

# Polygenic Risk Score (PRS)

## X-chromosome aware PRS - Advances and Challenges

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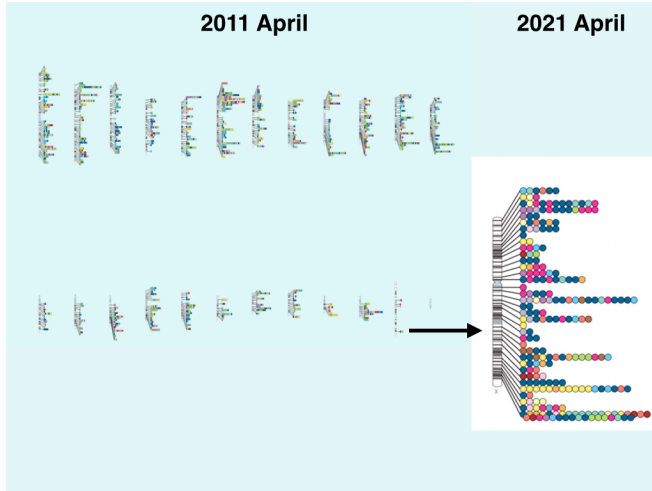
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# An X-inclusive trend towards GWAS results reporting



Wise et al. (2013). *American Journal of Human Genetics*. **eXclusion: toward integrating the X chromosome in genome-wide association analyses.**

# From autosomes to the X-chromosome (Xchr)

## *Quick facts:*

- ▶ Xchr accounts for 5% of the nuclear genome (155Mb)
- ▶ 800 protein-coding genes
- ▶ Many X-linked (recessive) diseases

## *Characteristics:*

- ▶ Female (two copies) vs. Male (one copy from mother);
- ▶ Dosage compensation via random Xchr-inactivation (XCI);
- ▶ Two pseudo-autosomal regions (PAR1 & PAR2) at termini of Xchr.

# Dosage compensation and Xchr-inactivation (XCI)

*Dosage compensation*: a process to equalize expression of genes between sexes through **XCI** in females.

- ▶ Random XCI: randomly silence one copy of XX in females
- ▶ No XCI: some genes escape silencing
- ▶ Skewed XCI: preferentially silence one X over the other

Carrel et al (2005). *Nature*. X-inactivation profile reveals extensive variability in X-linked gene expression in females.

Tukiainen et al. (2017). *Nature*. Landscape of X chromosome inactivation across human tissues.

The same 'weighted sum' approach in principle, **BUT**

$$X\text{-}PRS_i \stackrel{?}{=} \sum_{j=1}^J \hat{\beta}_j G_{ij}$$

- ▶  **$X\text{-}PRS_i$** : Xchr PRS for individual  $i$   
(sex-specific? combining males and females)
- ▶  **$J$** : the total number of "relevant" bi-allelic (common) SNPs  
(which **Xchr association test**; under no, random, or skewed XCI?)
- ▶  **$G_{ij}$** : the number of copies of the risk allele  
(counting  $a$  or  $A$ : **coding choices?**)
- ▶  **$\hat{\beta}_j$** : **estimated effect size of SNP  $j$**   
(sex-specific or combined? **effect size interpretation** in the context of XCI uncertainty and  $G \times \text{Sex}$  interaction effect!)
- ▶  **$\sum$** : (how to measure **LD** for Xchr?)

## Coding choices: what does it mean analytically for $G_{ij}$ ?

$G_j^M$ : **Genotype of a (single allele, male Xchr) SNP  $j$**

- ▶  $a$  = the reference allele
- ▶  $A$  = the alternative allele  
(often the minor allele with MAF of  $p$ )
- ▶ coded  $(0, 1)$  for  $(a, A)$

$G_j^F$ : **Genotype of a (bi-allelic, female Xchr) SNP  $j$**

- ▶ **no XCI**:  $(0, 1, 2)$  for  $(aa, Aa, AA)$
- ▶ **random XCI**:  $(0, 0.5, 1)$  for  $(aa, Aa, AA)$

male  $G=(0, 2)$  and female  $G=(0, 1, 2)$ , the PLINK default and  
male  $G=(0, 1)$  and female  $G=(0, 0.5, 1)$  are statistically the same.

- ▶ **skewed XCI?**

# Modelling considerations in Xchr-inclusive GWAS

- ▶ status of XCI
- ▶  $G_A$ : baseline allele  $a$  vs  $A$
- ▶  $S$ : sex as a confounder
- ▶  $G_A \times S$ : interaction?
- ▶  $G_D$ : dominance effect

Chen et al. (2021). *Genetic Epidemiology*. The X factor: a robust and powerful approach to X-chromosome-inclusive whole-genome association studies.

