

Astrocytic intensity and area coverage analysis

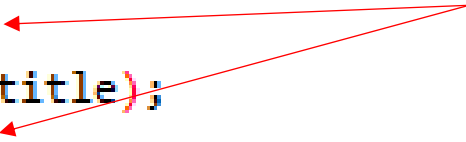
Automatized pipeline

Preprocessing

- Open ImageJ and open the macro *GLT-1_GFAP.ijm*

```
if (channels == 4) {  
    run("Split Channels");  
    selectWindow ("C1-" + title);  
    run("Close");  
    selectWindow ("C2-" + title);  
    run("Close");  
    selectWindow ("C3-" + title);  
    rename("GLT");  
    selectWindow ("C4-" + title);  
    rename("GFAP");  
}
```

Please change channel name of GFAP and GLT depending on which channel hold this information. If you have less than 4 channel, change it in the accroding line.

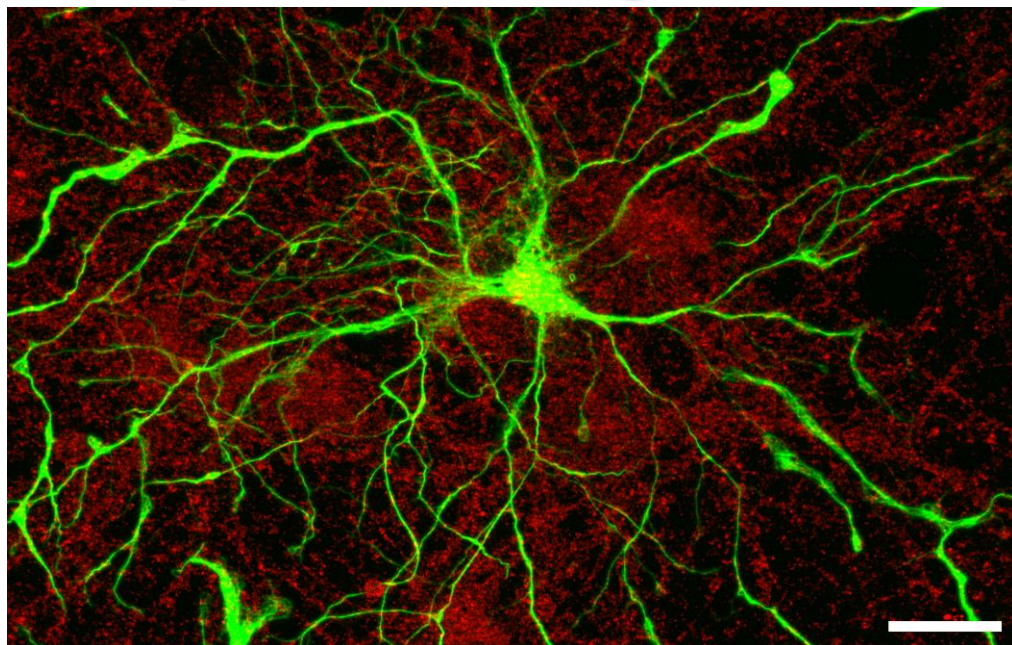


Preprocessing

```
// Thresholding of Astrocytes  
selectWindow("GFAP_threshold");  
setAutoThreshold("Huang dark");  
setOption("BlackBackground", false);  
run("Convert to Mask");  
selectWindow("GFAP_threshold");  
run("Create Selection");
```

If your images are not good with the same thresholding algorithm, you can change the algorithm used here.

