# Reservoir Gradient: Microbial Communities

Jay T. Lennon, Megan L. Larsen, Mario E. Muscarella 10 September, 2015

Project looking at microbial composition and processes along a reservoir gradient

### **Initial Setup**

```
rm(list=ls())
setwd("~/GitHub/ReservoirGradient")
source("./bin/MothurTools.R")

## Loading required package: reshape

require("vegan")

## Loading required package: vegan
## Loading required package: permute
## Loading required package: lattice
## This is vegan 2.2-1

se <- function(x, ...){sd(x, na.rm = TRUE)/sqrt(length(na.omit(x)))}</pre>
```

## Import Shared and Design Files

```
# Define Inputs
# Design = general design file for experiment
# shared = OTU table from mothur with sequence similarity clustering
# Taxonomy = Taxonomic information for each OTU
design <- ".../data/UL.design.txt"
shared <- ".../data/UL.bac.final.shared"
taxon <- ".../data/UL.bac.final.0.03.taxonomy"

# Import Design
design <- read.delim(design, header=T, row.names=1)

# Import Shared Files
OTUs <- read.otu(shared = shared, cutoff = "0.03")  # 97% Similarity

# Import Taxonomy
OTU.tax <- read.tax(taxonomy = taxon, format = "rdp")</pre>
```

#### **Data Transormations**

```
# Remove OTUs with less than two occurances across all sites
OTUs <- OTUs[, which(colSums(OTUs) >= 2)]

# Make Relative Abundence Matrices
OTUsREL <- OTUs
for(i in 1:dim(OTUs)[1]){
    OTUsREL[i,]<- OTUs[i,]/sum(OTUs[i,])
}

# Log Transform Relative Abundances
OTUsREL.log <- decostand(OTUs, method="log")

# Remove Low Coverage Samples
coverage <- rowSums(OTUs)</pre>
```

### Calculate Alpha Diversity

```
# Observed Richness
S.obs \leftarrow rowSums((OTUs > 0) * 1)
# Good's Coverage
C \leftarrow function(x = "") \{
  1 - (sum(x == 1) / rowSums(x))
C(OTUs)
## ULSoil_01 ULSoil_02 ULSoil_03 UL_01_DNA UL_01_cDNA UL_02_DNA
## 0.9541560 0.9537302 0.9389795 0.7123476 0.9521973 0.7763052
## UL_02_cDNA UL_03_DNA UL_03_cDNA UL_04_DNA UL_04_cDNA UL_05_DNA
## 0.9500715 0.7293705 0.9464016 0.7562412 0.9409259 -0.9708085
## UL_05_cDNA UL_06_DNA UL_06_cDNA UL_07_DNA UL_07_cDNA UL_08_DNA
## 0.8688629 0.9381043 -0.3017887 0.7216556 0.9220278 0.9156897
## UL_08_cDNA UL_09_DNA UL_09_cDNA UL_10_DNA UL_10_cDNA UL_11_DNA
## 0.9475004 0.7773052 0.9121607 0.9363162 0.8771606 0.9381341
## UL_11_cDNA UL_12_DNA UL_12_cDNA UL_13_DNA UL_13_cDNA UL_14_DNA
## 0.9493440 0.9452848 0.9369904 0.9456289 0.9431292 0.9236055
## UL_14_cDNA UL_15_DNA UL_15_cDNA UL_16_DNA UL_16_cDNA UL_17_DNA
## 0.9564206 0.9089748 0.9232507 0.9075687 0.9518381 0.8627176
## UL 17 cDNA UL 18 DNA UL 18 cDNA
## 0.9315844 0.9318416 0.9592666
# Simpson's Evenness
SimpE \leftarrow function(x = ""){
 x <- as.data.frame(x)</pre>
 D <- diversity(x, "inv")</pre>
 S \leftarrow sum((x > 0) * 1)
 E \leftarrow (D)/S
```

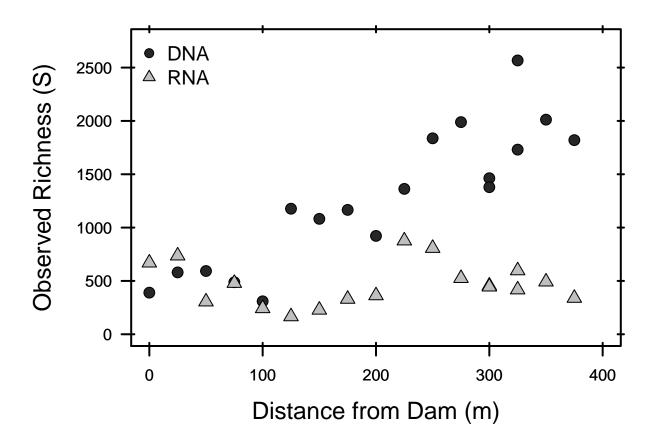
```
return(E)
}
simpsE <- round(apply(OTUs, 1, SimpE), 3)</pre>
# Diversity
H \leftarrow function(x = ""){
 x \leftarrow x[x>0]
 H = 0
 for (n_i in x){
   p = n_i / sum(x)
   H = H - p*log(p)
 }
 return(H)
shan <- round(apply(OTUs, 1, H), 2)</pre>
diversity(OTUs, index = "shannon")
## ULSoil_01 ULSoil_02 ULSoil_03 UL_01_DNA UL_01_cDNA UL_02_DNA
## 6.5548875 6.4528725 6.5691267 4.0795977 3.1684072 4.2357046
## UL_02_cDNA UL_03_DNA UL_03_cDNA UL_04_DNA UL_04_cDNA UL_05_DNA
## 3.4726269 4.1754273 1.9109298 4.2176218 0.8091232 4.2789561
## UL_05_cDNA UL_06_DNA UL_06_cDNA UL_07_DNA UL_07_cDNA UL_08_DNA
## 1.5800722 4.1328886 1.3139208 4.4590932 1.4832583 4.2888036
## UL_08_cDNA UL_09_DNA UL_09_cDNA UL_10_DNA UL_10_cDNA UL_11_DNA
## 2.8562167 4.6499951 1.6225924 4.7479989 4.3440328 5.0381606
## UL_11_cDNA UL_12_DNA UL_12_cDNA UL_13_DNA UL_13_cDNA UL_14_DNA
## 3.6131264 5.4000309 3.0954874 4.0250790 2.6736182 4.2285382
## UL_14_cDNA UL_15_DNA UL_15_cDNA UL_16_DNA UL_16_cDNA UL_17_DNA
## 3.1389321 5.2472604 3.0578964 3.5985282 0.8868672 4.9131101
## UL_17_cDNA UL_18_DNA UL_18_cDNA
## 2.3490868 5.5273013 1.5167291
alpha.div <- cbind(design, S.obs, simpsE, shan)
```