# Coding and Modelling choice **critical** for Xchr!

**Not including Sex as a covariate** leads to

- ▶ Type I error issue; sex is an inherent confounder for Xchr
- ▶ Biased  $\beta$  estimation

**Misspecified XCI** leads to

- ▶ Inaccurate  $\beta$  testing
- ▶ Biased  $\beta$  estimation



## This 3 d.f. test resolves above issues simultaneously

$$g(E(Y)) = \beta_0 + \beta_S \text{Sex} + \beta_A G_{\text{Additive}} + \beta_D G_{\text{Dominance}} + \beta_{GS} G \times \text{Sex},$$

$$H_0 : \beta_A = \beta_D = \beta_{GS} = 0.$$

- ▶ Code the  $G_A$  whichever way you want, and the association testing results will be the same!
- ▶ XCI uncertainty is statistically equivalent to  $G \times \text{Sex}$ .
- ▶ XCI skewness is statistically equivalent to a dominance effect.

Model, $g(E(Y)) =$	Testing $H_0 :$	a vs A & S	XCI and $G \times S$	skewed XCI and $G_D$
$M_0 : \beta_0 + \beta_A G_A$	$\beta_A = 0$	×	×	×
$M_1 : \beta_0 + \beta_S S + \beta_A G_A$	$\beta_A = 0$	✓	×	×
$M_2 : \beta_0 + \beta_S S + \beta_A G_A + \beta_D G_D$	$\beta_A = \beta_D = 0$	✓	×	✓
$M_3 : \beta_0 + \beta_S S + \beta_A G_A + \beta_{GS} GS$	$\beta_A = \beta_{GS} = 0$	✓	✓	×
$M_4 : \beta_0 + \beta_S S + \beta_A G_A + \beta_D G_D + \beta_{GS} GS$	$\beta_A = \beta_D = \beta_{GS} = 0$	✓	✓	✓

Chen et al. (2021). *Genetic Epidemiology*. The X factor: a robust and powerful approach to X-chromosome-inclusive whole-genome association studies.

# Trouble in paradise

Chen et al. (2021). *Genetic Epidemiology*. The X factor: a robust and powerful approach to X-chromosome-inclusive whole-genome association studies.

**“GWAS alone cannot identify the underlying true genetic model,** because we have shown, for example, XCI uncertainty is analytically equivalent to a gene-sex interaction effect, while XCI skewness is analytically equivalent to dominance effect.”

Song et al. (2021) *Genetic Epidemiology*. Testing and estimation of X-chromosome SNP effects: Impact of model assumptions.

**“We demonstrated sex and SNP coefficient biases in several situations,** particularly if the assumptions about XCI made by the coding scheme used and the assumptions made about sex differences in SNP effect of the fitted model were incorrect.”

## Numerical demonstration of the biased $\beta$ estimates

Consider the commonly used model *without*  $G_D$  or  $G \times \text{Sex}$ ,

$$g(E(Y)) = \beta_0 + \beta_S \text{Sex} + \beta_A G_{\text{Additive}}$$

		Estimated $\beta_A$				
		Bias		SD		
	$\beta_S$	$\beta_A$	XCI fit	no XCI fit	XCI fit	no XCI fit
XCI	0	0.75	0.006	0.012	0.074	0.110
	0.1	0.5	0.006	0.040	0.079	0.109
	0.2	0.2	0.012	0.081	0.083	0.120
	0.5	0.1	0.005	0.179	0.073	0.106
	0.75	0	0.009	0.278	0.090	0.128
no XCI	0	0.75	-0.002	-0.008	0.076	0.104
	0.1	0.5	-0.026	0.011	0.078	0.110
	0.2	0.2	-0.077	-0.013	0.075	0.106
	0.5	0.1	-0.175	-0.012	0.078	0.115
	0.75	0	-0.251	0.009	0.076	0.108

Song et al. (2021) *Genetic Epidemiology*. Testing and estimation of X-chromosome SNP effects: Impact of model assumptions.

# Xchr association analysis: only partially solved

Recommendation for association **testing**:

$$g(E(Y)) = \beta_0 + \beta_S \text{Sex} + \beta_A G_{\text{Additive}} + \beta_D G_{\text{Dominance}} + \beta_{GS} G \times \text{Sex},$$

$$3 \text{ d.f. test of } H_0 : \beta_A = \beta_D = \beta_{GS} = 0$$

Recommendation for association **reporting**:

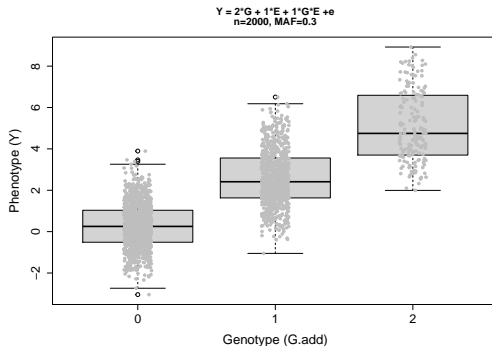
Wang, Sun, Paterson (working paper). Challenges with X-chromosome analyses and reporting in genome-wide association studies.

