using other scoring schemes with **procare_rescoring.py**

```
(procare)$ python utils/procare_rescoring.py  
-s cfpfh_2rh1_cavity.mol2 -t 5d6l_cavity.mol2 -d 2
```

Outputs:

- **procare_rescoring.tsv**: score file

Help and options:

```
(procare)$ python procare_rescoring.py -help
```

```
(procare)$ python procare_rescoring.py [options] <values>
```

Input options

-s, --source <cavity>	source cavity (mobile) mol2
-t, --target <cavity>	target cavity (stationary) mol2
-d, --distance	distance (Å) threshold for scoring
default: 1.5	

Output options

-o, --ofile	output rescoring file
default: procare_rescoring.tsv	

5. References

1. Eguida, M.; Rognan, D. A Computer Vision Approach to Align and Compare Protein Cavities: Application to Fragment-Based Drug Design. *J. Med. Chem.* **2020**, *63*, 7127–7142.
2. Rusu, R. B.; Cousins, S. 3D Is Here: Point Cloud Library (PCL). In *2011 IEEE International Conference on Robotics and Automation*; IEEE, 2011; Vol. I, pp 1–4.
3. Rusu, R. B.; Blodow, N.; Beetz, M. Fast Point Feature Histograms (FPFH) for 3D Registration. *2009 IEEE Int. Conf. Robot. Autom.* **2009**, 3212–3217.
4. Zhou, Q.-Y.; Park, J.; Koltun, V. Open3D: A Modern Library for 3D Data Processing. *arXiv:1801.09847* **2018**.
5. Desaphy, J.; Azdimousa, K.; Kellenberger, E.; Rognan, D. Comparison and Druggability Prediction of Protein–Ligand Binding Sites from Pharmacophore-Annotated Cavity Shapes. *J. Chem. Inf. Model.* **2012**, *52*, 2287–2299.

6. Case studies

1. Eguida, M.; Rognan, D. Unexpected Similarity between HIV-1 Reverse Transcriptase and Tumor Necrosis Factor Binding Sites Revealed by Computer Vision. *J. Cheminform.* **2021**, *13*, 1–13
2. Eguida, M.; Schmitt-Valencia, C.; Hibert, M.; Villa, P.; Rognan, D. Target-Focused Library Design by Pocket-Applied Computer Vision and Fragment Deep Generative Linking. *J. Med. Chem.* **2022**, *65*, 13771–13783.