## Alpha Diversity Plots

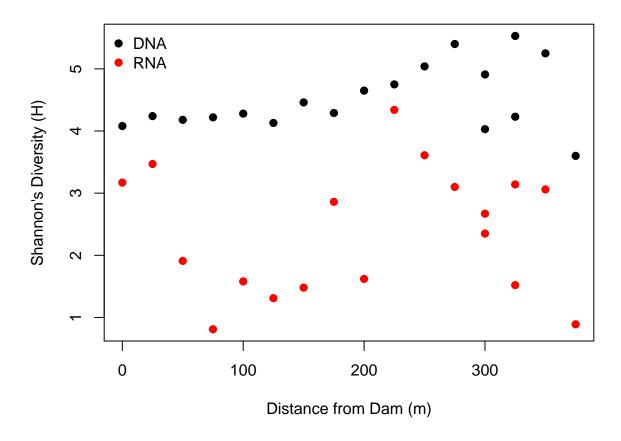
```
# Seperate data based on lake and soil samples
lake <- alpha.div[-c(1:3),]
soil <- alpha.div[c(1:3),]

# Richness across Reservoir Gradient
opar <- par()
par(mar = c(5,6, 1, 1))
mol <- rep(NA, length(lake$molecule))
for (i in 1:length(mol)){
   if (lake$molecule[i] == "DNA"){
      mol[i] <- 21
   } else {
      mol[i] <- 24</pre>
```

```
}
 }
cols <- rep(NA, length(lake$molecule))</pre>
  for (i in 1:length(cols)){
    if (lake$molecule[i] == "DNA"){
      cols[i] <- "gray15"</pre>
    } else {
      cols[i] <- "gray75"</pre>
    }
  }
plot(lake$S.obs ~ lake$distance, col= "black", bg = cols, pch=mol, las = 1,
     xlim = c(0,400), ylim = c(0, 2750), cex = 1.5,
     xlab="", ylab="")
axis(side = 1, lwd.ticks = 2, cex.axis = 1, las = 1)
axis(side = 2, lwd.ticks = 2, cex.axis = 1, las = 1)
axis(side = 3, lwd.ticks = 2, tck=-0.01, labels = F, cex.axis = 2, las = 1)
axis(side = 4, lwd.ticks = 2, tck=-0.01, labels = F, cex.axis = 2, las = 1)
axis(side = 1, lwd.ticks = 2, tck=0.01, labels = F, cex.axis = 2, las = 1)
axis(side = 2, lwd.ticks = 2, tck=0.01, labels = F, cex.axis = 2, las = 1)
axis(side = 3, lwd.ticks = 2, tck=0.01, labels = F, cex.axis = 2, las = 1)
axis(side = 4, lwd.ticks = 2, tck=0.01, labels = F, cex.axis = 2, las = 1)
mtext("Distance from Dam (m)" , side = 1, line = 3, cex=1.5)
mtext("Observed Richness (S)" , side = 2, line = 4, cex=1.5)
box(1wd=2)
legend("topleft", legend = levels(lake$molecule), pch=c(21, 24),
       pt.bg = c("gray15", "gray75"), bty='n', cex = 1.25)
abline(h = mean(soil$S.obs), lty=2, col="blue")
```



```
mean(soil$S.obs)
## [1] 7168.667
```



## **Beta Diversity**

```
# Calculate Bray-Curtis
UL.db <- vegdist(OTUsREL, method = "bray")</pre>
UL.pcoa <- cmdscale(UL.db, eig = TRUE, k = 3)</pre>
explainvar1 <- round(UL.pcoa$eig[1] / sum(UL.pcoa$eig), 3) * 100
\verb|explainvar2| <- round(UL.pcoa\$eig[2] / sum(UL.pcoa\$eig), 3) * 100|\\
explainvar3 <- round(UL.pcoa$eig[3] / sum(UL.pcoa$eig), 3) * 100
sum.eig <- sum(explainvar1, explainvar2, explainvar3)</pre>
# Define Plot Parameters
par(mar = c(5, 5, 1, 2) + 0.1)
# Initiate Plot
plot(UL.pcoa$points[ ,1], UL.pcoa$points[ ,2], ylim = c(-0.8, 0.8),
     xlab = paste("PCoA 1 (", explainvar1, "%)", sep = ""),
     ylab = paste("PCoA 2 (", explainvar2, "%)", sep = ""),
     pch = 16, cex = 2.0, type = "n", cex.lab = 1.5, cex.axis = 1.2, axes = FALSE)
# Add Axes
axis(side = 1, labels = T, lwd.ticks = 2, cex.axis = 1.2, las = 1)
```