- ▶ specify the testing model, and if not the 3 d.f. model above
- ▶ specify the baseline allele
- ▶ specify the coding choice and the XCI assumption if invoked
- ▶ provide sex-specific estimates

Recommendation for effect size **estimation: An Open Problem!**

# Back to association testing (not parameter estimation)

$G \times E$  interaction leads to variance heterogeneity in  $Y$ :

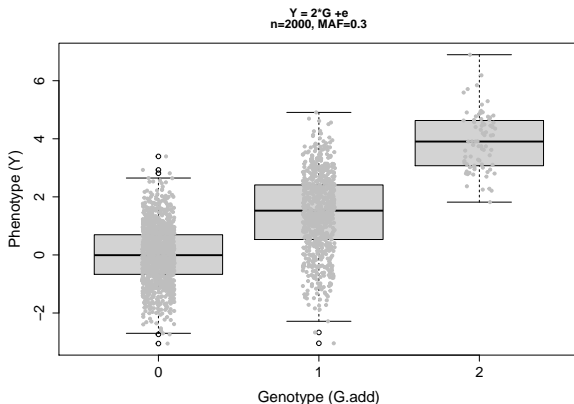


**When  $E$  is not available**, can we leverage the variance heterogeneity observation to **indirectly detect the  $G \times E$  interaction**?

Soave and Sun (2017). *Biometrics*. A generalized Levene's scale test for variance heterogeneity in the presence of sample correlation and group uncertainty

# Another source of variance heterogeneity specific to Xchr

A higher phenotypic variance in **heterozygous** females when true model is under XCI:



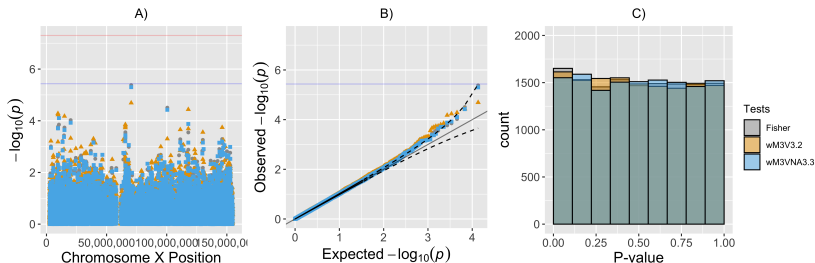
```
## Var(Y|G=0) Var(Y|G=1) Var(Y|G=2)
## 0.9603048 1.8152005 0.9969926
```

Ma et al. (2015). *BMC Genomics*. X-inactivation informs variance-based testing for X-linked association of a quantitative trait.

# UK Biobank and MESA Xchr application

Variance heterogeneity of *height* in UKB ( $n > 300k$ )

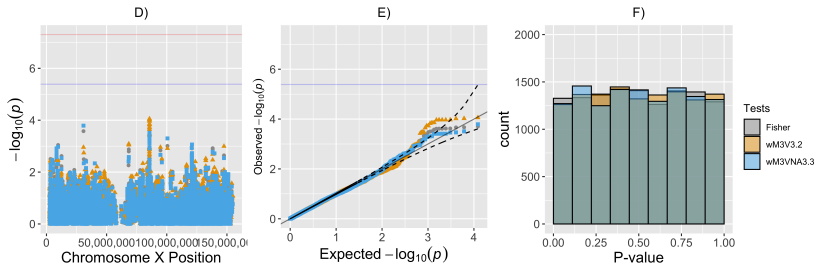
No Xchr-wide signals at 0.05/13,621



Deng et al. (2019). *Genetic Epidemiology*. Analytical strategies to include the X-chromosome in variance heterogeneity analyses: Evidence for trait-specific polygenic variance structure

# UK Biobank and MESA Xchr application (cont'd)

Variance heterogeneity of *height* in MESA ( $n = 2k$ )