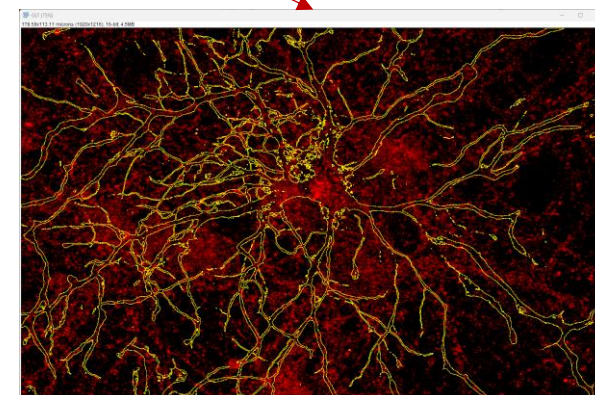
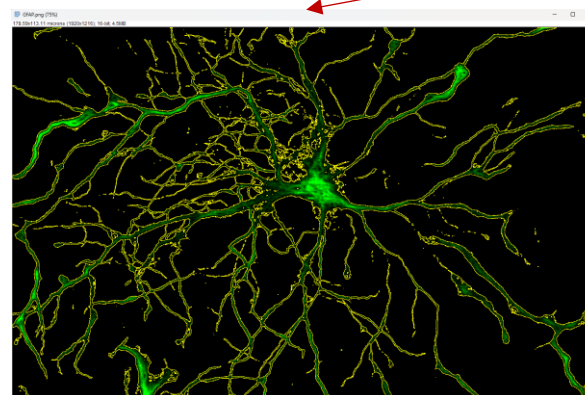
Preprocessing



thresholding
Huang dark



Selection overlay



Postprocessing of morphological data

- Use the *AstrocyteAnalysis.py* code to process you data

Important: you have to decode your data if they are coded


```
14 #####
15 #
16 # Please decode your conditions (or leave this empty if its already decoded)
17 #
18 #####
19 global replacements
20 replacements = {
21     'A1' : 'Male_ctrl_24h_+MG_A_',
22     'A2' : 'Male_H7N7_24h_+MG_A_',
23     'A3' : 'Male_ctrl_24h_-MG_A_',
24     'A4' : 'Female_ctrl_24h_-MG_A_',
25     'A5' : 'Female_ctrl_24h_+MG_A_',
26     'A6' : 'Female_H7N7_24h_+MG_A_',
27     'A7' : 'Male_ctrl_6h_-MG_A_',
28     'A8' : 'Female_ctrl_6h_-MG_A_',
29     'A9' : 'Female_ctrl_6h_+MG_A_',
30     'A10' : 'Female_H7N7_6h_+MG_A_',
31     'A11' : 'Male_H7N7_24h_-MG_A_',
32     'A12' : 'Male_ctrl_6h_+MG_A_',
33     'A13' : 'Male_H7N7_6h_+MG_A_',
34     'A14' : 'Male_H7N7_6h_-MG_A_',
35     'A15' : 'Female_H7N7_6h_-MG_A_',
36     'A16' : 'Female_H7N7_24h_-MG_A_',
37 }
```

Postprocessing of morphological data

If you would like to have an automatized normalization, you may have to adjust the round names (round_letter) and the time_points (if you have some)

```
132 #####
133 #
134 # Normalization
135 #
136 #####
137
138 df_data['Round'] = df_data['file'].str.extract(r'_([ABC])_')
139 df_data['Time'] = df_data['file'].str.extract(r'_([0-9]+h)_')
140
141
142 normalized_data = df_data.copy()
143
144 for round_letter in ['A', 'B', 'C']:
145     for time_point in ['6h', '24h']:
146         # Subset current group
147         current_group = df_data[(df_data['Round'] == round_letter) & (df_data['Time'] == time_point)]
148
```

What you get after running the code

 hyperexcitability.csv

 hyperexcitability_normalized.csv

	A	B	C	D	E	F
1	file	GFAP_area	GFAP_mean	GLT-1_mean	GLT-1_mean_int_whole	
2	MAX_Male_ci	1312.83	199.186	77.386	64.779	
3	MAX_Male_ci	1979.437	110.76	61.517	52.438	
4	MAX_Male_ci	1292.429	362.056	74.476	65.262	
5	MAX_Male_ci	1996.127	384.861	69.125	54.002	
6	MAX_Male_ci	1523.756	387.29	77.027	67.689	
7	MAX_Male_ci	1826.739	309.369	81.972	66.208	
8	MAX_Male_ci	2076.322	421.51	71.03	58.234	
9	MAX_Male_ci	1519.664	114.974	79.808	63.738	
10	MAX_Male_ci	1951.751	260.464	70.393	56.434	
11	MAX_Male_ci	1849.32	238.218	83.268	65.808	

Despite the used intensity staining for GLT-1, each staining within astrocytes can be analyzed with this code!