

# High-throughput bio-part characterization using combinatorial library assembly approach



Seong-Kun Bak<sup>1, 2</sup>, Wonjae Seong<sup>1, 2</sup>, A-young Park<sup>1, 3</sup>, Eugene Rha<sup>1</sup>, Haseong Kim<sup>1, 2</sup>, and Seung-Goo Lee<sup>1, 2</sup>

<sup>1</sup> Synthetic Biology and Bioengineering Research Center, Korea Research Institute of Bioscience and Biotechnology (KRIBB), Daejeon 34141, Republic of Korea <sup>2</sup> Department of Biosystems and Bioengineering, KRIBB School of Biotechnology, University of Science and Technology, Daejeon 34113, Republic of Korea

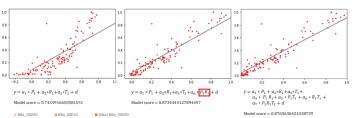
<sup>3</sup> Department of Chemical Engineering and Applied Chemistry, Chungnam National University, Daejeon 34134, Republic of Korea

### **ABSTRACT**

For the successful design of a genetic circuit, simulation techniques are required for selecting an appropriate circuit that shows the desired output among various combinations of parts. The prediction technique reduces the time and effort required for design, build, and test unnecessary part combinations. To do this, quantitatively characterized bio-parts are essential. In conventional methods to quantify the parts, the strength of every part is measured by the expression ratio of green fluorescence protein involving the target part, and the reference red fluorescent protein. However, this approach has a problem in that the measured quantity can be varied depending on the combination of other parts in the same circuit. Also, the quantification process takes time as the number of parts increases. In this study, we built the combinatorial library of parts using the Golden gate assembly technique and quantified the parts in the combinatorial library by measuring their fluorescence in the colony state. The genotype of the colonies was obtained by a long-read sequencing technique using tagging primer. The proposed technique can calculate quantitative values of the DNA parts considering the interaction among the parts, and quantify multiple parts in a high-throughput manner with a single experiment with combinatorial libraries.

## High-throughput Fluorescence analysis Introduction Image analysis Multiple plate reader $R^2 = 0.900547$ puthon Phenotype (Image analysis) Master plate(Tray plate) made by FACS sorting Captured by Fluorescence microscope Captured GFP, RFP in same condition with specific wave length filter High throughput Phenotyping using fluorescence microscope image and computational analysis Fluorescence(y) = f(P1, R1, T1)Genotype (Nanopore sequencing) Nanopore sequencing ■ Construction of Bio-part Library 0.61 Bio-part Library Bsal Genotype - Phenotype linkage 0.82 \* 102 part module library High-throughput Genotyping Relative fluorescence = RFP using tagged PCR Nanopore sequencing

## ■ Conclusions & Discussion



# 88a\_00030 # 88a\_00032 # 88a\_00032 # 88a\_00030 # 88a\_

Linear regression analysis using representative values confirmed the Interaction between specific Promoter and RiboJ

\* y = relative fluorescence

- 0.82\*10<sup>2</sup> part module library constructed by golden gate assembly
- We confirmed the correlation between computational Image analysis and multiple plate reader.
- Representative bio-part value is measured as mean of variable library.
- Using linear regression analysis confirmed the interaction between specific Promoter and RBS with insulator RiboJ
- For extended characterization, Two or three module circuit should be considered

### References

- Kelly JR, Rubin AJ, Davis JH, et al. Measuring the activity of BioBrick promoters using an in vivo reference standard. J Biol Eng. 2009;3:4. Published 2009 Mar 20. doi:10.1186/1754-1611-3-4
- Nuñez I, Matute T, Herrera R, et al. Low cost and open source multi-fluorescence imaging system for teaching and research in biology and bioengineering. PLoS One. 2017;12(11):e0187163. Published 2017 Nov 15. doi:10.1371/journal.pone.0187165.
- doi:10.1371/journal.pone.0187163

  Registry of Standard Biological Parts[http://parts.igem.org]