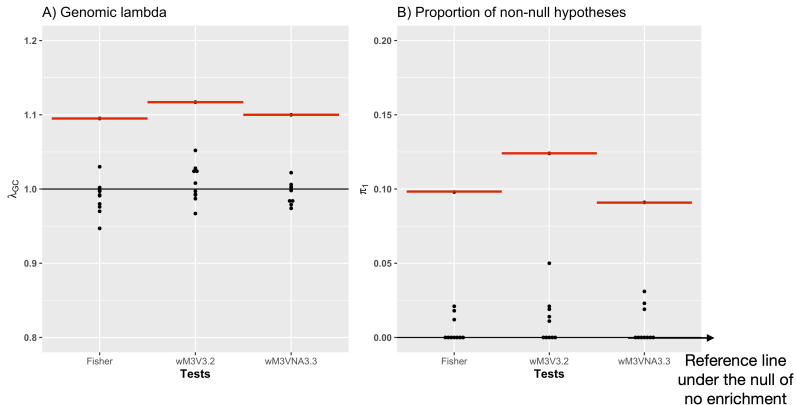


Deng et al. (2019). *Genetic Epidemiology*. Analytical strategies to include the X-chromosome in variance heterogeneity analyses: Evidence for trait-specific polygenic variance structure



# Height potentially enriched for $G \times E$ interactions (UKB)

A permutation analysis for variance of height (UKB): **evidence for excess of variance heterogeneity**

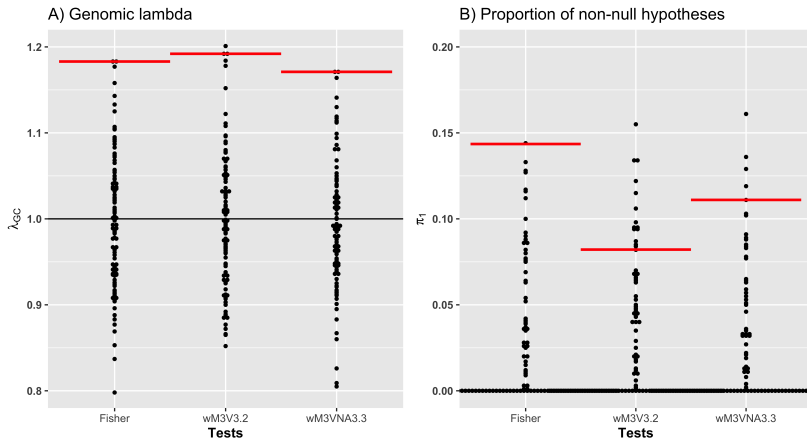


**Permuted statistics**

**Observed statistics**

- ▶  $\lambda_{GC} = \frac{\text{observed median}}{\text{theoretical median}}$
- ▶  $\pi_1$ ; proportion of non-null hypothesis

# Height potentially enriched for $G \times E$ interactions (MESA)



Deng et al. (2019). *Genetic Epidemiology*. Analytical strategies to include the X-chromosome in variance heterogeneity analyses: Evidence for trait-specific polygenic variance structure

# Recap the goal of this talk: a better understanding of

- ▶ Why association testing of an Xchr variant is actually 'easy'.  
advances in both location (mean) and scale (variances) methods  
“=” boosted power for discovery
- ▶ Why **genetic effect estimation is hard for Xchr!**  
GWAS data alone cannot identify the true Xchr genetic model.
- ▶ Why  $\hat{\beta}$  is not meaningful for an Xchr variant.  
**Good Xchr-inclusive PRS analysis is still an open question!**

# Xchr open but **consequential** questions

## Improving data quality

- ▶ Xchr-aware calling of sequencing data
- ▶ Xchr-aware imputation
- ▶ Xchr-aware QC steps

## Inference issues

- ▶ Xchr association **testing** problem solved, BUT
- ▶ **Which effect estimates to use for Xchr-PRS?**
- ▶ Sex-stratified, sex-combined effects,  $G \times S$  interaction effects?

## Other issues

- ▶ How to measure **LD for the Xchr?**
- ▶ Is it okay to use autosomes-derived PCA for the Xchr?
- ▶ How to perform **PCA for the Xchr?** How to do LD pruning?
- ▶ How to estimate  $h^2$  **for the Xchr?** Particularly in the context of the  $\beta$  estimation issue!

# References

- ▶ Carrel et al (2005). *Nature*. X-inactivation profile reveals extensive variability in X-linked gene expression in females.
- ▶ Tukiainen et al. (2017). *Nature*. Landscape of X chromosome inactivation across human tissues.
- ▶ Clayton (2008). *Biostatistics*. Testing for association on the X chromosome.
- ▶ Wise et al. (2013). *American Journal of Human Genetics*. eXclusion: toward integrating the X chromosome in genome-wide association analyses.
- ▶ Konig et al. (2014). *Genetic Epidemiology*. How to include chromosome X in your genome-wide association study.
- ▶ Wang et al. (2014). *Genetic Epidemiology*. X-chromosome genetic association test accounting for X-inactivation, skewed X-inactivation, and escape from X-inactivation.
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- ▶ Ma et al. (2015). *BMC Genomics*. X-inactivation informs variance-based testing for X-linked association of a quantitative trait.
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